

Supporting Information for

Carboxylation and Mitsunobu Reaction of Amines to Give Carbamates: Retention vs Inversion of Configuration is Substituent-Dependent

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General Experimental Information: Proton NMR spectra were recorded on Varian Unity or Varian Plus (500 or 400 MHz) spectrometers. Chemical shifts are reported in parts per million (δ) downfield from tetramethylsilane as an internal standard. Low resolution electrospray mass spectra were recorded on a Micromass ZMD spectrometer using a Waters 2690 HPLC (2.1 x 100 mm C18 column, H₂O/MeCN w/0.05% TFA) for sample introduction. Samples provided for accurate mass measurement were taken up in acetonitrile:water:glacial acetic acid (50:50:0.1% v/v). The solutions were analyzed by use of electrospray ionization (ESI) or atmospheric pressure chemical ionization (APCI) on either a Bruker Daltonics 3T or 7T Fourier transform ion cyclotron resonance (FTICR) mass spectrometer. External calibration was accomplished with polypropylene glycol (425 or 750). Silica gel 60 (230-400) mesh from EM Science was used for flash column chromatography, and analytical thin layer chromatography was performed on EM Science Kieselgel 60 F254 plates. Solvents and reagents were obtained from commercial sources and used without further purification. The reported yields are the actual isolated yields of purified material and are not optimized.

2-[(2-Naphthylmethyl)amino]ethanol (4a): To a solution of ethanolamine (41.4 g, 678 mmol) in 600 mL of acetonitrile at room temperature was added dropwise a solution of 2-(bromomethyl)naphthalene (10.0 g, 45.2 mmol) in 125 mL of acetonitrile. After 2 hours, the solution was concentrated *in vacuo*, and the residue partitioned between dichloromethane and saturated aqueous NaHCO₃ solution. The aqueous layer was extracted three times with dichloromethane, and the combined organics were dried (Na₂SO₄), filtered, and concentrated *in vacuo*. The crude product was recrystallized from hot EtOAc-hexane to provide **4a** as a white crystalline solid (5.88g, 65% yield). Analytical LCMS: single peak (1.00 min, CH₃CN/H₂O/0.05% TFA, 4 min gradient) >95% pure by UV (215 nm); ¹H NMR (500 MHz, CDCl₃) δ 7.79-7.88 (m, 4H), 7.46-7.53 (m, 3H), 4.04 (s, 2H), 3.72 (t, *J* = 5.0 Hz, 2H), 2.89 (t, *J* = 5.0 Hz, 2H), 2.16 (br s, 2H); *m/z* (ES⁺) = 202.3 (MH⁺).

General procedure for carboxylation/Mitsunobu cyclization (Table 2):

3-(2-Naphthylmethyl)-1,3-oxazolidin-2-one (6a): To a solution of 2-[(2-naphthylmethyl)amino]ethanol (**4a**) (100 mg, 0.497 mmol, 1.0 equiv.) in 2.5 mL of anhydrous acetonitrile was added 1,8-diazabicyclo[5.4.0]undec-7-ene (7 μ L, 0.05 mmol, 0.1 equiv.). The resulting heterogeneous solution was then stirred under an atmosphere of carbon dioxide gas *via* a balloon, during which the mixture turned homogeneous momentarily and then heterogeneous. After 45 minutes, the reaction was cooled to 0 °C. *n*-Bu₃P was added (0.260 mL, 1.04 mmol, 2.1 equiv.), followed by a solution of di-*tert*-butyl azodicarboxylate (240 mg, 1.04 mmol, 2.1 equiv.) in 1 mL of acetonitrile in a dropwise manner. Over a period of 20 minutes the system became homogeneous. Removal of the solvent *in vacuo* afforded a clear yellow residue which was purified by flash chromatography (30-50% EtOAc/hexanes) to give 1-(2-naphthylmethyl)-2-oxazolidinone **6a** (104 mg, 92%) as a white solid. Analytical LCMS: single peak (2.22 min, CH₃CN/H₂O/0.05% TFA, 4 min gradient) >97% pure by UV (215 nm); ¹H-NMR (500 MHz, CDCl₃) δ 7.81-7.86 (m, 3H), 7.73 (br s, 1H), 7.48-7.52 (m, 2H), 7.42 (d, *J* = 8.5 Hz, 1H), 4.60 (s, 2H), 4.31 (t, *J* = 7.1 Hz, 2H) and 3.45 (t, *J* = 7.1 Hz, 2H) ppm; *m/z* (ES⁺) = 228.2 (MH⁺); HRMS (APCI) exact mass calcd for C₁₄H₁₄NO₂ (M+H⁺): 228.1019; found 228.1032.

3-Phenyl-1,3-oxazolidin-2-one (6b):¹ Analytical LCMS: single peak (1.64 min, CH₃CN/H₂O/0.05% TFA, 4 min gradient) >99% pure by UV (215 nm); ¹H NMR (500 MHz, CDCl₃) δ 7.56 (br d, *J* = 9 Hz, 2H), 7.39 (t, *J* = 7.5 Hz, 2H), 7.15 (t, *J* = 7.5 Hz, 1H), 4.49 (t, *J* = 8 Hz, 2H), 4.07 (t, *J* = 8 Hz, 2H); *m/z* (ES⁺) = 164.2 (MH⁺); HRMS (APCI) exact mass calcd for C₉H₁₀NO₂ (M+H⁺): 164.0706; found 164.0712.

(4R)-4-Benzyl-1,3-oxazolidin-2-one (6c):² Analytical HPLC: single peak (1.73 min, CH₃CN/7.4 mM aq NH₄OH, 5 min gradient) 95.8% pure by UV (254 nm); ¹H NMR (500 MHz, CDCl₃) δ 7.38 (t, *J* = 7.1 Hz, 2H), 7.30 (t, *J* = 7.1 Hz, 1H), 7.20 (br d, *J* = 7 Hz, 2H), 4.95 (br s, 1H), 4.51 (dd, *J* = 9 and 8 Hz, 1H), 4.18 (dd, *J* = 8.8 and 5.6 Hz,

1H), 4.10 (m, 1H), 2.91 (dd, $J = 13.4$ and 5.6 Hz, 1H), 2.86 (dd, $J = 13.4$ and 8.3 Hz, 1H); m/z (ES^+) = 178.0 (MH^+); HRMS (APCI) exact mass calcd for $C_{10}H_{12}NO_2$ ($M+H^+$): 178.0863; found 178.0869.

(4S)-4-Phenyl-1,3-oxazolidin-2-one (6d):³ Analytical LCMS: single peak (1.07 min, $CH_3CN/H_2O/0.05\%$ TFA, 4 min gradient) 97.3% pure by UV (215 nm); 1H NMR (500 MHz, $CDCl_3$) δ 7.33-7.42 (m, 5H), 5.97 (br s, 1H), 4.96 (br t, $J = 8$ Hz, 1H), 4.74 (t, $J = 8.6$ Hz, 1H), 4.19 (dd, $J = 8.6$ and 6.9 Hz, 1H); m/z (ES^+) = 164.3 (MH^+); HRMS (APCI) exact mass calcd for $C_9H_{10}NO_2$ ($M+H^+$): 164.0706; found 164.0712.

(5R)-5-Phenyl-1,3-oxazolidin-2-one (6e):⁴ Analytical LCMS: single peak (1.10 min, $CH_3CN/H_2O/0.05\%$ TFA, 4 min gradient) >95% pure by UV (215 nm); 1H NMR (500 MHz, $CDCl_3$) δ 7.36-7.43 (m, 5H), 5.68 (br s, 1H), 5.63 (br t, $J = 8.2$ Hz, 1H), 3.98 (t, $J = 8.6$ Hz, 1H), 3.54 (t, $J = 8$ Hz, 1H); m/z (ES^+) = 164.2 (MH^+); HRMS (APCI) exact mass calcd for $C_9H_{10}NO_2$ ($M+H^+$): 164.0706; found 164.0709; $[\alpha]_D^{22} = -24.6^\circ$ (c 0.045, EtOH); literature value for opposite enantiomer: $[\alpha]_D^{25} = +23^\circ$ (c 4.3, EtOH).

(4R,5R)-4-(Methoxymethyl)-5-phenyl-1,3-oxazolidin-2-one (6f):⁵ Analytical LCMS: single peak (1.29 min, $CH_3CN/H_2O/0.05\%$ TFA, 4 min gradient) >95% pure by UV (215 nm); 1H NMR (500 MHz, $CDCl_3$) δ 7.35-7.40 (m, 5H), 5.77 (br s, 1H), 5.24 (d, $J = 5.6$ Hz, 1H), 3.89 (br q, $J = 6$ Hz, 1H), 3.55 (dd, $J = 9.5$ and 4.6 Hz, 1H), 3.51 (dd, $J = 9.5$ and 6.9 Hz, 1H), 3.41 (s, 3H); m/z (ES^+) = 208.3 (MH^+); HRMS (APCI) exact mass calcd for $C_{10}H_{14}NO$ ($M+H^+-CO_2$): 164.1075; found 164.1075. Proof of structure was provided by hydrolysis to give (1R,2R)-2-amino-3-methoxy-1-phenylpropan-1-ol (**4f**). To a solution of **6f** (87 mg, 0.42 mmol) in 1 mL of ethanol was added 0.5 mL of 20% aqueous KOH solution. The mixture was heated at $60^\circ C$ for 3 days, then cooled to room temperature. The solution was partitioned between $CHCl_3$ and water, and the aqueous layer was extracted three times with $CHCl_3$. The combined organic layers were dried (Na_2SO_4), filtered, and concentrated *in vacuo*. Purification by flash column chromatography (7-10% MeOH/ CH_2Cl_2) gave **4f** as a colorless oil (38.8 mg, 50% yield), which was spectroscopically identical (1H NMR, Analytical LCMS) to commercial material, and had the correct sign of optical rotation: $[\alpha]_D^{22} = +19.2^\circ$ (c 0.030, $CHCl_3$); literature value: $[\alpha]_D^{25} = +25^\circ$ (c 10.6, $CHCl_3$).

(4R,5S)-4-Methyl-5-phenyl-1,3-oxazolidin-2-one (6g):⁶ Analytical HPLC: single peak (1.92 min, $CH_3CN/7.4$ mM aq NH_4OH , 5 min gradient) >99% pure by UV (254 nm); 1H NMR (500 MHz, $CDCl_3$) δ 7.36-7.43 (m, 3H), 7.32 (br d, $J = 8$ Hz, 1H), 5.74 (d, $J = 7.8$ Hz, 1H), 5.08 (br s, 1H), 4.22 (dq, $J = 7$ and 7 Hz, 1H), 0.83 (d, $J = 7.8$ Hz, 3H); m/z (ES^+) = 178.2 (MH^+); HRMS (APCI) exact mass calcd for $C_{10}H_{12}NO_2$ ($M+H^+$): 178.0863; found 178.0870.

(4R,5R)-3,4-Dimethyl-5-phenyl-1,3-oxazolidin-2-one (6h):⁷ Analytical HPLC: single peak (2.14 min, $CH_3CN/7.4$ mM aq NH_4OH , 5 min gradient) >99% pure by UV (254 nm); 1H NMR (500 MHz, $CDCl_3$) δ 7.37-7.43 (m, 5H), 4.92 (d, $J = 7.8$ Hz, 1H), 3.55 (dq, $J = 7$ and 7 Hz, 1H), 2.88 (s, 3H), 1.38 (d, $J = 7.8$ Hz, 3H); m/z (ES^+) = 192.0 (MH^+); HRMS (APCI) exact mass calcd for $C_{11}H_{14}NO_2$ ($M+H^+$): 192.1019; found 192.1024; $[\alpha]_D^{22} = -30^\circ$ (c 0.040, $CHCl_3$).

(4R,5S)-4,5-Diphenyl-1,3-oxazolidin-2-one (6i):⁸ Analytical LCMS: single peak (1.73 min, $CH_3CN/H_2O/0.05\%$ TFA, 4 min gradient) >97% pure by UV (215 nm); 1H NMR (400 MHz, $CDCl_3$) δ 7.01-7.07 (m, 6H), 6.87-6.92 (m, 4H), 5.89 (d, $J = 8.3$ Hz, 1H), 5.73 (br s, 1H), 5.13 (d, $J = 8.3$ Hz, 1H); m/z (ES^+) = 240.3 (MH^+); HRMS (APCI) exact mass calcd for $C_{15}H_{14}NO_2$ ($M+H^+$): 240.1019; found 240.1025; $[\alpha]_D^{22} = +80.8^\circ$ (c 0.013, $CHCl_3$) (measured $[\alpha]_D^{22} = -80.4^\circ$ (c 0.013, $CHCl_3$) for authentic commercially available opposite enantiomer).

(4S,5S)-3-Benzyl-4,5-diphenyl-1,3-oxazolidin-2-one (6j): Analytical LCMS: single peak (3.20 min, $CH_3CN/H_2O/0.05\%$ TFA, 4 min gradient) 94% pure by UV (215 nm); 1H NMR (500 MHz, $CDCl_3$) δ 7.38-7.41 (m, 3H), 7.26-7.29 (m, 3H), 7.22-7.24 (m, 6H), 7.17 (m, 1H), 7.10 (m, 1H), 7.07 (m, 1H), 5.22 (d, $J = 7.4$ Hz, 1H), 4.90 (d, $J = 14.9$ Hz, 1H), 4.25 (d, $J = 7.4$ Hz, 1H), 3.67 (d, $J = 14.9$ Hz, 1H); m/z (ES^+) = 330.1 (MH^+); HRMS (APCI) exact mass calcd for $C_{22}H_{20}NO_2$ ($M+H^+$): 330.1489; found 330.1479.

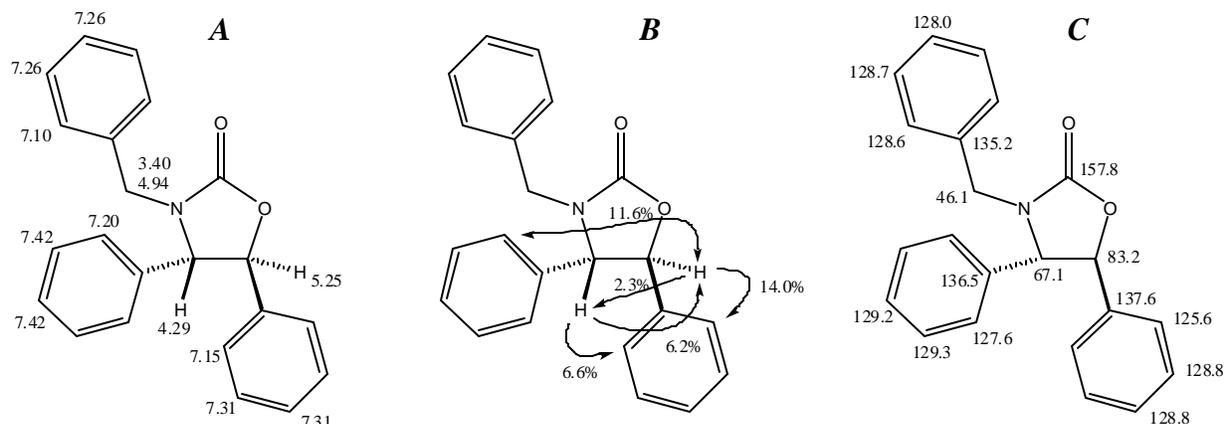


Figure 1. NMR spectral assignments for **6j** derived from ^1H NMR (600 MHz, CDCl_3), HMQC, qHMBC and difference NOE experiments. **A:** ^1H NMR assignments (ppm); **B:** NOE signal enhancements (%); **C:** ^{13}C NMR assignments (ppm).

Support for the structure assignment was provided by hydrolysis to give (1*S*,2*S*)-2-(benzylamino)-1,2-diphenylethanol. To a solution of **6j** (20 mg, 0.061 mmol) in 1 mL of ethanol was added 0.5 mL of 20% aqueous KOH solution. The mixture was heated to 60 °C for 24 hours, after which another portion of 20% aqueous KOH solution was added. After 1 hour the solution was cooled, then extracted three times with CH_2Cl_2 . The combined organic layers were dried (Na_2SO_4), filtered, and concentrated *in vacuo* to give an oil (m/z (ES^+) = 304.2 (MH^+)), the ^1H NMR spectrum of which does not match commercially available (1*R*,2*S*)-2-(benzylamino)-1,2-diphenylethanol (**4j**).

(3*aR*,7*aS*)-3-Benzylhexahydro-1,3-benzoxazol-2(3*H*)-one (6k):⁹ Analytical LCMS: single peak (2.31 min, $\text{CH}_3\text{CN}/\text{H}_2\text{O}/0.05\%$ TFA, 4 min gradient) >97% pure by UV (215 nm); ^1H NMR (500 MHz, CDCl_3) δ 7.30-7.38 (m, 5H), 4.80 (d, J = 15.1 Hz, 1H), 4.48 (q, J = 5 Hz, 1H), 4.06 (d, J = 15.1 Hz, 1H), 3.51 (q, J = 7 Hz, 1H), 1.95 (m, 1H), 1.77 (m, 2H), 1.58 (m, 2H), 1.50 (m, 1H), 1.44 (m, 1H), 1.24 (m, 1H); m/z (ES^+) = 232.1 (MH^+); HRMS (APCI) exact mass calcd for $\text{C}_{14}\text{H}_{18}\text{NO}_2$ ($\text{M}+\text{H}^+$): 232.1332; found 232.1341. Support for the structure assignment was provided by hydrolysis to give (1*S*,2*R*)-2-(benzylamino)cyclohexanol. To a solution of **6k** (20 mg, 0.086 mmol) in 1 mL of ethanol was added 0.5 mL of 20% aqueous KOH solution. The mixture was heated to 60 °C for 24 hours, after which another portion of 20% aqueous KOH solution was added. After 48 hours, the solution was cooled, then extracted three times with CH_2Cl_2 . The combined organic layers were dried (Na_2SO_4), filtered, and concentrated *in vacuo* to give an oil (m/z (ES^+) = 206.3 (MH^+)), the ^1H NMR spectrum of which does not match commercially available (1*R*,2*R*)-2-(benzylamino)cyclohexanol (**4k**).

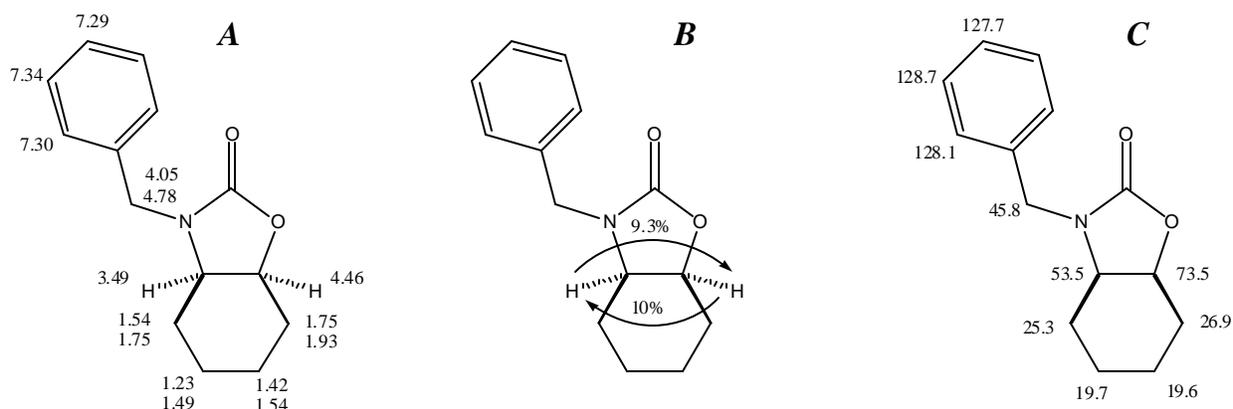


Figure 2. NMR spectral assignments for **6k** derived from ^1H NMR (600 MHz, CDCl_3), HMQC, GCOSY and difference NOE experiments. **A:** ^1H NMR assignments (ppm); **B:** NOE signal enhancements (%); **C:** ^{13}C NMR assignments (ppm).

(±)-4-Phenyl-1,3-oxazinan-2-one (9):¹⁰ Analytical LCMS: single peak (1.04 min, $\text{CH}_3\text{CN}/\text{H}_2\text{O}/0.05\%$ TFA, 4 min gradient) >97% pure by UV (215 nm); ^1H NMR (500 MHz, CDCl_3) δ 7.39 (br t, J = 8 Hz, 2H), 7.31-7.35 (m, 3H),

5.55 (br s, 1H), 4.66 (t, $J = 6.1$ Hz, 1H), 4.28-4.34 (m, 2H), 2.26 (m, 1H), 1.98 (m, 1H); m/z (ES^+) = 178.3 (MH^+); HRMS (APCI) exact mass calcd for $C_{10}H_{12}NO_2$ ($M+H^+$): 178.0863; found 178.0870.

Intermolecular carboxylation/Mitsunobu reaction (eq 4):

Benzyl 3,4-dihydroisoquinoline-2(1H)-carboxylate (23):¹¹ To a solution of 1,2,3,4-tetrahydroisoquinoline (**22**) (203 mg, 0.152 mmol, 1.0 equiv.) and benzyl alcohol (0.237 mL, 2.29 mmol, 1.5 equiv.) in 7 mL of anhydrous acetonitrile was added 1,8-diazabicyclo[5.4.0]undec-7-ene (23 μ L, 0.15 mmol, 0.1 equiv.). The resulting solution was then stirred under an atmosphere of carbon dioxide gas *via* a balloon, during which the mixture remained homogeneous. After 45 minutes, the reaction was cooled to 0 °C. *n*-Bu₃P was added (0.799 mL, 3.20 mmol, 2.1 equiv.), followed by a solution of di-*tert*-butyl azodicarboxylate (737 mg, 3.20 mmol, 2.1 equiv.) in 2 mL of acetonitrile in a dropwise manner. After 20 minutes, removal of the solvent *in vacuo* afforded a clear yellow residue which was purified by flash chromatography (50-80% CH₂Cl₂/hexanes) to give benzyl 3,4-dihydroisoquinoline-2(1H)-carboxylate **23** (365 mg, 90%) as a colorless oil. Analytical LCMS: single peak (2.49 min, CH₃CN/H₂O/0.05% TFA, 4 min gradient) 97% pure by UV (215 nm); ¹H-NMR (500 MHz, CDCl₃) δ 7.33-7.42 (m, 4H), 7.14-7.21 (m, 5H), 5.20 (s, 2H), 4.67 (s, 2H), 3.75 (br t, $J = 5$ Hz, 2H) and 2.87 (br s, 2H) ppm; m/z (ES^+) = 268.2 (MH^+); m/z (EI) = 176 ($C_{10}H_{10}NO_2$), 132 ($C_9H_{10}N$), 91 (C_7H_7).

Procedure for carboxylation/Mitsunobu cyclization using ¹⁸O-labeled CO₂ (eq 2 and 3):

(Compounds 10 and 11): A mixture of aminoalcohol (either **4g** or **4h**, 0.33 mmol, 1.0 equiv.) and 1,8-diazabicyclo[5.4.0]undec-7-ene (5 μ L, 0.03 mmol, 0.1 equiv.) in 3 mL of anhydrous acetonitrile was degassed by bubbling argon through the solution for 20 minutes. The resulting solution was then stirred under an atmosphere of ¹⁸O-labeled carbon dioxide gas (97 atom% ¹⁸O; ISOTEC lot no. CR0916) *via* balloon, during which the mixture remained homogeneous. After 45 minutes, the reaction was cooled to 0 °C. *n*-Bu₃P was added (0.173 mL, 0.69 mmol, 2.1 equiv.), followed by a solution of di-*tert*-butyl azodicarboxylate (160 mg, 0.69 mmol, 2.1 equiv.) in 0.5 mL of degassed acetonitrile in a dropwise manner. After 20 minutes, LCMS analysis indicated the presence of a single product peak. Removal of the solvent *in vacuo* afforded a clear yellow residue which was analyzed directly by LCMS (ES), HRMS (APCI) and ¹H NMR spectroscopy. The ¹H NMR (500 MHz) spectrum of the crude reaction mixture containing **10** was identical to that for crude **6g**, and the corresponding spectrum of the crude reaction mixture containing **11** was identical to that for crude **6h**. Copies of the LCMS and HRMS spectra from these experiments are shown at the end of this document.

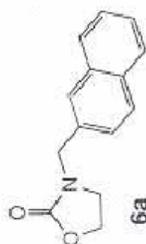
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Following Pages:

Copies of the ¹H NMR spectra for compounds **6a-k**, **9** and **23**:

stovrilite =



INDEX	SHIFT (PPM)	FREQ. (HZ)	HEIGHT
1	7.860	3331.27	48.0
2	7.843	3322.97	67.8
3	7.836	3319.55	48.6
4	7.831	3316.62	26.9
5	7.823	3312.96	15.7
6	7.818	3310.28	28.5
7	7.732	3267.06	51.8
8	7.522	3262.32	8.5
9	7.512	3257.13	45.5
10	7.509	3255.97	48.1
11	7.502	3252.07	53.1
12	7.494	3248.16	48.3
13	7.491	3246.70	29.7
14	7.484	3243.03	7.4
15	7.481	3241.81	8.3
16	7.433	3217.64	35.5
17	7.430	3216.18	28.1
18	7.416	3209.10	33.0
19	7.263	3632.68	135.6
20	7.260	3631.46	202.8
21	4.801	2301.34	193.7
22	4.836	2168.77	41.9
23	4.834	2167.79	57.3
24	4.820	2160.95	76.7
25	4.818	2159.73	82.8
26	4.816	2158.51	04.3
27	4.804	2157.30	48.7
28	4.802	2151.68	59.4
29	3.860	1730.52	45.5
30	3.858	1729.54	60.7
31	3.844	1727.71	75.7
32	3.842	1721.49	86.5
33	3.828	1714.85	44.7
34	3.826	1713.43	57.2
35	1.865	782.76	66.1
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ppm

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3

4

5

6

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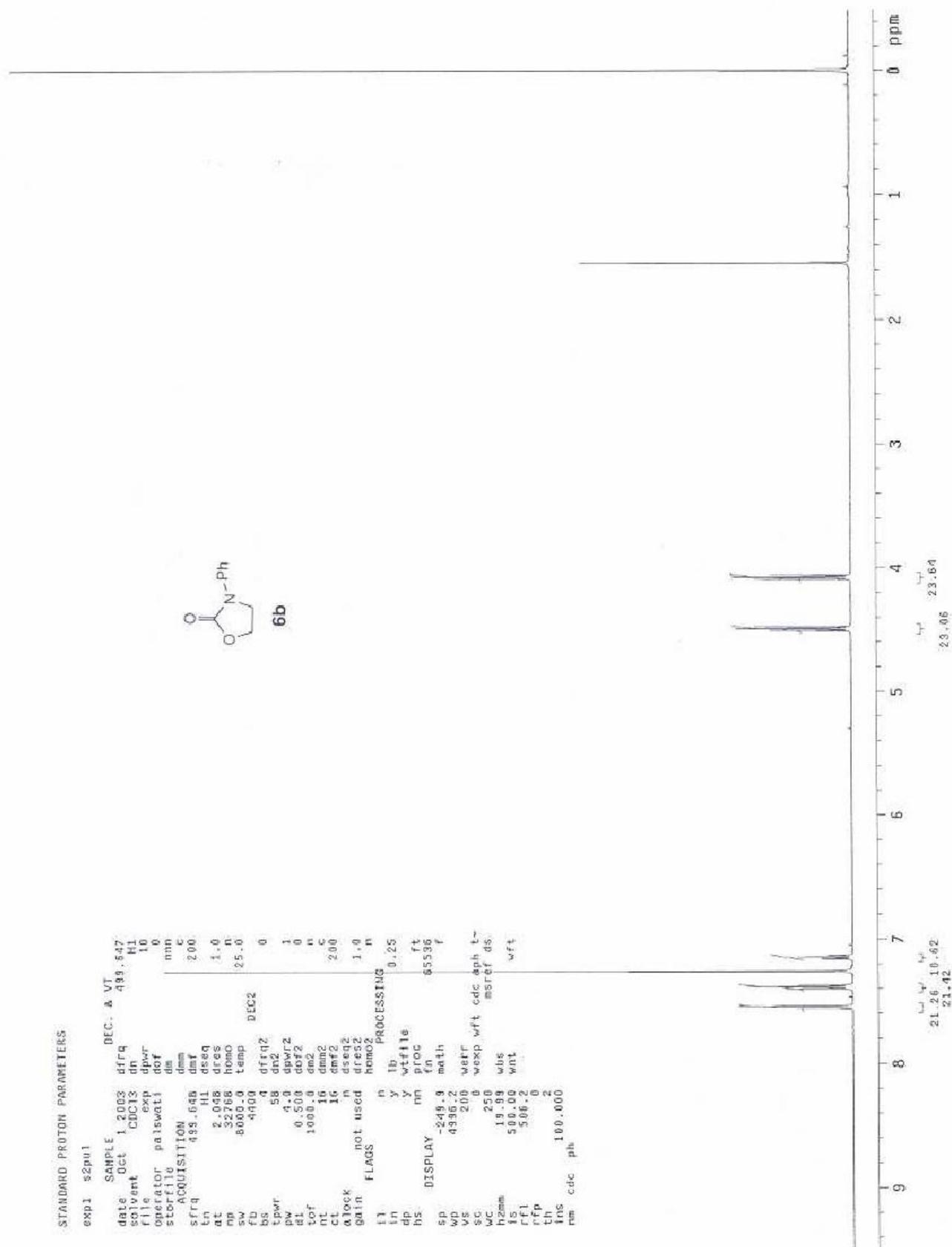
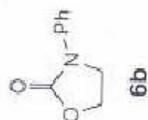
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STANDARD PROTON PARAMETERS

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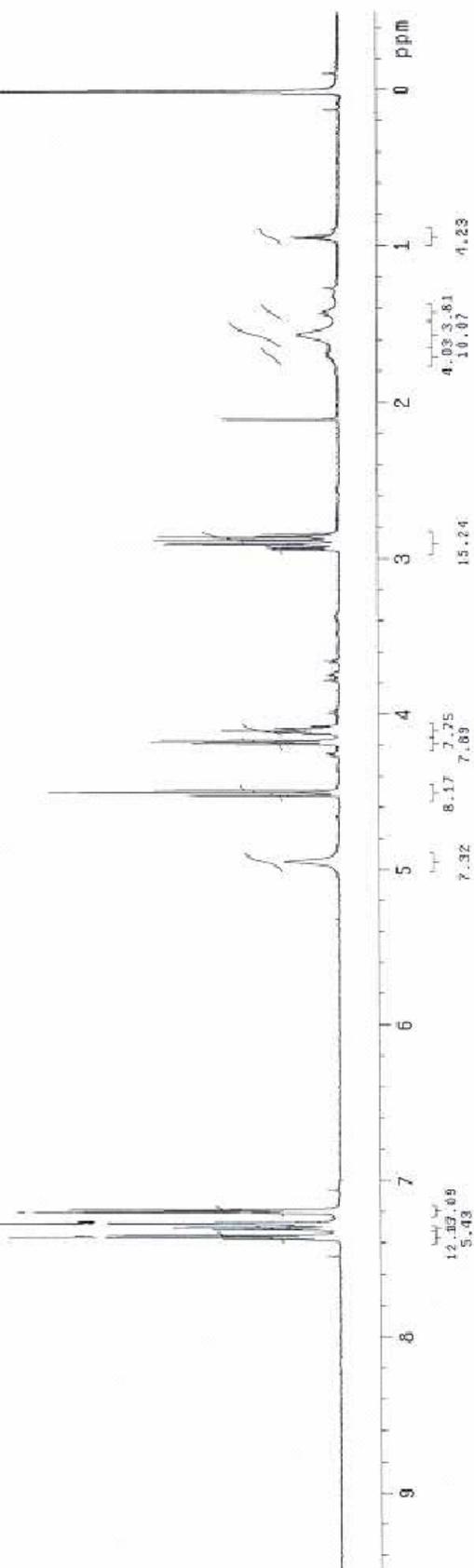
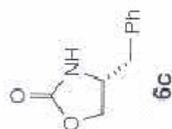
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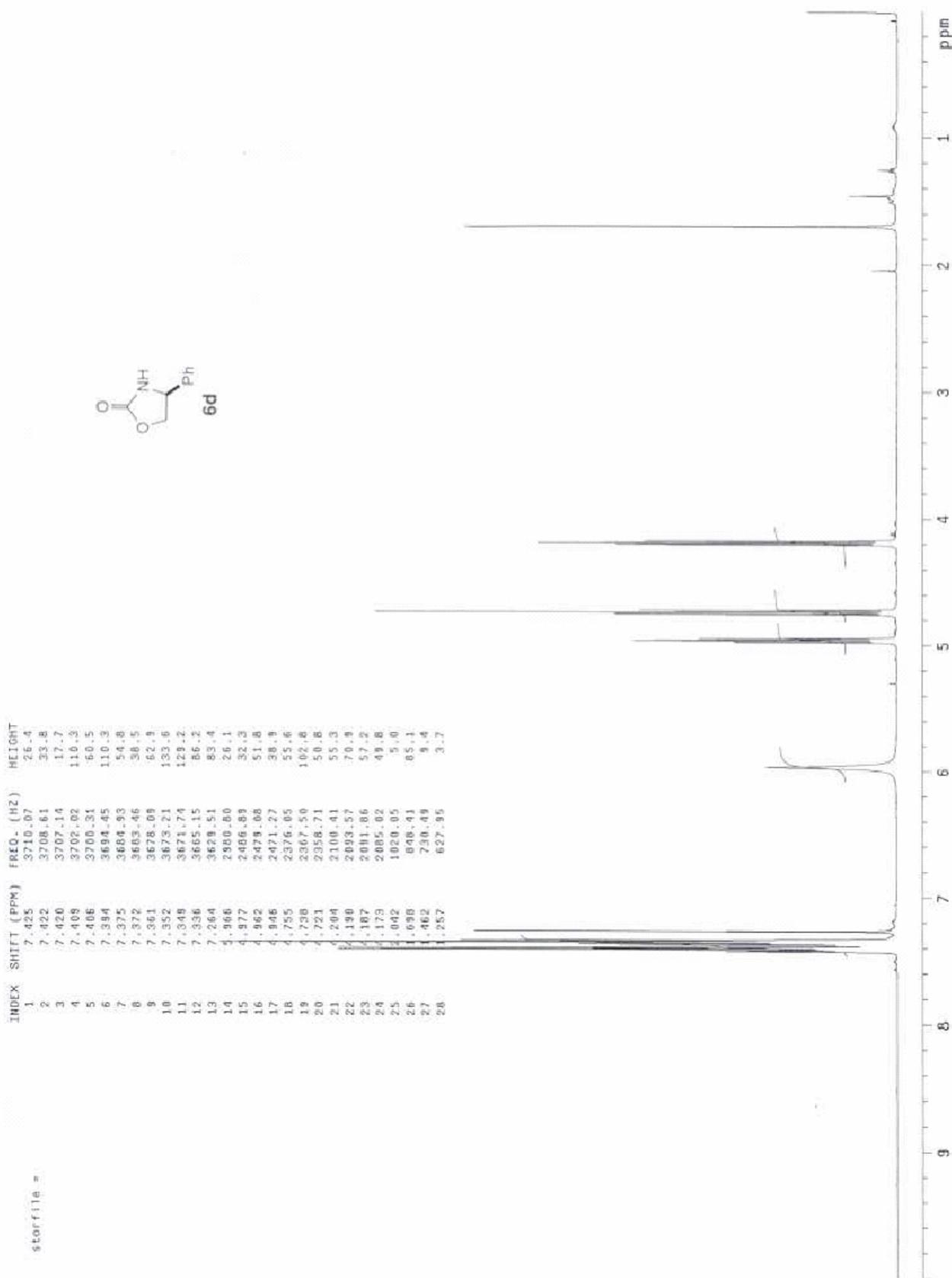


STANDARD PROTON PARAMETERS

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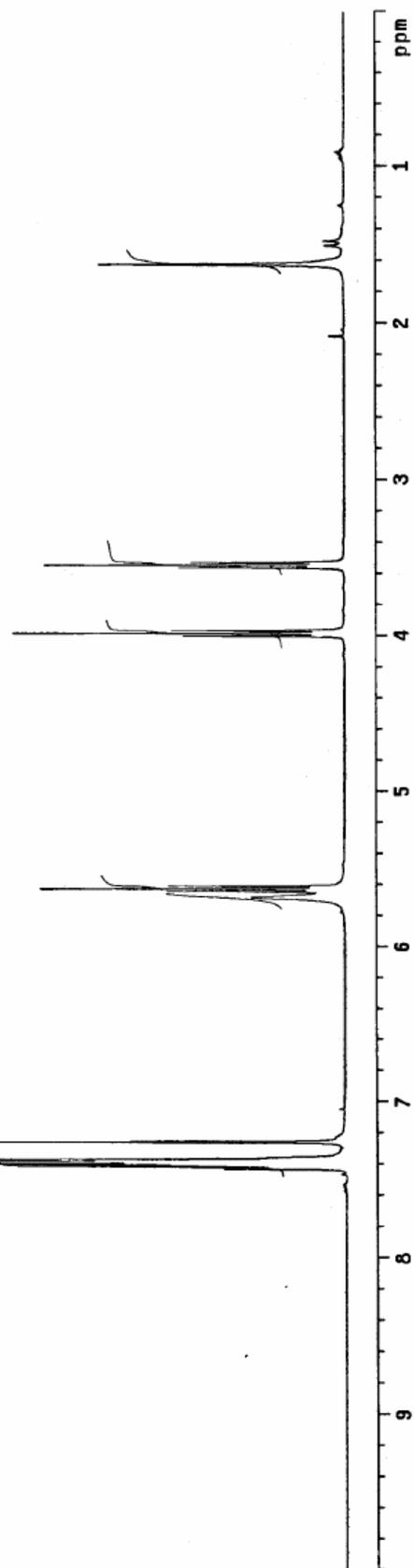
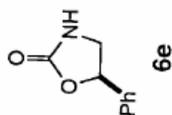
exp1  s2pu1
SAMPLE      DEC. & VT
date: Sep 24 2003  dfrq 500.171
solvent CDC13      dn   H1
file      exp      g    0
operator paiswati dof  0
storfile  dm       mm   200
ACQUISITION
sfrq      500.172  dmf  0
tn        H1      dseq  1
at        2.000   dres  0
pp        32000   homo  0
sw        8000.0 temp  25.0
fb        4000   drc2  0
bs        4      dfrq2 0
spwr      58     dn2    1
pw        4.0   dppwr2 1
d1        0.500 dof2   0
tof       1000.0 dm2    0
at        16    dmm2   0
ct        16    dmf2   0
a1ock.    n     dseq2  1
gain      not used dres2 1.0
          homo2   n
          flags   n
          lb      0.25
          y       y
          wf file ft
          mm     fn  B5536
          math    F
DISPLAY
sp      -250.3
wp      5001.5
vs      500
sc      250
wc      250
h2mm    20.01
ls      1000.00
rf1     495.0
rfp     0
th      5
ins     100.000
nm      cdc  ph
  
```

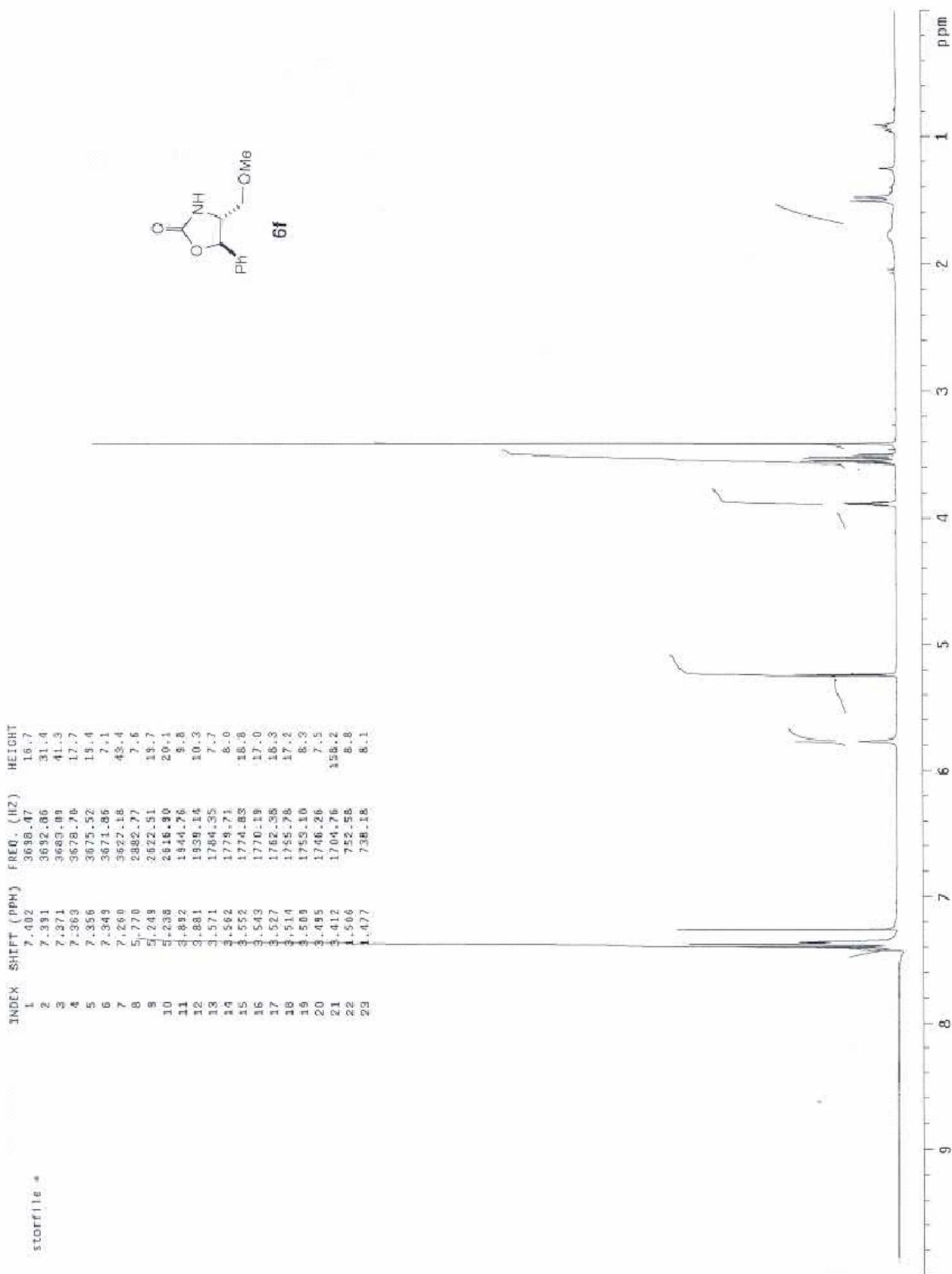




storfile =

INDEX	SHIFT (PPM)	FREQ. (HZ)	HEIGHT
1	7.430	3712.14	19.3
2	7.426	3710.19	14.5
3	7.415	3704.82	54.5
4	7.412	3703.60	54.1
5	7.408	3701.16	28.7
6	7.401	3697.74	94.9
7	7.388	3691.15	158.2
8	7.373	3684.07	65.6
9	7.364	3679.18	14.2
10	7.358	3676.25	12.6
11	7.259	3626.94	134.8
12	5.685	2840.29	15.0
13	5.646	2821.00	28.3
14	5.530	2812.95	48.3
15	5.513	2804.65	28.0
16	4.003	1999.93	25.6
17	3.986	1991.39	52.5
18	3.968	1982.60	27.4
19	3.565	1781.18	26.3
20	3.548	1772.63	47.4
21	3.532	1764.82	24.3
22	1.628	813.37	38.7



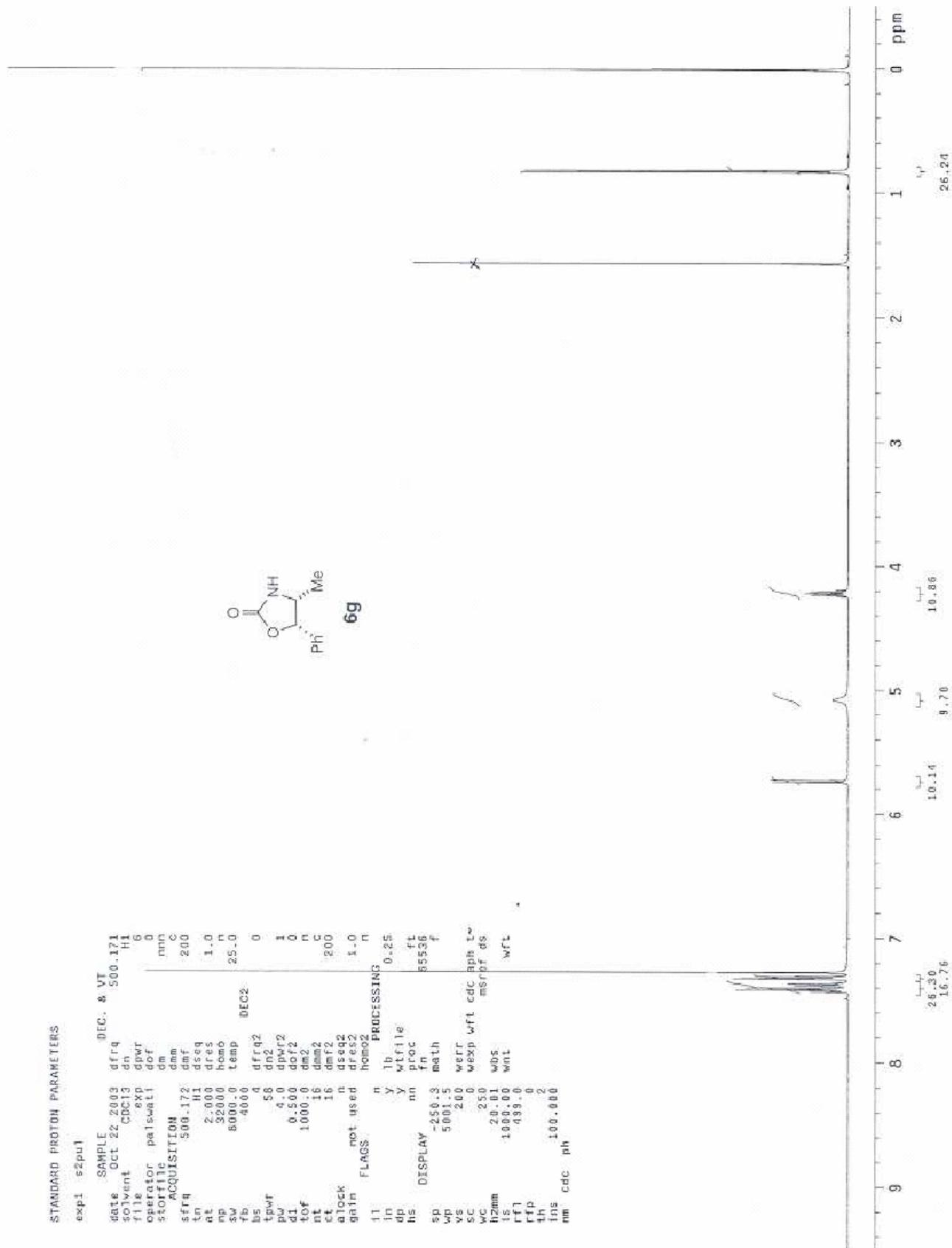
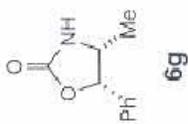


STANDARD PROTON PARAMETERS

```

exp1 s2pu1
SAMPLE DEC. & UT
date OCT 22 2003 dfrq 500.171
solvent CDCl3 dn HI
file exp dprw 0
operator paliswal dn 0
STORAGE dnm 200
ACQUISITION dnm C
sfrq 500.172 dmf 1.0
tn HI dseq 25.0
at 2.000 dres n
np 32000 homo n
sw 6000.0 temp DEC2
fb 4000 d dfrq2 0
ls d dn2 58 dnt 1
tpwr 4.0 dprw2 1
pw 0.500 dof2 0
dl 1000.0 dm2 n
fof 16 dnm2 C
nt 16 dmf2 200
ct 16 dseq2 n
alock n dres2 1.0
gain not used homo2 n
FLAGS n
ll n y
in y lb 0.25
dp y wfile
hs nn proc fl
bs 85536 fn f
DISPLAY -250.3 math
sp 5001.5 werr
vp 200 wexp wft cdc bph tw
vs 250 msrpf ds
sc 20.01 wds
b2mm 1000.00 wnt
rf1 439.0
rfp 0
rh 2
ins 100.000
nm cdc ph

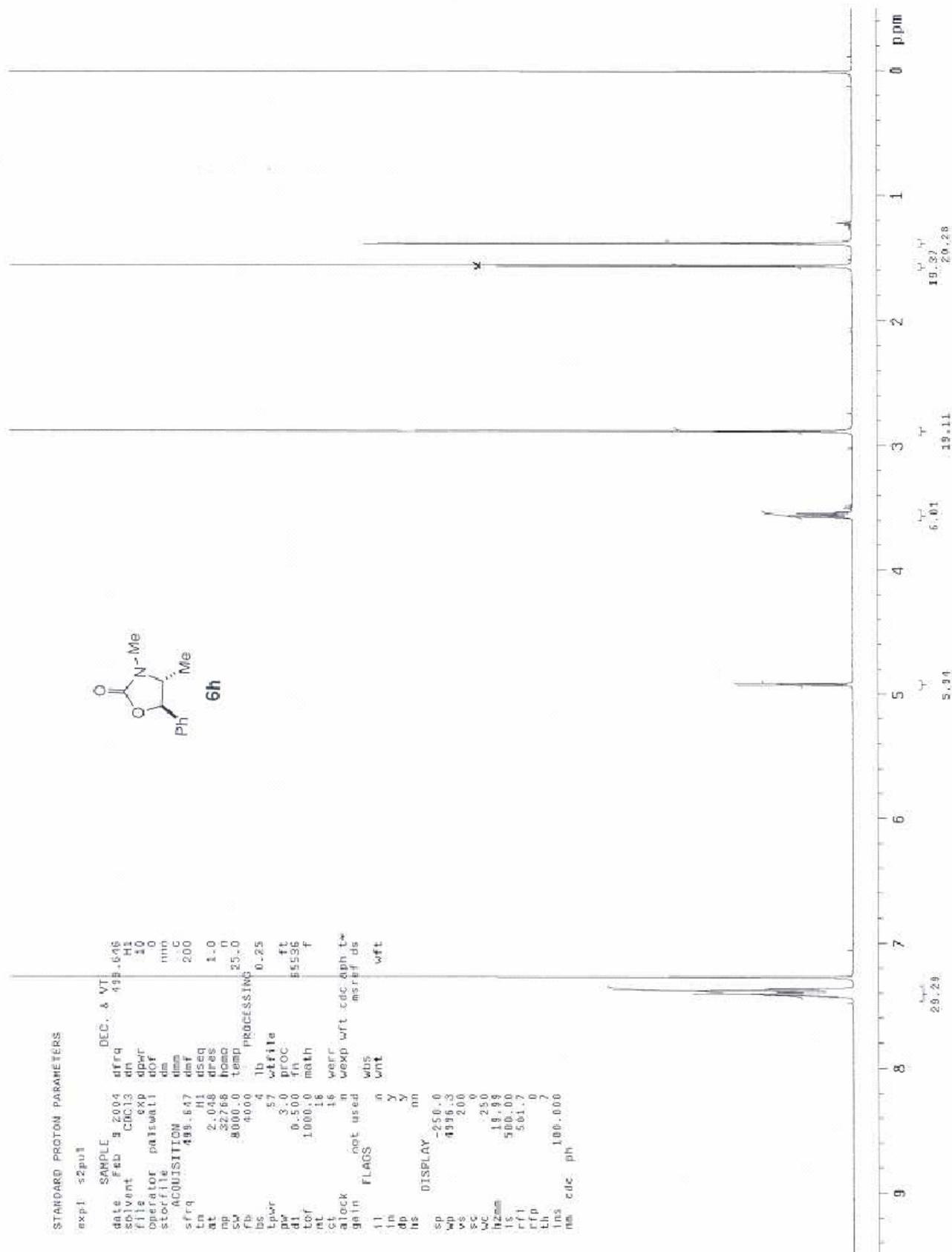
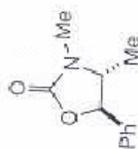
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STANDARD PROTON PARAMETERS

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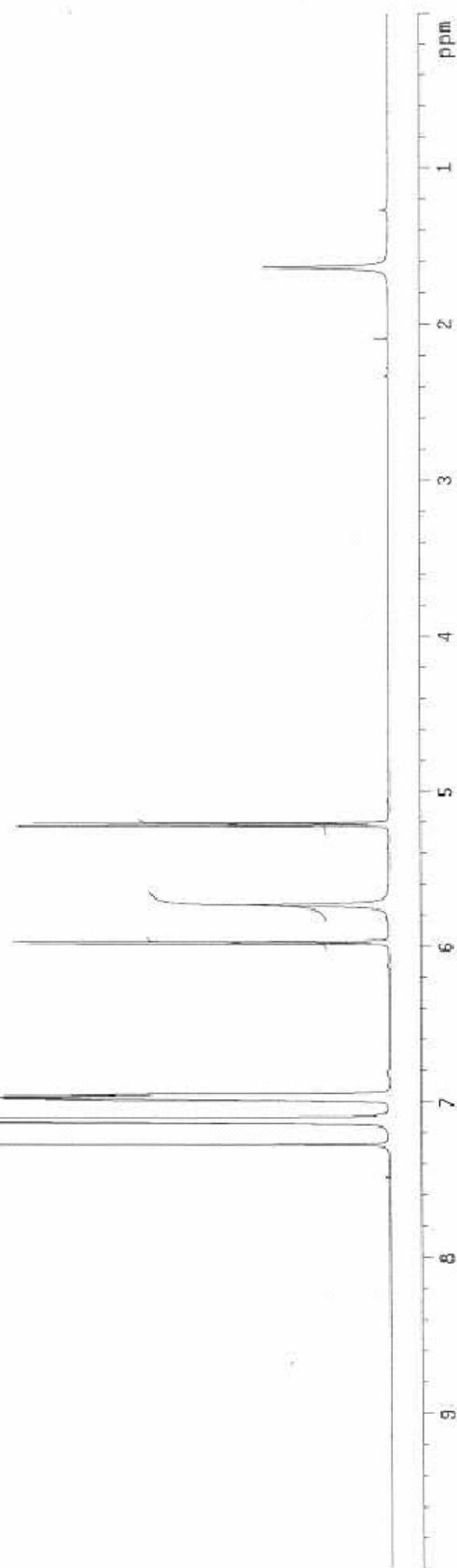
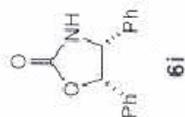
exp1 s2pu1
SAMPLE
date Feb 9 2004
solvent CDC13
File exp
operator patiswall
storfile
ACQUISITION
sfrq 499.647
in H1
at 2.048
np 32768
sw 8000.0
fb 4000
bs 4
lpr 57
pw 3.0
d1 0.500
tof 1000.0
at 16
ct 16
alock not used
gain not used
wexp wft
wus msref
wnt ds
wft
DEC. & VT
dfrq 499.646
dn Hs
exp 30
dm 0
nm 0
temp 25.0
PROCESSING
lb 0.25
wifile
proc ft
fn 51536
math f
werr
wexp wft
cdc dph t*
msref ds
wft
FLAGS
l1 n
l2 y
l3 y
l4 n
l5 n
DISPLAY
sp -250.0
wp 4336.3
vs 200
sc 0
ac 250
hzmax 18.99
ls 500.00
rf1 561.7
rfp 0
lh 7
lms 100.000
ma cdc
ph
  
```



INDEX	SHIFT (PPM)	FREQ. (HZ)	HEIGHT
1	7.278	3640.13	430.0
2	7.146	3574.21	42.6
3	7.138	3570.30	109.1
4	7.133	3567.62	117.7
5	7.126	3563.96	107.5
6	7.113	3500.54	421.4
7	7.112	3557.36	115.8
8	7.106	3554.19	108.4
9	6.981	3496.57	51.8
10	6.987	3494.62	55.1
11	6.960	3480.56	61.7
12	6.972	3487.05	92.7
13	6.966	3485.10	69.7
14	6.960	3481.19	61.6
15	6.953	3477.77	44.6
16	5.981	2991.45	57.6
17	5.965	2983.39	60.3
18	5.730	2865.96	23.0
19	5.226	2610.83	59.4
20	5.208	2602.53	57.0
21	1.638	819.33	19.3

solvent =

(DMSO-d₆)



STANDARD PROTON PARAMETERS

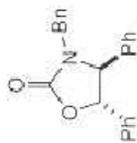
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expt s2pu1
SAMPLE
date Mar 5 2004
solvent CDCl3
file exp
operator palswat1
storfile
ACQUISITION
sfreq 399.867
in h1
at 2.735
nd 32768
sw 5891.2
fb -3000
ds 4
tpwr 58
pw 6.0
di 0.500
fof 600.0
nt 16
ct 16
alock not used
gain FLAGS
l1 n
l2 y
ds nn
DISPLAY
sp -200.0
wp 3888.6
ws 200
sc 0
mc 250
h2mm 15.99
ls 500.00
fft 356.0
rff 0
th 2
ins cdc ph
ns cdc ph
  
```

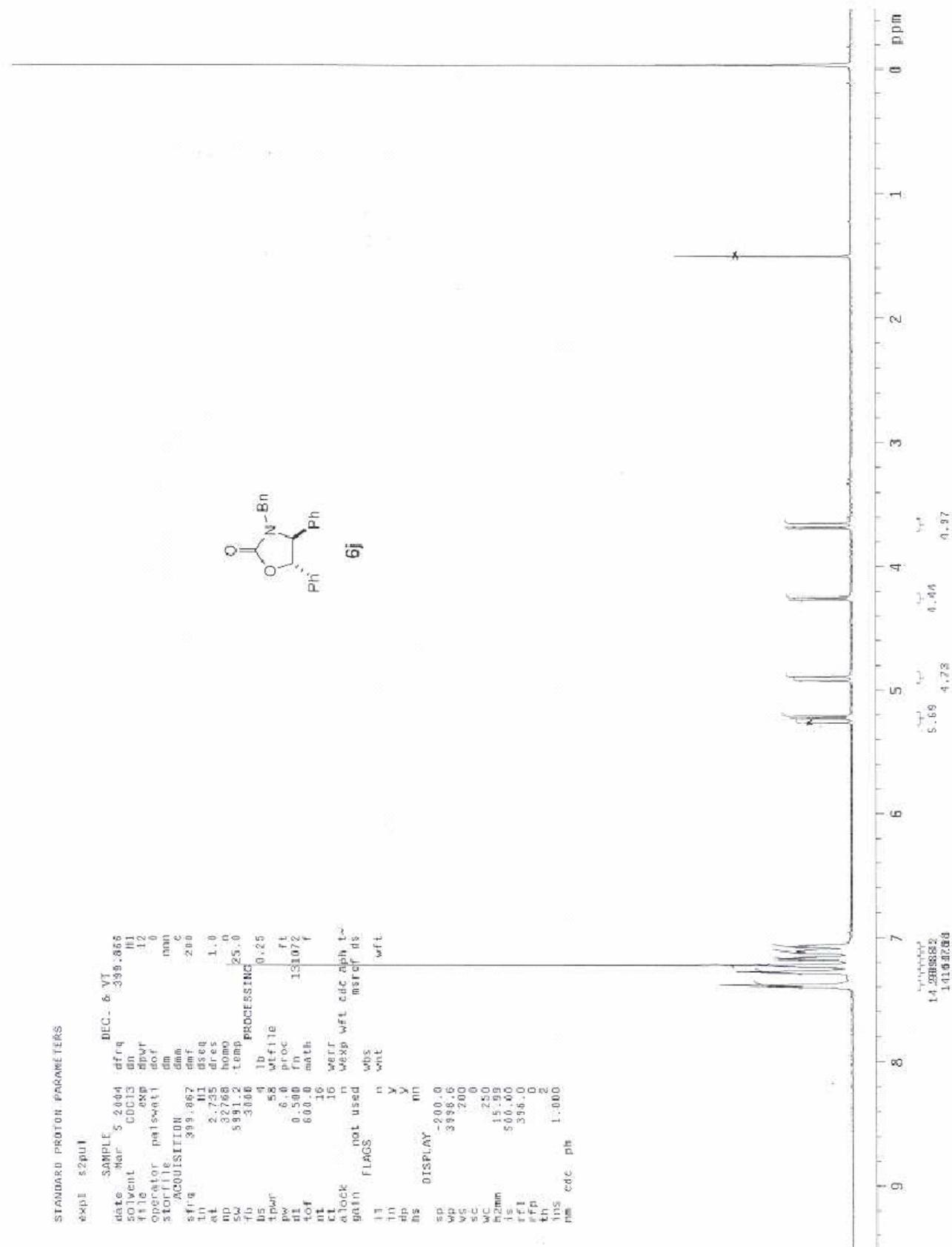
DEC. 6-VT

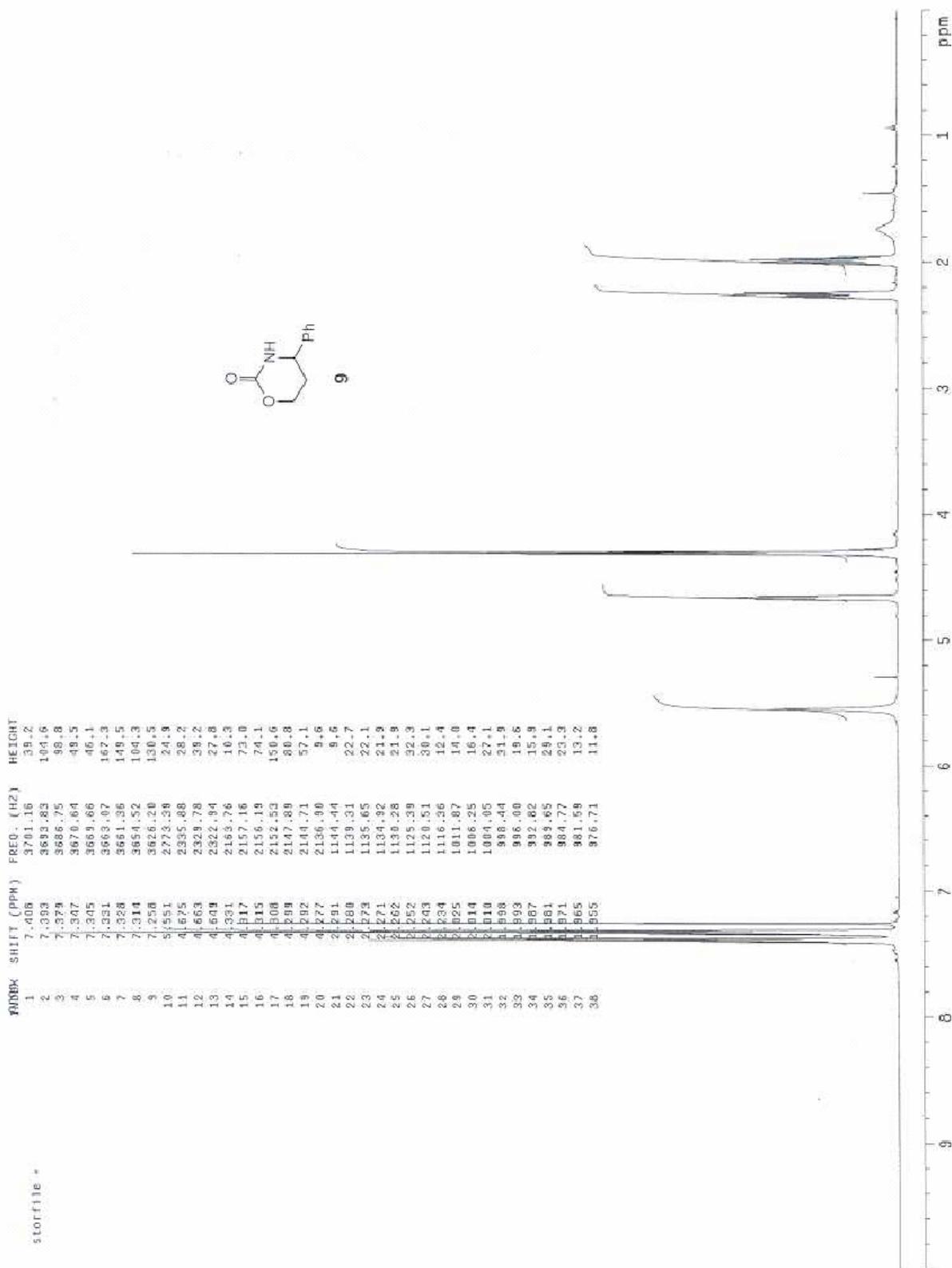
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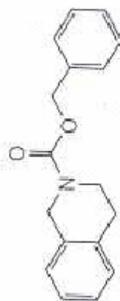
dfrq 399.865
dn h1
dppr 12
dof 0
dm mm
dmf -200
dseq 1.0
dres 0
homo 25.0
temp
PROCESSING
lb 0.25
wf file
proc ft
fn 131072
math f
wfft
wexp wft
cac apb t
msrcf ds
wbs
wnt wft
  
```



6j



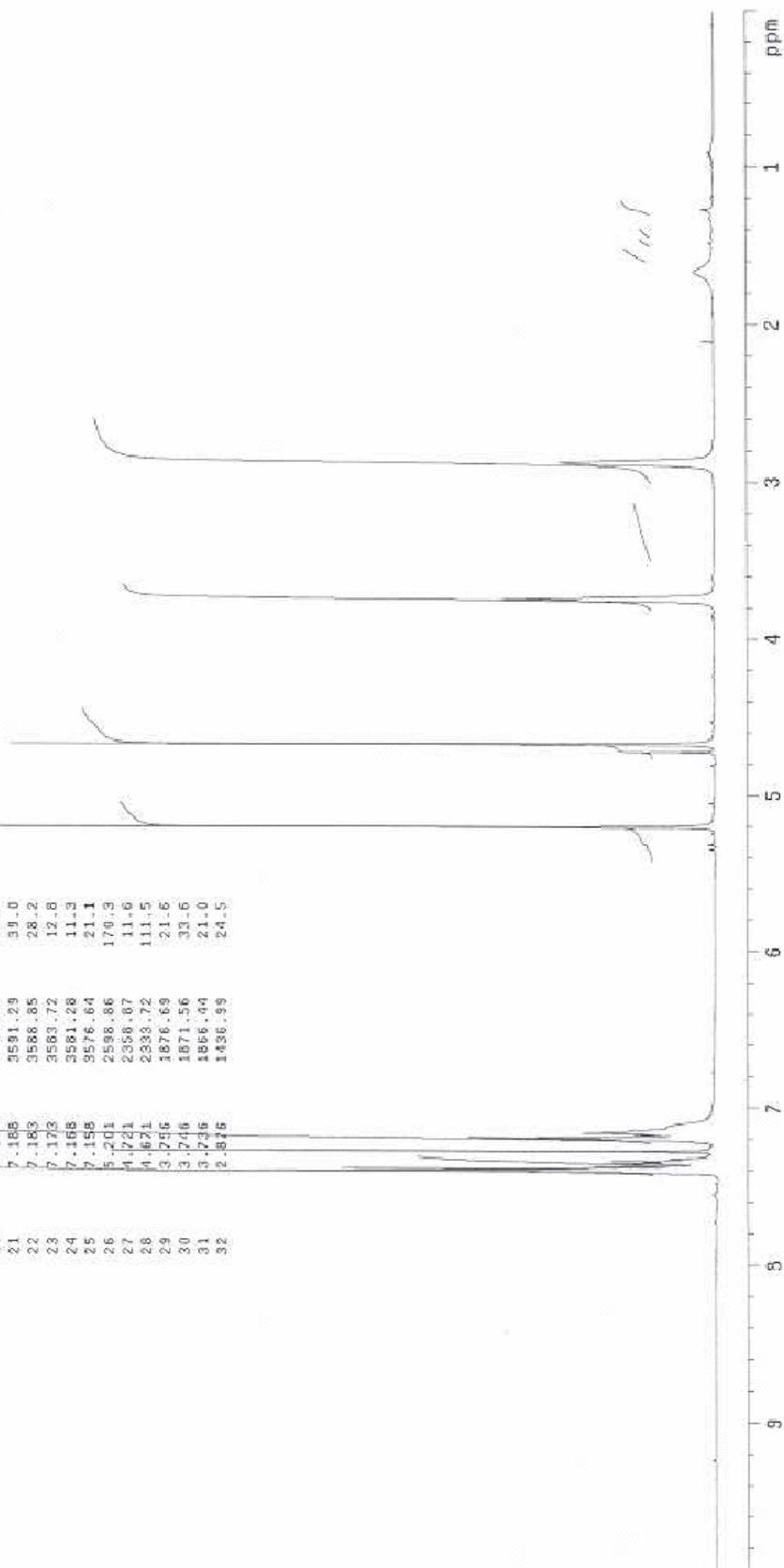




23

storfite =

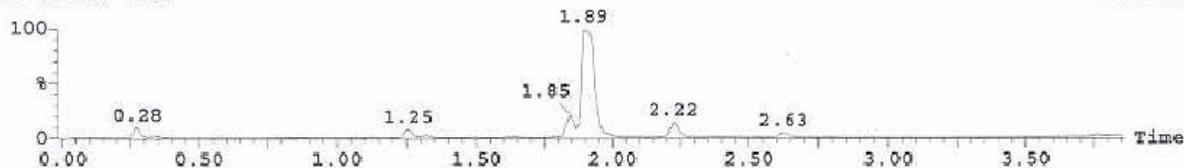
INDEX	SHIFT (PPM)	FREQ. (HZ)	HEIGHT
1	7.418	3706.28	10.1
2	7.414	3704.33	14.9
3	7.400	3697.25	93.2
4	7.398	3696.52	92.3
5	7.392	3693.34	28.3
6	7.386	3690.41	57.1
7	7.385	3689.92	55.1
8	7.381	3687.97	21.1
9	7.379	3683.82	15.0
10	7.370	3682.00	19.1
11	7.355	3674.79	13.5
12	7.351	3672.83	17.4
13	7.346	3670.15	11.0
14	7.344	3669.42	11.5
15	7.337	3666.00	16.4
16	7.325	3659.65	7.4
17	7.290	3637.43	10.4
18	7.279	3634.02	95.7
19	7.201	3598.13	26.8
20	7.194	3594.96	36.8
21	7.188	3591.29	39.0
22	7.183	3588.85	28.2
23	7.173	3583.72	12.8
24	7.168	3581.28	11.3
25	7.158	3576.64	21.1
26	5.201	2598.86	170.3
27	4.221	2358.87	11.6
28	4.071	2333.72	111.5
29	3.246	1876.69	21.6
30	3.246	1871.56	33.6
31	3.236	1866.44	21.0
32	2.815	1456.99	24.5



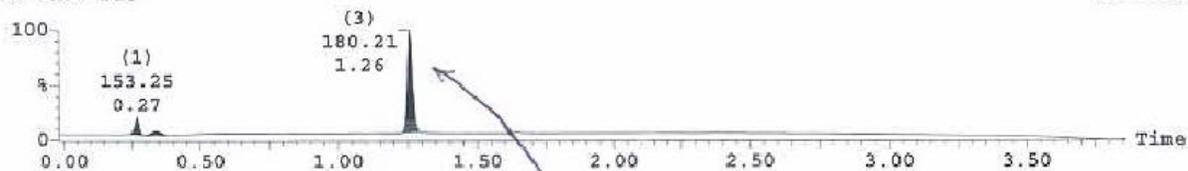
Copies of the LCMS and HRMS spectra from the carboxylation/Mitsunobu cyclization reaction of 4g using ^{18}O -labeled CO_2 to give 10 (eq 2):

Openlynx Report Page 1
 Sample: 1 Vial:1:83 Date:20-Feb-2004
JobCode:dinsmore2196
 Printed: Fri Feb 20 13:42:16 2004

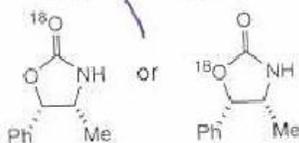
1: MS ES+ :TIC 4.8e+006



2: DAD: 215 6.3e+005

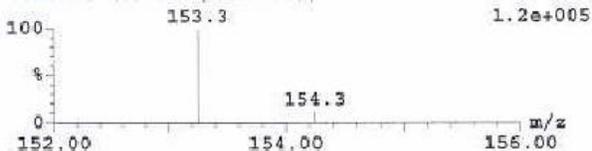


Peak Number	Time	Area %Total
1	0.27	12.28
2	0.34	6.62
3	1.26	81.10



Peak ID	Time	Area %BP
1	0.27	15

Combine (19:29 - (1:47:52))

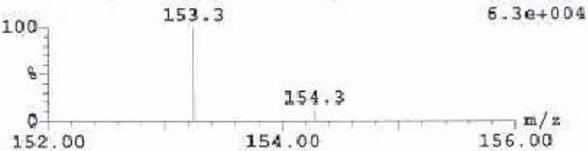


10

1:MS ES+
1.2e+005

Peak ID	Time	Area %BP
2	0.34	8

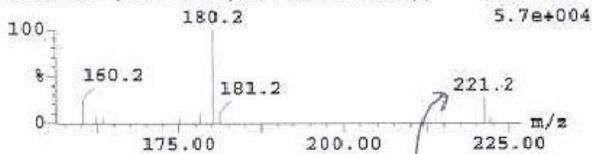
Combine (26:36 - (2:7+57:62))



1:MS ES+
6.3e+004

Peak ID	Time	Area %BP
3	1.26	100

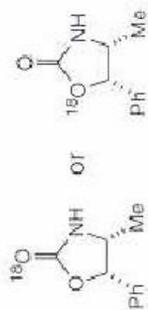
Combine (118:128 - (95:100+147:152))



$n\text{-Bu}_3\text{P}^{18}\text{O}$

C. J. Dinmore

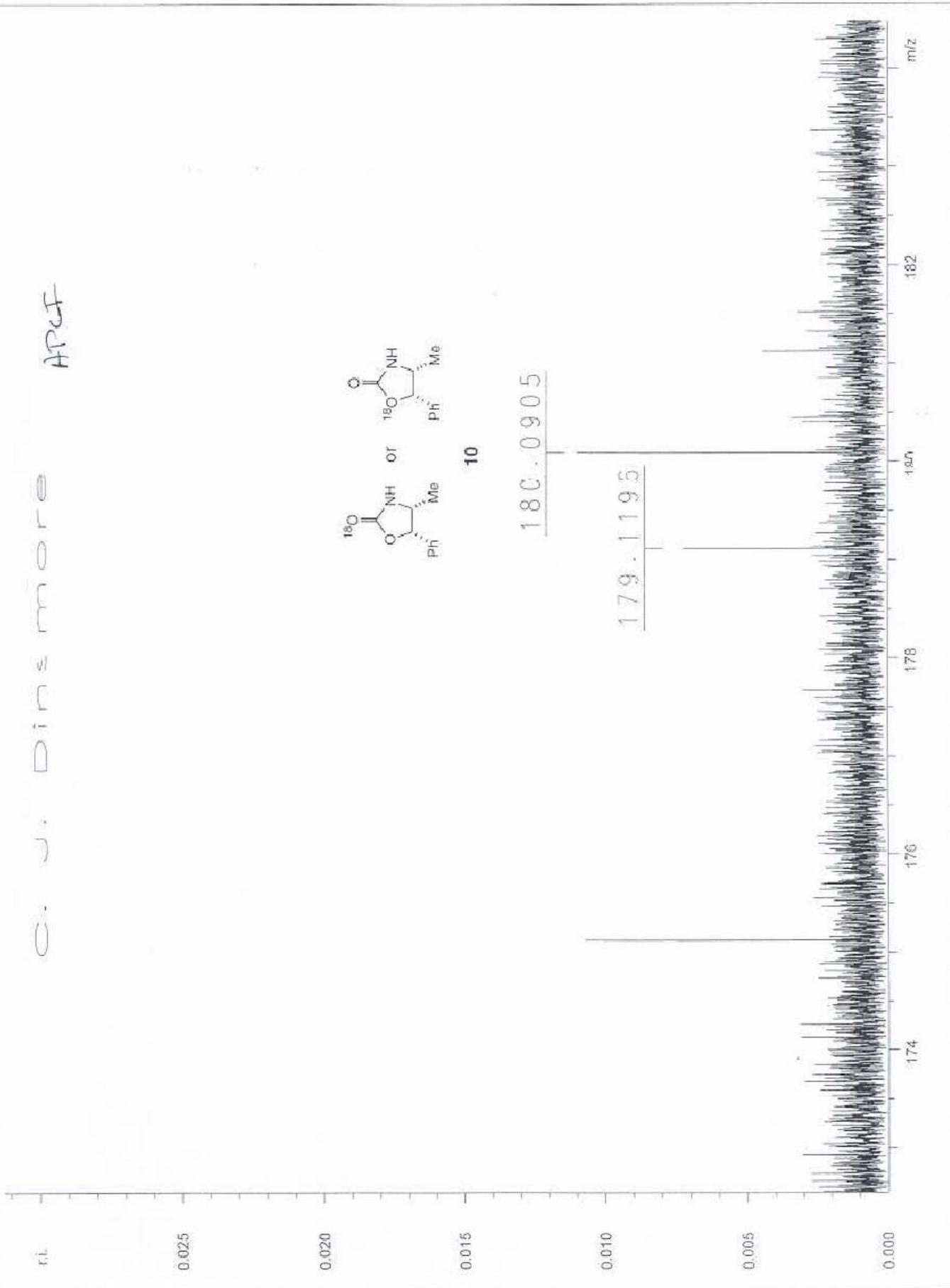
APCF

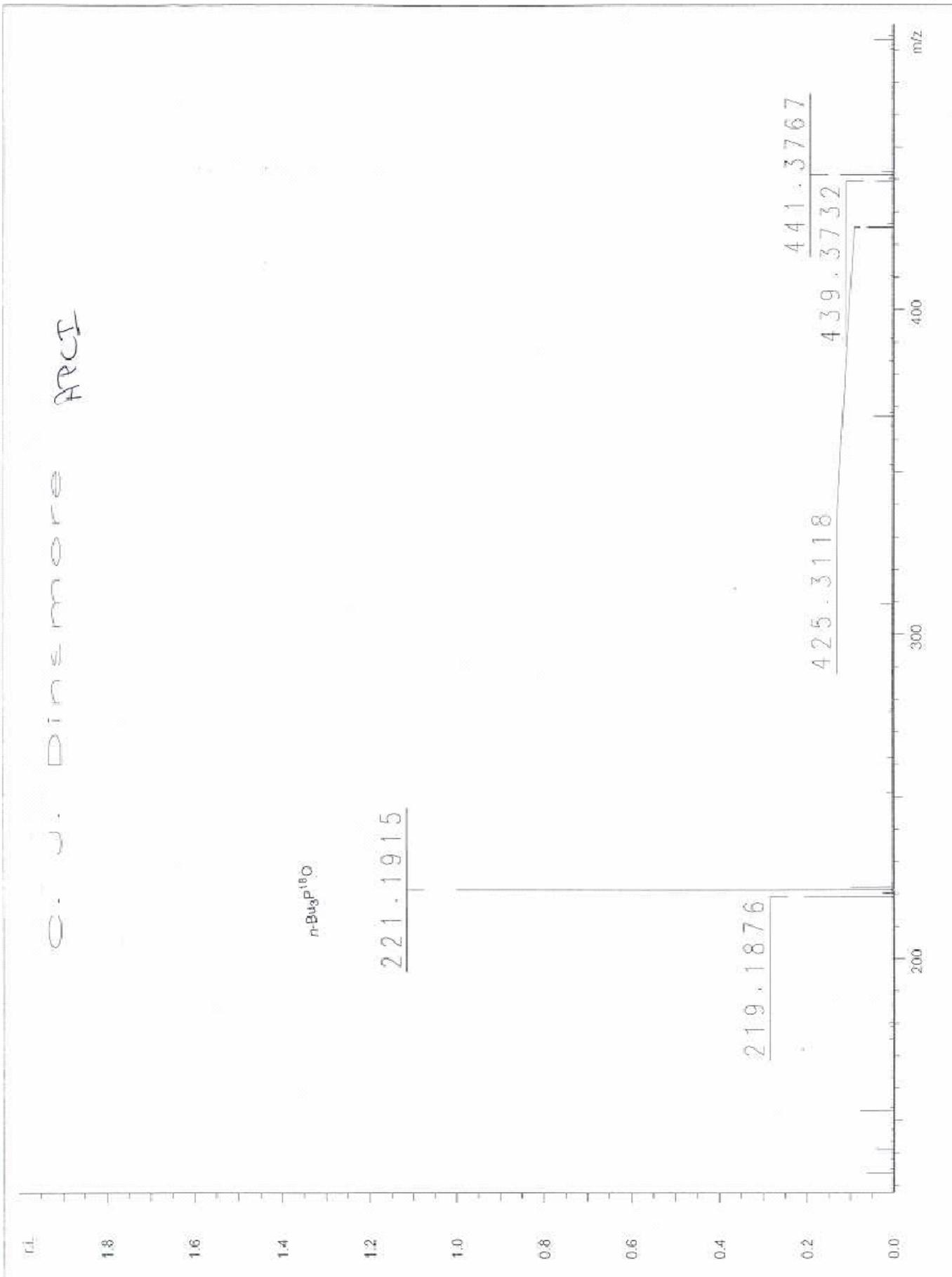


10

180.0905

179.1195

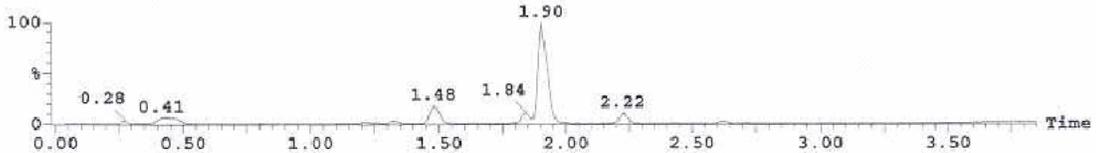




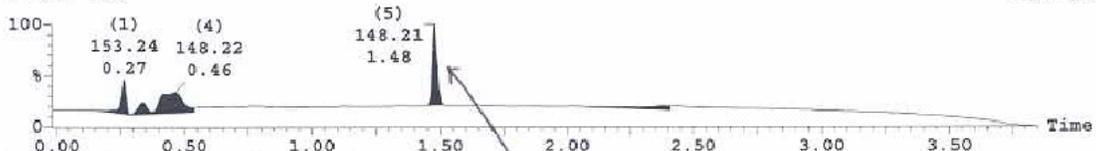
Copies of the LCMS and HRMS spectra from the carboxylation/Mitsunobu cyclization reaction of 4h using ¹⁸O-labeled CO₂ to give 11 (eq 3):

Openlynx Report Page 1
 Sample: 2 Vial:1:84 Date:20-Feb-2004
JobCode:dinsmore2198
 Printed: Fri Feb 20 13:47:58 2004

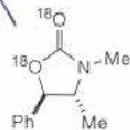
1: MS ES+ :TIC 5.0e+006



2: DAD: 215 2.0e+005



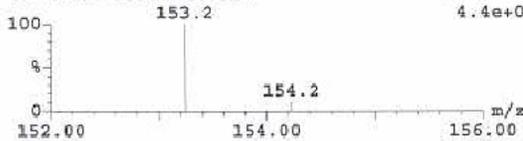
Peak Number	Time	Area %Total
1	0.27	15.08
2	0.34	8.90
3	0.42	12.64
4	0.46	22.41
5	1.48	30.36
6	2.38	10.60



11

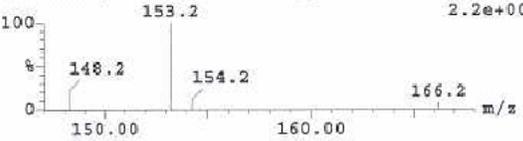
Peak ID	Time	Area %BP
1	0.27	50

Combine (19:29-46:51)



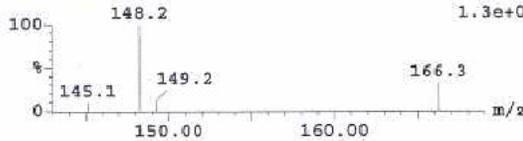
Peak ID	Time	Area %BP
2	0.34	29

Combine (26:36-(2:7+55:60))



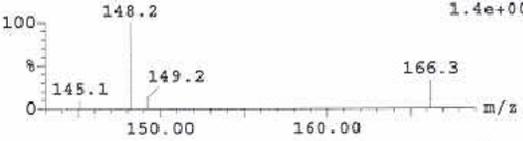
Peak ID	Time	Area %BP
3	0.42	42

Combine (34:44-(10:15+61:66))



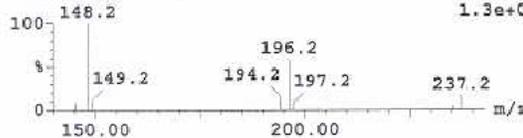
Peak ID	Time	Area %BP
4	0.46	74

Combine (39:49-(16:21+72:77))



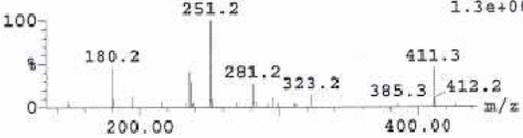
Peak ID	Time	Area %BP
5	1.48	100

Combine (140:148-(117:122+167:172))



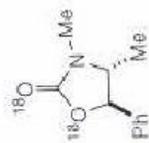
Peak ID	Time	Area %BP
6	2.38	35

Combine (229:239-(176:181+257:262))



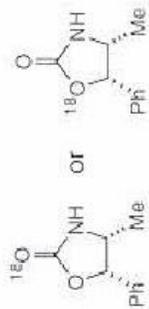
C. U. Dime more

APCS



11

196.1106



10

194.1063

192.1236

rel.

0.050

0.040

0.030

0.020

0.010

0.000

m/z

201

199

197

195

193

191

ARG

O. J. Dinimore

