

Click Chemistry as a Route to Cyclic Tetrapeptide Analogs: Synthesis of *cyclo*-[Pro-Val- ψ (triazole)-Pro-Tyr]

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Experimental Methods

General: The following general procedures were used in all reactions unless otherwise noted. Oxygen- and moisture-sensitive reactions were carried out using standard Schlenk techniques under a nitrogen atmosphere. Similarly sensitive liquids and solutions were transferred via gas-tight syringe or cannula. Reactions were stirred with an oven-dried, Teflon-covered magnetic stir bar. Removal of solvents was accomplished by evaporation on a Buchi rotary evaporator (water bath 40 °C). Tetrahydrofuran and diethyl ether were freshly distilled from sodium/benzophenone. Dry DMF and CH₂Cl₂ were freshly distilled from CaH₂. All commercially available reagents were used as received, unless indicated otherwise.

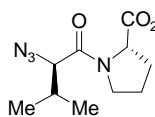
Chromatography: Analytical TLC chromatography was performed on 250 μ m silica gel 60 plates with 254 nm fluorescent indicator. Chromatographic purification refers to flash chromatography¹ using the indicated solvent (mixture) and Acros silica gel (0.035–0.070 mm). Analytical reverse phase HPLC was carried out on C₁₈ columns using gradients between 95 : 5 : 0.01 (water / acetonitrile / formic acid) and 5 : 95 : 0.01 (water / acetonitrile / formic acid).

Physical and Spectroscopic Measurements: NMR spectra were recorded in Fourier Transform mode on a Bruker ARX 400 (¹H at 400 MHz, ¹³C at 100 MHz) magnetic resonance spectrophotometer. ¹H NMR spectra are reported as chemical shifts in parts per million (ppm) downfield from a tetramethylsilane internal standard (0.00 ppm). Spin multiplicity is described by the following abbreviations: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, dd = doublet of doublets, and br = broad. Coupling constants (*J*) are reported in Hertz (Hz). ¹³C NMR spectra are reported as chemical shifts in ppm with the solvent resonance as the internal standard (CDCl₃: 77.07 ppm; MeOD: 49.00 ppm) and were recorded with complete heterodecoupling as APT (attached proton test) spectra. Infrared spectra were obtained from CDCl₃ solutions on a Bruker IFS 28 Fourier Transform spectrometer (FTIR) and are reported in wavenumbers (cm⁻¹). Fast Atom Bombardment (FAB) mass spectrometry was carried out using a JEOL JMS SX/SX 102A four-sector mass spectrometer, coupled to a JEOL MS-MP9021D/UPD system program. Samples were loaded in a matrix solution (3-nitrobenzyl alcohol) on a stainless steel probe and bombarded with xenon atoms with an energy of 3keV. During the high resolution FAB-MS measurements a resolving power of 10,000 (10% valley definition) was used. MALDI-TOF mass spectra were recorded on a Micromass ToFSpec 2EC (Micromass, Whytenshawe, UK). Samples (50 pmol) were dissolved in EtOAc, mixed with a concentrated solution of DHB in EtOAc and spotted directly on the stainless steel MALDI target. Silver nitrate was used for the ionization of the molecules.

Abbreviations: DHB, 2,5-dihydroxybenzoic acid, DIPEA, diisopropylethylamine; EDC, 1-(3-(dimethylamino)propyl)-3-ethyl-carbodiimide hydrochloride; HATU, *O*-(7-azabenzotriazole-1-yl)-

1,1,3,3-tetramethyluronium hexafluorophosphate; HOBt, *N*-hydroxybenzotriazole; MALDI-TOF: matrix assisted laser desorption/ionization–time of flight; MeOD, methanol-*d*₄; PE, petroleum ether; PyAOP, (7-azabenzotriazol-1-yloxy)-tris(pyrrolidino)phosphonium hexafluorophosphate

(*S*)-*tert*-butyl 1-((*S*)-2-azido-3-methylbutanoyl)pyrrolidine-2-carboxylate (5)



To a 100 mL round-bottomed flask equipped with a CaCO₃ drying tube and charged with azido valine **7**² (0.740 g, 5.17 mmol, 1 equiv) in freshly distilled CH₂Cl₂ (20 mL) was added EDC (1.09 g, 5.69 mmol, 1.1 equiv) and HOBt (0.734 g, 5.43 mmol, 1.05 equiv). *L*-proline *t*-butyl ester (0.927 g, 5.43 mmol, 1.05 equiv) was then added in freshly distilled CH₂Cl₂ (5 mL). After 16 h, this solution was diluted with CHCl₃ (40 mL) and washed with H₂O (1 × 70 mL), satd aq NaHCO₃ (1 × 70 mL), and 1N HCl(aq) (1 × 70 mL). The combined organics were then dried over Na₂SO₄, filtered, and concentrated in vacuo to yield azido valine-proline *t*-butyl ester **5** (1.447 g, 3.10 mmol, 60%) as a light yellow solid. This solid was carried on without further purification.

Data for compound **5**:

¹H NMR (CDCl₃, 400 MHz) δ 4.52–4.31 (m, 1H, Pro CH_α), 3.78–3.59 (m, 2H, Pro NCH₂), 3.39–3.19 (m, 1H, Val CH_α), 2.35–1.97 (m, 5H, Pro NCH₂CH₂; Pro CH_αCH₂; Val CH_αCH), 1.48 (s, 9H, C(CH₃)₃), 1.12 (d, *J* = 6.7 Hz, 3H, one of two Val CH(CH₃)₂), 1.07 (d, *J* = 6.7 Hz, 3H, one of two Val CH(CH₃)₂) ppm.

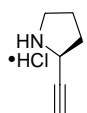
¹³C NMR (CDCl₃, 100 MHz) δ 170.77, 170.59, 168.70, 168.35, 82.60, 81.25, 65.99, 65.48, 60.00, 59.62, 47.01, 46.40, 42.74, 31.26, 30.71, 30.19, 29.48, 28.87, 27.76, 27.64, 24.58, 22.06, 19.01, 18.98, 18.90, 18.76 ppm.

IR 3448, 2974, 2876, 2079, 1736, 1654, 1432, 1392, 1368, 1258, 1226, 1154, 1093, 1044, 1014, 979, 953, 882, 847 cm⁻¹.

HMRS (FAB) Calculated for C₁₄H₂₅N₄O₃ (MH⁺): 297.1928
Found: 297.1917

[α]_D²⁰ –2.13 (*c* 1.97, CHCl₃)

(*S*)-2-ethynylpyrrolidine hydrochloride (9)



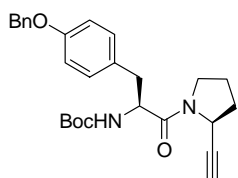
To a 100 mL round-bottomed flask charged with Boc-protected alkyne **8**³ (3.10 g, 15.86 mmol, 1 equiv) in Et₂O (48 mL) was added concentrated HCl(aq) (4.1 mL, 47.59 mmol, 3 equiv). After 24 h, TLC indicated consumption of the starting materials, so the volatiles were evaporated in vacuo to yield deprotected amino alkyne **9** (2.23 g) as a sticky, red oil. This oil was carried on without further purification.

Data for compound **9**:

¹H NMR (MeOD, 400 MHz) δ 4.42 (m, 1H, Pro H_α), 3.37 (m, 3H, NCH₂ and CCH), 2.40 (m, 1H, one of two NCH₂CH₂CH₂), 2.20 (m, 1H, one of two NCH₂CH₂CH₂), 2.10 (m, 2H, NCH₂CH₂CH₂) ppm.

IR 3388, 3230, 2957, 2739, 2548, 2125, 1646, 1455, 1408, 1195, 1050, 1020, 934 cm⁻¹.

***tert*-butyl (*S*)-3-(4-(benzyloxy)phenyl)-1-((*S*)-2-ethynylpyrrolidin-1-yl)-1-oxopropan-2-ylcarbamate (6)**



To a 100 mL round-bottomed flask equipped with a CaCO₃ drying tube and charged with deprotected amino alkyne **9** (1.676 g, 12.73 mmol, 1 equiv) in freshly distilled CH₂Cl₂ (20 mL) was added DIPEA (2.2 mL, 12.73 mmol, 1 equiv). After 10 min of stirring, this red solution was added to a 250 mL oven-dried flask equipped with a CaCO₃ drying tube and charged with Boc-Tyr(Bn)-OH (4.97 g, 13.37 mmol, 1.05 equiv), EDC (2.44 g, 12.73 mmol, 1 equiv), HOBt (1.72 g, 12.73

mmol, 1 equiv) and freshly distilled CH₂Cl₂ (15 mL). After 16 h of stirring at room temperature, this solution was diluted with CHCl₃ (90 mL) and washed with H₂O (1 × 100 mL), satd aq NaHCO₃ (1 × 100 mL), and 1N HCl(aq) (1 × 100 mL). The organics were then dried over Na₂SO₄, filtered, and concentrated in vacuo to yield a red solid. The product was purified via flash chromatography (6:4 PE:EtOAc) to afford Boc-protected tyrosine-proline alkyne **6** (3.48 g, 7.77 mmol, 61%) as a white solid.

Data for compound **6**:

¹H NMR (CDCl₃, 400 MHz) δ 7.38 (m, 5H, OCH₂Ph), 7.17 (m, 2H, Tyr ArH), 6.89 (d, *J* = 8.4 Hz, 2H, Tyr ArH), 5.43 (m, 1H, NH), 5.06 (s, 2H, OCH₂Ph), 4.74 (m, 1H, Tyr H_α), 4.50 (m, 1H, Pro H_α), 3.55–2.71 (m, 4H, Tyr CH_αCH₂ and Pro NCH₂), 2.29 (m, 1H, CCH), 2.04–1.83 (m, 4H, Pro NCH₂CH₂ and Pro CH_αCH₂), 1.43 (s, 9H, C(CH₃)₃) ppm.

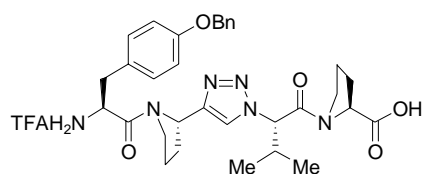
¹³C NMR (CDCl₃, 100 MHz) δ 169.59, 157.27, 154.75, 154.25, 136.77, 136.65, 130.53, 130.09, 128.61, 128.25, 128.18, 127.52, 127.00, 114.57, 114.45, 82.53, 81.67, 79.03, 72.31, 70.22, 69.45, 53.61, 53.22, 47.43, 47.12, 45.60, 45.20, 39.84, 38.81, 33.32, 31.55, 28.05, 24.36, 22.17 ppm.

IR 3429, 3302, 2978, 2876, 1711, 1644, 1612, 1583, 1511, 1446, 1391, 1366, 1295, 1242, 1174, 1110, 1024, 907, 863, 824 cm⁻¹.

HMRS (FAB) Calculated for C₂₇H₃₃N₂O₄ (MH⁺): 449.2442
Found: 449.2438

[α]_D²⁰ -53.8 (*c* 1.02, CHCl₃)

(S)-1-((S)-2-(4-((S)-1-((S)-2-amino-3-(4-(benzyloxy)phenyl)propanoyl)pyrrolidin-2-yl)-1H-1,2,3-triazol-1-yl)-3-methylbutanoyl)pyrrolidine-2-carboxylic acid triflic acid salt (3**):**



A 25 mL round-bottomed flask containing MeCN (5 mL) and THF (1 mL) was degassed with argon for thirty minutes. Azido valine-proline *t*-butyl ester **5** (0.296 mmol, 1.00 mmol, 1 equiv) and Boc-protected tyrosine-proline alkyne **6** (0.448 g, 1.00 mmol, 1 equiv) were added, followed by DIPEA (0.35 mL, 2.00 mmol, 2 equiv), 2,6-lutidine (0.23 mL, 2.00 mmol, 2 equiv), and CuI (0.019 g, 0.1 mmol,

0.1 equiv). After 16 h stirring under argon, the reaction was quenched by addition of satd aq NH₄Cl (10 mL), and the solution became a cloudy white. The volatiles were evaporated in vacuo, and the resulting aqueous slurry was extracted with CH₂Cl₂ (3 × 20 mL). The combined organics were washed with satd aq NH₄Cl (3 × 20 mL), 0.5 M KHSO₄ (1 × 20 mL), H₂O (1 × 20 mL), and brine (1 × 20 mL), dried over Na₂SO₄, filtered, and concentrated in vacuo. The product was then purified via flash chromatography (4:6 PE:EtOAc) to afford a white solid. This triazole product was dissolved in TFA (4 mL) and CHCl₃ (4mL) and stirred for five hours. The solvents were then removed in vacuo and subsequently coevaporated with toluene (2 × 10 mL) and CHCl₃ (2 × 10 mL) to afford triazole-substituted tetrapeptide **3** (0.460 g, 0.742 mmol, 74%) as a white solid. This solid was carried on without further purification.

Data for compound **3**:

¹H NMR (CDCl₃, 400 MHz) δ 8.23–7.61 (m, 4H), 7.44–6.88 (m, 9H), 5.23–4.99 (m, 2H), 4.79 (m, 1H), 4.51–4.07 (m, 3H), 3.97–3.59 (m, 4H), 3.35 (m, 1H), 3.02 (m, 1H), 3.73–1.57 (m, 9H), 1.31–1.01 (m, 3H), 0.81–0.65 (m, 3H) ppm.

¹³C NMR (CDCl₃, 100 MHz) δ 176.38, 169.19, 169.00, 168.91, 168.46, 167.51, 195.85, 159.79, 159.71, 159.51, 138.11, 132.41, 132.14, 131.90, 130.42, 130.03, 130.00, 129.93, 129.81, 129.61, 129.46, 129.44, 129.28, 129.06, 128.87, 128.79, 128.69, 127.13, 126.99, 126.68, 122.99, 116.98, 116.88, 116.75, 71.33, 71.29, 71.01, 69.39, 69.11, 68.94, 61.97,

61.40, 61.33, 56.08, 55.70, 55.44, 55.15, 55.00, 54.44, 49.80, 48.59, 48.52, 48.28, 48.13, 47.95, 38.80, 38.75, 34.92, 34.75, 34.52, 33.69, 33.19, 32.75, 31.08, 30.12, 26.58, 26.47, 25.40, 23.83, 23.66, 23.18, 30.71, 20.45, 19.75, 19.51, 19.25 ppm.

IR 2967, 2359, 1729, 1650, 1512, 1456, 1371, 1181, 1140, 1048, 1023, 911, 836 cm^{-1} .

Attempted Cyclization of Triazole-substituted Tetrapeptide **3** via EDC/HOBt-Mediated Coupling

To a 250 mL round-bottomed flask equipped with a CaCO_3 drying tube and charged with triazole-substituted tetrapeptide **3** (0.072 g, 0.100 mmol, 1 equiv) in freshly distilled CH_2Cl_2 (100 mL) was added DIPEA (0.034 mL, 0.100 mmol, 1 equiv). After 10 min of stirring, EDC (0.0191 g, 0.100 mmol, 1 equiv) and HOBt (0.0135 g, 0.100 mmol, 1 equiv) were added. After 16 h, this solution was quenched by addition of H_2O (50 mL) and concentrated in vacuo to remove the volatiles. The resulting aqueous slurry was diluted with CHCl_3 (50 mL), and the layers were separated. The organic layer was then washed with satd aq NaHCO_3 (1×50 mL), 1N HCl(aq) (1×50 mL), and H_2O (1×50 mL). The combined organics were dried over Na_2SO_4 , filtered, and concentrated in vacuo to yield a yellow solid (0.075 g). MALDI-TOF analysis indicated that the isolated products were dimers and higher oligomers.

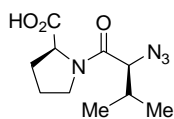
Attempted Cyclization of Triazole-substituted Tetrapeptide **3** via HATU-Mediated Coupling

To a 250 mL round-bottomed flask equipped with a CaCO_3 drying tube and charged with triazole-substituted tetrapeptide **3** (0.035 g, 0.0499 mmol, 1 equiv) in freshly distilled DMF (75 mL) was added HATU (0.019 mg, 0.100 mmol, 1 equiv) and DIPEA (0.035 mL, 0.200 mmol, 4 equiv). After 16 h, this solution was concentrated in vacuo and diluted with EtOAc (50 mL). The organic layers was then washed 0.5 M KHSO_4 (1×50 mL), H_2O (1×50 mL), and brine (1×50 mL). The combined organics were then dried over Na_2SO_4 , filtered, and concentrated in vacuo to yield a yellow solid (0.028 g). MALDI-TOF analysis indicated that the isolated products were dimers and higher oligomers.

Attempted Cyclization of Triazole-substituted Tetrapeptide **3** via PyOAP-Mediated Coupling

To a 250 mL round-bottomed flask equipped with a septum and $\text{N}_2(\text{g})$ inlet and charged with triazole-substituted tetrapeptide **3** (0.0726 g, 0.103 mmol, 1 equiv) in freshly distilled CH_2Cl_2 (100 mL) was added DIPEA (0.034 mL, 0.206 mmol, 2 equiv). After 10 min of stirring, PyAOP (0.0521 g, 0.103 mmol, 1 equiv) were added. After 16 h, this solution was concentrated in vacuo to yield a yellow solid (0.082 g). MALDI-TOF analysis indicated that the isolated products were dimers and higher oligomers.

(S)-1-((S)-2-azido-3-methylbutanoyl)pyrrolidine-2-carboxylic acid (**11**):



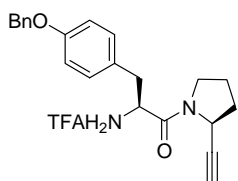
To a 100 mL round-bottomed flask charged with *t*Bu-protected valine-proline azide **5** (1.32 g, 4.45 mmol, 1 equiv) was added TFA (20 mL) and CHCl_3 (5 mL), and the mixture was stirred at rt. After 1 h, reaction progress was assessed by TLC. Since the starting material had been consumed, the mixture was concentrated in vacuo and subsequently coevaporated with CHCl_3 (4×10 mL) and toluene (2×10 mL) to afford valine-proline azide **11** (1.215 g, 4.15 mmol, quantitative) as a black oil. This oil was carried on without further purification.

Data for compound **11**:

$^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 8.80 (br s, 1H, CO_2H), 4.70–4.55 (m, 1H, Pro CH_α), 3.68 (t, $J = 7.3$ Hz, 2H, Pro NCH_2), 3.48–3.39 (m, 1H, Val CH_α), 2.39–2.01 (m, 5H, Pro NCH_2CH_2 ; Pro $\text{CH}_\alpha\text{CH}_2$; Val $\text{CH}_\alpha\text{CH}_2$), 1.12 (d, $J = 6.6$ Hz, 3H, one of two Val $\text{CH}(\text{CH}_3)_2$), 1.03 (d, $J = 6.7$ Hz, 3H, one of two Val $\text{CH}(\text{CH}_3)_2$) ppm.

IR 2973, 2881, 2550, 2099, 1786, 1723, 1626, 1602, 1448, 1346, 1218, 1155, 1047, 974, 910, 847 cm^{-1} .

(S)-2-amino-3-(4-(benzyloxy)phenyl)-1-((S)-2-ethynylpyrrolidin-1-yl) propan-1-one triflic acid salt (12)



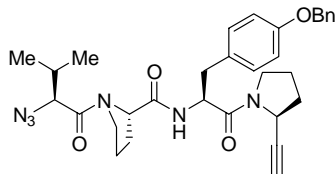
To a 25 mL round-bottomed flask charged with Boc-protected tyrosine-proline alkyne **6** (1.990 g, 4.45 mmol, 1 equiv) was added TFA (5 mL) and CHCl_3 (5 mL), and the mixture was stirred at rt. After 2 h, reaction progress was assessed by TLC. Since the starting material had been consumed, the mixture was concentrated in vacuo, and upon Et_2O (5 mL) addition, a white solid formed. The solid was filtered and washed in Et_2O (2×5 mL) to afford tyrosine-proline TFA-salt **12** (1.915 g, 4.14 mmol, 93%). This solid was carried on without further purification.

Data for compound **12**:

$^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 7.61–7.32 (m, 5H, OCH_2Ph), 7.20 (m, 2H, Tyr ArH), 6.91 (d, $J = 8.3$ Hz, 2H, Tyr ArH), 5.06 (s, 2H, OCH_2Ph), 4.67 (m, 1H, Tyr H_α), 4.26 (m, 1H, Pro H_α), 3.56–3.00 (m, 4H, Tyr $\text{CH}_\alpha\text{CH}_2$ and Pro NCH_2), 2.47 (m, 1H, CCH), 2.02–1.65 (m, 4H, Pro NCH_2CH_2 and Pro $\text{CH}_\alpha\text{CH}_2$) ppm.

IR 3288, 2880, 2287, 1660, 1612, 1510, 1454, 1365, 1329, 1242, 1202, 1175, 1135, 1040, 1026, 912, 861, 835 cm^{-1} .

(S)-1-((S)-2-azido-3-methylbutanoyl)-N-((S)-3-(4-(benzyloxy)phenyl)-1-((S)-2-ethynylpyrrolidin-1-yl)-1-oxopropan-2-yl) pyrrolidine-2-carboxamide (4)



To a 100 mL round-bottomed flask equipped with a CaCO_3 drying tube and charged with tyrosine-proline alkyne **6** (1.915 g, 4.15 mmol, 1 equiv) in freshly distilled CH_2Cl_2 (5 mL) was added DIPEA (0.75 mL, 4.15 mmol, 1 equiv). After 10 min of stirring, this solution was added to a 250 mL oven-dried flask equipped with a CaCO_3 drying tube and charged with proline-valine azide **5** (1.215 g, 4.45 mmol, 1.07 equiv), EDC (0.795 g, 4.15 mmol, 1 equiv), HOBt (0.560 g, 4.15 mmol, 1 equiv) and freshly distilled CH_2Cl_2 (10 mL). After 16 h, this black solution was diluted with CHCl_3 (50 mL) and washed with H_2O (1×100 mL), satd aq NaHCO_3 (1×100 mL), 1N $\text{HCl}(\text{aq})$ (1×100 mL), and H_2O (1×100 mL). The combined organics were then dried over Na_2SO_4 , filtered, and concentrated in vacuo to yield a brown solid. The product was purified via flash chromatography (100% EtOAc) to afford azido valine-proline-tyrosine-proline alkyne **4** (1.664 g, 2.90 mmol, 70%) as a white solid.

Data for compound **4**:

$^1\text{H NMR}$ (CDCl_3 , 400 MHz) δ 7.44–7.30 (m, 5H), 7.28–7.12 (m, 2H), 6.89–6.87 (m, 2H) 5.05 (m, 3H), 4.71 (m, 1H), 4.62 (m, 1H), 3.61–2.68 (m, 8H), 2.37–1.66 (m, 9H), 1.12 (m, 3H), 1.07–0.99 (m, 3H) ppm.

$^{13}\text{C NMR}$ (CDCl_3 , 100 MHz) δ 170.43, 170.15, 170.06, 169.56, 169.52, 168.99, 157.60, 157.50, 157.47, 136.91, 136.84, 136.77, 130.84, 130.70, 130.40, 130.31, 128.62, 128.35, 128.06, 127.75, 127.69, 127.22, 126.91, 114.72, 114.62, 82.51, 81.75, 72.29, 70.46, 70.31, 69.66, 69.64, 65.93, 65.52, 60.65, 60.06, 60.19, 59.61, 53.02, 52.88, 52.46, 52.37, 47.60, 47.55, 47.34, 45.99, 45.83, 45.59, 39.50, 38.54, 38.07, 33.54, 33.44, 31.96, 31.86, 30.64, 30.27, 30.24, 28.04, 28.35, 24.82, 24.75, 24.53, 22.33, 22.21, 20.87, 19.14, 19.11, 18.97, 18.78 ppm.

IR 3297, 2971, 2875, 2096, 1645, 1510, 1447, 1386, 1299, 1238, 1178, 1109, 1023, 911, 837 cm^{-1} .

HMRS (FAB) Calculated for $\text{C}_{32}\text{H}_{39}\text{N}_6\text{O}_4$ (MH^+):

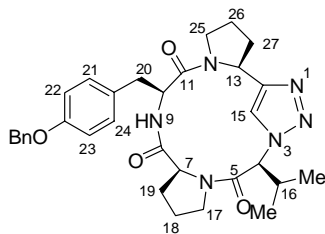
571.3035

Found:

571.3032

$[\alpha]_D^{20}$ -41.0 (*c* 1.40, CHCl₃)

Bn-protected cyclic peptide 10



To a 250 mL round-bottomed flask charged with alkyne azide **4** (0.111 g, 0.195 mmol, 1 equiv) in toluene (195 mL) was added DBU (87 μ L, 0.584 mmol, 3 equiv). The solution was degassed with argon for thirty minutes and then heated to reflux while flushing with argon. At reflux, CuBr (5.6 mg, 0.039 mmol, 0.2 equiv) was added, and the solution was stirred at reflux under argon for 16 h. The mixture was then cooled to rt and poured through a 2 in pad of Celite. The Celite pad was washed with MeOH (3 \times 25 mL). The filtrate was concentrated in vacuo to provide a blue-green oil. The product was purified via flash chromatography (3% MeOH in CH₂Cl₂) to afford Bn-protected cyclic peptide **10** (0.0782 g, 0.136 mmol, 70% yield) as a white solid.

Data for compound **10**:

¹H NMR (MeOD, 400 MHz) δ 7.63 (s, 1H, C₁₅H), 7.28–7.12 (m, 5H, OCH₂Ph), 7.14 (d, *J* = 8.6 Hz, 2H, Tyr C₂₁H and C₂₄H), 6.88 (d, *J* = 8.7 Hz, 2H, Tyr C₂₂H and C₂₃H), 5.49 (d, *J* = 6.8 Hz, 1H, Val C₄H), 5.07 (m, 3H, OCH₂Ph and Tyr C₁₀H), 4.45 (m, 1H, Pro C₁₃H), 4.31 (m, 1H, Pro C₇H), 3.71 (m, 2H, one of two Pro C₁₇H₂ and one of two Pro C₂₅H₂), 3.59 (m, 1H, one of two Pro C₂₅H₂), 3.45 (m, 1H, one of two Pro C₁₇H₂), 2.96 (dd, *J* = 13.1 and 9.1 Hz, 1H, one of two Tyr C₂₀H₂), 2.82 (dd, *J* = 13.1 and 5.4 Hz, 1H, one of two Tyr C₂₀H₂), 2.64 (m, 1H, Val C₁₆H), 2.42 (m, 1H, one of two Pro C₂₇H₂), 2.04 (m, 2H, one of two Pro C₂₇H₂ and one of two Pro C₁₉H₂), 1.90 (m, 3H, Pro C₂₆H₂ and one of two Pro C₁₈H₂), 1.78 (m, 1H, one of two Pro C₁₉H₂), 1.68 (m, 1H, one of two Pro C₁₈H₂), 1.08 (d, *J* = 6.6 Hz, 3H, one of two Val C₁₆(CH₃)₂), 0.81 (d, *J* = 6.8 Hz, 3H, one of two Val C₁₆(CH₃)₂) ppm.

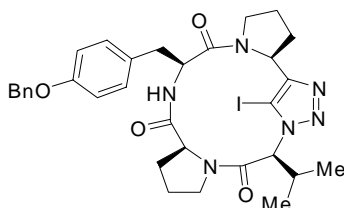
¹³C NMR (MeOD, 100 MHz) δ 172.41, 170.33, 168.41, 158.96, 152.34, 138.82, 131.52, 130.80, 129.47, 128.82, 128.56, 124.07, 115.79, 71.52, 70.90, 68.81, 63.25, 56.50, 55.46, 37.55, 35.36, 33.01, 31.02, 23.31, 22.27, 20.76, 18.37 ppm.

IR 3305, 2966, 1649, 1612, 1507, 1444, 1414, 1380, 1230, 1179, 1106, 1012 cm⁻¹.

HMRS	Calculated for C ₃₂ H ₃₉ N ₆ O ₄ (MH ⁺):	571.3035
(FAB)	Found:	571.3032
(ESI)	Found:	571.31

$[\alpha]_D^{20}$ -85.9 (*c* 0.92, CHCl₃)

Iodotriazole 13



Data for compound **13**:

¹H NMR (CDCl₃, 400 MHz) δ 7.44–7.31 (m, 5H), 7.19(d, *J* = 8.5 Hz, 2H), 6.87 (d, *J* = 9.0 Hz, 2H), 5.30 (d, *J* = 7.0 Hz, 1H), 5.04 (s, 2H), 4.82 (dd, *J* = 5.5 and 1.5 Hz, 1H), 4.62 (d, *J* = 9.5 Hz, 1H), 4.57–4.51 (m, 1H), 4.30 (d, *J* = 8.5 Hz, 1H), 4.10–4.05 (m, 1H), 3.72–3.65 (m, 1H), 3.42–3.34 (m, 2H), 3.07–3.01 (m, 2H), 2.84 (dd, *J* = 13.0 and 4.5,

1H), 2.36–2.30 (m, 2H), 2.13–2.07 (m, 2H), 1.95–1.91 (m, 3H), 1.85–1.80 (m, 1H), 1.17 (d, $J = 6.0$ Hz, 3H), 0.96 (d, $J = 6.5$, 3H) ppm.

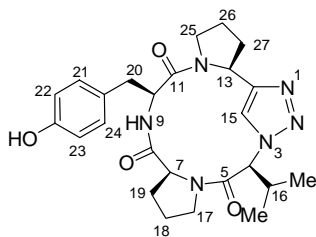
^{13}C NMR (MeOD, 100 MHz) δ 170.8, 168.2, 167.2, 157.4, 153.0, 137.1, 130.7, 129.5, 128.5, 127.9, 127.5, 114.5, 80.8, 69.9, 66.2, 62.4, 55.1, 54.4, 47.9, 47.1, 37.5, 33.2, 32.5, 30.5, 22.0, 21.9, 20.6, 18.9 ppm.

IR 3305, 2966, 1649, 1612, 1507, 1444, 1414, 1380, 1230, 1179, 1106, 1012 cm^{-1} .

MS	Calculated for $\text{C}_{32}\text{H}_{38}\text{IN}_6\text{O}_4$ (MH^+):	697.20
(FAB)	Found:	697.22
	Calculated for $\text{C}_{32}\text{H}_{37}\text{IN}_6\text{O}_4\text{Na}$ (MNa^+):	719.18
	(FAB) Found:	

719.21

Cyclo-[Pro-Val- ψ (triazole)-Pro-Tyr] 2



To a 50 mL round-bottomed flask charged with Bn-protected cyclic peptide **10** (13.7 mg, 0.024 mmol, 1 equiv) in methanol (5 mL) was added 10% Pd/C (7 mg). The resulting mixture was subjected to a three-cycle of vacuum- H_2 and was stirred at rt under a H_2 balloon for 16 h. The catalyst was then removed by filtration through Celite, and the filtrate concentrated in vacuo to afford cyclic peptide **2** (10.5 mg, 0.022 mmol, 93% yield) as a white solid.

Data for compound **2**:

^1H NMR (MeOD, 400 MHz) δ 7.62 (s, 1H, C_{15}H), 7.03 (d, $J = 8.4$ Hz, 2H, Tyr C_{21}H and C_{24}H), 6.65 (d, $J = 8.5$ Hz, 2H, Tyr C_{22}H and C_{23}H), 5.47 (d, $J = 6.8$ Hz, 1H, Val C_4H), 5.06 (m, 1H, Tyr C_{10}H), 4.41 (m, 1H, Pro C_{13}H), 4.32 (m, 1H, Pro C_7H), 3.68 (m, 2H, one of two Pro C_{17}H_2 and one of two Pro C_{25}H_2), 3.57 (m, 1H, one of two Pro C_{25}H_2), 3.44 (m, 1H, one of two Pro C_{17}H_2), 2.91 (dd, $J = 13.1$ and 9.2 Hz, 1H, one of two Tyr C_{20}H_2), 2.78 (dd, $J = 13.1$ and 5.4 Hz, 1H, one of two Tyr C_{20}H_2), 2.64 (m, 1H, Val C_{16}H), 2.42 (m, 1H, one of two Pro C_{27}H_2), 2.10 (m, 1H, one of two Pro C_{19}H_2), 2.03 (m, 1H, one of two Pro C_{27}H_2), 1.91 (m, 3H, Pro C_{26}H_2 and one of two Pro C_{18}H_2), 1.79 (m, 1H, one of two Pro C_{19}H_2), 1.69 (m, 1H, one of two Pro C_{18}H_2), 1.07 (d, $J = 6.7$ Hz, 3H, one of two Val $\text{C}_{16}(\text{CH}_3)_2$), 0.78 (d, $J = 6.8$ Hz, 3H, one of two Val $\text{C}_{16}(\text{CH}_3)_2$) ppm.

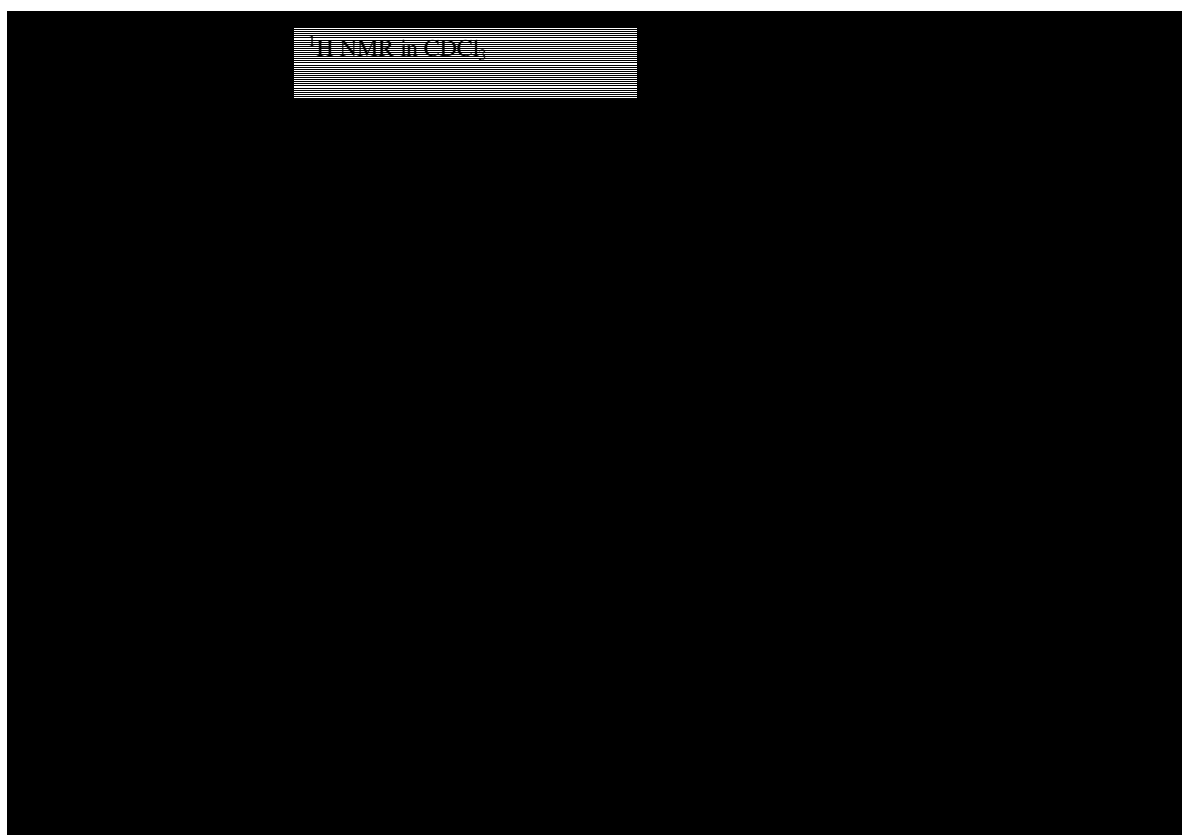
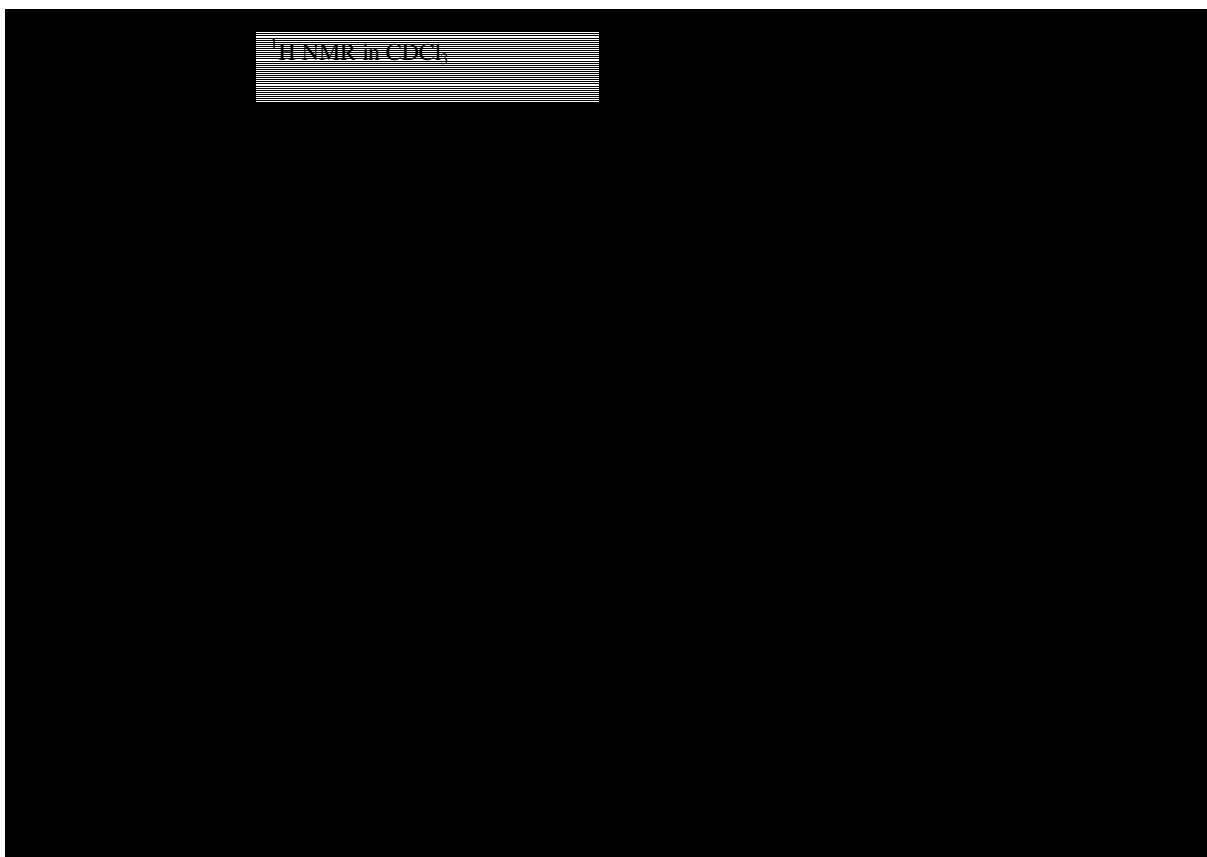
^{13}C NMR (MeOD, 100 MHz) δ 179.39, 170.37, 168.40, 157.17, 152.32, 131.47, 129.19, 124.09, 116.04, 68.78, 63.28, 56.51, 55.58, 49.70, 32.99, 31.01, 23.31, 22.26, 20.77 ppm.

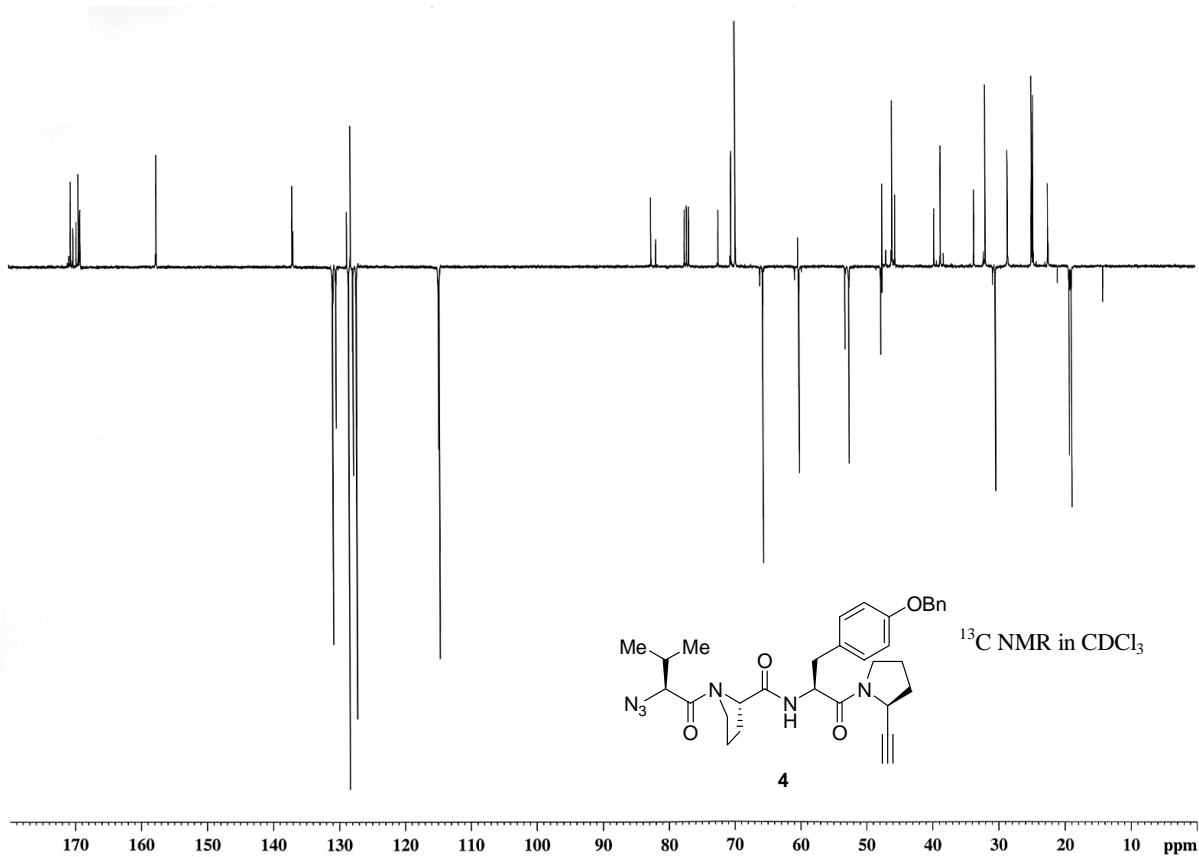
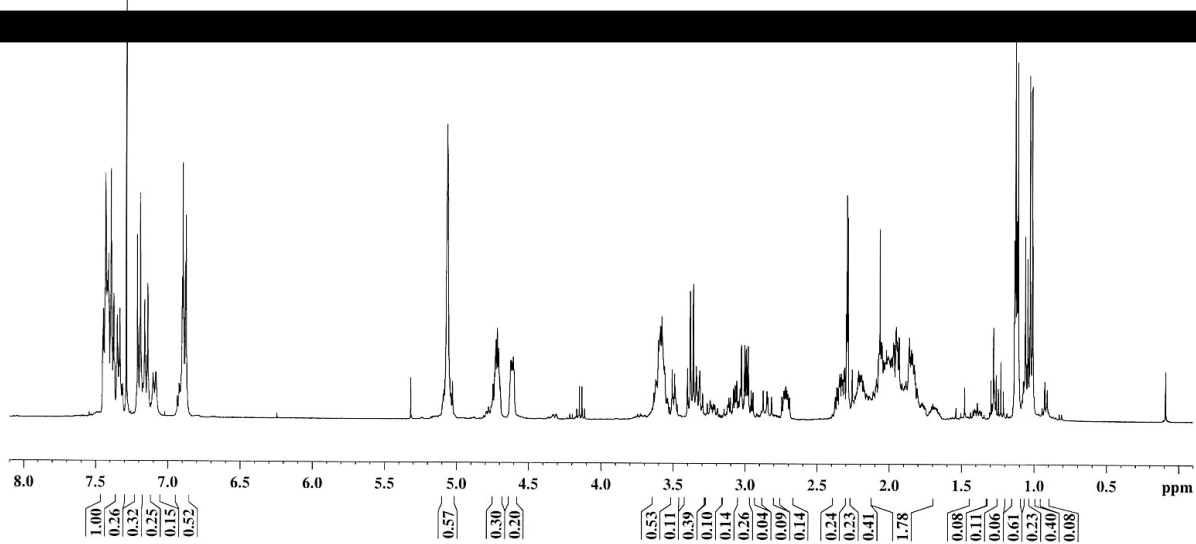
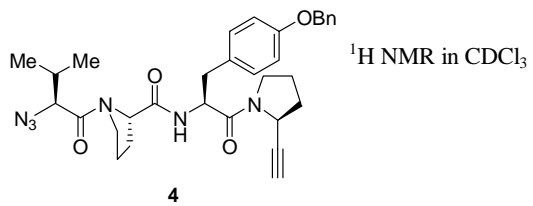
IR 3486, 2966, 2924, 2541, 1645, 1616, 1514, 1444, 1261, 1230, 1183, 1105, 1051, 974 cm^{-1} .

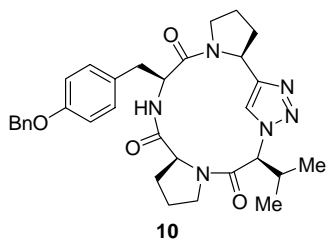
HMRS (FAB)	Calculated for $\text{C}_{25}\text{H}_{33}\text{N}_6\text{O}_4$ (MH^+):	481.2565
	Found:	481.2564

$[\alpha]_{\text{D}}^{20} -83.6$ (c 0.95, CHCl_3)

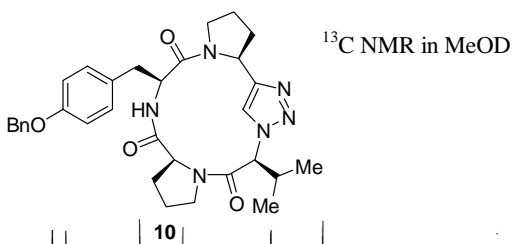
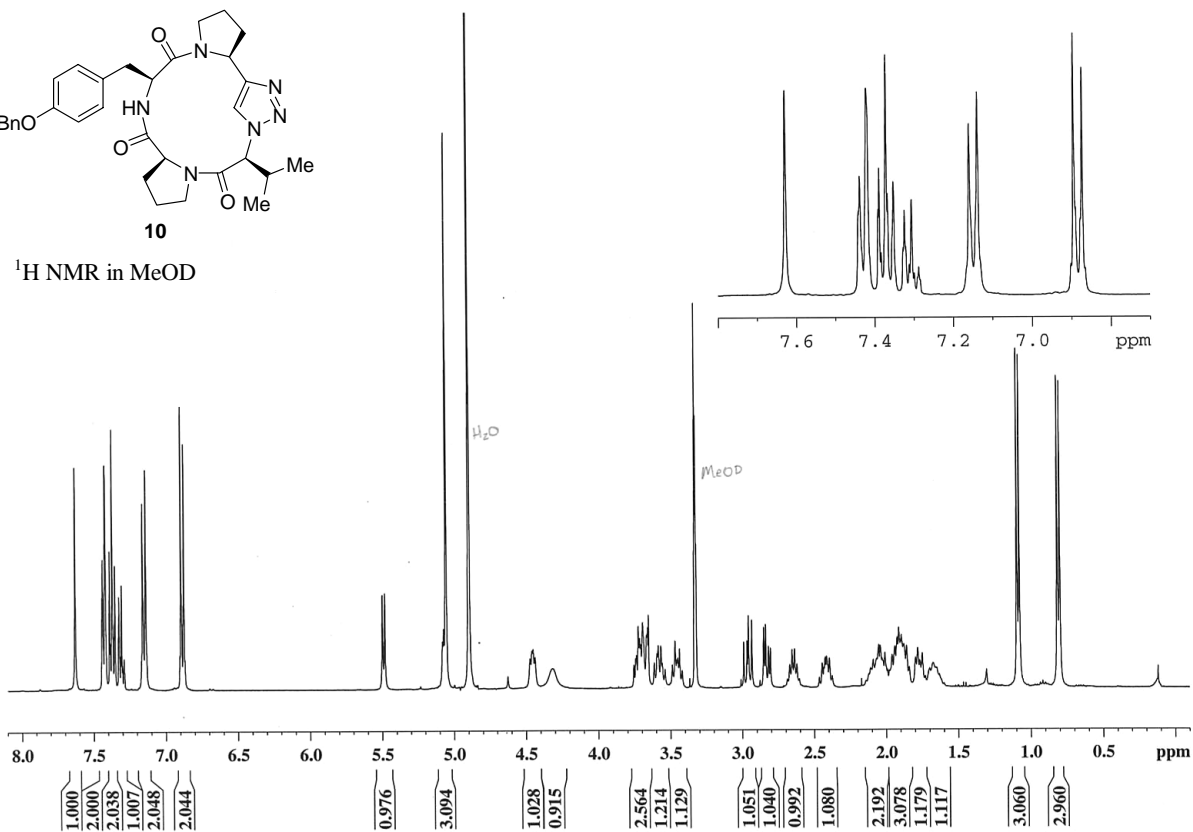
NMR Characterization



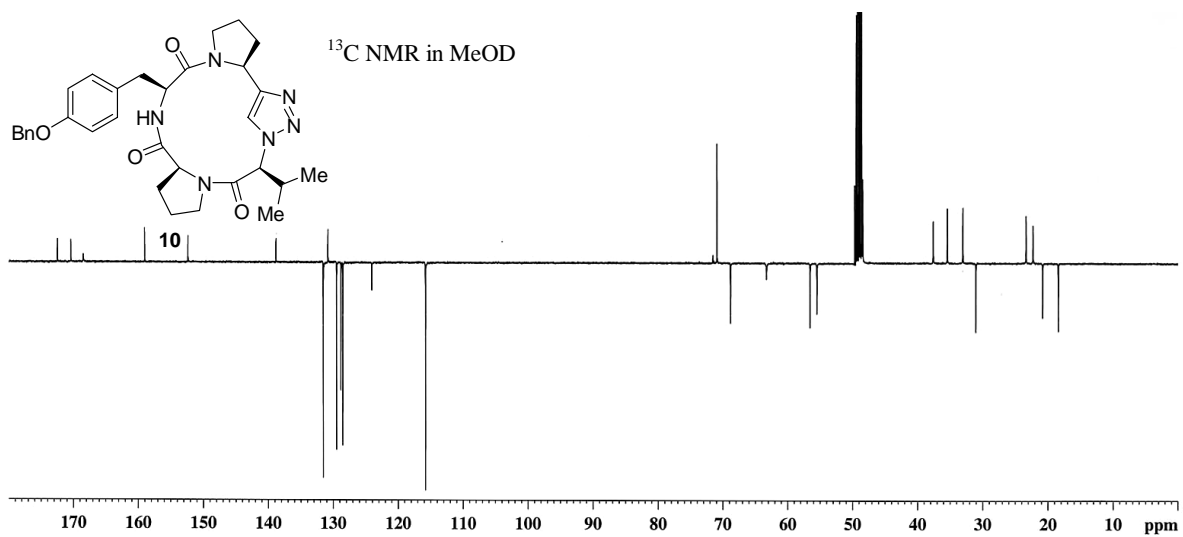


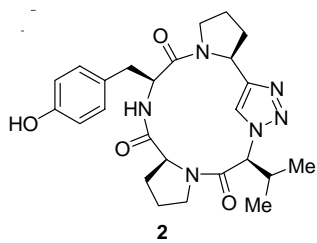


¹H NMR in MeOD

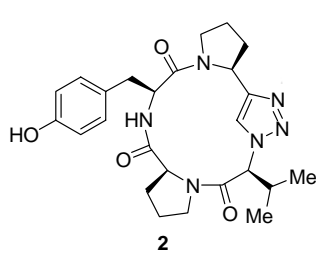
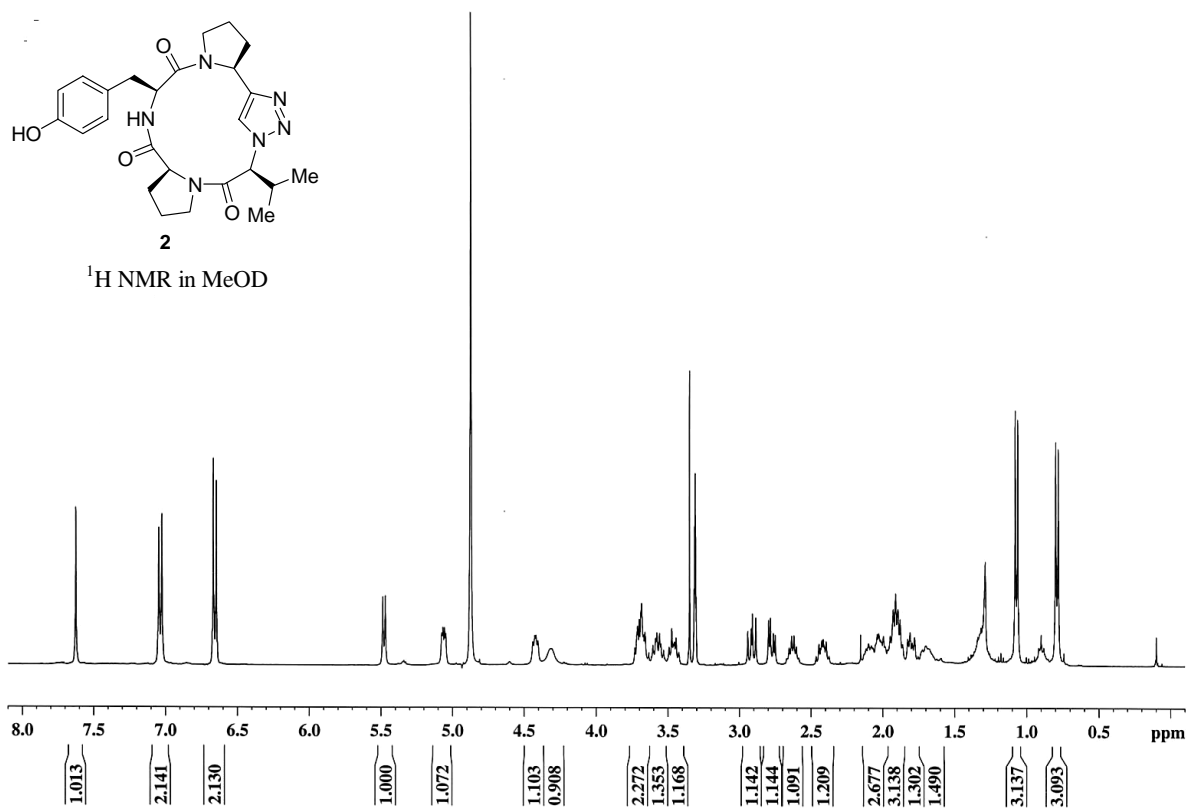


¹³C NMR in MeOD

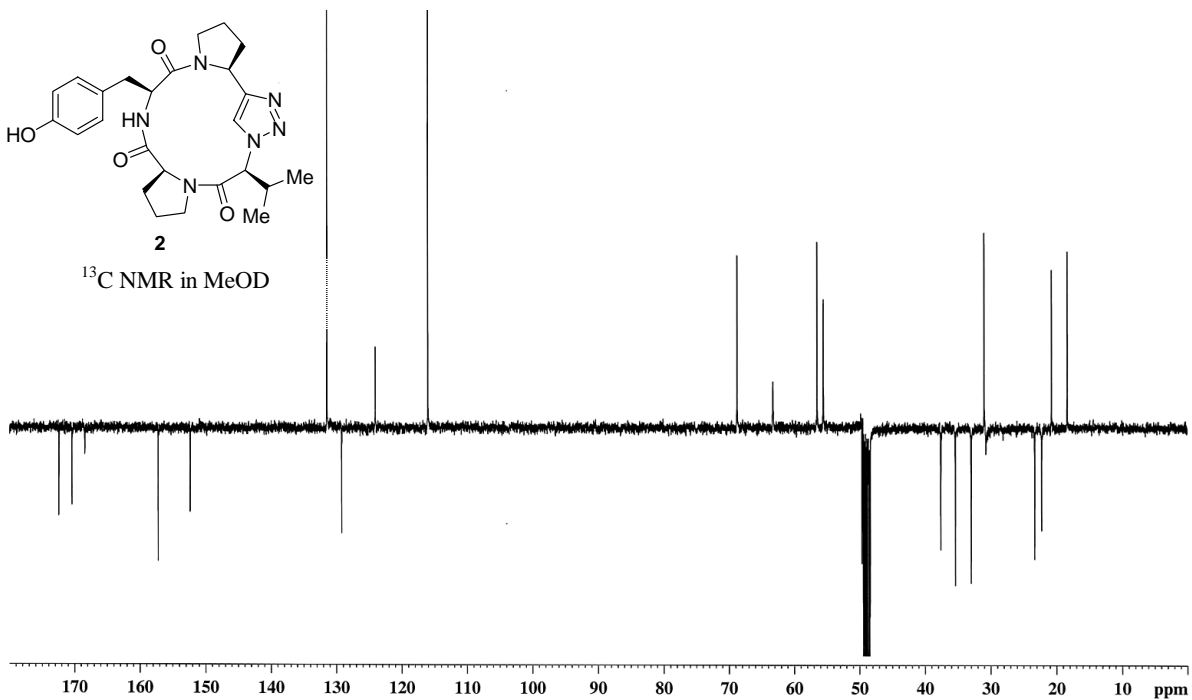


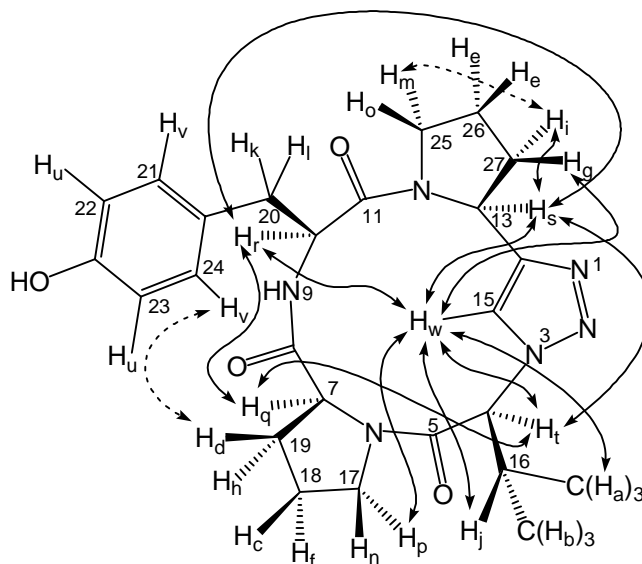


2
¹H NMR in MeOD



2
¹³C NMR in MeOD





SI Figure 1. Important NOESY correlations in cyclic peptide **2** in MeOD. Solid and dashed lines indicate observed strong and weak NOESY cross-peaks, respectively. See SI Table 1 for peak assignments.

SI Table 1. ^1H NMR peak assignments of cyclic peptide **2** in MeOD. Peaks were assigned via H,H-COSY and C,H-COSY (HETCOR) experiments.

δ (ppm)	Corresponding proton(s)	Multiplicity	Integration
0.78	H _a	d	3H
1.07	H _b	d	3H
1.69	H _c	m	1H
1.79	H _d	m	1H
1.91	H _e and H _f	m	3H
2.03	H _g	m	1H
2.10	H _h	m	1H
2.42	H _i	m	1H
2.64	H _j	m	1H
2.78	H _k	dd	1H
2.91	H _l	dd	1H
3.44	H _m	m	1H
3.57	H _n	m	1H
3.68	H _o and H _p	m	2H
4.32	H _q	m	1H
4.41	H _r	m	1H
5.06	H _s	m	1H
5.47	H _t	d	1H
6.65	H _u	d	2H
7.03	H _v	d	2H
7.62	H _w	s	1H

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- ¹ Still, W. C.; Kahn, M.; Mitra, A. *J. Org. Chem.* **1978**, *43*, 2923–2925.
- ² Lundquist, J. T., IV; Pelletier, J. C. *Org. Lett.* **2001**, *3*, 781–783.
- ³ Dickson, H. D.; Smith, S. C.; Hinkle, K. W. *Tetrahedron Lett.* **2004**, *45*, 5597–5599.