

Supporting Information

Electron-Donating Perylene-Tetracarboxylic Acids for Dye-Sensitized Solar Cells

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S1. Experimental Details.

General. ^1H and ^{13}C NMR spectra were recorded on a JEOL AL300 (300 MHz for ^1H) and a JEOL ECX-400P (400 MHz for ^1H , 99.6 MHz for ^{13}C) spectrometer in CDCl_3 and acetic acid- d_4 and chemical shifts were noted in δ ppm with reference to internal tetramethylsilane peak for CDCl_3 and internal residual solvent peak (CHD_2COOD , 2.03 ppm) for acetic acid- d_4 . Silica gel column chromatography was performed using UltraPure Silicagel (230-400 mesh, SiliCycle inc.). Alumina column chromatography was carried out using activated alumina (300 mesh, Wako). Reversed-phase column chromatography was performed using Cosmosil 76C₁₈-OPN (Nacalai). Thin layer chromatography (TLC) was carried out on aluminum plates coated with silica gel 60 F₂₅₄ (Merck), aluminium oxide 60 F₂₅₄ (Merck), or glass plates coated with functionalized silica gel (RP-18 F₂₅₄S, Merck). Infrared (IR) spectra were recorded in KBr pellet by using FT/IR-470Plus (JAS.CO). High-resolution mass spectra (HRMS) were recorded on a JEOL JMS-HX110A spectrometer. All reactions were carried out under nitrogen.

N,N'-bis(2,6-diisopropylphenyl)-1,7-dibromoperylene-3,4:9,10-tetracarboxylic acid bisimide (**1**),ⁱ *N*-cyclohexyl-1,7-bis(pyrrolidin-1-yl)-3,4:9,10-tetracarboxylic acid-3,4-anhydride-9,10-imide (**Cy-PMI**),ⁱⁱ and *tert*-butyl 4-aminobenzoateⁱⁱⁱ were synthesized as described before.

Synthesis.

***N,N'*-bis(2,6-diisopropylphenyl)-1,7-bis(pyrrolidin-1-yl)perylene-3,4:9,10-tetracarb**

oxylic acid bisimide (2). A solution of **1** (1.00 g, 1.16 mmol) in pyrrolidine (30 mL) was heated at 50 °C (external temperature) with an oil bath for 14 h. After evaporation, the residue was subjected to the silica gel column chromatography (CH₂Cl₂/AcOEt = 30/1, R_f (CH₂Cl₂) = 0.44), followed by the subjection to the alumina column chromatography (hexane/CHCl₃ = 1/2, R_f (CH₂Cl₂) = 0.50) to give **2** (408 mg, 0.480 mmol, 41% yield) as green solids; IR (KBr); 2963, 2929, 2870, 1698, 1662, 1592, 1580, 1560, 1507, 1485, 1467, 1453, 1417, 1382, 1342, 1309, 1245, 1231, 1200, 1180, 1128, 1098, 1058, 972, 946, 894, 870, 842, 810, 791, 772, 755, 737, 719, 695, 665, 650, 680, 606, 580, 550, 532, 496, 483, 475, 458, 442, 425, 415; ¹H NMR (300 MHz, CDCl₃) δ 8.58 (s, 2H), 8.52 (d, *J* = 8.1 Hz, 2H), 7.89 (d, *J* = 8.1 Hz, 2H), 7.48 (t, *J* = 7.4 Hz, 2H), 7.35 (d, *J* = 7.4 Hz, 4H), 3.92-3.72 (m, 4H), 3.00-2.75 (m, 4H), 2.80 (sept, *J* = 6.9 Hz, 4H), 2.20-1.90 (m, 8H), 1.19 (d, *J* = 6.9 Hz, 24H); ¹³C NMR (99.6 MHz, CDCl₃) δ 164.24, 164.20, 146.77, 145.70, 134.83, 131.19, 130.51, 129.41, 127.26, 123.99, 122.95, 121.99, 121.30, 119.20, 118.41, 52.45, 29.13, 25.91, 24.09, 24.04; HRMS (FAB, positive mode) found 848.4329, C₅₆H₅₆N₄O₄ requires 848.4302.

***N*-(2,6-diisopropylphenyl)-1,7-bis(pyrrolidin-1-yl)-3,4:9,10-tetracarboxylic**

acid-3,4-anhydride-9,10-imide (iPr-PMI). A solution of **2** (163 mg, 0.192 mmol) and KOH (718 mg, 12.8 mmol) in *tert*-butyl alcohol (33 mL) was heated to reflux. After stirred for 1 h, the reaction mixture was poured into a mixture of acetic acid (41 mL) and 1N HCl aq (22 mL) and stirred for 5 h at room temperature. The resulting mixture was poured into a biphasic mixture of CH₂Cl₂ (100 mL) and H₂O (50 mL).

After separation, the organic layer was washed with brine (100 mL) and dried over MgSO₄. After filtration and evaporation, the residue was subjected to the silica gel column chromatography (CH₂Cl₂, then CH₂Cl₂/AcOEt = 100/1, then 50/1, R_f (CH₂Cl₂) = 0.30). The reprecipitation from CH₂Cl₂/MeOH gave **iPr-PMI** (60.8 mg, 0.0881 mmol, 46% yield) as green solids. IR (KBr); 2963, 2929, 2868, 1764, 1730, 1700, 1664, 1592, 1579, 1559, 1543, 1507, 1456, 1419, 1345, 1311, 1244, 1228, 1201, 1145, 1128, 1101, 1009, 940, 867, 840, 804, 768, 739, 717, 692, 668, 649, 602, 578, 552, 531, 520, 509, 473, 440, 430, 413; ¹H NMR (300 MHz, CDCl₃) δ 8.57 (s, 1H), 8.50 (d, *J* = 7.8 Hz, 1H), 8.49 (s, 1H), 8.45 (d, *J* = 7.8 Hz, 1H), 7.78 (d, *J* = 7.8 Hz, 1H), 7.63 (d, *J* = 7.8 Hz, 1H), 7.49 (d, *J* = 7.5 Hz, 1H), 7.34 (d, *J* = 7.5 Hz, 2H), 3.90-3.72 (m, 4H), 3.00-2.80 (m, 4H), 2.77 (sept, *J* = 6.9 Hz, 2H), 2.20-1.95 (m, 8H), 1.18 (d, *J* = 6.9 Hz, 12H); ¹³C NMR (99.6 MHz, CDCl₃) δ 164.09, 164.03, 161.40, 160.84, 147.17, 146.42, 145.65, 136.02, 134.15, 131.03, 130.74, 130.41, 129.48, 128.80, 127.03, 124.76, 124.43, 124.01, 123.82, 122.79, 122.56, 122.51, 121.36, 119.83, 119.50, 117.46, 117.32, 114.52, 52.63, 52.48, 29.14, 25.89, 24.06, 24.01; HRMS (FAB, positive mode) found 689.2904, C₄₄H₃₉N₃O₅ requires 689.2890; UV-vis-NIR (λ_{max} (ε)) 708.0 nm (45600), 433.5 nm (17800).

***N*-(4-*tert*-butoxycarbonylphenyl)-*N'*-(2,6-diisopropylphenyl)-1,7-bis(pyrrolidin-1-yl)perylene-3,4:9,10-tetracarboxylic acid bisimide (3).** A mixture of **iPr-PMI** (32.4 mg, 0.0470 mmol), *tert*-butyl 4-aminobenzoate (25.5 mg, 0.132 mmol), zinc acetate dihydrate (6.0 mg, 0.027 mmol), and imidazole (500 mg) was heated at 140 °C in a

sealed Schlenk tube. After stirred for 2 h, the reaction mixture was cooled to room temperature. The resulting solids were dispersed in 1N HCl aq (6 mL) and sonicated for 30 min. The resulting mixture was poured into a biphasic mixture of 1N HCl aq (14 mL) and CHCl₃ (20 mL). After separation, the aqueous layer was extracted with CHCl₃ (20 mL). The combined organic layer was washed with brine (20 mL) and dried over MgSO₄. After filtration and evaporation, the residue was subjected to the silica gel column chromatography (CH₂Cl₂, then CH₂Cl₂/AcOEt = 100/1, then 50/1, then 40/1, R_f (CH₂Cl₂/AcOEt = 50/1) = 0.43). The reprecipitation from CH₂Cl₂/MeOH gave **3** (29.0 mg, 0.0335 mmol, 71% yield) as green solids. IR (KBr); 2963, 2927, 2868, 1698, 1665, 1592, 1579, 1560, 1508, 1455, 1443, 1342, 1307, 1290, 1246, 1231, 1198, 1180, 1166, 1117, 1057, 1020, 972, 946, 893, 860, 840, 828, 807, 772, 753, 740, 715, 688, 662, 642, 633, 622, 594, 550, 530, 521, 513, 500; ¹H NMR (300 MHz, CDCl₃) δ 8.58 (s, 1H), 8.56 (s, 1H), 8.53-8.50 (m, 2H), 8.20 (d, *J* = 7.5 Hz, 2H), 7.80 (d, *J* = 8.1 Hz, 1H), 7.76 (d, *J* = 8.1 Hz, 1H), 7.49 (t, *J* = 8.1 Hz, 1H), 7.41 (d, *J* = 8.1 Hz, 2H), 7.35 (d, *J* = 7.5 Hz, 2H), 3.95-3.70 (m, 4H), 3.00-2.80 (m, 4H), 2.79 (sept, *J* = 6.6 Hz, 2H), 2.20-1.95 (m, 8H), 1.64 (s, 9H), 1.19 (d, *J* = 6.6 Hz, 12H); ¹³C NMR; (99.6 MHz, CDCl₃) δ 165.12, 164.21, 164.10, 146.84, 146.63, 145.69, 139.58, 135.09, 134.65, 132.25, 131.16, 130.57, 130.47, 129.43, 128.78, 127.24, 124.21, 123.99, 122.92, 122.75, 122.15, 121.71, 121.26, 119.38, 118.95, 118.64, 118.16, 81.28, 52.43, 29.14, 28.23, 25.90, 24.08, 24.03; HRMS (FAB, positive mode) found 864.3875, C₅₅H₅₂N₄O₆ requires 864.3887.

***N*-(4-carboxyphenyl)-*N'*-(2,6-diisopropylphenyl)-1,7-bis(pyrrolidin-1-yl)perylene-3,4:9,10-tetracarboxylic acid bisimide (*iPr*-PBI).** A mixture of **3** (29.0 mg, 0.0335 mmol), trifluoroacetic acid (10 mL), 5wt% H₂SO₄ aq (7.5 mL), and CHCl₃ (10 mL) was heated to reflux. After stirred for 24 h, the reaction mixture was cooled to room temperature. The resulting biphasic mixture was separated and the aqueous layer was extracted with CHCl₃ (20 mL). The combined organic layer was washed with saturated NaHCO₃ aq (20 mL), saturated NH₄Cl aq (20 mL), and brine (50 mL) and dried over MgSO₄. After filtration and evaporation, the residue was subjected to the silica gel column chromatography (AcOEt/MeOH = 10/1, R_f = 0.40), followed by the subsection to the reversed-phase column chromatography (AcOEt/MeOH = 1/4, R_f = 0.53). The reprecipitation from CH₂Cl₂/hexane gave ***iPr*-PBI** (22.3 mg, 0.0276 mmol, 82% yield) as green solids. IR (KBr); 3440, 2963, 2930, 2869, 1697, 1664, 1592, 1578, 1559, 1507, 1455, 1415, 1384, 1343, 1309, 1246, 1231, 1200, 1128, 1100, 1057, 1021, 972, 946, 843, 825, 807, 775, 752, 739, 718, 693, 669, 657, 611, 600, 582, 567, 557, 545, 534, 519, 512; ¹H NMR (300 MHz, acetic acid-*d*₄) δ 8.64 (s, 1H), 8.59 (d, *J* = 8.4 Hz, 1H), 8.54 (s, 1H), 8.51 (d, *J* = 8.4 Hz, 1H), 8.29 (d, *J* = 8.4 Hz, 2H), 7.83-7.80 (m, 2H), 7.57 (d, *J* = 8.4 Hz, 2H), 7.47 (t, *J* = 7.8 Hz, 1H), 7.35 (d, *J* = 7.8 Hz, 2H), 3.95-3.75 (m, 4H), 2.95-3.75 (m, 4H), 2.83 (sept, *J* = 6.9 Hz, 1H), 2.20-1.90 (m, 8H), 1.14 (d, *J* = 6.9 Hz, 12H); HRMS (FAB, positive mode) found 808.3221, C₅₁H₄₄N₄O₆ requires 808.3261; UV-vis-NIR (λ_{max} (ε)) 709.0 nm (42000), 435.5 nm (16800). ¹³C NMR spectrum could not be measured because of the low solubility.

***N*-(4-*tert*-butoxycarbonylphenyl)-*N'*-cyclohexyl-1,7-bis(pyrrolidin-1-yl)perylene-3,4:9,10-tetracarboxylic acid bisimide (4).** A mixture of **Cy-PMI** (33.8 mg, 0.0553 mmol), *tert*-butyl 4-aminobenzoate (26.0 mg, 0.135 mmol), zinc acetate dihydrate (5.0 mg, 0.023 mmol), and imidazole (400 mg) was heated at 140 °C in a sealed Schlenk tube. After stirred for 2 h, the reaction mixture was cooled to room temperature. The resulting solids were dispersed in 1N HCl aq (6 mL) and sonicated for 30 min. The resulting mixture was poured into a biphasic mixture of 1N HCl aq (14 mL) and CHCl₃ (20 mL). After separation, the aqueous layer was extracted with CHCl₃ (20 mL). The combined organic layer was washed with brine (40 mL) and dried over MgSO₄. After filtration and evaporation, the residue was subjected to the silica gel column chromatography (CH₂Cl₂, then CH₂Cl₂/AcOEt = 100/1, then 50/1, then 40/1, then 30/1, R_f (CH₂Cl₂/AcOEt = 40/1) = 0.40). The reprecipitation from CH₂Cl₂/MeOH gave **4** (37.4 mg, 0.0475 mmol, 86% yield) as green solids. IR (KBr); 2970, 2929, 2853, 1693, 1654, 1593, 1578, 1560, 1507, 1483, 1453, 1415, 1339, 1306, 1291, 1257, 1245, 1230, 1213, 1190, 1164, 1120, 1020, 981, 949, 927, 895, 867, 839, 828, 806, 739, 767, 754, 716, 702, 679, 654, 633, 617, 585, 558, 530, 521, 511; ¹H NMR (400 MHz, CDCl₃) δ 8.51 (s, 1H), 8.48 (s, 1H), 8.44 (d, *J* = 7.6 Hz, 1H), 8.42 (d, *J* = 7.6 Hz, 1H), 8.19 (d, *J* = 8.4 Hz, 2H), 7.75 (d, *J* = 7.6 Hz, 1H), 7.66 (d, *J* = 7.6 Hz, 1H), 7.41 (d, *J* = 8.4 Hz, 2H), 5.13-5.05 (m, 1H), 3.85-3.65 (m, 4H), 2.95-2.70 (m, 4H), 2.65-2.55 (m, 2H), 2.15-1.87 (m, 10H), 1.80-1.70 (m, 3H), 1.63 (s, 9H), 1.55-1.33 (m, 3H); ¹³C NMR (99.6 MHz, CDCl₃) δ 165.15, 164.52, 164.11, 146.78, 146.38, 139.61, 134.92, 133.94, 132.20, 130.55, 130.24, 129.97, 128.79, 127.16, 126.58, 124.19, 123.68, 122.67, 122.63, 122.20,

121.41, 121.11, 120.80, 119.91, 118.78, 118.68, 117.62, 81.25, 53.80, 52.27, 29.20, 28.23, 26.63, 25.83, 25.55; HRMS (FAB, positive mode) found 786.3421, C₄₉H₄₆N₄O₆ requires 786.3417.

***N*-(4-carboxylphenyl)-*N*'-cyclohexyl-1,7-bis(pyrrolidin-1-yl)perylene-3,4:9,10-tetra carboxylic acid bisimide (Cy-PBI).** A mixture of **4** (10.3 mg, 0.0131 mmol), trifluoroacetic acid (10 mL), 5wt% H₂SO₄ aq (7.5 mL), and CHCl₃ (10 mL) was heated to reflux. After stirred for 43 h, the reaction mixture was cooled to room temperature. The resulting biphasic mixture was separated and the aqueous layer was extracted with CHCl₃ (20 mL). The combined organic layer was washed with saturated NaHCO₃ aq (20 mL), saturated NH₄Cl aq (20 mL), and brine (50 mL) and dried over MgSO₄. After filtration and evaporation, the residue was subjected to the silica gel column chromatography (AcOEt/MeOH = 10/1, then 5/1, then 1/1, R_f (AcOEt/MeOH = 10/1) = 0.30), followed by the subjection to the reversed-phase column chromatography (AcOEt/MeOH = 1/4, R_f = 0.44). The reprecipitation from CH₂Cl₂/hexane gave **Cy-PBI** (6.1 mg, 0.0083 mmol, 64% yield) as green solids. IR (KBr); 3450, 2929, 2853, 1692, 1651, 1592, 1579, 1560, 1507, 1483, 1453, 1416, 1339, 1309, 1245, 1230, 1214, 1198, 1124, 1021, 980, 948, 927, 881, 860, 907, 770, 715, 660, 626, 617, 605, 592, 556, 532, 509, 499; ¹H NMR (400 MHz, acetic acid-*d*₄) δ 8.51-8.32 (m, 4H), 8.30 (d, *J* = 8.6 Hz, 2H), 7.68-7.45 (m, 2H), 7.61 (d, *J* = 8.6 Hz, 2H), 5.12-5.03 (m, 1H), 3.80-3.60 (m, 4H), 2.90-2.60 (m, 6H), 2.20-1.70 (m, 13H), 1.55-1.25 (m, 3H); HRMS (FAB, positive mode) found 730.2784, C₄₅H₃₈N₄O₆ requires 730.2791; UV-vis-NIR (λ_{max} (ε))

704.5 nm (42300), 434.0 nm (16700). ^{13}C NMR spectrum could be measured because of the low solubility.

Spectral measurements. Steady-state absorption spectra were measured with a Lambda 900 (PerkinElmer) UV/VIS/NIR spectrometer with a data interval of 0.5 nm. These spectra were taken with about 10^{-4} - 10^{-6} M solutions in a quartz cell with pathlength of 1 cm and 1 mm. Steady-state fluorescence spectra were acquired on a SPEX FluoroMax-3 spectrometer with a data interval of 1 nm. These spectra were taken with about 10^{-6} M solutions in a quartz cell with pathlength of 1 cm. The solution was degassed by bubbling with Ar for 30 min before measurements. Spectral grade CH_2Cl_2 (Wako) was used without further purification for these measurements.

Electrochemical measurements. Differential pulse voltammetry measurements were performed on an ALS660A electrochemical analyzer in deaerated CH_2Cl_2 containing 0.1 M TBAPF₆ as a supporting electrolyte. A conventional three-electrode cell was used with a grassy carbon working electrode and a platinum wire as a counter electrode. The measured potentials were recorded with respect to the Ag/AgNO₃ reference electrode (Fc/Fc⁺, 0.20 V vs Ag/AgNO₃). Ferrocene/ferrocenium (Fc/Fc⁺, 0.64 V vs NHE) was used as an internal standard for all measurements.

	<i>i</i> Pr-PMI	Cy-PMI	<i>i</i> Pr-PBI	Cy-PBI
E_{ox} (V) ^a	0.26	0.27	0.21	0.24

^a First oxidation potential (E_{ox}) vs Fc/Fc⁺.

Preparation of dye-modified TiO₂ electrode.^{iv} Nanoporous TiO₂ films were prepared from colloidal suspension of TiO₂ nanoparticles (P25, Nippon Aerogel) dispersed in deionized water and Triton X-100. The suspension was deposited on a transparent conducting glass (Asahi Glass, SnO₂: F, 9.4 ohm/sq) by using doctor blade technique. The films were annealed at 723 K for 1 h, followed by similar deposition and annealing (723 K, 1 h) for the 13- μ m-thick TiO₂ films. The thickness of the films was determined using surface roughness/profile measuring instrument (SURFCOM 130A, ACCRETECH). The TiO₂ electrodes were immersed into each of the 0.15 mM solution of dyes at room temperature. After dye adsorption, the dye-coated electrodes were copiously rinsed with the same solvent used for the adsorption.

Photovoltaic measurements. The photovoltaic measurements were performed in a sandwich cell consisting of the dye-sensitized TiO₂ electrode as the working electrode and a platinum-coated conducting glass as the counter electrode. The two electrodes were placed on top of each other using a thin transparent film of Surlyn polymer (Dupont) as a spacer to form the electrolyte space. A thin layer of electrolyte (0.1 M LiI, 0.05 M I₂, 0.6 M 2,3-dimethyl-1-propylimidazolium iodide, and 0.5 M 4-*t*-butylpyridine in acetonitrile) was introduced into the interelectrode space. The IPCE values and photocurrent-voltage characteristics were determined by using a potentiostat (Bunko-Keiki Co., Ltd., Model HCSSP-25) irradiated with simulated AM 1.5 solar light (100 mW cm⁻², Bunko-Keiki Co., Ltd., Model CEP-2000). All the experimental values were given as an average from three independent measurements.

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- (i) Chao, C.-C.; Leung, M.-k.; Su, Y. O.; Chiu, K.-Y.; Lin, T.-H.; Shieh, S.-J.; Lin, S.-C. *J. Org. Chem.* **2005**, *70*, 4323.
- (ii) Würthner, F.; Stepanenko, V.; Chen, Z.; Saha-Möller, C. R.; Kocher, N.; Stalke, D. *J. Org. Chem.* **2004**, *69*, 7933.
- (iii) Tayler, E. C.; Fletcher, S. R.; Sabb, A. L. *Synth. Commun.* **1984**, *14*, 921.
- (iv) (a) Imahori, H.; Hayashi, S.; Umeyama, T.; Eu, S.; Oguro, A.; Kang, S.; Matano, Y.; Shishido, T.; Ngamsinlapasathian, S.; Yoshikawa, S. *Langmuir* **2006**, *22*, 11405.
- (b) Eu, S.; Hayashi, S.; Umeyama, T.; Oguro, A.; Kawasaki, M.; Kadota, N.; Matano, Y.; Imahori, H. *J. Phys. Chem. C* **2007**, *111*, 3528.

S2. NMR Spectra.

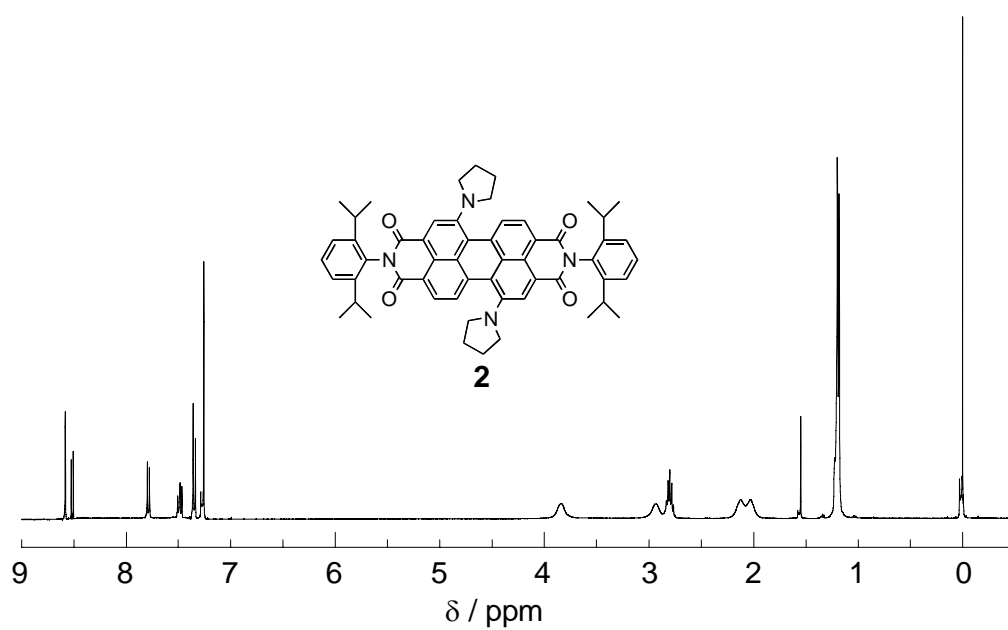


Figure S2-1. ^1H NMR spectrum of **2** in CDCl_3 .

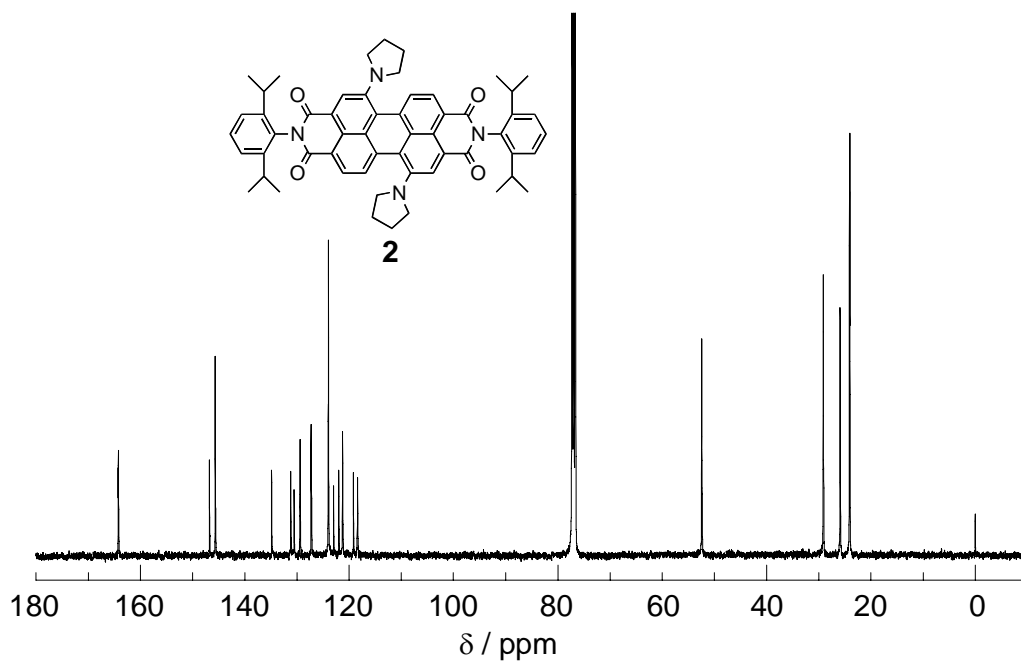


Figure S2-2. ^{13}C NMR spectrum of **2** in CDCl_3 .

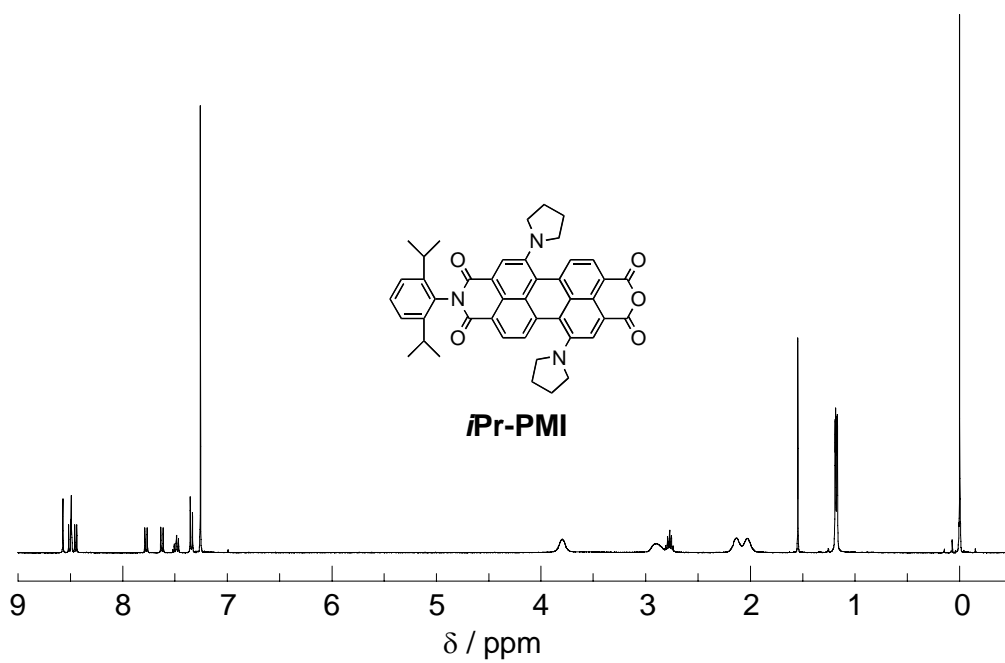


Figure S2-3. ^1H NMR spectrum of *iPr-PMI* in CDCl_3 .

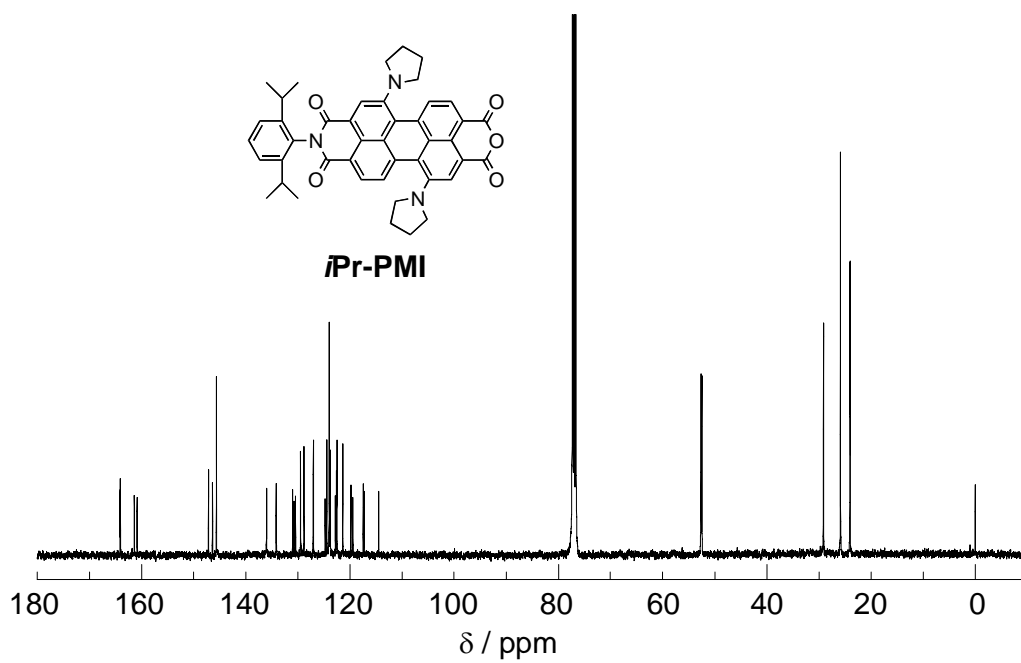


Figure S2-4. ¹³C NMR spectrum of *iPr-PMI* in CDCl₃.

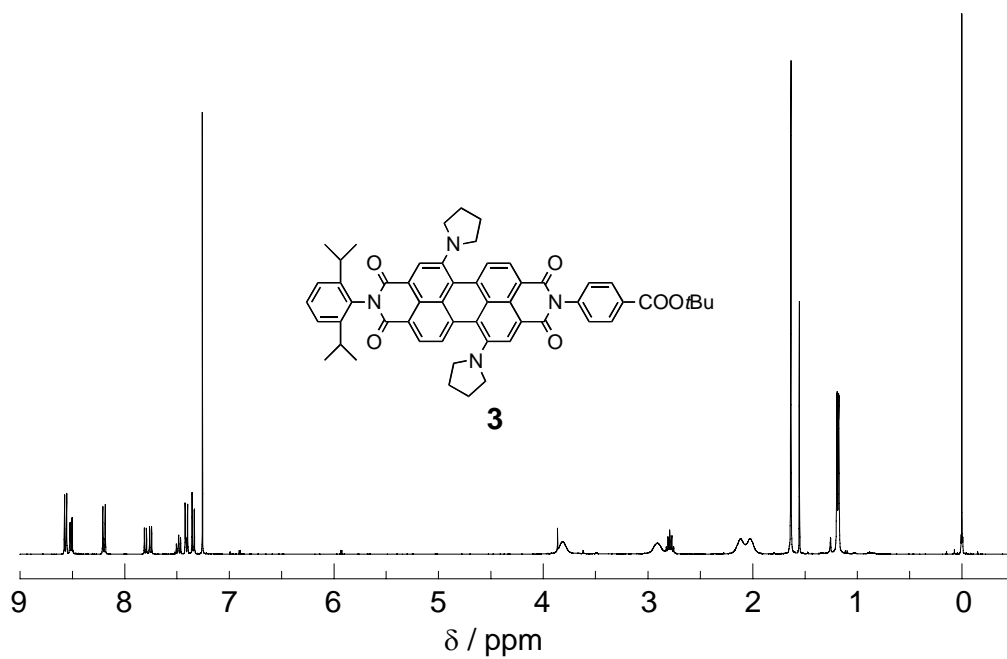


Figure S2-5. ¹H NMR spectrum of **3** in CDCl₃.

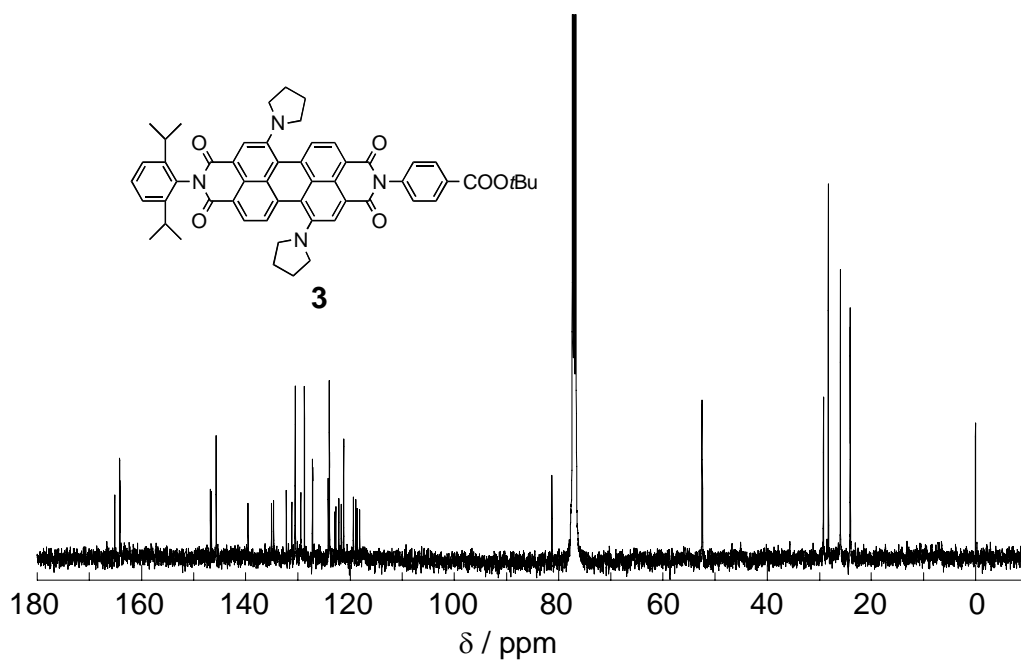


Figure S2-6. ^{13}C NMR spectrum of **3** in CDCl_3 .

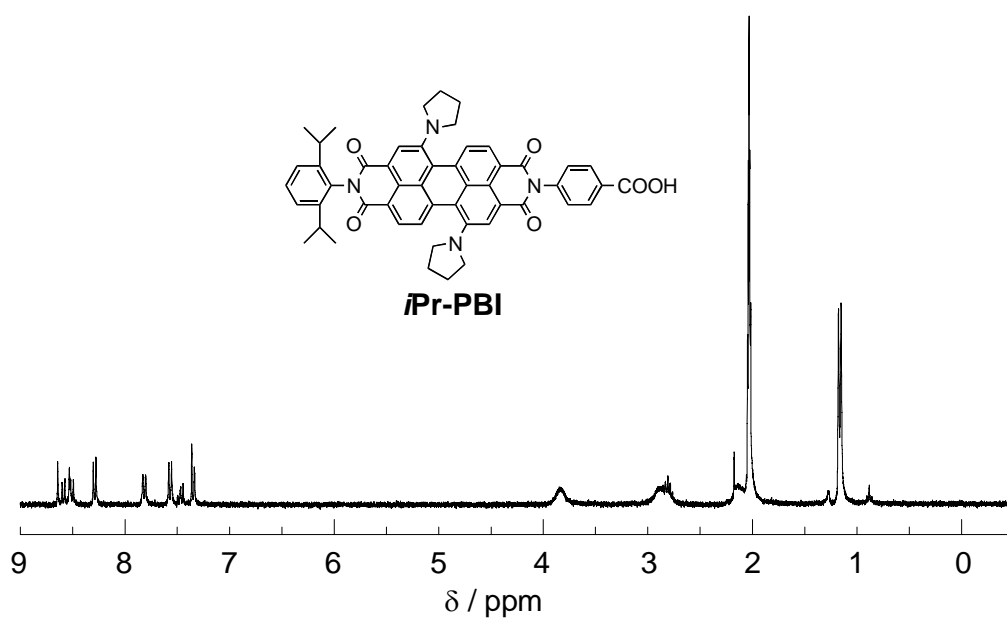


Figure S2-7. ^1H NMR spectrum of ***iPr-PBI*** in acetic acid- d_4 .

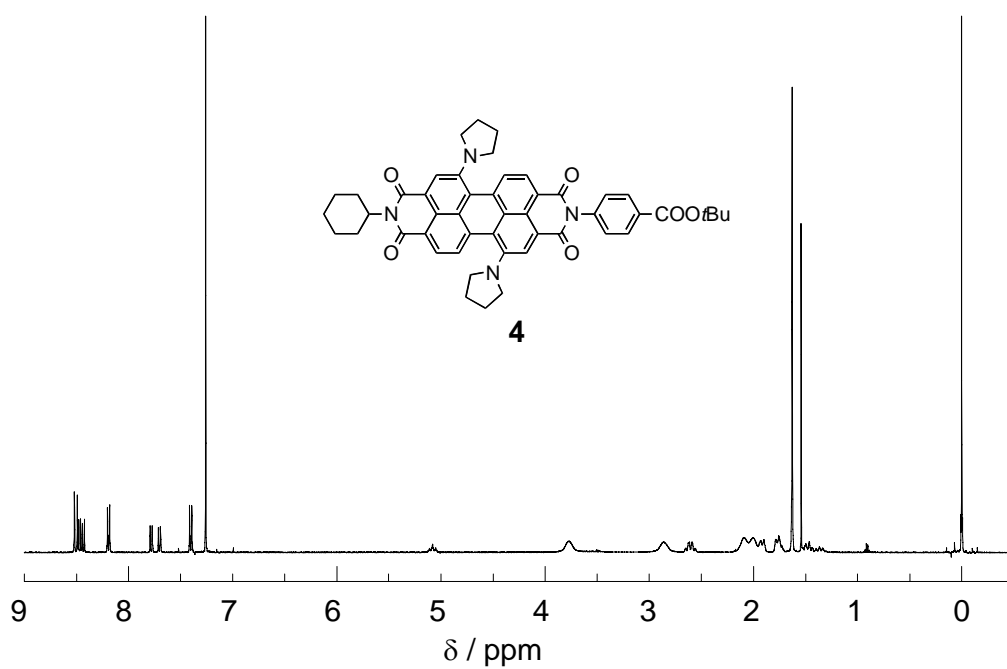


Figure S2-8. ^1H NMR spectrum of **4** in CDCl_3 .

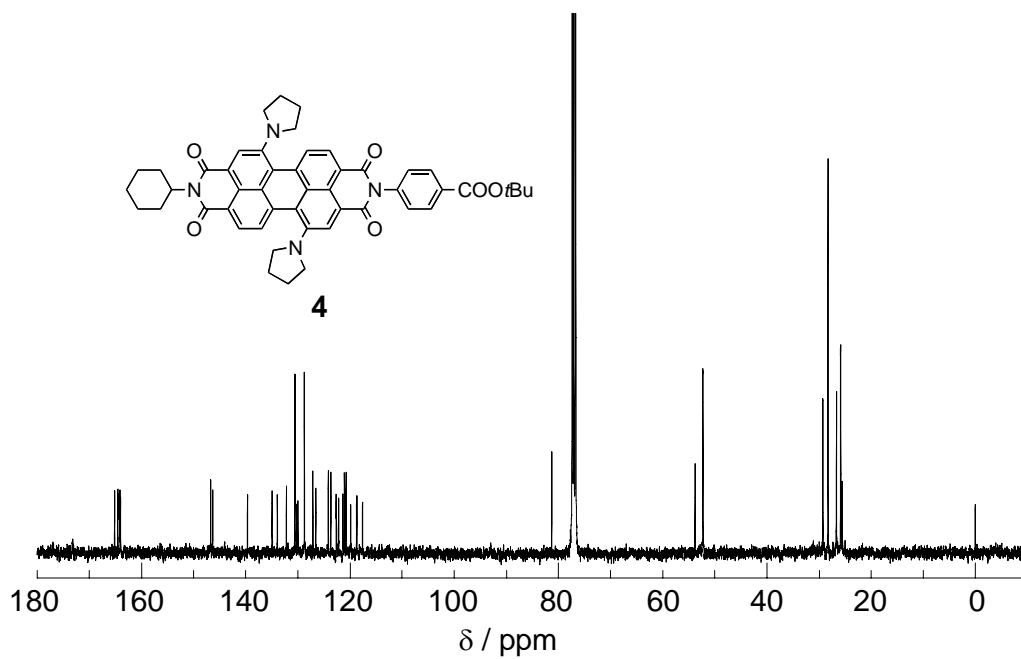


Figure S2-9. ^{13}C NMR spectrum of **4** in CDCl_3 .

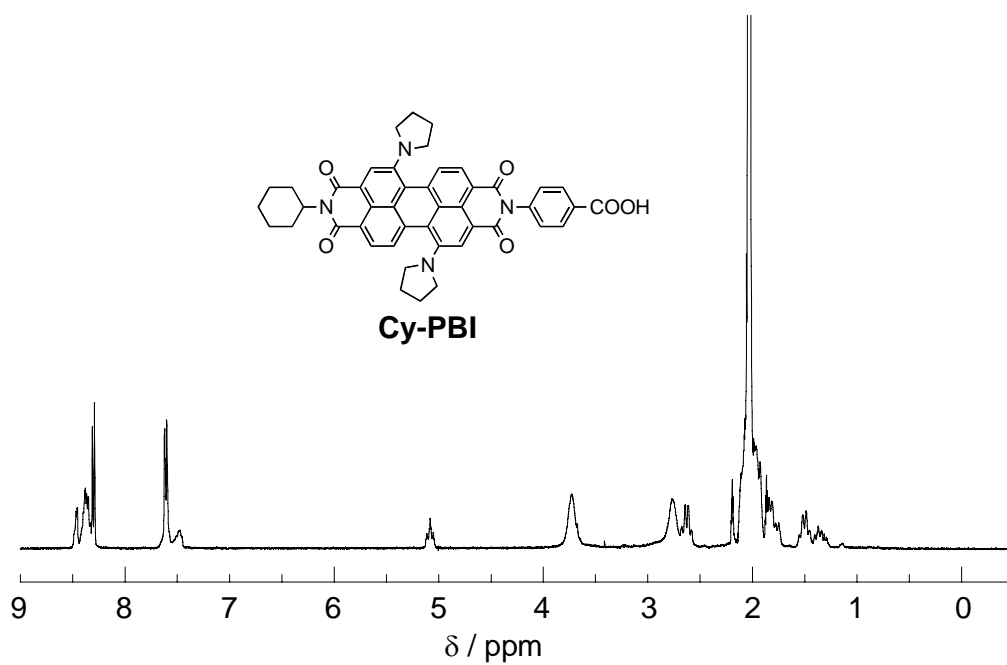


Figure S2-10. ¹H NMR spectrum of **Cy-PBI** in acetic acid-*d*₄.

S3.

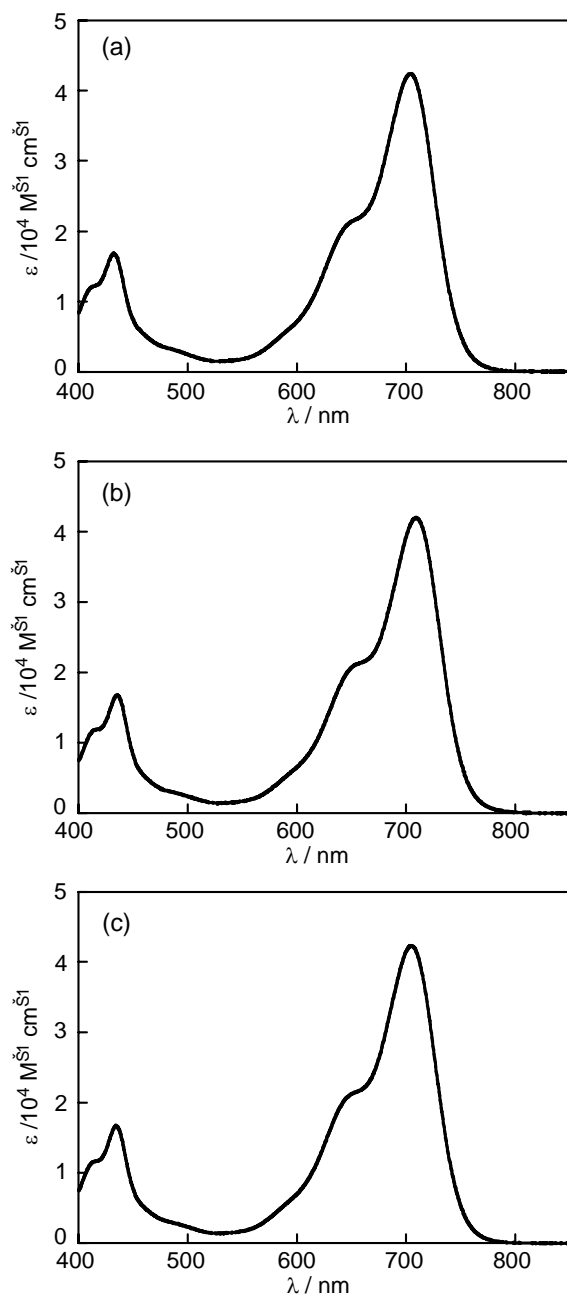


Figure S3. Absorption spectra of (a) **Cy-PMI**, (b) *iPr*-**PBI**, and (c) **Cy-PBI** in CH_2Cl_2 .

S4.

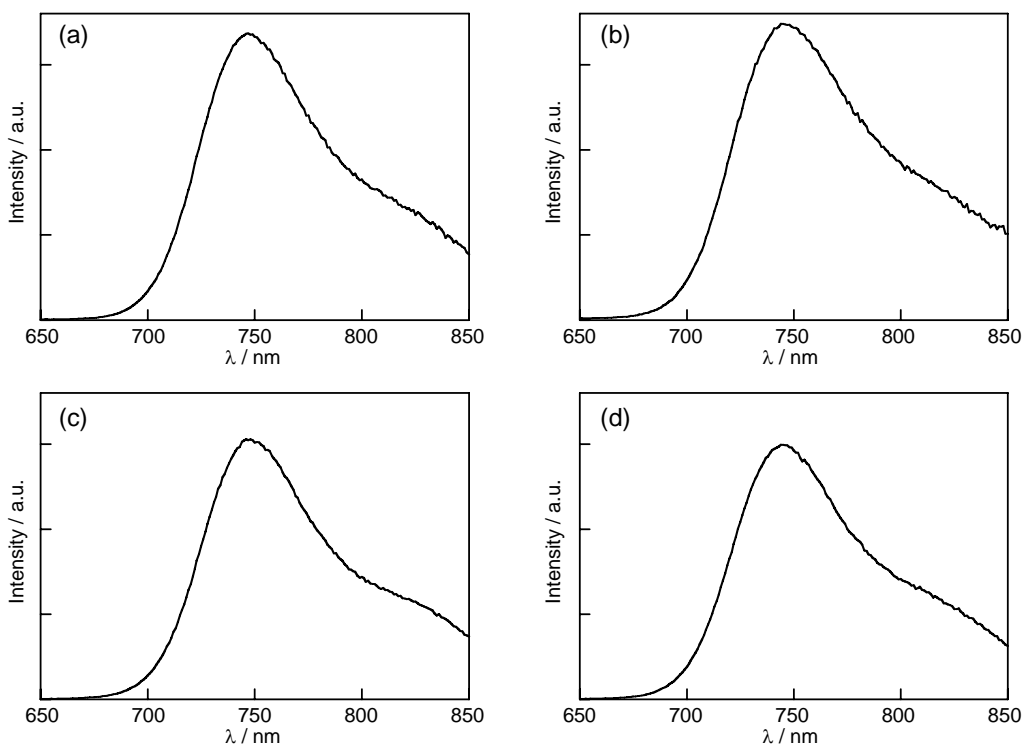


Figure S4. Fluorescence spectra of (a) *iPr-PMI*, (b) **Cy-PMI**, (c) *iPr-PBI*, and (d) **Cy-PBI** in CH_2Cl_2 with an excitation wavelength of 430 nm where the absorbances of the compounds are identical.

S5.

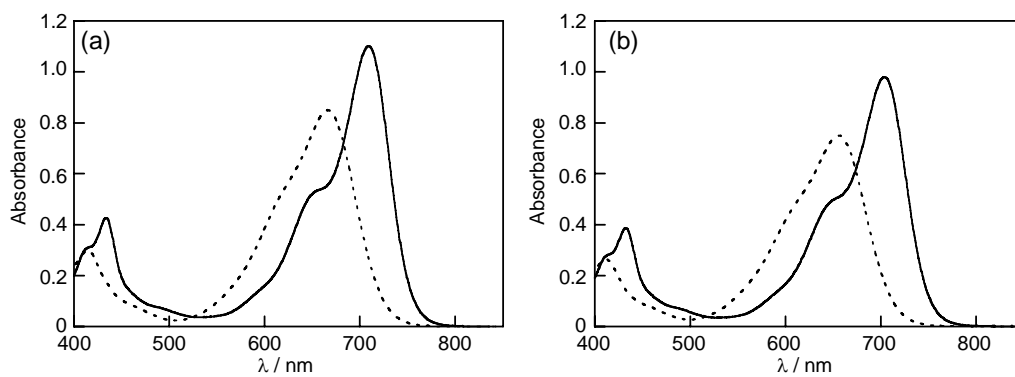


Figure S5. (a) Absorption spectra of 0.11 mM of *iPr-PMI* in CH_2Cl_2 before (solid line) and after (dashed line) addition of 100 equivalent of Bu_4NOH . (b) Absorption spectra of 0.069 mM of **Cy-PMI** in CH_2Cl_2 before (solid line) and after (dashed line) addition of 90 equivalent of Bu_4NOH . Further addition of Bu_4NOH did not change the spectra.

S6.

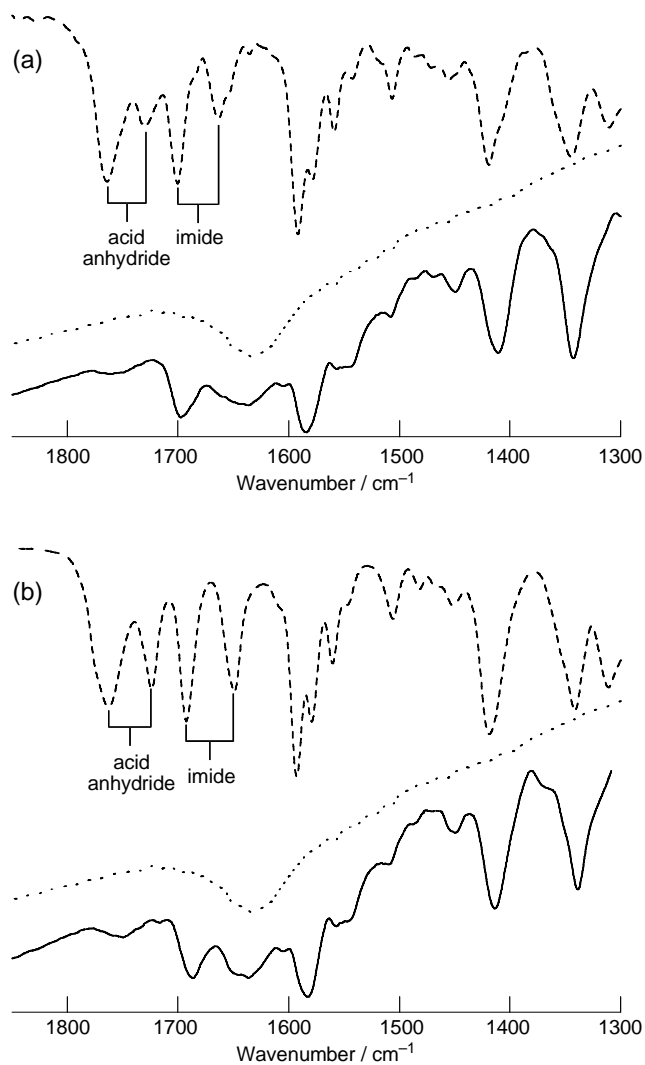


Figure S6. IR spectra of (a) *iPr-PMI* and (b) *Cy-PMI* in KBr pallet (dashed line) and on surface of TiO_2 (solid line). IR spectra of TiO_2 (dotted line) are also shown for comparison. The peaks at around $1770\text{-}1720\text{ cm}^{-1}$ and $1700\text{-}1660\text{ cm}^{-1}$ are assigned to symmetric and asymmetric vibration of C=O of acid anhydride and that of imide, respectively. The peaks due to the acid anhydride disappear after adsorption of the dyes on the TiO_2 electrode, while those due to the imide remain intact.

S7.

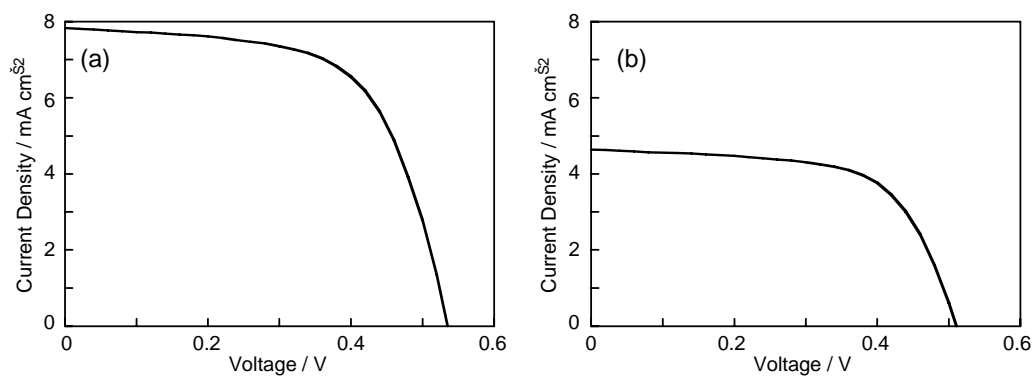


Figure S7. Current-voltage characteristics of (a) *iPr*-PMI and (b) *Cy*-PMI-sensitized TiO₂ cells. Conditions: electrolyte 0.1 M LiI, 0.05 M I₂, 0.6 M 2,3-dimethyl-1-propyl imidazolium iodide, and 0.5M 4-*t*-butylpyridine in CH₃CN; input power: AM 1.5 under simulated solar light (100 mW cm⁻²).