

# A Copper-Catalyzed Regiospecific Synthesis of *N*-Alkyl Benzimidazoles

*Nan Zheng and Stephen L. Buchwald\**

Department of Chemistry, Massachusetts Institute of Technology

Cambridge, Massachusetts, 02139.

## Experimental Section

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### **General Considerations**

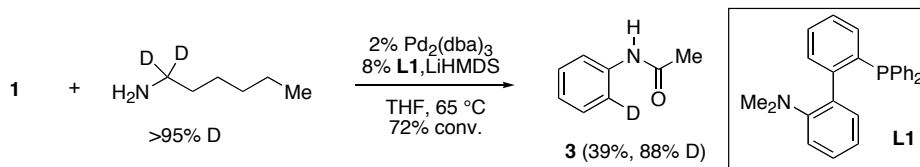
All reactions were carried out under an argon atmosphere in a Schlenk tube, with a stir bar and capped with a Teflon screw-cap. EtOAc was purchased from Mallinckrodt (ACS grade) and used as received. Anhydrous *t*-BuOH, DMF, and 1,4-dioxane were purchased from Aldrich in a Sure/Seal™ bottle, used as received and stored under Argon. Anhydrous granular K<sub>3</sub>PO<sub>4</sub> was purchased from Fluka. K<sub>2</sub>CO<sub>3</sub> powder (~325 mesh), NaOt-Bu and LHMDs (1M solution in THF) were purchased from Sigma-Aldrich. Cs<sub>2</sub>CO<sub>3</sub> was a gift from Chemetall. Bulk quantities of K<sub>2</sub>CO<sub>3</sub>, K<sub>3</sub>PO<sub>4</sub> and Cs<sub>2</sub>CO<sub>3</sub> were stored in a nitrogen-filled glovebox. Small portions (1-2 g) were removed from the glovebox in glass vials and stored in a desiccator filled with anhydrous calcium sulfate. 4Å molecular sieves (purchased from Aldrich, activated powder, 5 μm) were activated by heating under vacuum prior to use.

$\text{Pd}(\text{OAc})_2$  was a gift from Englehard.  $\text{Pd}_2(\text{dba})_3$  and  $\text{CuI}$  were purchased from Strem. Diamine ligands, *N,N'*-dimethylethylenediamine (**L3**) and *rac-trans-N,N'*-dimethylcyclohexane-1,2-diamine (**L2**), were purchased from Aldrich and used as received. 2-(Diphenylphosphino)-2'-(*N,N*-di-methylamino)biphenyl (**L1**) was synthesized following a published procedure.<sup>1</sup>

2-Bromo-*N*-methylaniline **13**, 2-bromoacetanilide **1**, and 2-bromo-4-methylacetanilide were purchased from Aldrich. 2-Bromo-4-trifluoromethylacetanilide was purchased from Acros. 2-Iodo-(4-trifluoromethyl)aniline was purchased from Alfa Aesar. The rest of starting materials were purchased from either Aldrich or Acros and used as received unless specified otherwise. *n*-Hexylamine was purified by distillation over  $\text{CaH}_2$  and stored in a nitrogen-filled Schlenk tube. 2-Iodotrifluoroacetanilide **8**<sup>2</sup> and labeled *n*-hexylamine<sup>3</sup> were prepared based on reported procedures. 2-Iodoacetanilide **6**<sup>4</sup> was synthesized using a published procedure.<sup>5</sup>

All new compounds were characterized by  $^1\text{H}$  NMR,  $^{13}\text{C}$  NMR, and IR spectroscopy, in addition to elemental analysis performed by Atlantic Microlabs Inc., Norcross, GA and/or low resolution mass spectroscopy. For starting materials, copies of the  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra are attached for those compounds of which a satisfactory elemental analysis was not obtained. Copies of the  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra are attached for all products. Nuclear Magnetic Resonance spectra were recorded on a Varian Mercury 300 or Varian Inova 500 instrument. All  $^1\text{H}$  NMR experiments are reported in  $\delta$  units, parts per million (ppm) and were measured relative to the signals for residual chloroform (7.27 ppm) and methanol (3.31 ppm) in the deuterated solvents. All  $^{13}\text{C}$  NMR spectra (obtained with  $^1\text{H}$  decoupling) are reported in ppm relative to deuteriochloroform (77.23 ppm) and deuterodimethyl sulfoxide (39.51 ppm). Infrared spectra were recorded using a Perkin-Elmer 2000 FT-IR. Melting points (uncorrected) were obtained on a Mel-Temp capillary melting point apparatus. Gas chromatography analyses were performed on an Agilent 6890 instrument with a FID detector and an Agilent DB-1 column (10 m x 0.1 mm i.d.). Flash column chromatography was performed manually or using a Biotage SP4 Flash Purification System with KP-Sil silica cartridges (methylene chloride was used to transfer the crude product onto the silica gel samplet).

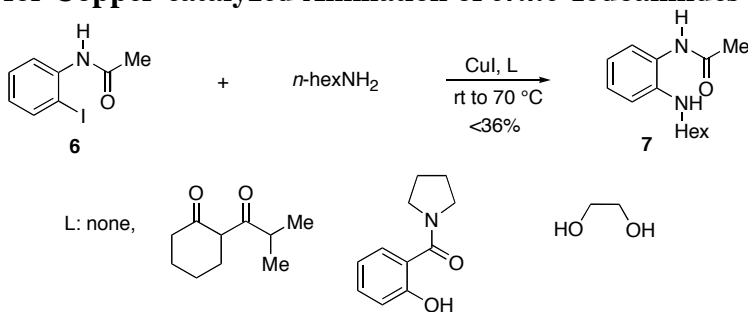
The yields for products (**9**, **12a-d**, **15**, and **18a-h**) are isolated yields (average of two runs). Yields for the preparation of starting materials refer to a single experiment. All compounds isolated were estimated to be  $\geq 95\%$  pure as determined by  $^1\text{H}$  NMR and GC analysis and/or combustion analysis.



***N*-phenylacetamide  $d_1^6$  (Scheme 1, 3)** An oven-dried Schlenk tube containing a stir bar was charged with  $\text{Pd}_2\text{dba}_3$  (9.2 mg, 0.01 mmol, 4.0 mol % Pd), **L1** (0.04 mmol, 8 mol %), and 2-bromoacetanilide **1** (107.1 mg, 0.5 mmol). The Schlenk tube was capped with a rubber septum and then evacuated and backfilled with argon (3 cycles). LiHMDS (1.0 M in THF, 1.5 mL) was added to the Schlenk tube through the septum via syringe at room temperature. The reaction mixture was stirred at room temperature for 5 min. Then 2,2-dideuteriohexylamine (86  $\mu\text{L}$ , 0.65 mmol) was added to the Schlenk tube through the septum via syringe. The septum was replaced with a Teflon screw cap, and the Schlenk tube was sealed and put into a pre-heated oil bath at 65 °C. After stirring for 18 h, the reaction mixture was allowed to cool to 0 °C. 1N HCl (1.5 mL)

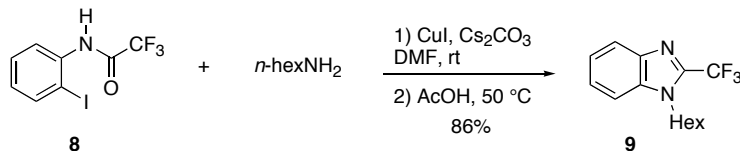
was added dropwise. 10 min later, saturated aqueous NaHCO<sub>3</sub> solution (2 mL) was added. The mixture was extracted with EtOAc (4 mL x 3). The combined EtOAc layers were dried over Na<sub>2</sub>SO<sub>4</sub>. The EtOAc extracts were analyzed by GC first (113  $\mu$ L of dodecane was added as an internal standard) and then concentrated under vacuum. The residual was purified by flash column chromatography on silica gel (gradient elution: 2/1, and then 1.5/1 hexanes/EtOAc) to provide **3** (26.2 mg, 39 %, 88% D).

### Ligand Screening for Copper-catalyzed Amination of *ortho*-Iodoanilides (Scheme 3)



A representative procedure for screening no added ligand, 2-isobutyrylcyclohexanone, and *N*-(2-hydroxybenzoyl)pyrrolidine is as follows: an oven-dried Schlenk tube containing a stir bar was charged with CuI (9.5 mg, 0.05 mmol, 5.0 mol % Cu), *ortho*-iodoacetanilide **6** (261 mg, 1 mmol), and Cs<sub>2</sub>CO<sub>3</sub> (652 mg, 2.0 mmol) (*N*-(2-hydroxybenzoyl)pyrrolidine was added here). The Schlenk tube was capped with a rubber septum and then evacuated and backfilled with argon (3 cycles). Ligand (0.20 mmol, 20 mol%), *n*-hexylamine (0.20 mL, 1.5 mmol), and DMF (0.5 mL) were added through the septum via syringe. The septum was replaced with a Teflon screw cap, and the Schlenk tube was sealed and stirred at rt or 70 °C for 12 h.

A representative procedure for screening ethylene glycol is as follows: an oven-dried Schlenk tube containing a stir bar was charged with CuI (19.0 mg, 0.10 mmol, 10 mol % Cu), *ortho*-iodoacetanilide **6** (261 mg, 1 mmol), and K<sub>3</sub>PO<sub>4</sub> (425 mg, 2.0 mmol). The Schlenk tube was capped with a rubber septum and then evacuated and backfilled with argon (3 cycles). Ethylene glycol (0.11 mL, 2 mmol), *n*-hexylamine (0.20 mL, 1.5 mmol), and *i*-PrOH (1 mL) were added through the septum via syringe. The septum was replaced with a Teflon screw cap, and the Schlenk tube was sealed and stirred at 70 °C for 12 h.



**1-hexyl-2-(trifluoromethyl)-1*H*-benzimidazole<sup>7</sup> (Scheme 3, 9)** To an oven-dried Schlenk tube containing a stir bar were added CuI (9.5 mg, 0.05 mmol, 5.0 mol % Cu), *ortho*-iodotrifluoroacetanilide **8** (315 mg, 1 mmol), and preactivated 4Å molecular sieves (150 mg). The tube was evacuated and transferred to a nitrogen-filled glovebox. Cs<sub>2</sub>CO<sub>3</sub> (652 mg, 2 mmol), *n*-hexylamine (0.20 mL, 1.5 mmol) and DMF (0.50 mL) were added. The Schlenk tube was removed from the glove box and the reaction mixture was stirred at room temperature for 18 h. The reaction mixture was diluted with EtOAc (4 mL) and then filtered through Celite with the aid of EtOAc. The filtrate was concentrated under reduced pressure to give a crude mixture that was dissolved in AcOH (1 mL). The formed solution was heated to 50 °C and stirred at 50 °C for 2 h. After cooling to room temperature, AcOH was removed by vacuum. The residual material was partitioned between dichloromethane (3 mL) and saturated aqueous NaHCO<sub>3</sub>.

solution (3 mL). The aqueous layer was separated and extracted by dichloromethane (3 mL x 2). The combined dichloromethane layers were dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under vacuum. The residual was purified by flash chromatography on silica gel (Biotage, 3-28% ethyl acetate in hexanes gradient) to provide the title compound as a brown oil (235 mg, 87%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ: 7.89 (d, *J* = 7.6 Hz, 1H), 7.48-7.34 (m, 3H), 4.29 (t, *J* = 7.9 Hz, 2H), 1.93-1.82 (m, 2H), 1.46-1.27 (m, 6H), 0.90 (t, *J* = 7.1 Hz, 3H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ: 141.3, 140.5 (q, *J* = 38.4 Hz), 135.5, 125.3, 123.6, 121.7, 119.3 (q, *J* = 271.2 Hz), 110.6, 45.3, 31.3, 30.0, 26.5, 22.5, 13.9.

### General Procedures for Cu-Catalyzed Synthesis of *N*-Alkyl Benzimidazoles (Scheme 4 and Scheme 5)

Procedure A: an oven-dried Schlenk tube containing a stir bar was charged with CuI (9.5 mg, 0.05 mmol, 5.0 mol % Cu), *ortho*-iodo-*N*-alkylaniline (1 mmol), amide (1.2 mmol), and Cs<sub>2</sub>CO<sub>3</sub> (652 mg, 2.0 mmol). The Schlenk tube was capped with a Teflon screw cap and then evacuated and backfilled with argon (3 cycles). *N,N'*-Dimethylethylenediamine **L3** (11 μL, 0.1 mmol, 10 mol%) was added via syringe to the Schlenk tube followed by the addition of 1,4-dioxane (1 mL) under a positive flow of Ar. The Schlenk tube was sealed and put into a pre-heated oil bath at 110 °C. After stirring for 17 h, the reaction mixture was allowed to cool to room temperature.

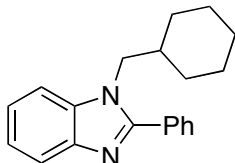
Procedure B: an oven-dried Schlenk tube containing a stir bar was charged with CuI (9.5 mg, 0.05 mmol, 5.0 mol % Cu), *ortho*-iodo-*N*-alkylaniline (1 mmol), amide (1.2 mmol), and Cs<sub>2</sub>CO<sub>3</sub> (652 mg, 2.0 mmol). The Schlenk tube was capped with a Teflon screw cap and then evacuated and backfilled with argon (3 cycles). *rac-trans-N,N'*-Dimethylcyclohexane-1,2-diamine **L2** (32 μL, 0.2 mmol, 20 mol%) was added via syringe to the Schlenk tube followed by the addition of 1,4-dioxane (1 mL) under a positive flow of Ar. The Schlenk tube was sealed and put into a pre-heated oil bath at 90 °C. After stirring for 2 h, the reaction mixture was allowed to cool to room temperature.

Depending on the amides used, two procedures were developed for the cyclodehydration step.

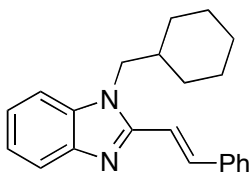
Procedure C: when benzamide and (*E*)-cinnamamide were used, the Schlenk tube was opened under a positive Ar flow. K<sub>3</sub>PO<sub>4</sub> (318 mg, 1.5 mmol) was added followed by the addition of *t*-BuOH (1 mL). The Schlenk tube was resealed and put into a pre-heated oil bath at 110 °C. After stirring for 4 h or 8 h, the reaction mixture was allowed to cool to room temperature. The reaction mixture was first diluted with dichloromethane (4 mL) and then filtered through Celite with the aid of dichloromethane. The filtrate was concentrated under reduced pressure. The residual material was purified by flash chromatography on silica gel.

Procedure D, in the case of cyclohexanecarboxamide and hexanamide, the reaction mixture was diluted with dichloromethane (4 mL) first. Then, the resulting mixture was filtered through Celite with the aid of dichloromethane. The filtrate was concentrated under reduced pressure. The residual material was dissolved in AcOH (1 mL) and then heated to 75 °C. After stirring for 2 h, the reaction mixture was allowed to cool to room temperature. AcOH was removed under vacuum. The residual material was partitioned between dichloromethane (3 mL) and saturated aqueous NaHCO<sub>3</sub> solution (3 mL) (if an emulsion formed, 2 mL of brine was added to aid in the separation). The aqueous layer was separated and extracted by dichloromethane (3 mL x 2). The combined dichloromethane layers were dried over Na<sub>2</sub>SO<sub>4</sub> and

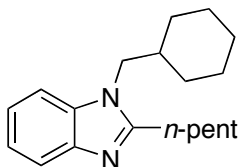
concentrated under reduced pressure. The residual material was purified by flash column chromatography on silica gel.



**1-(cyclohexylmethyl)-2-phenyl-1H-benzimidazole (Scheme 4, 12a)** Following procedure A, *ortho*-iodo-*N*-alkylaniline **10** (240 mg, 0.76 mmol) was coupled with benzamide (111 mg, 0.91 mmol). The cyclodehydration step was accomplished using procedure C (243 mg of  $K_3PO_4$ , 0.8 mL of *t*-BuOH, 4 h). The crude product was purified by flash chromatography on silica gel (gradient elution: 4/1, and then 3/1 hexanes/EtOAc) to provide the title compound as a red solid (150 mg, 68%). mp 97-98 °C.  $^1H$  NMR (500 MHz,  $CDCl_3$ )  $\delta$ : 7.85-7.82 (m, 1H), 7.71-7.67 (m, 2H), 7.55-7.49 (m, 3H), 7.45-7.41 (m, 1H), 7.33-7.29 (m, 2H), 4.12 (d,  $J$  = 7.5 Hz, 2H), 1.85-1.76 (m, 1H), 1.62-1.55 (m, 3H), 1.45 (d,  $J$  = 12.8 Hz, 2H), 1.11-0.99 (m, 3H), 0.81-0.74 (m, 2H).  $^{13}C$  NMR (126 MHz,  $CDCl_3$ )  $\delta$ : 154.3, 143.1, 135.9, 131.1, 129.60, 129.57, 128.7, 122.6, 122.3, 120.0, 110.7, 50.9, 38.1, 30.8, 26.1, 25.6. IR (neat,  $cm^{-1}$ ): 3061, 2925, 2851, 1448, 1390, 1356, 1327, 1273, 774, 742, 699. Anal. Calcd for  $C_{20}H_{22}N_2$ : C, 82.72; H, 7.64. Found: C, 82.45; H, 7.63.

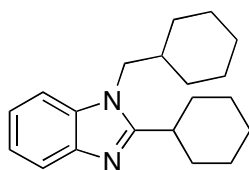


**1-(cyclohexylmethyl)-2-[(*E*)-2-phenylethenyl]-1H-benzimidazole (Scheme 4, 12b)** Following procedure A, *ortho*-iodo-*N*-alkylaniline **10** (218 mg, 0.69 mmol) was coupled with (*E*)-cinnamamide (122 mg, 0.83 mmol). The cyclodehydration step was accomplished using procedure C (220 mg of  $K_3PO_4$ , 0.7 mL of *t*-BuOH, 4 h). The crude product was purified by flash chromatography on silica gel (gradient elution: 5/1, and then 4/1 hexanes/EtOAc) to provide the title compound as a yellow solid (131 mg, 60%). mp 165-167 °C.  $^1H$  NMR (500 MHz,  $CDCl_3$ )  $\delta$ : 8.02 (d,  $J$  = 15.8 Hz, 1H), 7.80-7.78 (m, 1H), 7.63 (d,  $J$  = 7.3 Hz, 2H), 7.43 (t,  $J$  = 7.4 Hz, 2H), 7.38-7.33 (m, 2H), 7.30-7.24 (m, 2H), 7.07 (d,  $J$  = 15.8 Hz, 1H), 4.08 (d,  $J$  = 7.4 Hz, 2H), 1.94-1.85 (m, 1H), 1.75-1.69 (m, 5H), 1.24-1.05 (m, 5H).  $^{13}C$  NMR (126 MHz,  $CDCl_3$ )  $\delta$ : 151.1, 143.2, 137.0, 136.2, 136.0, 129.1, 129.0, 127.4, 122.6, 122.5, 119.4, 113.4, 109.9, 49.9, 39.1, 31.3, 26.2, 25.8. IR (neat,  $cm^{-1}$ ): 3059, 2926, 2852, 1635, 1500, 1449, 1405, 1329, 967, 755, 741, 700.

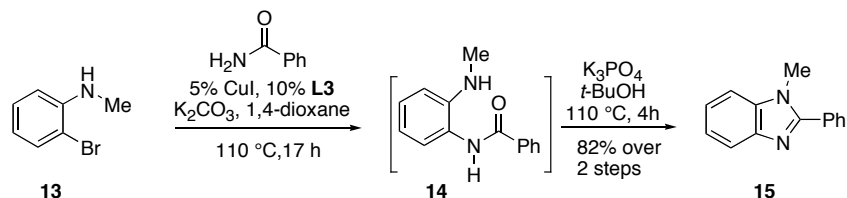


**1-(cyclohexylmethyl)-2-pentyl-1H-benzimidazole (Scheme 4, 12c)** Following procedure B, *ortho*-iodo-*N*-alkylaniline **10** (204 mg, 0.65 mmol) was coupled with hexanamide (89.6 mg, 0.78 mmol). The cyclodehydration step was accomplished using procedure D (1 mL of AcOH). The crude product was purified by flash chromatography on silica gel (Biotage, 8-66% ethyl acetate

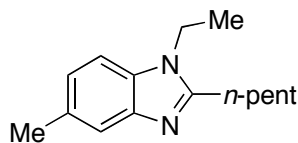
in hexanes gradient) to provide the title compound as a brown oil (160 mg, 87%).  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$ : 7.76-7.70 (m, 1H), 7.32-7.28 (m, 1H), 7.25-7.19 (m, 2H), 3.92 (d,  $J = 7.4\text{Hz}$ , 2H), 2.84 (dd,  $J = 7.9, 8.0\text{Hz}$ , 2H), 1.98-1.80 (m, 3H), 1.75-1.62 (m, 5H), 1.50-1.34 (m, 4H), 1.24-0.98 (m, 5H), 0.93 (t,  $J = 7.1\text{Hz}$ , 3H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$ : 155.6, 142.8, 135.6, 121.8, 121.6, 119.2, 109.7, 50.1, 38.7, 31.9, 31.2, 27.7, 27.6, 26.3, 25.8, 22.6, 14.2. IR (neat,  $\text{cm}^{-1}$ ): 2926, 2853, 1615, 1508, 1458, 1409, 1328, 1285, 740. Anal. Calcd for  $\text{C}_{17}\text{H}_{18}\text{N}_2$ : C, 80.23; H, 9.92. Found: C, 79.97; H, 9.92.



**2-cyclohexyl-1-(cyclohexylmethyl)-1H-benzimidazole (Scheme 4, 12d)** Following procedure B, *ortho*-iodo-*N*-alkylaniline **10** (156 mg, 0.49 mmol) was coupled with cyclohexanecarboxamide (75.4 mg, 0.59 mmol). The cyclodehydration step was accomplishing using procedure D (1 mL of AcOH, 12 h at 75 °C instead of the standard 2 h). The crude product was purified by flash chromatography on silica gel (Biotage, 5-40% ethyl acetate in hexanes gradient) to provide the title compound as an off-white solid (122 mg, 83%). mp 90-91 °C.  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$ : 7.76-7.73 (m, 1H), 7.32-7.28 (m, 1H), 7.23-7.20 (m, 2H), 3.94 (d,  $J = 7.5\text{Hz}$ , 2H), 2.81 (tt,  $J = 11.6, 3.4\text{Hz}$ , 1H), 1.95-1.63 (m, 13H), 1.47-1.37 (m, 3H), 1.24-1.14 (m, 3H), 1.09-1.01 (m, 2H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$ : 159.7, 142.9, 135.3, 121.8, 121.7, 119.4, 109.9, 49.9, 38.9, 36.6, 32.3, 31.3, 26.7, 26.4, 25.96, 25.95. IR (neat,  $\text{cm}^{-1}$ ): 2927, 2852, 1503, 1459, 1420, 1347, 1272, 742.

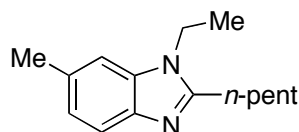


**1-methyl-2-phenyl-1H-benzimidazole<sup>8</sup> (Scheme 4, 15)** Following procedure A, 2-Bromo-*N*-methylaniline **13** (123  $\mu\text{L}$ , 1.0 mmol) was coupled with benzamide (145 mg, 1.2 mmol). The cyclodehydration step was accomplishing using procedure C (318 mg of  $\text{K}_3\text{PO}_4$ , 1 mL of *t*-BuOH, 4 h). The crude product was purified by flash chromatography on silica gel (gradient elution: 1.5/1, 1/1, and then 1/1.5 hexanes/ ethyl acetate) to provide **15** as a brown solid (171 mg, 82 %). mp 97-99 °C (lit. 92-94 °C).<sup>8</sup>  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$ : 7.88-7.81 (m, 1H), 7.80-7.74 (m, 2H), 7.57-7.50 (m, 3H), 7.43-7.30 (m, 3H), 3.86 (s, 3H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$ : 153.8, 143.0, 136.6, 130.3, 129.8, 129.5, 128.7, 122.8, 122.5, 119.9, 109.7, 31.7. Calcd for  $\text{C}_{14}\text{H}_{12}\text{N}_2$ : C, 80.74; H, 5.81. Found: C, 80.48; H, 5.84.

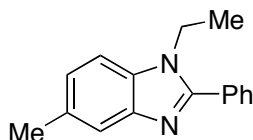


**1-ethyl-5-methyl-2-pentyl-1H-benzimidazole (Scheme 5, 18a)** Following procedure B, *ortho*-iodo-*N*-alkylaniline **16a** (242 mg, 0.93 mmol) was coupled with hexanamide (128 mg, 1.1 mmol). The cyclodehydration step was accomplishing using procedure D (1 mL of AcOH). The

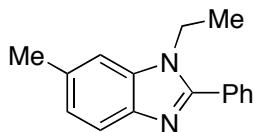
crude product was purified by flash chromatography on silica gel (gradient elution: 2/1, and then 1.5/1 hexanes/ ethyl acetate) to provide the title compound as a light yellow solid (166 mg, 78%). mp 48-50 °C.  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$ : 7.51 (s, 1H), 7.19 (d,  $J = 8.2\text{Hz}$ , 1H), 7.06 (dd,  $J = 8.2, 1.6\text{Hz}$ , 1H), 4.14 (q,  $J = 7.3\text{Hz}$ , 2H), 2.84 (t,  $J = 7.9\text{Hz}$ , 2H), 2.47 (s, 3H), 1.94-1.83 (m, 2H), 1.48-1.36 (m, 4H), 1.40 (t,  $J = 7.2\text{Hz}$ , 3H), 0.92 (t,  $J = 7.1\text{Hz}$ , 3H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$ : 154.8, 143.2, 132.8, 131.2, 123.2, 119.1, 108.6, 38.3, 31.8, 27.7, 27.5, 22.5, 21.6, 15.2, 14.1. IR (neat,  $\text{cm}^{-1}$ ): 2956, 2931, 2871, 1508, 1456, 1410, 1324, 1277, 1107, 967, 791, 605. Anal. Calcd for  $\text{C}_{15}\text{H}_{22}\text{N}_2$ : C, 78.21; H, 9.63. Found: C, 78.30; H, 9.63.



**1-ethyl-6-methyl-2-pentyl-1H-benzimidazole (Scheme 5, 18b)** Following procedure B, *ortho*-iodo-*N*-alkylaniline **16b** (231 mg, 0.88 mmol) was coupled with hexanamide (122 mg, 1.1 mmol). The cyclodehydration step was accomplishing using procedure D (1 mL of AcOH). The crude product was purified by flash chromatography on silica gel (gradient elution: 2/1, and then 1.5/1 hexanes/ ethyl acetate) to provide the title compound as a brown solid (133 mg, 65%). mp 48-49 °C.  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$ : 7.60 (d,  $J = 8.2\text{Hz}$ , 1H), 7.10 (s, 1H), 7.05 (dd,  $J = 8.2, 1.6\text{Hz}$ , 1H), 4.14 (q,  $J = 7.3\text{Hz}$ , 2H), 2.84 (t,  $J = 7.9\text{Hz}$ , 2H), 2.50 (s, 3H), 1.94-1.84 (m, 2H), 1.49-1.33 (m, 4H), 1.41 (t,  $J = 7.2\text{Hz}$ , 3H), 0.92 (t,  $J = 7.1\text{Hz}$ , 3H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$ : 154.4, 141.0, 135.0, 131.8, 123.2, 118.8, 109.2, 38.3, 31.9, 27.7, 27.6, 22.6, 21.9, 15.2, 14.1. IR (neat,  $\text{cm}^{-1}$ ): 2956, 2930, 2860, 1515, 1471, 1413, 1332, 1282, 1097, 967, 808, 606. Anal. Calcd for  $\text{C}_{15}\text{H}_{22}\text{N}_2$ : C, 78.21; H, 9.63. Found: C, 78.46; H, 9.82.

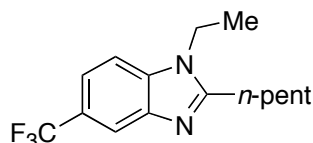


**1-ethyl-5-methyl-2-phenyl-1H-benzimidazole (Scheme 5, 18c)** Following procedure B, *ortho*-iodo-*N*-alkylaniline **16a** (168 mg, 0.61 mmol) was coupled with benzamide (88.6 mg, 0.73 mmol). The cyclodehydration step was accomplishing using procedure C (194 mg of  $\text{K}_3\text{PO}_4$ , 0.6 mL of *t*-BuOH, 8 h). The crude product was purified by flash chromatography on silica gel (gradient elution: 3/1, and then 2/1 hexanes/ ethyl acetate) to provide the title compound as a yellow solid (120 mg, 79%). mp 90-92 °C.  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$ : 7.75-7.70 (m, 2H), 7.63-7.62 (m, 1H), 7.56-7.49 (m, 3H), 7.32 (d,  $J = 8.2\text{Hz}$ , 1H), 7.15 (dd,  $J = 8.2, 1.6\text{Hz}$ , 1H), 4.26 (q,  $J = 7.2\text{Hz}$ , 2H), 2.52 (s, 3H), 1.46 (t,  $J = 7.2\text{Hz}$ , 3H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$ : 153.4, 143.5, 133.5, 131.9, 130.7, 129.6, 129.2, 128.7, 124.1, 119.7, 109.5, 39.6, 21.6, 15.3. IR (neat,  $\text{cm}^{-1}$ ): 2976, 1471, 1446, 1386, 1348, 1325, 1153, 793, 771, 700. Anal. Calcd for  $\text{C}_{16}\text{H}_{16}\text{N}_2$ : C, 81.32; H, 6.82. Found: C, 81.18; H, 6.88.

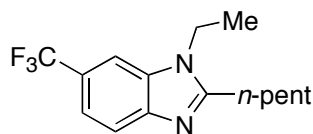


**1-ethyl-6-methyl-2-phenyl-1H-benzimidazole (Scheme 5, 18d)** Following procedure B, *ortho*-iodo-*N*-alkylaniline **16b** (165 mg, 0.63 mmol) was coupled with benzamide (91.6 mg, 0.76 mmol). The cyclodehydration step was accomplishing using procedure C (201 mg of  $\text{K}_3\text{PO}_4$ , 0.6

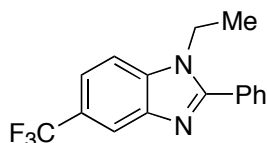
mL of *t*-BuOH, 8 h). The crude product was purified by flash chromatography on silica gel (gradient elution: 4/1, and then 3/1 hexanes/ ethyl acetate) to provide the title compound as a white solid (126 mg, 85%). mp 161-162 °C. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ: 7.74-7.72 (m, 3H), 7.55-7.51 (m, 3H), 7.23 (s, 1H), 7.15 (d, *J* = 8.2Hz, 1H), 4.27 (q, *J* = 7.2Hz, 2H), 2.55 (s, 3H), 1.48 (t, *J* = 7.2Hz, 3H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ: 153.1, 141.5, 135.7, 132.7, 130.8, 129.7, 129.3, 128.8, 124.0, 119.6, 110.0, 39.6, 22.1, 15.4. IR (neat, cm<sup>-1</sup>): 2981, 1467, 1399, 1356, 1330, 1269, 1098, 802, 782, 706, 699, 606. Anal. Calcd for C<sub>16</sub>H<sub>16</sub>N<sub>2</sub>: C, 81.32; H, 6.82. Found: C, 81.21; H, 6.84.



**1-ethyl-2-pentyl-5-(trifluoromethyl)-1H-benzimidazole (Scheme 5, 18e)** Following procedure B, *ortho*-iodo-*N*-alkylaniline **16c** (230 mg, 0.73 mmol) was coupled with hexanamide (101 mg, 0.88 mmol). The cyclodehydration step was accomplishing using procedure D (1 mL of AcOH). The crude product was purified by flash chromatography on silica gel (gradient elution: 4/1, and then 3/1 hexanes/ ethyl acetate) to provide the title compound as a gray solid (163 mg, 78%). mp 41-43 °C. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ: 8.00 (s, 1H), 7.49 (dd, *J* = 8.5, 1.6Hz, 1H), 7.39 (d, *J* = 8.4Hz, 1H), 4.21 (q, *J* = 7.3Hz, 2H), 2.88 (t, *J* = 7.9Hz, 2H), 1.95-1.89 (m, 2H), 1.49-1.37 (m, 4H), 1.44 (t, *J* = 7.3Hz, 3H), 0.94 (t, *J* = 7.1Hz, 3H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ: 157.1, 142.4, 136.8, 125.1 (q, *J* = 271.6Hz), 124.2 (q, *J* = 32.1Hz), 118.9 (q, *J* = 3.5Hz), 116.9 (q, *J* = 4.0Hz), 109.5, 38.7, 31.8, 27.6, 27.5, 22.5, 15.2, 14.1. IR (neat, cm<sup>-1</sup>): 2959, 2874, 1515, 1444, 1405, 1329, 1158, 1142, 1117, 1050, 929, 807. Anal. Calcd for C<sub>15</sub>H<sub>19</sub>F<sub>3</sub>N<sub>2</sub>: C, 63.37; H, 6.74. Found: C, 63.29; H, 6.75.

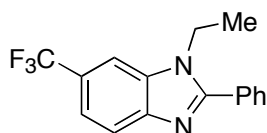


**1-ethyl-2-pentyl-6-(trifluoromethyl)-1H-benzimidazole (Scheme 5, 18f)** Following procedure B, *ortho*-iodo-*N*-alkylaniline **16d** (240 mg, 0.76 mmol) was coupled with hexanamide (105 mg, 0.91 mmol). The cyclodehydration step was accomplishing using procedure D (1 mL of AcOH). The crude product was purified by flash chromatography on silica gel (gradient elution: 4/1, and then 3/1 hexanes/ ethyl acetate) to provide the title compound as an off-white solid (189 mg, 87%). mp 57-59 °C. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ: 7.79 (d, *J* = 8.4Hz, 1H), 7.59 (t, *J* = 0.8Hz, 1H), 7.49 (dd, *J* = 8.4, 1.2Hz, 1H), 4.23 (q, *J* = 7.3Hz, 2H), 2.89 (t, *J* = 7.9Hz, 2H), 1.96-1.90 (m, 2H), 1.49-1.37 (m, 4H), 1.46 (t, *J* = 7.3Hz, 3H), 0.94 (t, *J* = 7.1Hz, 3H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ: 157.6, 145.2, 134.2, 125.1 (q, *J* = 271.6Hz), 124.1 (q, *J* = 32.1Hz), 119.5, 118.8 (q, *J* = 3.6Hz), 106.7 (q, *J* = 4.2Hz), 109.5, 38.6, 31.8, 27.5, 27.4, 22.5, 15.2, 14.0. IR (neat, cm<sup>-1</sup>): 2959, 2934, 2874, 1512, 1460, 1345, 1308, 1284, 1160, 1117, 1050, 823, 666. Anal. Calcd for C<sub>15</sub>H<sub>19</sub>F<sub>3</sub>N<sub>2</sub>: C, 63.37; H, 6.74. Found: C, 63.32; H, 6.77.



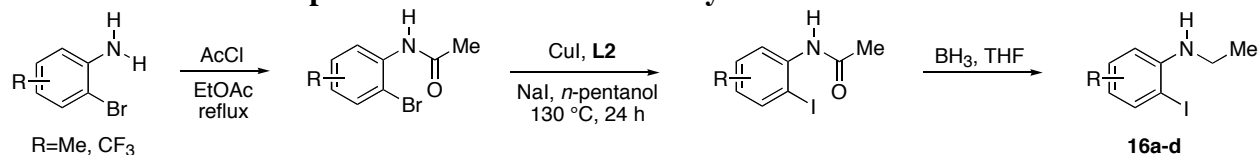


**1-ethyl-2-phenyl-5-(trifluoromethyl)-1H-benzimidazole (Scheme 5, 18g)** Following procedure B, *ortho*-iodo-*N*-alkylaniline **16d** (116 mg, 0.37 mmol) was coupled with benzamide (53.4 mg, 0.44 mmol). The cyclodehydration step was accomplished using procedure C (117 mg of  $K_3PO_4$ , 0.4 mL of *t*-BuOH, 8 h). The crude product was purified by flash chromatography on silica gel (gradient elution: 4/1, and then 3/1 hexanes/ ethyl acetate) to provide the title compound as a white solid (99.9 mg, 94%). mp 108-110 °C.  $^1H$  NMR (500 MHz,  $CDCl_3$ )  $\delta$ : 8.11 (s, 1H), 7.75-7.70 (m, 2H), 7.59-7.50 (m, 5H), 4.33 (q,  $J = 7.2$  Hz, 2H), 1.49 (t,  $J = 7.3$  Hz, 3H).  $^{13}C$  NMR (126 MHz,  $CDCl_3$ )  $\delta$ : 155.6, 142.8, 137.5, 130.3, 130.0, 129.3, 129.0, 125.0 (q,  $J = 271.8$  Hz), 124.9 (q,  $J = 32.1$  Hz), 119.7 (q,  $J = 3.5$  Hz), 117.7 (q,  $J = 4.2$  Hz), 110.5, 40.0, 15.3. IR (neat,  $cm^{-1}$ ): 1620, 1436, 1385, 1331, 1234, 1147, 1115, 1050, 885, 813, 775, 707. Anal. Calcd for  $C_{16}H_{13}F_3N_2$ : C, 66.20; H, 4.51. Found: C, 66.34; H, 4.55.

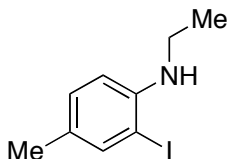


**1-ethyl-2-phenyl-6-(trifluoromethyl)-1H-benzimidazole (Scheme 5, 18h)** Following procedure B, *ortho*-iodo-*N*-alkylaniline **16d** (149 mg, 0.47 mmol) was coupled with benzamide (68.8 mg, 0.57 mmol). The cyclodehydration step was accomplished using procedure C (151 mg of  $K_3PO_4$ , 0.5 mL of *t*-BuOH, 8 h). The crude product was purified by flash chromatography on silica gel (gradient elution: 4/1, and then 3/1 hexanes/ ethyl acetate) to provide the title compound as a white solid (114 mg, 83%). mp 141-143 °C.  $^1H$  NMR (500 MHz,  $CDCl_3$ )  $\delta$ : 7.91 (dd,  $J = 8.6, 0.7$  Hz, 1H), 7.77-7.73 (m, 2H), 7.72 (t,  $J = 0.8$  Hz, 1H), 7.58-7.55 (m, 4H), 4.36 (q,  $J = 7.3$  Hz, 2H), 1.52 (t,  $J = 7.3$  Hz, 3H).  $^{13}C$  NMR (126 MHz,  $CDCl_3$ )  $\delta$ : 156.1, 145.5, 135.0, 130.4, 130.0, 129.4, 129.0, 125.0 (q,  $J = 272.0$  Hz), 124.9 (q,  $J = 32.1$  Hz), 120.5, 119.5 (q,  $J = 3.5$  Hz), 107.8 (q,  $J = 4.4$  Hz), 40.0, 15.5. IR (neat,  $cm^{-1}$ ): 1467, 1400, 1345, 1309, 1158, 1113, 869, 821, 780, 711, 700, 664. Anal. Calcd for  $C_{16}H_{13}F_3N_2$ : C, 66.20; H, 4.51. Found: C, 66.08; H, 4.51.

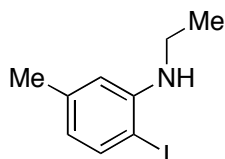
#### Procedure for the Preparation of *ortho*-Iodo-*N*-Alkylanilines **16a-d** Used in Scheme 5



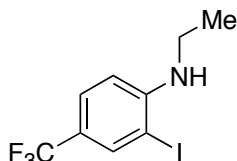
*ortho*-Iodo-*N*-alkylanilines **16a-d** were prepared via a three-step sequence (acetamide formation, copper-catalyzed halogen exchange, and borane reduction of acetamide) starting from the corresponding commercial available *ortho*-bromoanilines. The procedures for acetamide formation<sup>5</sup> and copper-catalyzed halogen exchange<sup>9</sup> have been previously disclosed. Full characterizations of **16a-d** are listed below.



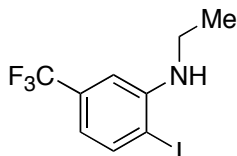
***N*-ethyl-2-iodo-4-methylaniline (16a)**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$ : 7.51 (d,  $J = 2.0\text{Hz}$ , 1H), 7.03 (dd,  $J = 8.3, 2.0\text{Hz}$ , 1H), 6.49 (d,  $J = 8.3\text{Hz}$ , 1H), 3.89 (bs, 1H), 3.18 (q,  $J = 7.1\text{Hz}$ , 2H), 2.22 (s, 3H), 1.31 (t,  $J = 7.1\text{Hz}$ , 3H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$ : 145.4, 139.3, 130.1, 127.9, 110.6, 85.5, 39.1, 20.0, 14.9. IR (neat,  $\text{cm}^{-1}$ ): 3389, 2968, 2867, 1608, 1514, 1312, 1273, 1163, 1029, 799, 664. Anal. Calcd for  $\text{C}_9\text{H}_{12}\text{IN}$ : C, 41.40; H, 4.63. Found: C, 41.68; H, 4.71.



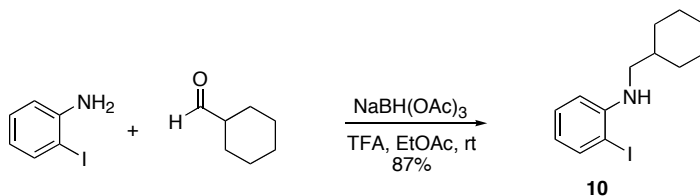
***N*-ethyl-2-iodo-5-methylaniline (16b)**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$ : 7.52 (d,  $J = 8.0\text{Hz}$ , 1H), 6.40 (d,  $J = 1.7\text{Hz}$ , 1H), 6.29 (ddd,  $J = 7.9, 2.0, 0.6\text{Hz}$ , 1H), 3.99 (bs, 1H), 3.19 (qd,  $J = 7.1, 5.2\text{Hz}$ , 2H), 2.28 (s, 3H), 1.32 (t,  $J = 7.1\text{Hz}$ , 3H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$ : 147.2, 139.5, 138.6, 119.6, 111.5, 81.6, 38.8, 21.6, 14.8. IR (neat,  $\text{cm}^{-1}$ ): 3391, 2968, 2869, 1591, 1505, 1419, 1304, 1187, 1004, 786, 584. Anal. Calcd for  $\text{C}_9\text{H}_{12}\text{IN}$ : C, 41.40; H, 4.63. Found: C, 41.16; H, 4.58.



***N*-ethyl-2-iodo-4-(trifluoromethyl)aniline (16c)**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$ : 7.88 (d,  $J = 1.5\text{Hz}$ , 1H), 7.45 (ddd,  $J = 8.6, 1.4, 0.7\text{Hz}$ , 1H), 6.54 (d,  $J = 8.6\text{Hz}$ , 1H), 4.44 (bs, 1H), 3.25 (qd,  $J = 7.1, 5.2\text{Hz}$ , 2H), 1.35 (t,  $J = 7.1\text{Hz}$ , 3H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$ : 149.8, 136.1 (q,  $J = 3.8\text{Hz}$ ), 126.9 (q,  $J = 3.8\text{Hz}$ ), 124.1 (q,  $J = 270.8\text{Hz}$ ), 119.8 (q,  $J = 33.0\text{Hz}$ ), 109.2, 83.7, 38.7, 14.5. IR (neat,  $\text{cm}^{-1}$ ): 3398, 2975, 2875, 1607, 1532, 1322, 1280, 1154, 1112, 1079, 805, 675. Anal. Calcd for  $\text{C}_9\text{H}_9\text{F}_3\text{IN}$ : C, 34.31; H, 2.88. Found: C, 34.66; H, 2.87.



***N*-ethyl-2-iodo-5-(trifluoromethyl)aniline (16d)**  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$ : 7.75 (dd,  $J = 8.1, 0.8\text{Hz}$ , 1H), 6.70 (s, 1H), 6.67 (ddd,  $J = 8.1, 2.1, 0.6\text{Hz}$ , 1H), 4.28 (bs, 1H), 3.24 (qd,  $J = 7.1, 5.2\text{Hz}$ , 2H), 1.36 (t,  $J = 7.1\text{Hz}$ , 3H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$ : 147.8, 139.5, 132.1 (q,  $J = 32.1\text{Hz}$ ), 124.4 (q,  $J = 272.4\text{Hz}$ ), 114.6 (q,  $J = 4.0\text{Hz}$ ), 106.4 (q,  $J = 4.0\text{Hz}$ ), 88.8 (q,  $J = 1.2\text{Hz}$ ), 38.8, 14.6. IR (neat,  $\text{cm}^{-1}$ ): 3398, 2974, 1597, 1580, 1513, 1434, 1331, 1280, 1167, 1124, 1082, 1006, 854.

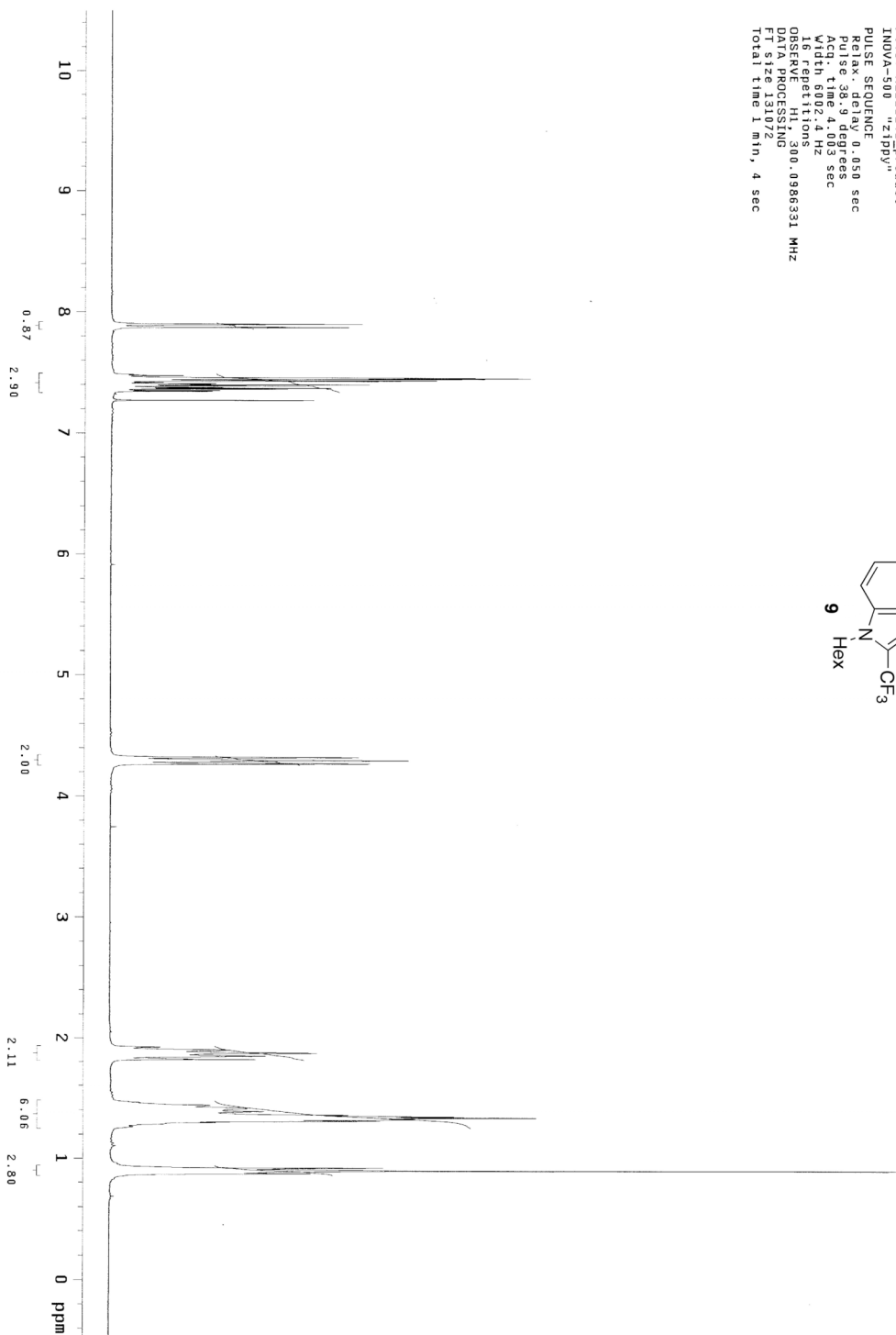
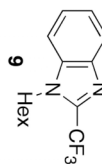


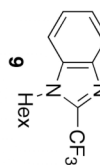
***N*-(cyclohexylmethyl)-2-iodoaniline** Following a published procedure,<sup>10</sup> 2-iodoaniline (2.0 g, 9.1 mmol) and cyclohexylcarboxyaldehyde (1.2 mL, 10.0 mmol) were converted to the title compound. The crude compound was purified by flash column chromatography on silica gel (gradient elution: hexanes and then 50/1 hexanes/ethyl acetate) to provide **10** as a light yellow oil (2.81 g, 87 %). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ: 7.65 (dd, *J* = 7.8, 1.6Hz, 1H), 7.20 (td, *J* = 8.2, 1.5Hz, 1H), 6.55 (dd, *J* = 8.2, 1.3Hz, 1H), 6.42 (td, *J* = 7.6, 1.5Hz, 1H), 4.24 (bs, 1H), 3.00 (t, *J* = 6.1Hz, 2H), 1.88-1.55 (m, 6H), 1.36-1.12 (m, 3H), 1.08-0.95 (m, 2H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ: 147.6, 139.1, 129.6, 118.3, 110.6, 85.6, 51.0, 37.4, 31.5, 26.7, 26.1. IR (neat, cm<sup>-1</sup>): 3400, 2922, 2850, 1591, 1507, 1449, 1426, 1320, 1004, 739. Anal. Calcd for C<sub>13</sub>H<sub>18</sub>IN: C, 49.54; H, 5.76. Found: C, 49.78; H, 5.73.

## References

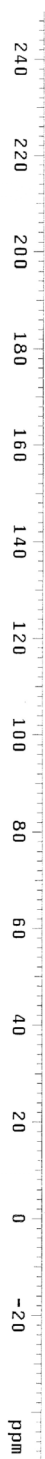
- <sup>1</sup> Tomori, H.; Fox, J. M.; Buchwald, S. L. *J. Org. Chem.* **2000**, *65*, 5334-5341.
- <sup>2</sup> Cironi, P.; Tulla-Puche, J.; Barany, G.; Albericio, F.; Alvarez, M. *Org. Lett.* **2004**, *6*, 1405-1408.
- <sup>3</sup> Nystrom, R. F. *J. Am. Chem. Soc.* **1955**, *77*, 2544-2545.
- <sup>4</sup> Kabalka, G. W.; Wang, L.; Pagni, R. M. *Tetrahedron* **2001**, *57*, 8017-8028.
- <sup>5</sup> Tsang, W. C. P.; Zheng, N.; Buchwald, S. L. *J. Am. Chem. Soc.* **2005**, *127*, 14560-14561.
- <sup>6</sup> Hesk, D.; Das, P. R.; Evans, B. *J. Labelled Compd. Radiopharm.* **1995**, *36*, 497-502.
- <sup>7</sup> Zou, B.; Yuan, Q.; Ma, D. *Angew. Chem. Int. Ed.* **2007**, *46*, 2598-2601.
- <sup>8</sup> Lewis, J. C.; Wiedeman, S. H.; Bergman, R. G.; Ellman, J. A. *Org. Letts.* **2004**, *6*, 35-38.
- <sup>9</sup> Klapars, A.; Buchwald, S. L. *J. Am. Chem. Soc.* **2002**, *124*, 14844-14845.
- <sup>10</sup> McLaughlin, M.; Palucki, M.; Davies, I. W. *Org. Letts.* **2006**, *8*, 3307-3310.

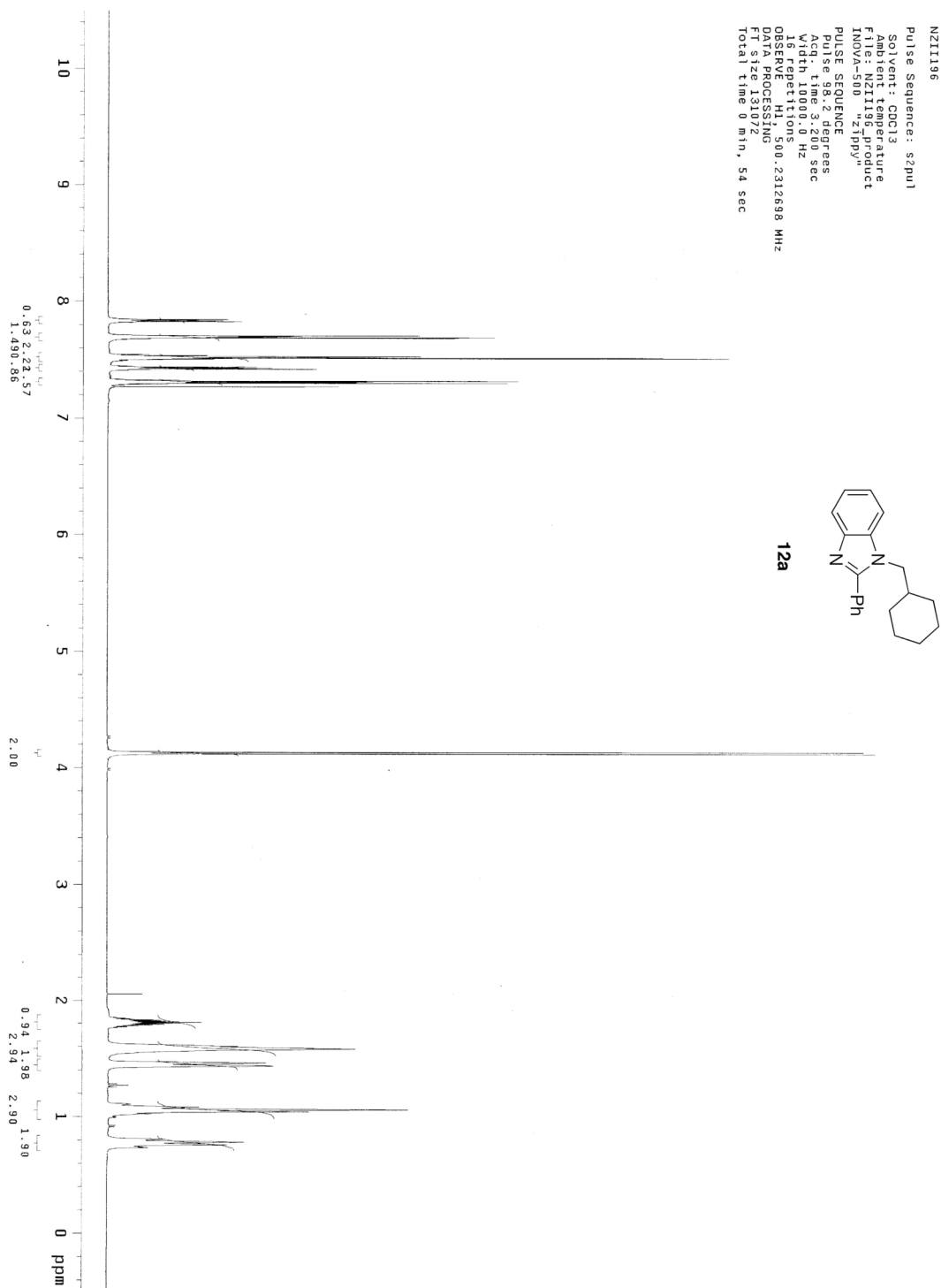
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F1 size 131072  
Total time 1 min, 4 sec

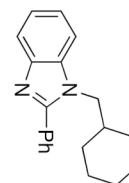




NZII1142  
Pulse Sequence: zgpg30  
Solvent: CDCl3  
Ambient temperature  
User: 1-11-97  
File: NZII1142.C13  
INSTR: spect  
PULPROG: zgpg30  
PULSE SEQUENCE  
P1: 12.00 sec  
P2: 0.76 sec  
P3: 0.76 sec  
Acq. time: 1.736 sec  
Width: 37735.8 Hz  
136 repetitions  
OBSERVE C13, 125.7832383 MHz  
DECUPLE H1, 500.232753 MHz  
SFO: 500.131072 MHz  
WALTZ-16 modulated  
continuously on  
DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 131072  
Total time 696 hr, 41 min, 12 sec

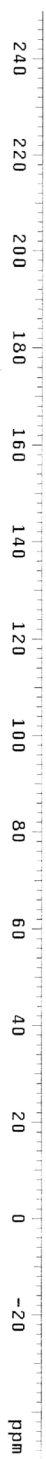


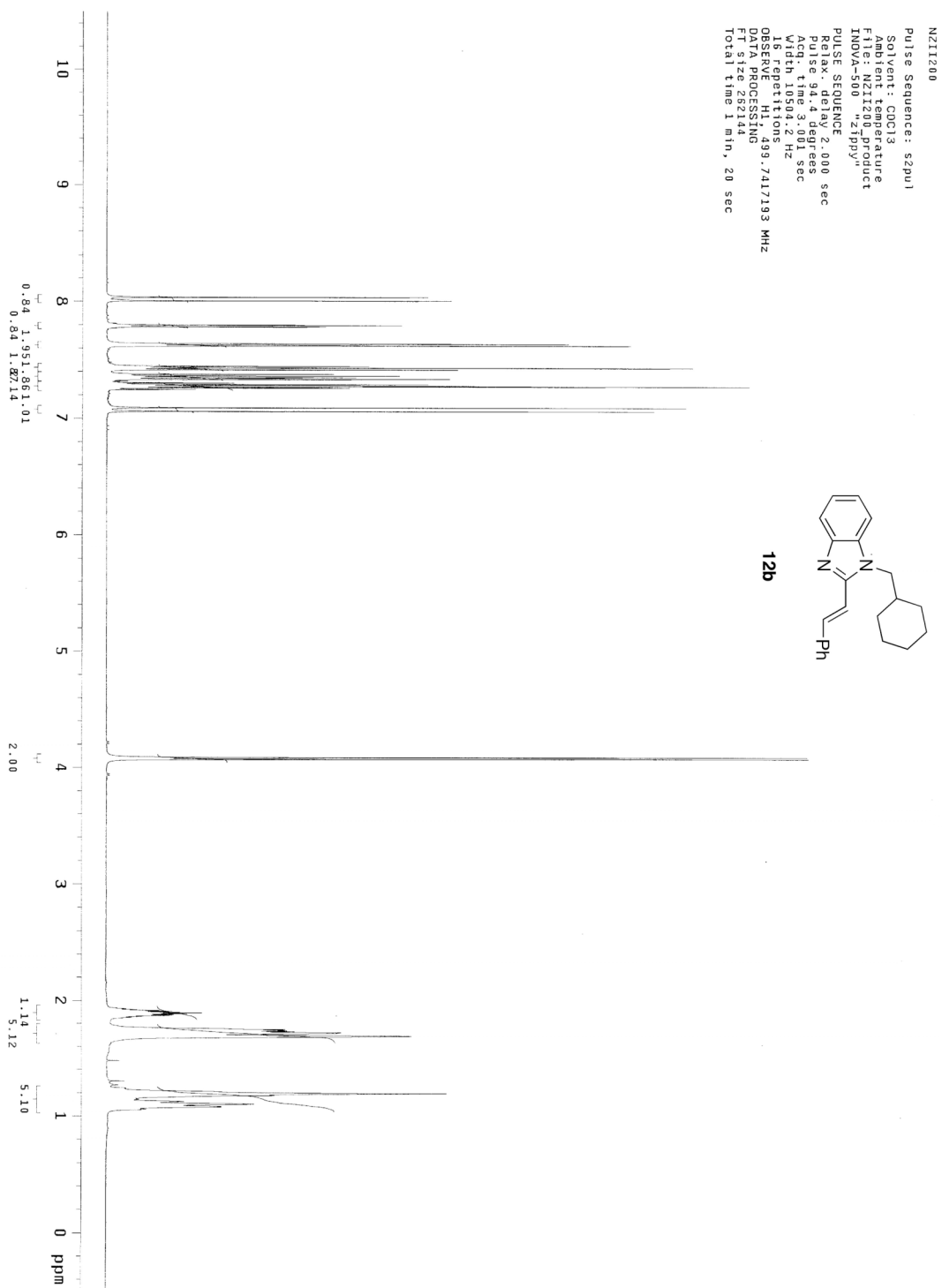




12a

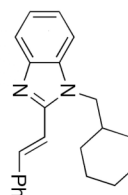
NZ11196  
Pulse Sequence: s2pu1  
Solvent: CDCl3  
Ambient temperature  
User: 1-14-87  
File: NZ11196\_product13  
INNOVA-500 2ippy  
PULSE SEQUENCE  
Relax: delay 0.763 sec  
Pulse: 4.1097 sec  
Acq: time 1.735 sec  
Width 3735.8 Hz  
10000 repetitions  
OBSERVE C13, 125.7632452 MHz  
DECOUPLE H1, 500.2332753 MHz  
Power 37.0 dB  
Spectrometer on  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 131072  
Total time 6 hr, 58 min, 3 sec



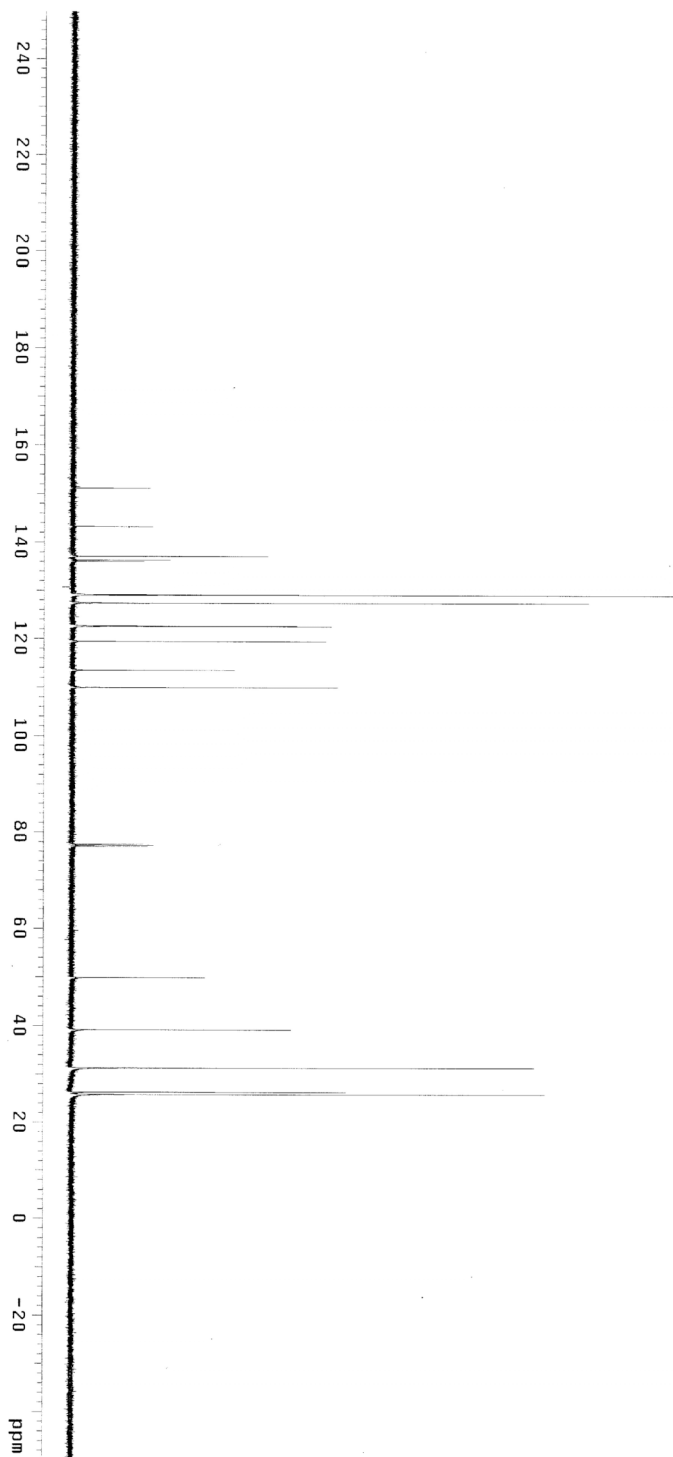




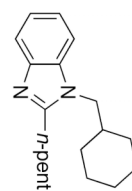
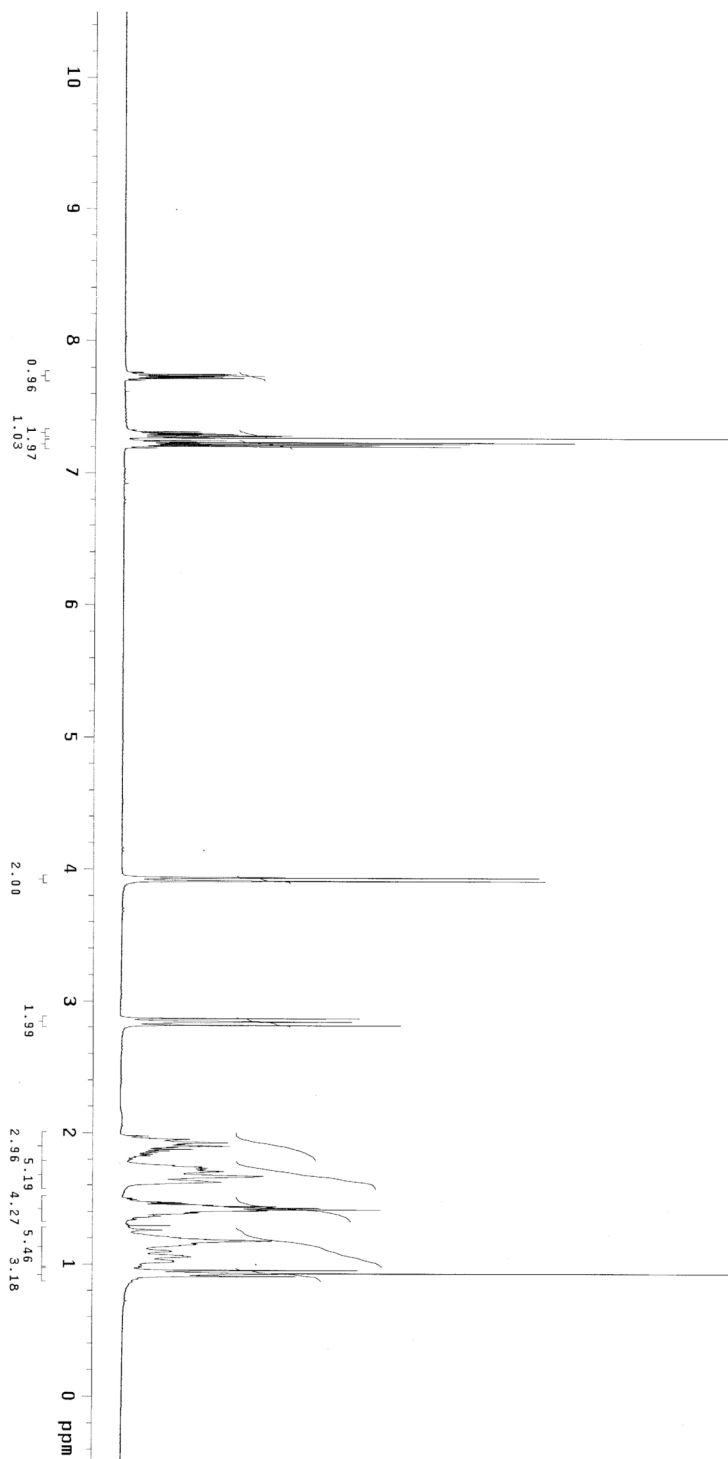
NZ11200  
Pulse Sequence: szpul  
Solvent: CDCl3  
Ambient temperature  
User: 1-14-87  
File: NZ11200\_1Cl3a  
INOVA-500 21ppm  
PULSE SEQUENCE  
Relax: delay 0.763 sec  
P1 1.736 sec  
Acq: time 1.736 sec  
Width 3735.8 Hz  
120 repetitions  
OBSERVE C13, 125.763423 MHz  
DECOUPLE H1, 500.2332753 MHz  
SOLVENT 125.763423 MHz  
Continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 131072  
Total time 69 hr, 40 min, 9 sec



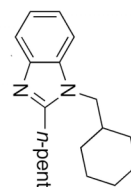
12b



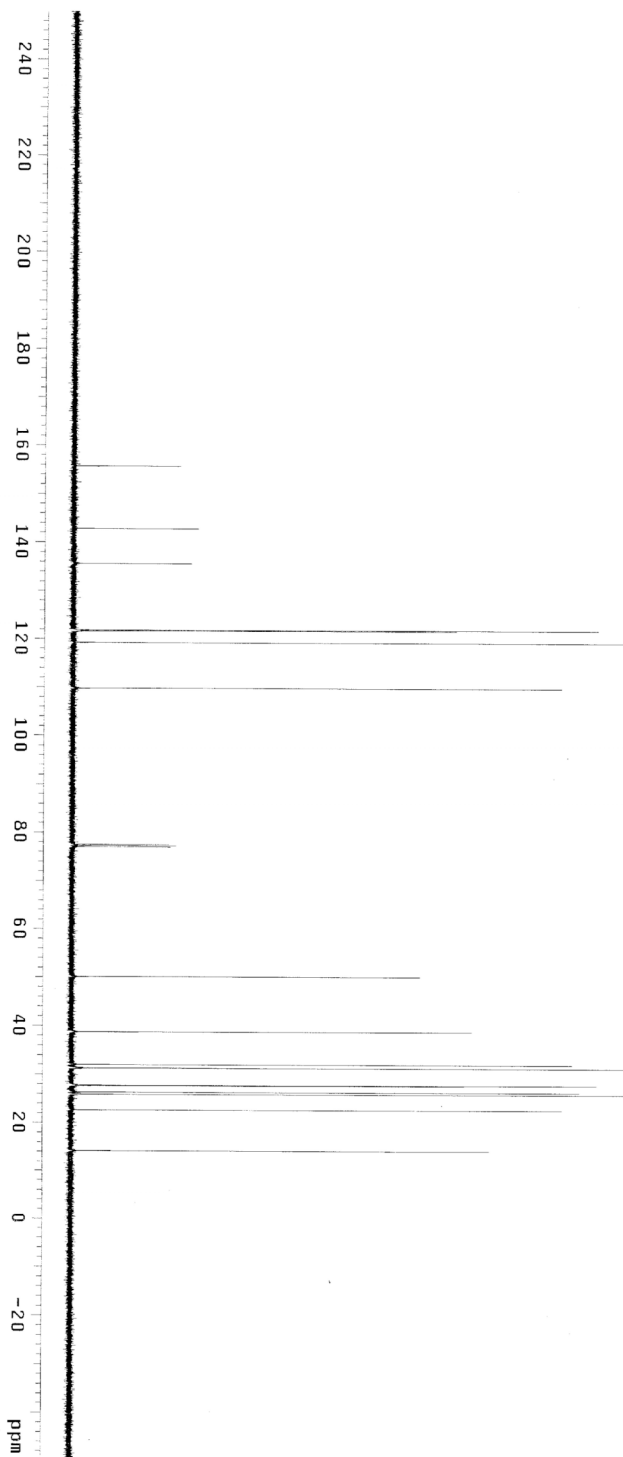
NZ11213  
Pulse Sequence: szpu1  
Solvent: CDCl3  
Ambient temperature  
File: NZ11213.product  
INOVA-500 "zippy"  
PULSE SEQUENCE  
Relax: delay 0.050 sec  
Pulse: 32.3 degrees  
Acq: 1.500 sec  
Width: 6002.4 Hz  
16 repetitions  
OBSERVE: H1, 300.0986330 MHz  
DATA PROCESSING  
FT size: 131072  
Total time 1 min, 4 sec

**12c**

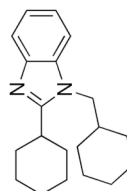
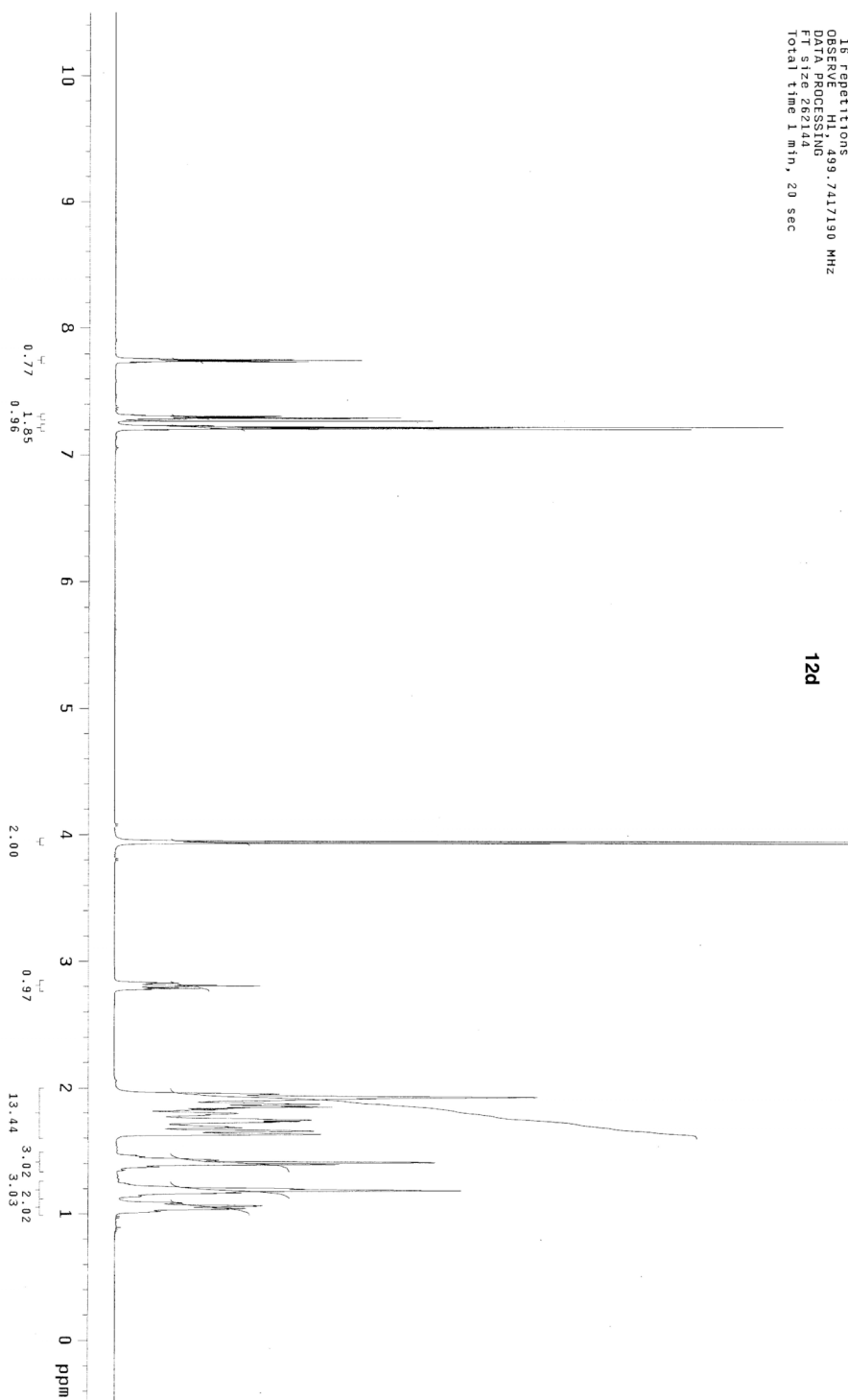
NZII1057  
Pulse Sequence: s2pul  
Solvent: CDCl3  
Ambient temperature  
User: 111-87  
Date: NZII1057  
INDVA-500 "z1py"  
PULSE SEQUENCE  
P3 0.1 sec, 1300.763 sec  
Pulse 65.4 degrees  
Acq. time 1.736 sec  
Width 37735.8 Hz  
64 repetitions  
OBSERVE C13, 125.7632389 MHz  
DECOUPL 1H, 500.2532753 MHz  
Power 157 dB  
continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 131072  
Total time 5 hr, 58 min, 3 sec



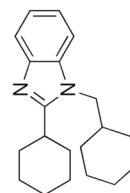
12c



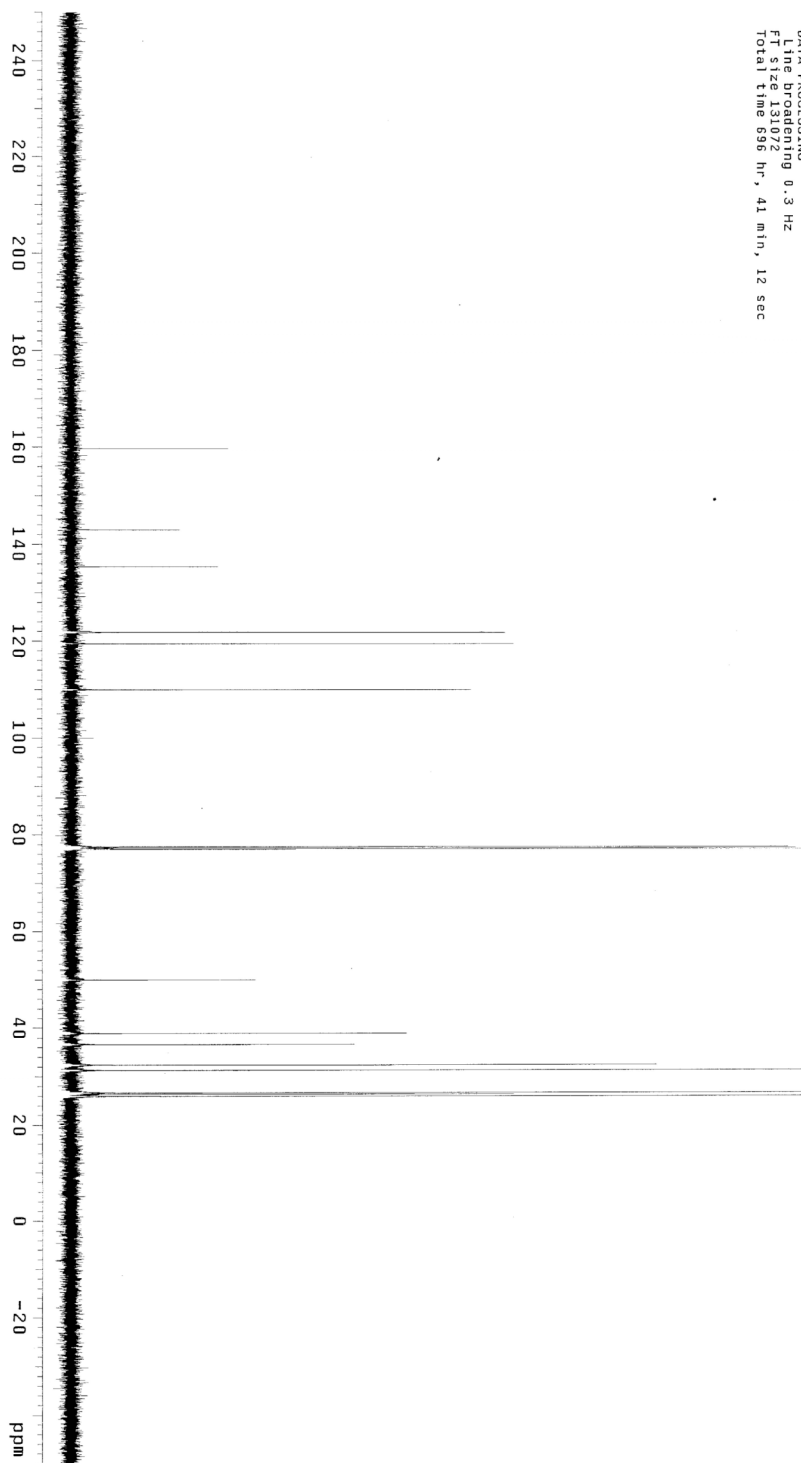
NZ111059  
Pulse Sequence: zgpg30  
Solvent: CDCl3  
Ambient temperature  
File: NZ111059\_Product  
INDY-500 "zgpg30"  
PULSE SEQUENCE  
Relax: delay 2.000 sec  
Pulse: 9.4 degrees  
Acq: 1.000 sec  
Width: 10504.2 Hz  
16 repetitions  
OBSERVE: H1, 499.7417190 MHz  
DATA PROCESSING  
FT size: 262144  
Total time: 1 min, 20 sec

**12d**

NZ111059  
Pulse Sequence: szpul  
Solvent: CDCl3  
Sample Name: 12d  
User: 11-14-87  
File: NZ111059.ProductC13  
INOVA-500 "2tppv"  
PULSE SEQUENCE  
Relax. delay 0.763 sec  
Pulse 77.6 degrees  
Acq. time 1.736 sec  
Width 37235.8 Hz  
NUC1 13C  
OBSERVE C13 125 763291 MHz  
DECOUPLE H1 500.2332753 MHz  
Power 44 dB  
continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
NUC2 13C  
FT size 131072  
Total time 696 hr, 41 min, 12 sec



12d



NZ111029

Pulse Sequence: szpu1

Solvent: CDCl3

Ambient temperature

File: NZ111029\_Product

INOVA-500 "z1ppy"

PULSE SEQUENCE

Relax. delay 0.050 sec

Pulse 36.9 degrees

Pulse 36.9 degrees

Width 6002.4 Hz

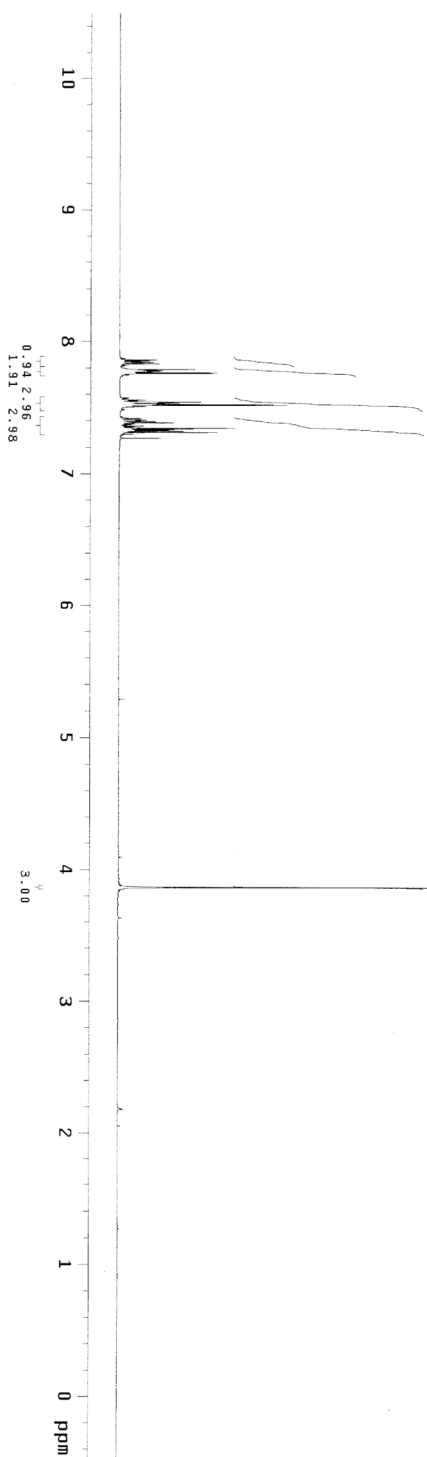
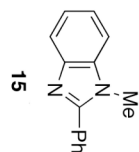
16 repetitions

OBSERVE H1, 300.0985328 MHz

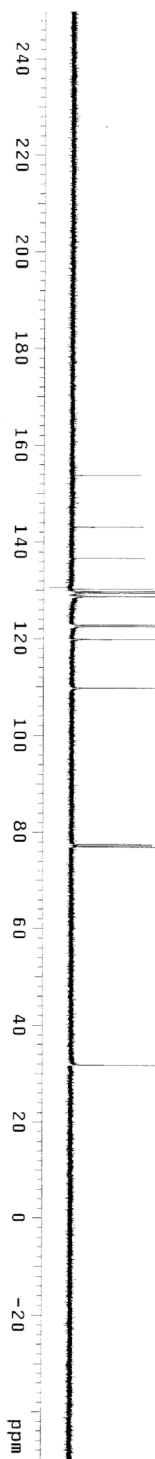
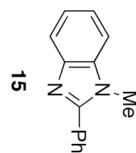
DATA PROCESSING

FT size 131072

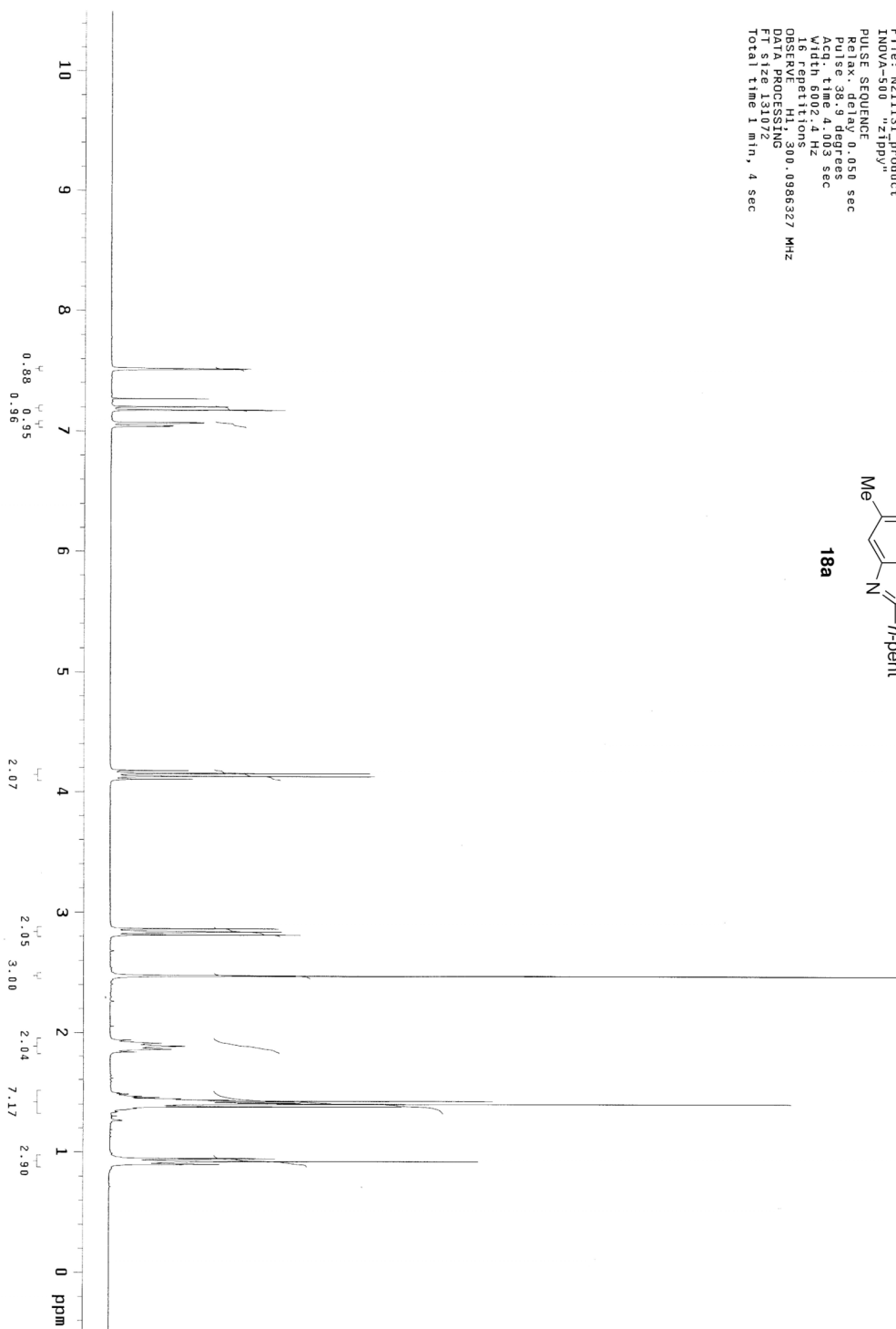
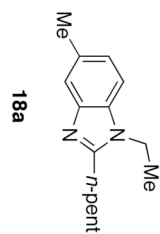
Total time 1 min, 4 sec



NZII1029  
Pulse Sequence: zgpg30  
Solvent: CDCl3  
Ambient temperature  
User: 11-14-87 C13  
Date: NZII1029 C13  
INOVA-500 "zgpg30"  
PULSE SEQUENCE  
zgpg30: 41.90.763 sec  
Pulse: 77.6 degrees  
Acq. time 1.736 sec  
Width 37735.8 Hz  
136 repetitions  
OBSERVE C13, 125.7832463 MHz  
DECOUPLE H1, 500.2532753 MHz  
Power 14.4 dB  
continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 0.3 Hz  
F1 size 13102  
Total time 636 hr, 41 min, 12 sec

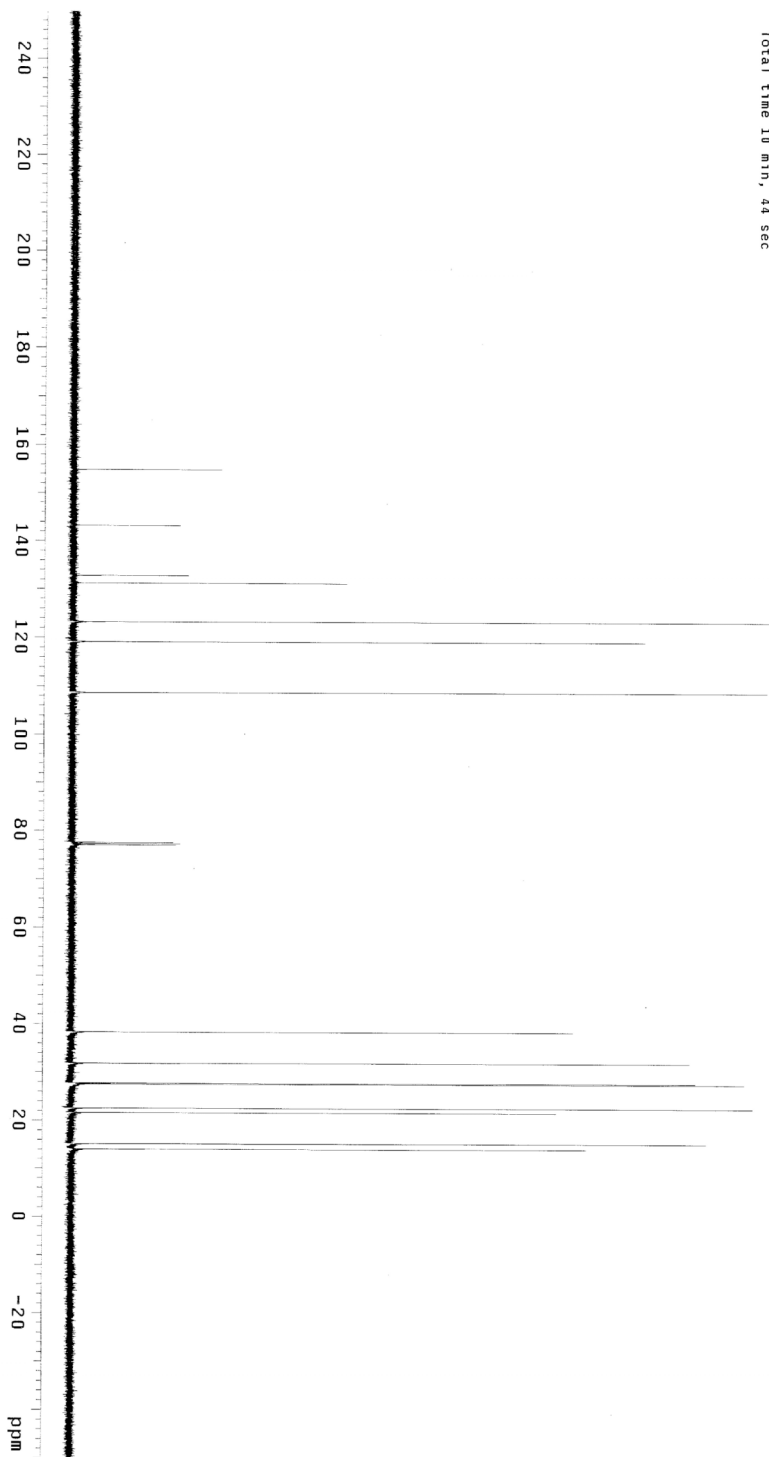
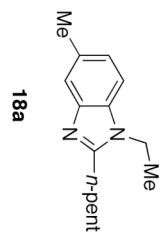


NZ111031  
Pulse Sequence: zgpg30  
Solvent: CDCl3  
Ambient temperature  
File: NZ11131product  
INOVA-500 "zgpg30"  
PULSE SEQUENCE  
Relax: delay 0.050 sec  
Acq: 300.136 MHz  
Acq: time 4.000 sec  
Width 6002.4 Hz  
16 repetitions  
OBSERVE H1, 300.0986327 MHz  
DATA PROCESSING  
F1 size 131072  
Total time 1 min, 4 sec

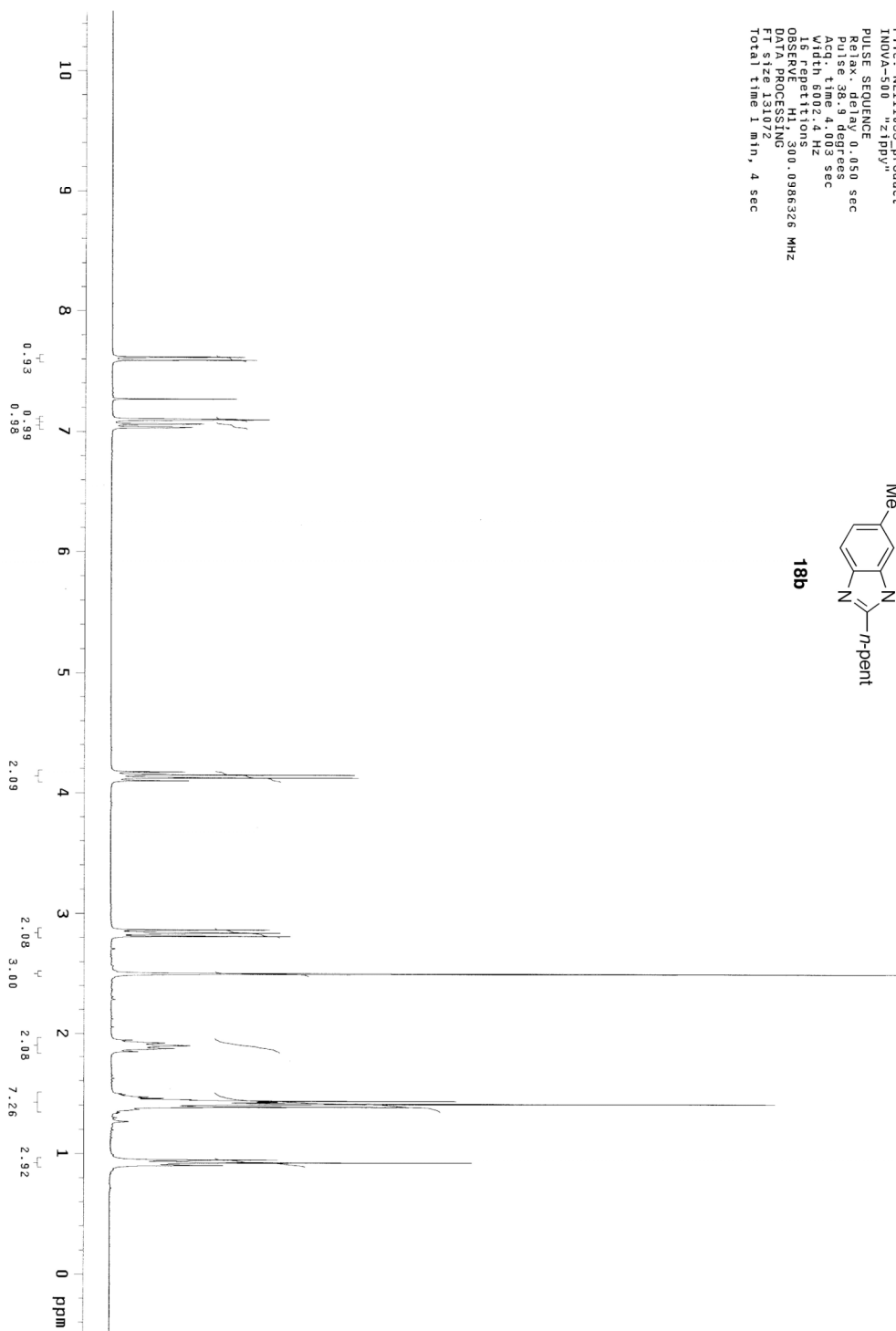
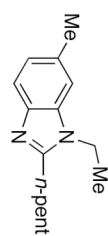




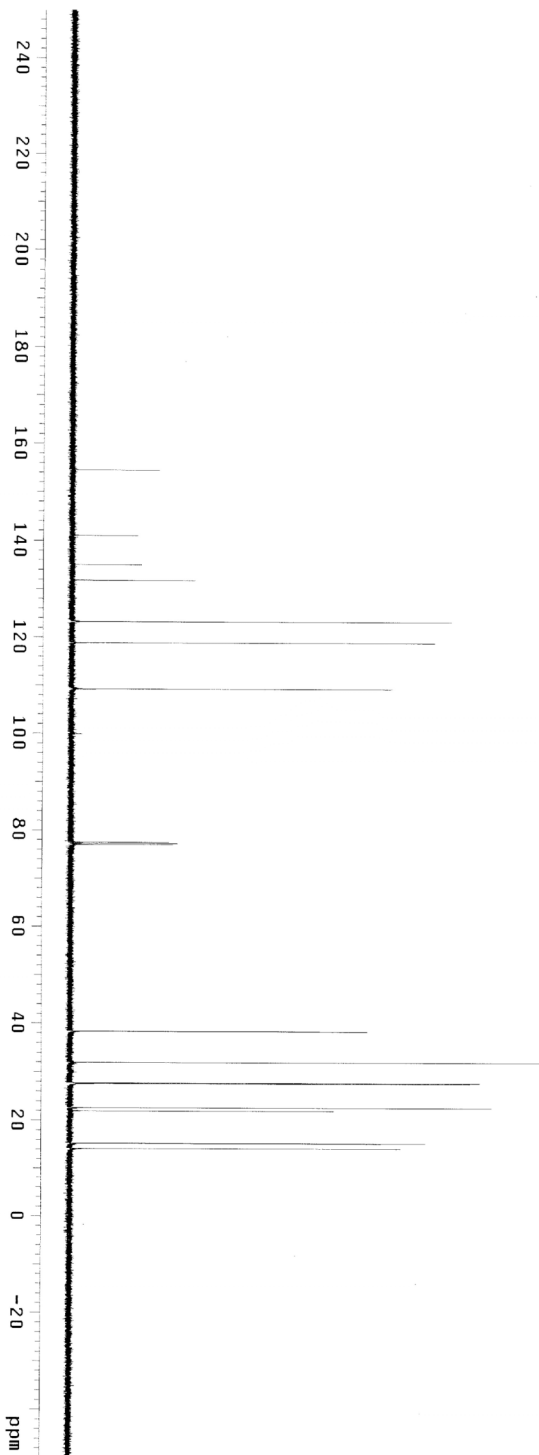
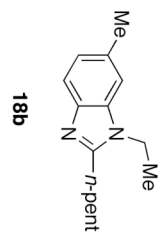
NZ11031  
Pulse Sequence: szpu1  
Solvent: CDCl3  
Ambient temperature  
User: 1-14-87  
File: NZ11031\_product13  
INOVA-500 Zippy  
PULSE SEQUENCE  
Relax: delay 0.763 sec  
Pulse: 6.10 sec  
Acq time: 1.736 sec  
Width: 3725.8 Hz  
256 repetitions  
OBSERVE: C13, 125.7632452 MHz  
DECUPLE: H1, 500.2332753 MHz  
Power: 37.48 W  
Spectrum only on  
DATA PROCESSING  
WAIT-16 modulated  
Line broadening 0.3 Hz  
FT size 131072  
Total time 10 min, 44 sec



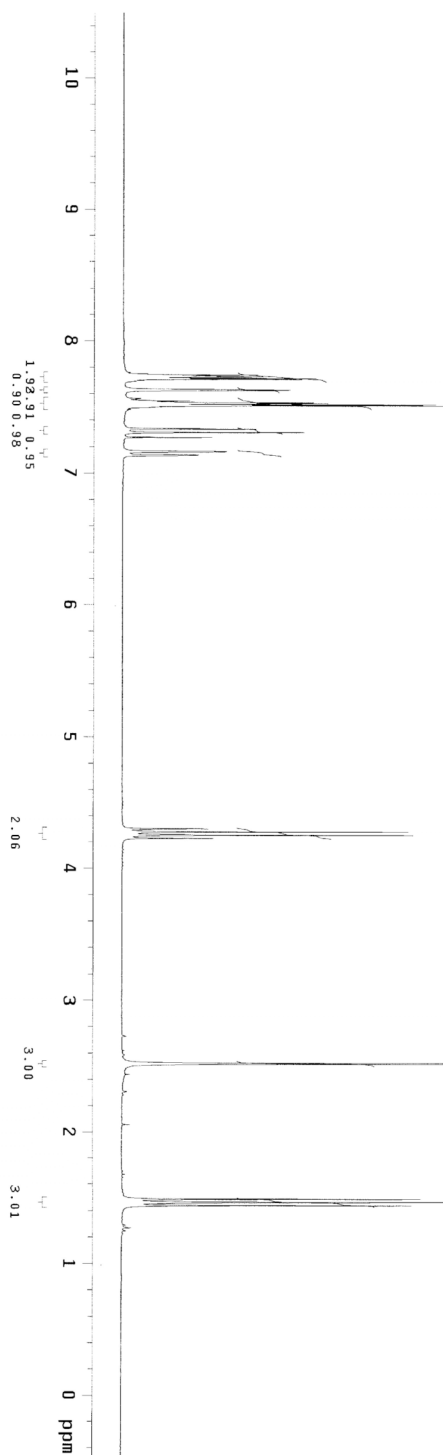
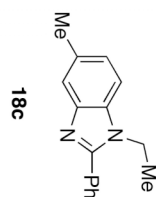
NZ111033  
Pulse Sequence: szpul  
Solvent: CDCl3  
Ambient temperature  
File: NZ111033\_product  
INOVA-500 zippy  
PULSE SEQUENCE  
Relax: delay 0.050 sec  
Acq: 3.33 sec  
Acq time 4.003 sec  
Width 6002.4 Hz  
16 repetitions  
OBSERVE H1, 300.0986326 MHz  
DATA PROCESSING  
F1 size 131072  
Total time 1 min, 4 sec



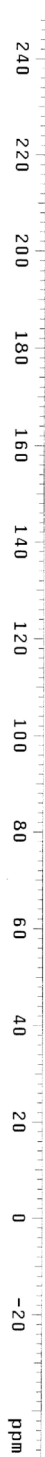
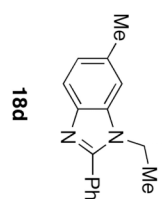
N211033  
Pulse Sequence: szpul  
Solvent: CDCl3  
Ambient temperature  
Date: N211033  
File: N211033.productC13  
INNOVA-500 "zippy"  
PULSE SEQUENCE  
Relax delay 0.763 sec  
Pulse 55.4 degrees  
Acq. time 1.736 sec  
Width 37735.8 Hz  
256 repetitions  
OBSERVE C13, 125.769337 MHz  
PULSE 4H, 500.2532753 MHz  
Power 37 dB  
continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 0.3 Hz  
F1 size 131072  
Total time 10 min, 44 sec



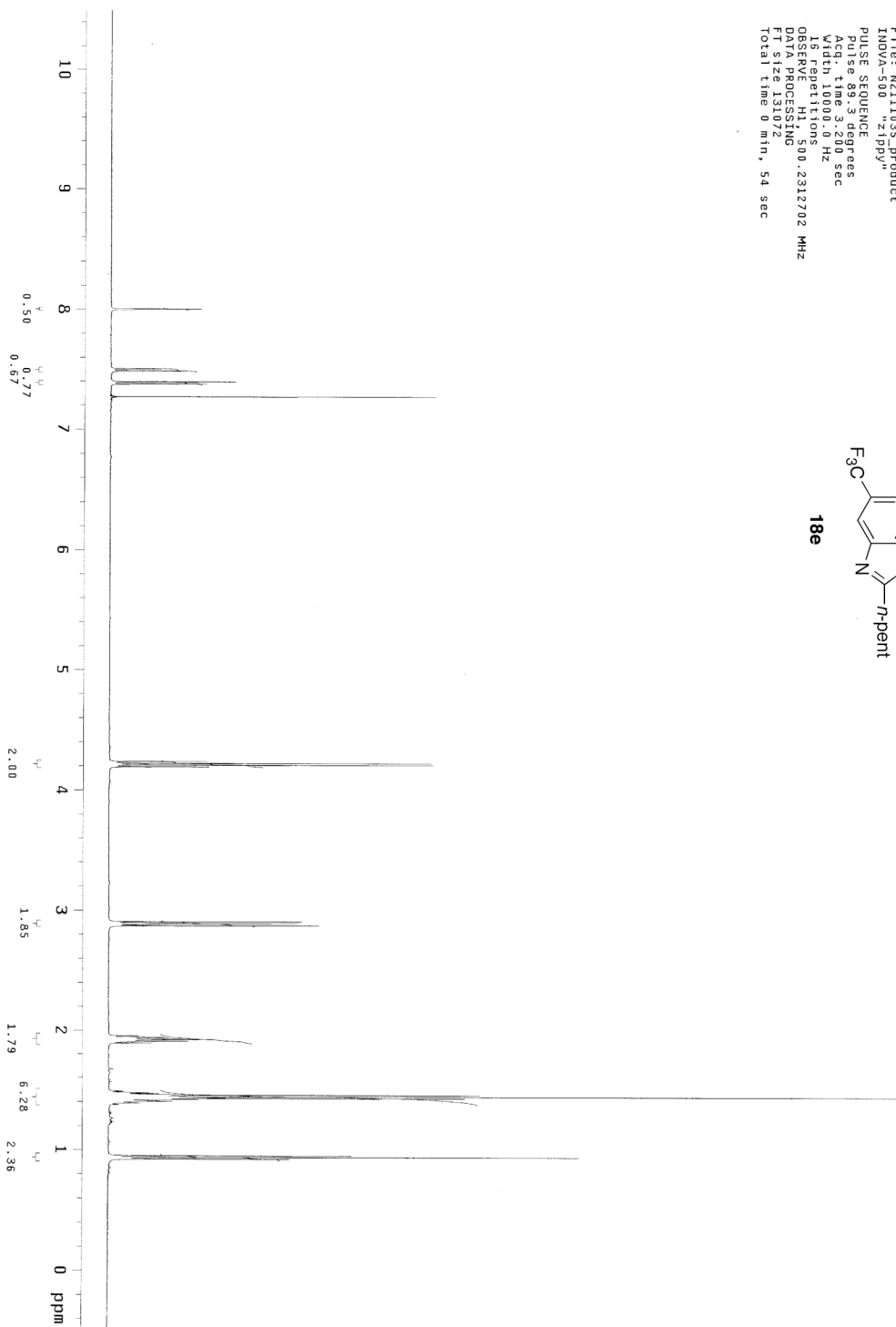
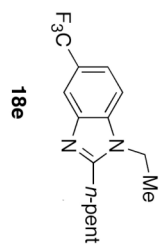
NZ1241  
Pulse Sequence: szpul  
Solvent: CDCl3  
Ambient temperature  
File: NZ1241\_product  
INNOVA-500 "zippy"  
PULSE SEQUENCE  
Relax: delay 0.050 sec  
Pulse: 32.9 degrees  
Acq: 120.0 sec  
Width 6002.4 Hz  
16 repetitions  
OBSERVE H1, 300.0986328 MHz  
DATA PROCESSING  
FT size 131072  
Total time 1 min, 4 sec



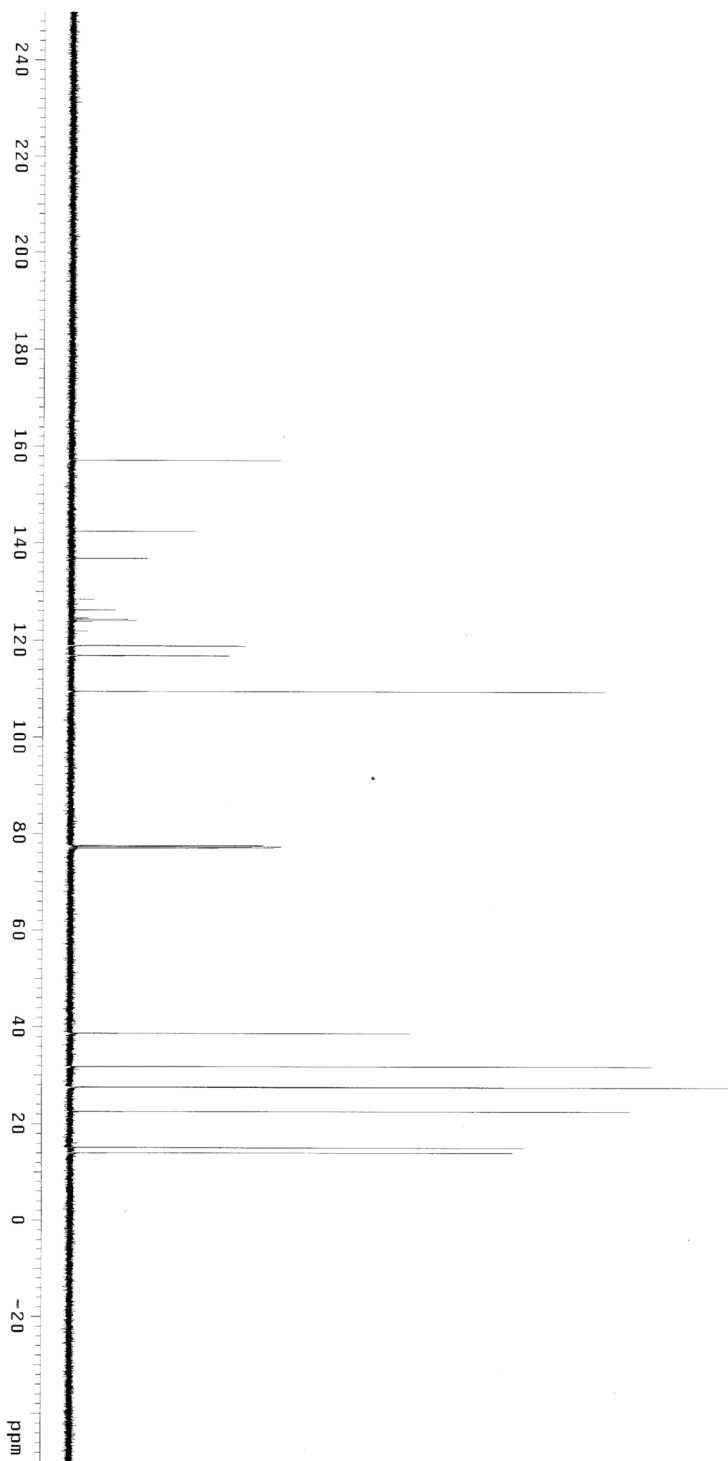
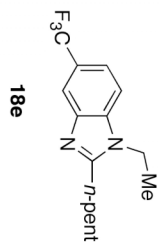
NZ11248  
Pulse Sequence: szpu1  
Solvent: CDCl3  
Ambient Temperature  
NUC1: 13C  
File: NZ11248\_product13  
INOVA-500 "zippy"  
PULSE SEQUENCE  
Relax delay 0.763 sec  
Pulse 65.4 degrees  
Acq. time 1.736 sec  
Width 37735.8 Hz  
88 repetitions  
OBSERVE CH: 125.763246 MHz  
DECOUPLE H1: 500.252753 MHz  
Power 37 dB  
continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
F1 time broadening 0.3 Hz  
F2 size 13102  
Total time 10 min, 44 sec



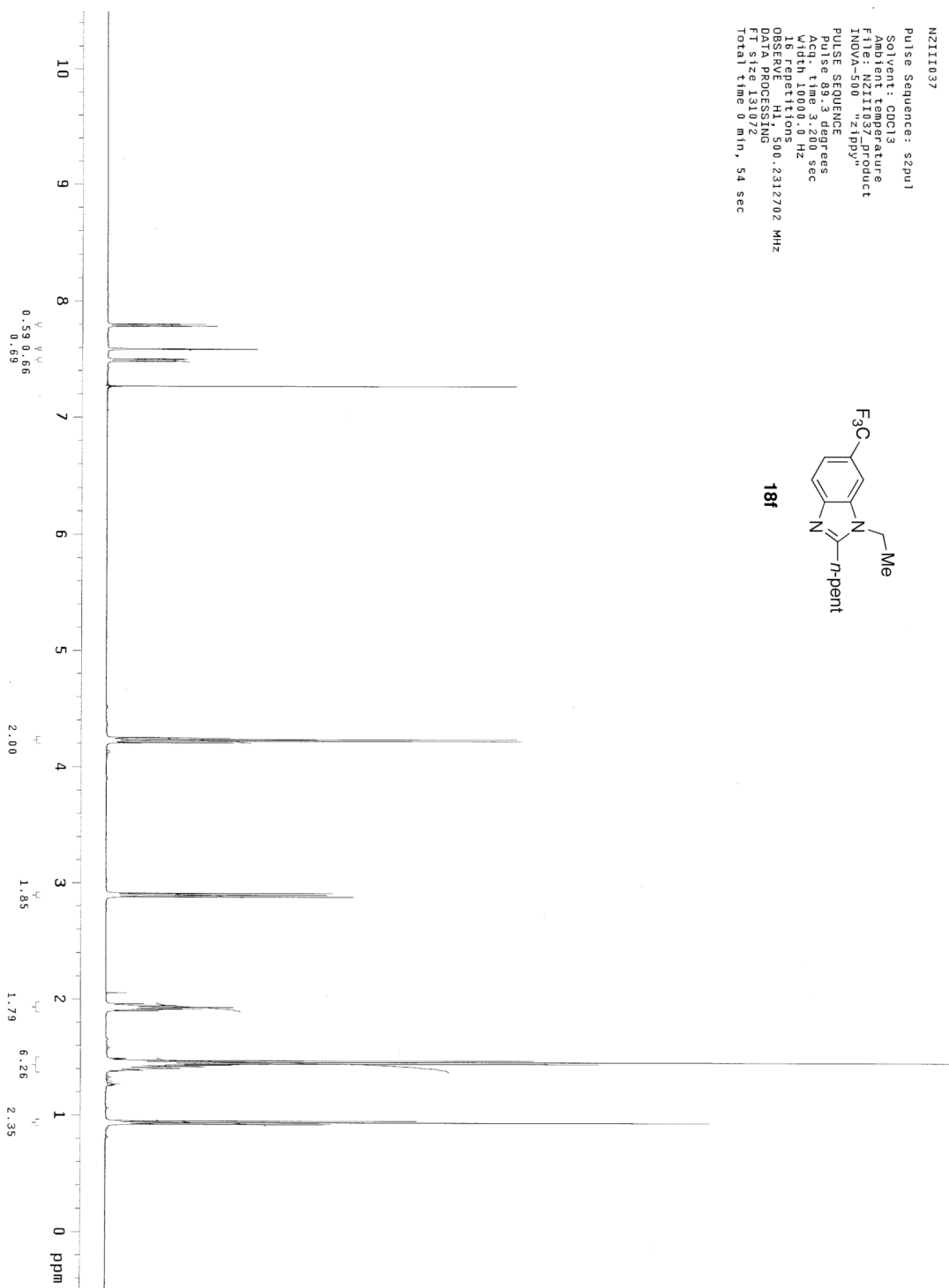
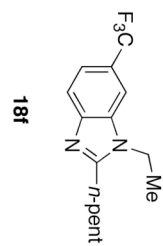
NZ111035  
Pulse Sequence: zgpg30  
Solvent: CDCl3  
Ambient temperature  
File: NZ111035-Product  
NOVA-500 "Zippy"  
PULSE SEQUENCE  
Pulse 89.3 degrees  
Pulse 14.0 degrees  
Width 1000.200 sec  
15 repetitions  
OBSERVE H1, 500.2312702 MHz  
DATA PROCESSING  
FT size 131072  
Total time 0 min, 54 sec



NZ111035  
Pulse Sequence: szpu1  
Solvent: CDCl3  
Ambient temperature  
User: 1111-87  
File: NZ111035\_ProductC13  
INOVA-500 "2ippy"  
PULSE SEQUENCE  
zgpg30: 120.763 sec  
Pulse: 65.4 degrees  
Acq. time: 1.736 sec  
Width: 37735.8 Hz  
144 repetitions  
OBSERVE C13, 125.763342 MHz  
DECOUPLE H1, 500.2532753 MHz  
Pulse: 138.00000000000000  
continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 0.3 Hz  
FT size: 131072  
Total time 69 hr, 40 min, 9 sec

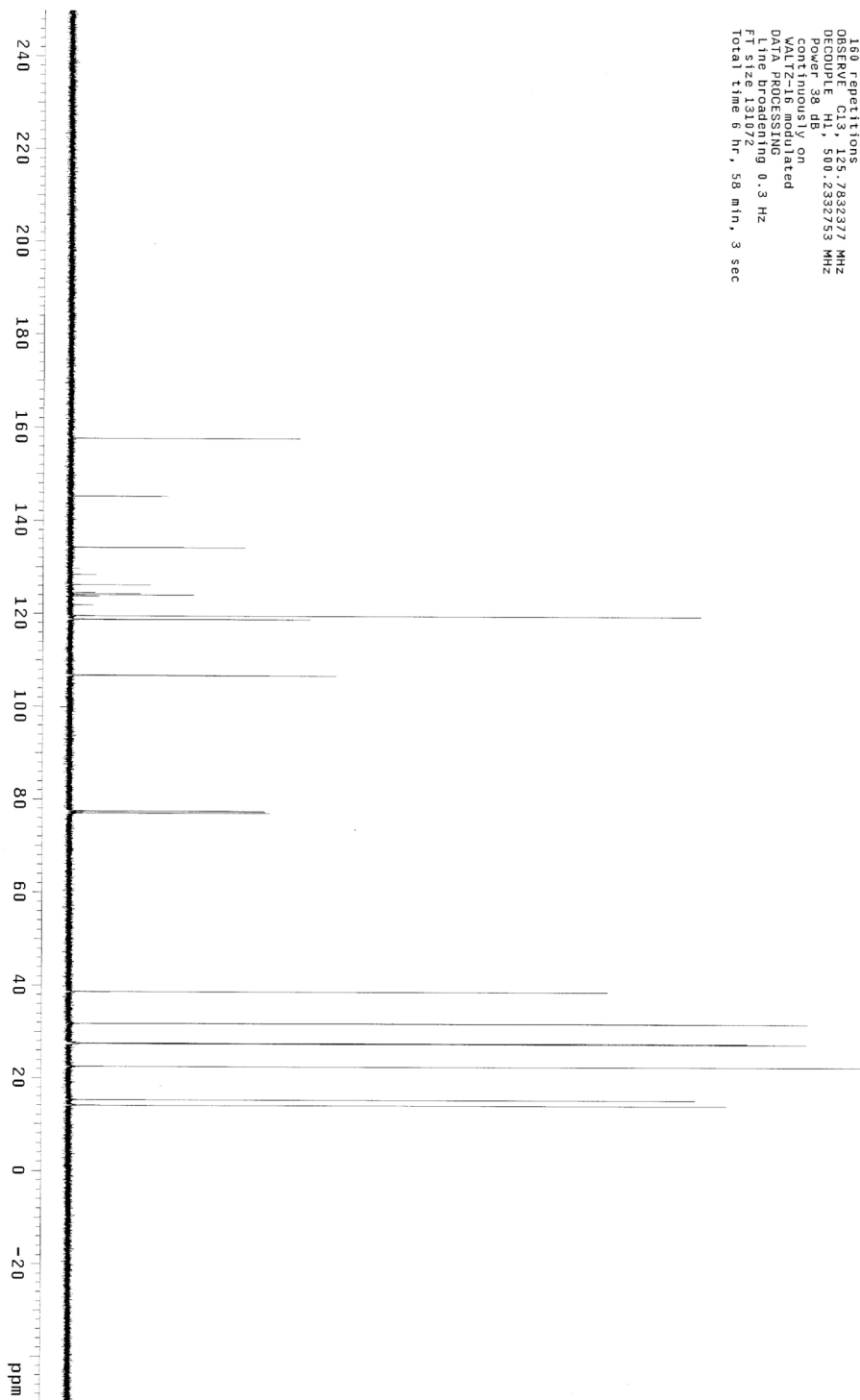
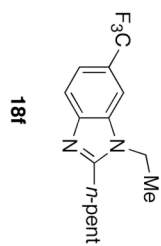


NZ11037  
Pulse Sequence: s2pu1  
Solvent: CDCl3  
Ambient temperature  
File: NZ11037\_Product  
INOVA-500 "zippy"  
PULSE SEQUENCE  
Pulse 89.3 degrees  
Acq. time 3.200 sec  
Acq. freq 500.130 MHz  
16 repetitions  
OBSERVE H1, 500.2312702 MHz  
DATA PROCESSING  
FT size 131072  
Total time 0 min, 54 sec

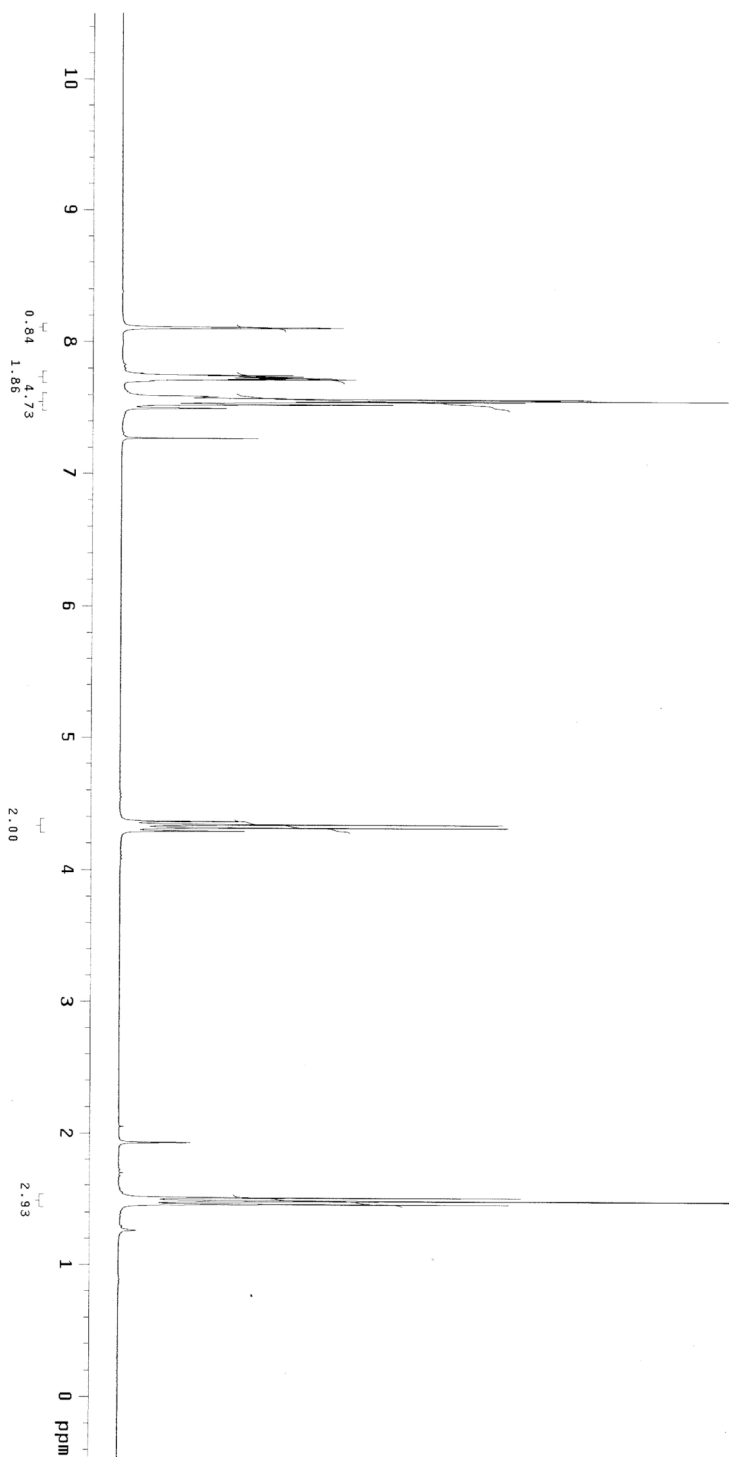
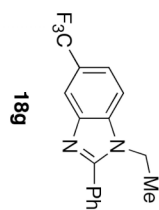




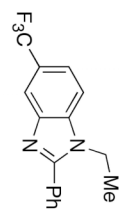
NZ111037  
Pulse Sequence: szpul  
Solvent: CDCl3  
Ambient Temperature  
User: 111-87  
File: NZ111037\_ProductC13  
INNOVA-500 "zippy"  
PULSE SEQUENCE  
NUC1: 13C, 125.763 MHz  
Acq. time 1.736 sec  
Pulse 55.4 degrees  
Width 3735.8 Hz  
160 repetitions  
OBSERVE C13, 125.763377 MHz  
DECOUPLE H1, 500.236753 MHz  
Pulse 38.41  
continuous on  
WALTZ-16 modulated  
DATA PROCESSING  
line broadening 0.3 Hz  
FT size 131072  
Total time 6 hr, 58 min, 3 sec



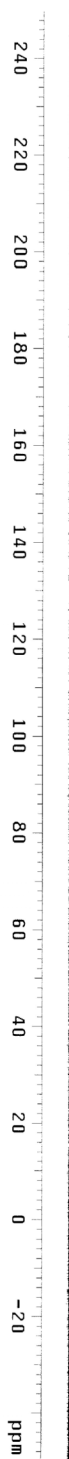
N2I1242  
Pulse Sequence: s2pul  
Solvent: CDCl3  
Ambient temperature  
File: N2I1242-product  
INSTRUM: spect  
PULSE SEQUENCE: zgpg30  
Relax: 3.00 sec  
Pulse: zgpg30  
Acq. time: 4.00 sec  
Width: 6002.4 Hz  
16 repetitions  
OBSERVE: H1, 300.0986326 MHz  
DATA PROCESSING  
F2: 300.13721072  
Total time: 1 min, 4 sec



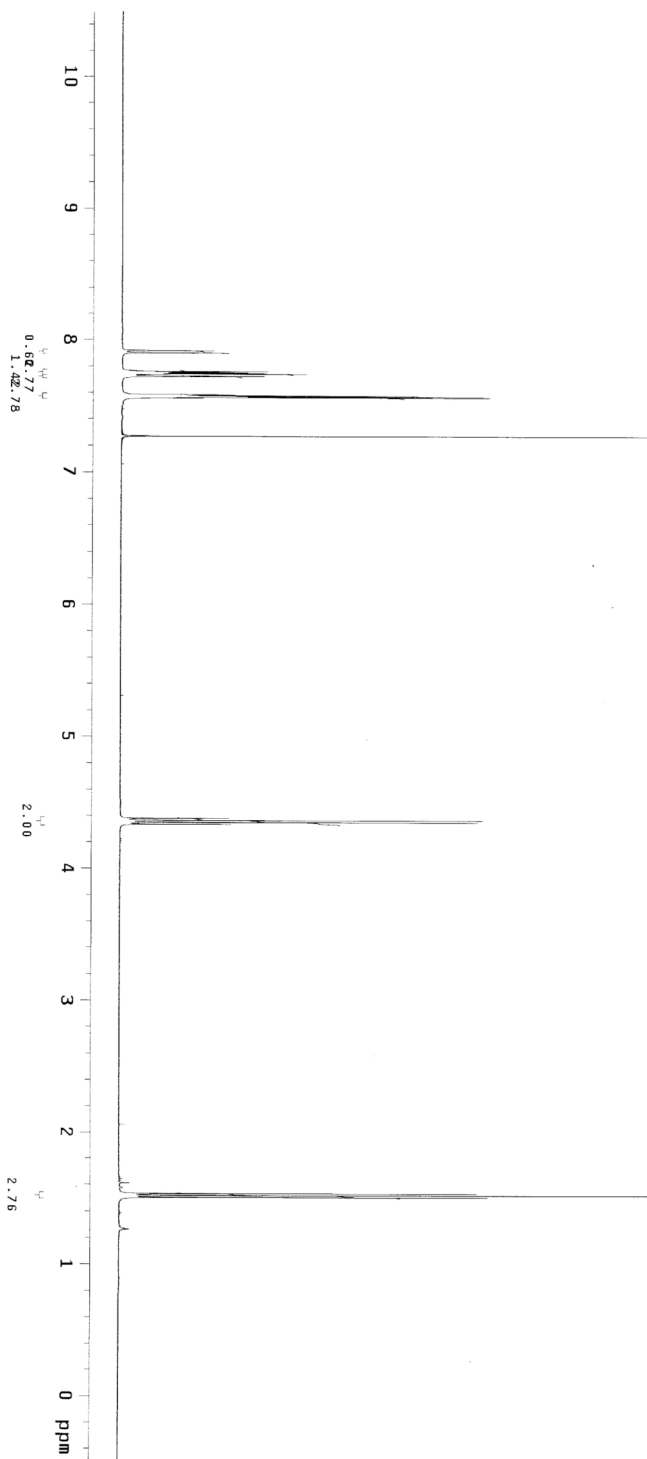
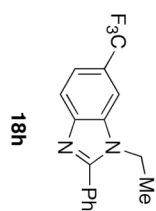
N2I1242  
Pulse Sequence: s2pul  
Solvent: CDCl3  
Ambient temperature  
User: J1148/  
Date: N2I1242 ProductCl3  
INNOVA-500 "z tppp"  
PULSE SEQUENCE  
Pulse delay 0.763 sec  
Pulse 65.4 degrees  
Acq. time 1.736 sec  
Width 37735.8 Hz  
1000 repetitions  
OBSERVE Cl3, 125.763366 MHz  
DECOUPLE H1, 500.2532753 MHz  
Power 157 dB  
continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 131072  
Total time 41 min, 50 sec



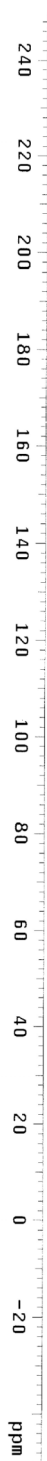
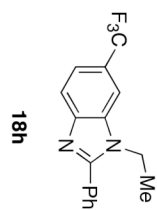
189



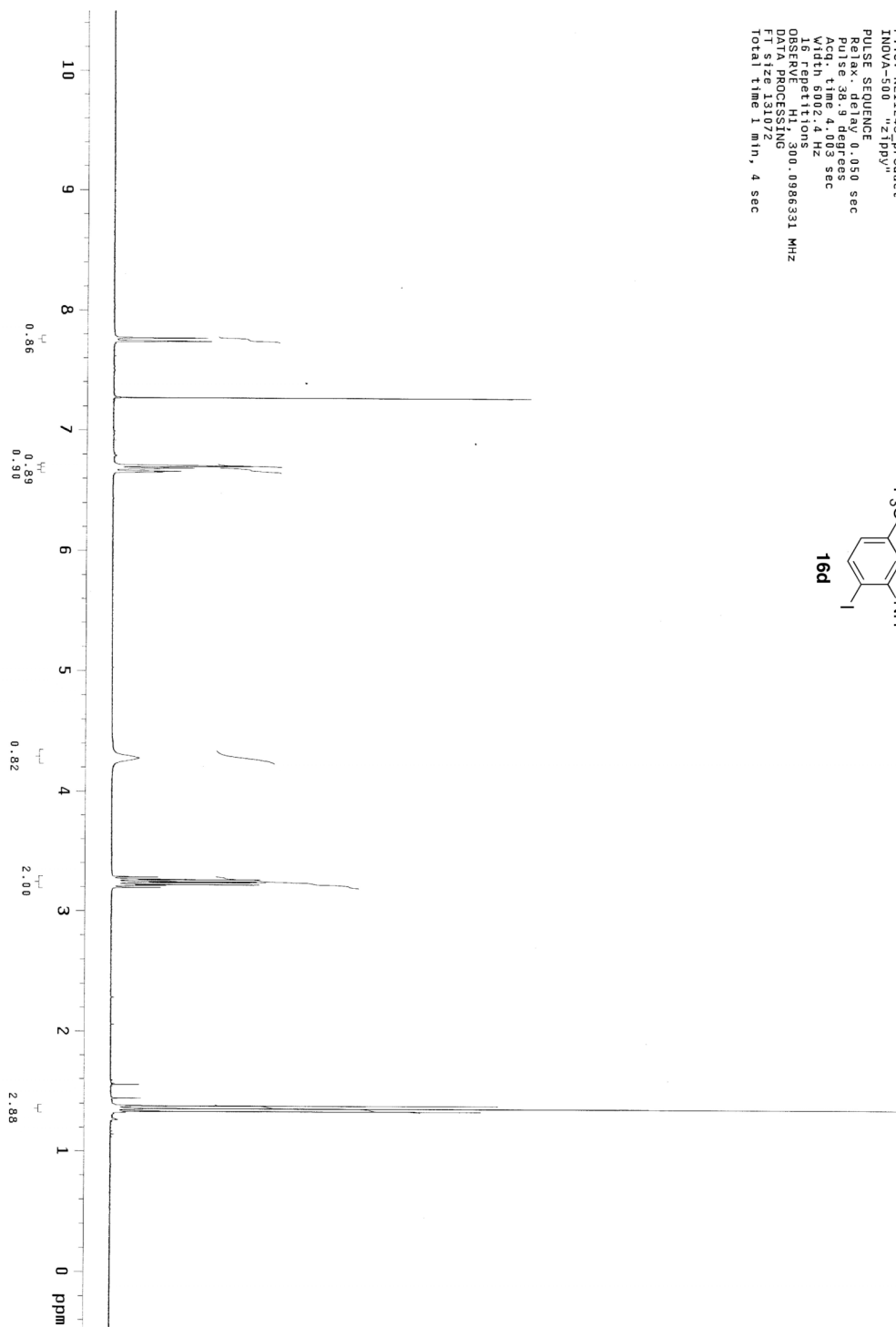
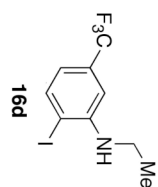
N2I1247  
Pulse Sequence: s2pu1  
Solvent: CDCl3  
Ambient Temperature  
File: N2I1247product  
INOVA-500 21ppv  
PULSE SEQUENCE  
Pulse 39.3 degrees  
Acq 129.3 degrees  
Width 1000.0 Hz  
16 repetitions  
OBSERVE H1, 500.2312702 MHz  
DATA PROCESSING  
FT size 131072  
Total time 0 min, 54 sec



NZ11247  
Pulse Sequence: szpu1  
Solvent: CDCl3  
Ambient temperature  
User: 11483  
File: NZ11247.product13  
INOVA-500 "zippy"  
PULSE SEQUENCE  
Relax. delay 0.763 sec  
Pulse 65.4 degrees  
Acq. time 1.736 sec  
Width 37735.8 Hz  
132 repetitions  
Observed Q13, 155.7832348 MHz  
DECOUPLE H1, 500.1332753 MHz  
Power 38 dB  
continuously on  
VALT2-16 modulated  
DATA PROCESSING  
Line broadening 0.3 Hz  
F1 size 32768  
Total time 696 hr, 41 min, 12 sec



NZ11243  
Pulse Sequence: szpu1  
Solvent: CDCl3  
Acquisition Date: 10/10/2001  
File: NZ11243product  
INOVA-500 "z1ppy"  
PULSE SEQUENCE  
Relax. delay: 0.050 sec  
Pulse: 38.9 degrees  
Acq. time: 4.003 sec  
Width: 6002.4 Hz  
16 repetitions  
Observed: 1300.0986331 MHz  
DATA PROCESSING  
FT size: 131072  
Total time: 1 min, 4 sec



N2I1243  
Pulse Sequence: s2pul  
Solvent: CDCl3  
Ambient temperature  
Date: 11-15-87  
File: N2I1243product13  
INOVA-500 "z1ppy"  
PULSE SEQUENCE  
Relax delay 0.763 sec  
Pulse 65.4 degrees  
Acq. time 1.736 sec  
Width 3735.8 Hz  
88 repetitions  
OBSERVE C13, 125.7032331 MHz  
PULSE C13, 500.2532753 MHz  
Power 37 dB  
continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 0.3 Hz  
F size 131072  
Total time 8 hr, 58 min, 3 sec

