Supporting Information

Synthesis and Evaluation of Vancomycin Aglycon

Analogues that Bear Modifications in the N-terminal

D-Leucyl Amino Acid

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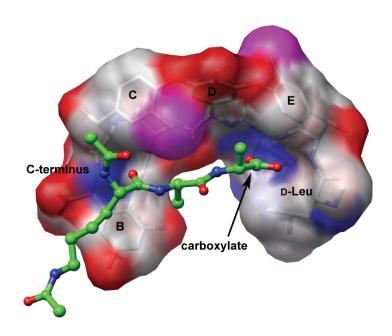


Figure 1SI. Surface representation of a single complex of the vancomycin aglycon (**5**) (surface area 767.16 Å²) co-crystallized with *N,N'*-Ac₂-L-Lys-D-Ala-D-Ala (**2**, PDB Code: 1FVM) as determined by X-ray crystallography to 1.8 Å-resolution.² The surface area at the *N*-terminal region of **5** is made up of the residue 3 asparagine and the residue 1 D-leucyl moieties, which generate a "concave" non-polar surface for carboxylate binding, thereby potentially reducing its desolvation penalty. Color code: ligand skeleton: green; vancomycin skeleton: grey; atoms: O: red; N: blue; H: white. Surface color code: by element (above). Figure and surface area calculations were generated with the molecular graphics program Chimera.³

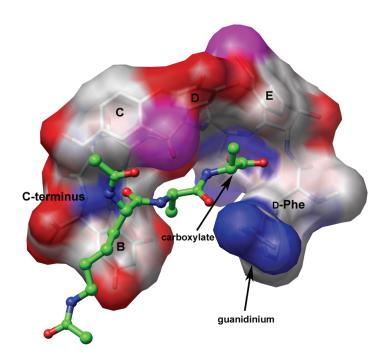
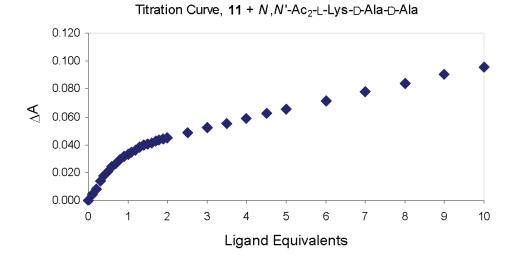


Figure 2SI. Surface representation of **11** (surface area 767.16 Å²) with bound *N,N'*-Ac₂-L-Lys-D-Ala-D-Ala (**2**) as determined by molecular modeling using the program MOLOC.¹ Molecular modeling was preformed using the X-ray crystal structure of **5** co-crystallized with **2** (PDB Code: 1FVM).² The *N*-terminal region of **11** is made up of the residue 3 asparagine and the residue 1 3-guanidyl-D-Phe. Unlike **5**, residues 3 and 1 do not interact to generate a "concave" shaped non-polar surface, rather the D-Phe has moved away from the aspargine, allowing the guanidine substituent to H-bond to the residue 2 carbonyl of **2**. Color code: ligand skeleton: green; vancomycin skeleton: grey; atoms: O: red; N: blue; H: white. Surface color code: by element (above). Figure and surface area calculations were generated with the molecular graphics program Chimera.³



Scatchard Analysis, 11 + N,N'-Ac₂-L-Lys-D-Ala-D-Ala (1.1-1.8 equiv.)

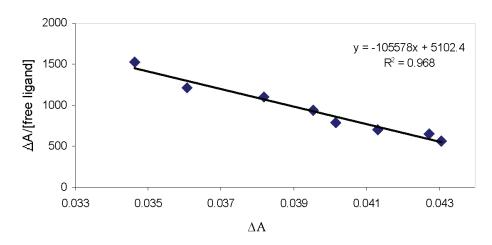
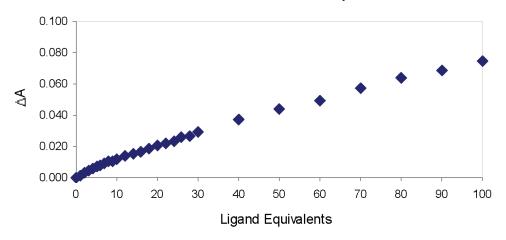


Figure 3SI. Representatice titration curve for the determination of the association constant (K_a) for the complex of **11** (1.1 × 10⁻⁴ M) + N_sN' -Ac₂-L-Lys-D-Ala-D-Ala (**2**) in 20 mM sodium citrate buffer, pH 5.1. (a) Saturation curve (0–10 equiv). (b) Scatchard analysis (1.1–1.8 equiv).

Titration Curve, 11 + N,N'-Ac₂-L-Lys-D-Ala-D-Lac



Scatchard Analysis, 11 + N,N'-Ac₂-L-Lys-D-Ala-D-Lac (5-18 equiv.)

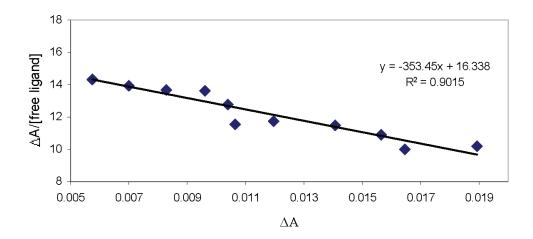


Figure 4SI. Representative titration curve for the determination of the association constant (K_a) for the complex of **11** (1.1 × 10⁻⁴ M) + N_sN' -Ac₂-L-Lys-D-Ala-D-Lac (**3**) in 20 mM sodium citrate buffer, pH 5.1. (a) Saturation curve (0–100 equiv). (b) Scatchard analysis (5–18 equiv).

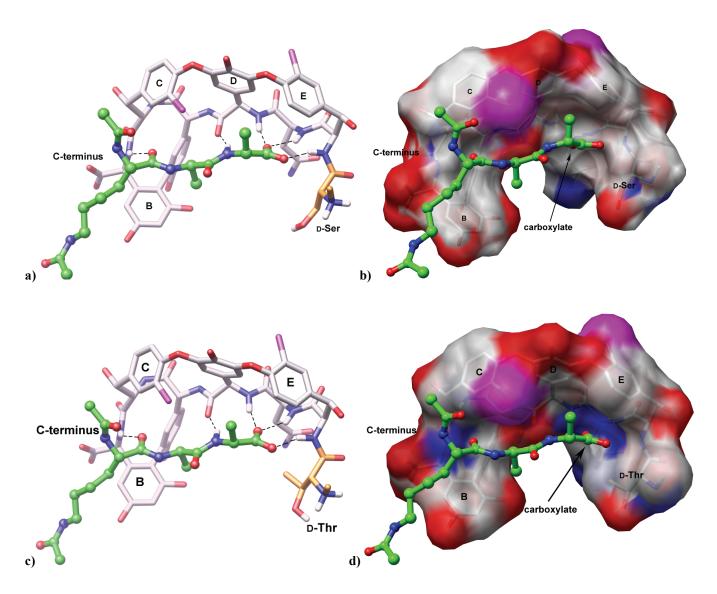


Figure 5SI. Proposed binding mode of target molecules **6** and **7** with ligand *N*,*N'*-Ac₂-L-Lys-D-Ala-D-Ala **(2)** as determined by molecular modeling using the program MOLOC.² Molecular modeling was preformed using the X-ray crystal structure of **5** co-crystallized with **2** (PDB Code: 1FVM).² The *N*-terminal region of **6** and **7** are made up of the residue 3 asparagine and the residue 1 D-Ser or D-Thr, respectively. Unlike **5**, residues 3 and 1 in the former do not generate a "concave" shaped non-polar surface (b), rather the additional methyl substituent of the latter compensates (d), thereby potentially reducing the desolvation penalty for carboxylate binding. (a) Ball and stick representation of **6**. (b) Surface representation of **6** (surface area 763.33 Å²) with bound **2**. (c) Ball and stick representation **7**. (d) Surface representation of **7** (surface area 766.22 Å²) with bound **3**. Black dashed lines represent

potential H-bonds. Color code: ligand skeleton: green; vancomycin skeleton: grey/orange; Cl: pink; amino acid skeleton: O: red; N: blue; H: white. Surface color code: by element above. All figures and surface area calculations were generated with the molecular graphics program Chimera.³

Table 1SI. Selected conditions examined for the reaction between hydrazine derivatives **13–15** and N,N'-Ac₂-L-Lys-D-Ala-D-Lac (3).

Entry	Compound	Ligand	[Compound]	Solvent	Temp (°C)	Time (h)	result
		(equiv)					
1	13	20	0.13 mM	H ₂ O	20	18	No product
2	13	20	0.13 mM	A^b	37	18	No product
3	13	20	0.13 mM	H ₂ O	20	18	No product
4	13	20	0.13 mM	A^b	37	18	No product
5	13	2	7 mM	МеОН	20	18	No product
6	13	2	6 mM	EtOH	20	72	No product
7	13	2	6 mM	EtOH	65	48	No product
8	13+NaHCO ₃ (2 equiv)	2	7 mM	EtOH	20	20	No product
9	13 +Et ₃ N (1 equiv)	1	0.02 м	EtOH/EtSH (99:1)	0-75	18	No product
10	13 +Et ₃ N (10 equiv)	1	0.02 м	EtOH/EtSH (99:1)	0-75	18	No product
11	14 +Et ₃ N(8 equiv)	0.8	0.07 м	H ₂ O	20-	24	No product
					75	48	
12	15	0.8	0.07 м	H ₂ O	20-	24	No product
					75	48	
13	14	0.9	0.06 м	H ₂ O	20	336	[14+3]
							complex
14	14 +NaHCO ₃ (10 equiv)	1.1	0.06 м	H ₂ O	20	72	No product
15	12 +NaHCO ₃ (20 equiv)	1.0	16 mM	<i>i</i> PrOH	20	72	No product
16	12 +NaHCO ₃ (20 equiv)	1.0	16 mM	МеОН	20	72	No product
17	12 +NaHCO ₃ (20 equiv)	1.0	16 mM	H ₂ O	20-	72-	[12+3]
					75	72h	complex

^a Crude reactions were monitored by LC–MS. ^b20 mM sodium citrate buffer (pH 5.1)

Experimental Section

General. Vancomycin aglycon (1) and desleucylvancomycin aglycon (16) were prepared according to literature procedures. 4,5 Reagents and solvents were purchased reagent-grade and used without further purification. Amino acid building blocks Boc-D-Thr and Boc-D-HoSer(OBn) were purchased from from Chem-Impex International, N-Boc-D-O-tert-butylserine from Novabiochem, and Boc-D-3-CN-Phe from Peptech. For the antimicrobial assays: Muller Hinton broth and BactoTM Brain heart infusion were purchased from Becton, Dickinson and Company. For the UV-difference titration assays: the ligands 2 and 3 were purchased from Bachem; and the buffer 0.2 mM sodium citrate (100 mL) was prepared from citric acid (0.131 g) and sodium citrate tribasic (0.388 g) and the pH was adjusted to 5.1 with 5 N NaOH. THF was freshly distilled from sodium benzophenone ketyl. All reactions were performed in oven-dried glassware under an Argon atmosphere, unless otherwise stated. Evaporation and concentration in vacuo was performed at 20 °C and ca. 760 m Hg. Further drying of the new compounds was carried out at ca. 5 millibar. TLC: pre-coated SiO₂ 60 F₂₅₄ glass plates from EMD, visualization by UV light (254 or 366 nm) or by staining with a solution of ninhydrin or bromocresol green. Column chromatography (FC): SiO₂ 60 (0.04-0.063 mm) from Fluka with a head pressure of ca. 0.2 bar (FC). Analytical and Preparative reverse-phase HPLC was performed using a Waters HPLC on a Waters Nova-Pac® HR C18 6 µm, 60 Å column (25 x 100 mm) using a gradient of MeCN/water with 0.07% TFA at flow rate of 5 mL min⁻¹ and with UV detection at $\lambda = 254$ and 280 nm. Uncorrected melting points (M.p.) were determined in an open capillary using a Mel-Temp II apparatus. NMR (¹H or ¹³C): Varian Inova-400, Bruker DRX-500 and Bruker DRX-600 NMR spectrophotometers at 298 K. Residual solvent peaks were used as an internal reference. Coupling constants (J) (H,H) are given in Hz. Coupling patterns are designated as singlet (s), doublet (d), triplet (t), quadruplet (q), quintuplet (qt), multiplet (m), or broad signal (br). IR spectra: Thermo Scientific Nicolet 380 FT-IR spectrophotometer measured neat. High resolution mass spectral data were acquired on an Agilent Technologies High Resolution LC/MSD-TOF and the detected masses are given as m/z with M representing the molecular ion.

In vitro Assays. One day before experiments were run fresh cultures of vancomycin-sensitive Staphlococcus aureus (strain ATCC 25923) in Mueller-Hinton broth and vancomycin-resistant Enterococcus faecalis (VanA, BM4166) in Brain-Heart Infusion were inoculated and warmed in an orbital shaker at 37 °C. After 24 h, the bacterial stock solutions were serial diluted with the culture medium (Muller–Hinton broth for S. aureus and Brain-Heart Infusion for E. faecalis-VanA) to achieve the turbidity equivalent of 1:100 dilution of 0.5 M Macfarland solution. This diluted bacterial stock solution was then inoculated into a well of a V-shaped 96-well microtiter plate, supplemented with serial diluted aliquots of the antibiotic solution in water or DMSO (1–5 μ L), to achieve a total assay volume of 0.1 mL. The plate was then incubated at 37 °C for 24 h, minimal inhibitory concentrations (MICs) were determined by monitoring the cell growth (observed as a pellet) in the wells. The lowest concentration of antibiotic (in μ g/mL) capable of eliminating the cell growth, in the wells, is reported as the MIC. The reported MIC values for the new antibiotics were determined against vancomycin or vancomycin aglycon as a standard in the first well, which have well-established MIC values.^{6,7}

UV-difference assay for the determination of association constants for the complexes of the natural product analogues and 2 and 3. The association constants (K_a) of the complexes between compounds 6–15 and the model ligands N,N'-Ac₂-L-Lys-D-Ala-D-Ala (2) and N,N'-Ac₂-L-Lys-D-Ala-D-Lac (3) were determined according to literature procedures. UV-difference experiments were carried out on a CARY 3E UV/Vis spectrometer in 1.5 mL quartz cuvettes (path length = 1 cm), and measured from $\lambda = 200$ to 345 nm. The UV absorbance scans were run after a baseline correction (0.02 M sodium citrate buffer) from $\lambda = 200$ to 345 nm. The initial absorbance (A_o) of a 1 mL solution of the antibiotic (1.1 × 10⁻⁴ M in 0.02 M sodium citrate buffer) was recorded ($\lambda = 200$ to 345 nm) with respect to reference cell containing only 1 mL of 0.02 M sodium citrate buffer. UV spectra were recorded after the

addition of 1 μ L of either **2** (0.011 M, 0.1–10 equivalents) or **3** (0.11 M, 1–100 equivalents) in 0.02 M sodium citrate buffer to each cell. The absorbance value at the λ_{max} (279 mm) was recorded and the running change in absorbance ΔA , at x-equivalents (A_x) of ligand, where $\Delta A = (A_o - A_x \text{ equiv})$, was measured. The number of ligand equivalents was plotted versus ΔA to provide a ligand saturation curve for each complex. The break point of this curve is the saturation point ($\Delta A_{\text{saturation}}$) of the system and its x,y-coordinates were determined by establishing the point intersection of the linear functions for the preand post-saturation parts of the titration curve. The concentration of free ligand ([free ligand]) in solution at each titration step was then calculated from $\Delta A_{\text{saturation}}$. A Scatchard plot of ΔA at x-equivalents versus ΔA /[free ligand] gave a linear function, the slope of which provides the association constant (K_a).

General Procedures (GP)

GP1 for the selective *N*-methylation of amino acids. NaH (3.3 equiv) was added portionwise to a stirred solution of the corresponding amino acid (1.0 equiv) in anhydrous THF (0.3 M) at 0 °C. After stirring for 10 min, MeI (3.0 equiv) and DMF (10% v/v) were added, and the mixture was allowed to stir for an additional hour at 0 °C and was then warmed to 20 °C. After 18 h, the reaction was quenched by the addition of water and extracted with EtOAc. The aqueous phase was then acidified to pH of 2–3 with aqueous 0.2 N NaHSO₄ and extracted with EtOAc, which was washed with aqueous 5% NaS₂O₄, and dried over Na₂SO₄ (see the experimental details).

GP2 for ester hydrolysis. An aqueous solution of 0.2 N LiOH (1.0–3.0 equiv) was added portionwise to a solution of the corresponding methyl ester (1.0 equiv) in THF (0.35 M) at 0 °C. After 2–4 h at 0 °C, the solution was acidified to a pH of 2–3 with aqueous 0.2 N NaHSO₄ and extracted with EtOAc (3 ×), and the combined organics layers were dried over Na₂SO₄, filtered and concentrated in vacuo.

GP3 for DCC mediated anhydride coupling. A solution of dicyclohexylcarbodiimide (2.0 equiv) in anhydrous CH_2Cl_2 was added to a solution of the corresponding amino acid (4.0 equiv) in anhydrous CH_2Cl_2 (0.14 M) at 0 °C. After 1 h at 0 °C, the volatiles were removed with a gentle stream on N_2 . The residue was re-dissolved in anhydrous DMF (0.15 M) and added dropwise to a solution of desleucylaglucovancomycin ($16^{4.5}$, 1.0 equiv) and NaHCO₃ (4.0 equiv) in anhydrous DMF (0.04 M) at 0 °C. After 1 h at 0 °C, the mixture was warmed to 20 °C. After 18 h, the volatiles were removed with a gentle stream of N_2 gas and the crude residue was purified by reverse-phase HPLC using a gradient of MeCN/water with 0.07% TFA. The fractions that contained product were combined and lyophilized. The resulting protected intermediates were isolated and directly carried to the next step without full characterization. The intermediates were treated with 10-50% TFA/CH₂Cl₂ at 0-20 °C for 5–40 min, the volatiles were removed with a gentle stream of N_2 and purified by reverse-phase HPLC (see the experimental details).

(R)-3-tert-Butoxy-2-(tert-butoxycarbonylamino)propanoic Acid (17)

Aqueous NaOH (3.6 mL, 0.47 mmol) followed by di-*tert*-butyl dicarbonate (390 μ L, 1.70 mmol) were added to a solution of H-D-Ser(tBu)-OH (250 mg, 1.55 mmol) in dioxane (3.6 mL) at 0 °C. After 1 h, the solution was warmed to 25 °C. After 24 h, the mixture was acidified to pH of 2–3 with aqueous 0.2 N NaHSO₄ and extracted with EtOAc (2 × 100 mL), which was dried over Na₂SO₄, filtered and concentrated. Column chromatography (26 g SiO₂, 0–10% MeOH/EtOAc) afforded **17** as a pale yellow oil (195 mg, theoretical 0.406 g, 48%): ¹H NMR (CDCl₃, 600 MHz) δ 8.08 (br s, 1H), 5.36 (br s, 1H),

4.39 (br s, 1H), 3.87 (d, 1H, J = 4.2 Hz), 3.56 (dd, 1H, J = 4.2, 8.4 Hz), 1.46 (s, 9H), 1.18 (s, 9H); ¹³C NMR (CDCl₃, 150 MHz) δ 174.3, 155.8, 80.2, 74.2, 61.7, 53.8, 28.3 (3C), 27.3 (3C); IR (neat) ν_{max} 2976m, 2361w, 1719s, 1506m, 1393m, 1366m, 1165s, 1103w, 1068w cm⁻¹; HR ESI-TOF m/z 284.1473 (M + Na⁺, C₁₂H₂₃NNaO₅⁺, requires 284.1468); $[\alpha]^{25}_{D}$ –21 (c 0.5, CH₂Cl₂).

(R)-3-tert-Butoxy-2-(tert-butoxycarbonyl(methyl)amino)propanoic Acid (19)

The title compound was prepared according to GPI with 17 (150 mg, 0.576 mmol), NaH (76 mg, 1.9 mmol), MeI (107 μ L, 1.73 mmol), THF (1.92 mL), and DMF (97 μ L). Column chromatography (10 g SiO₂, 10% MeOH/CH₂Cl₂) provided 19 (143 mg, theoretical 157 mg, 92%) as a pale yellow oil (rotomer ratio in CDCl₃ A/B = 10:9): ¹H NMR (CDCl₃, 600 MHz) δ 8.96 (br s, 2H), 4.65 (t, 1H_A, J = 5.4 Hz), 4.35 (t, 1H_B, J = 6.0 Hz), 3.83–3.71 (m, 4H), 2.95 (s, 3H_B), 2.92 (s, 3H_A), 1.46 (s, 9H_A), 1.43 (s, 9H_B), 1.19 (s, 18H); ¹³C NMR (CDCl₃, 150 MHz) δ 175.1/174.6, 157.2/156.1, 81.6/81.4, 75.1, 61.4/61.2, 61.5/60.1, 34.6, 29.20/29.16 (3C), 28.2 (3C); IR (neat) ν _{max} 2973m, 2928m, 1693s, 1452m, 1390m, 1366m, 1327w, 1148s, 1088m, 868m cm⁻¹; HR ESI-TOF m/z 276.1799 (M + H⁺, C₁₃H₂₆NO₅⁺, requires 276.1805); α ²⁵_D –20 (α 0.5, CH₂Cl₂).

1-(D-Serinyl)desleucylaglucovancomycin Trifluoroacetate (6) and 1-(*O-tert*-Butyl- D-serine)desleucylaglucovancomycin Trifluoroacetate (7)

The title compounds were prepared starting from 19 (7.80 mg, 0.028 mmol), and dicyclohexyl carbodiimide (2.92 mg, 0.014 mmol) in CH₂Cl₂ (0.2 mL), followed by treatment with NaHCO₃ (2.38 mg, 0.028 mmol) and desleucylaglucovancomycin (16, 8.00 mg, 0.007 mmol) in DMF (0.4 mL) according to GP3. The protected intermediate was isolated as a white solid (6.60 mg, 73%) by reversephase HPLC using a gradient of 35–95 % MeCN/water with 0.07% TFA ($t_R = 12.4$ min), and lyophilized, and taken directly to the next step without further characterization. The intermediate (6.60) mg, 0.0051 mmol) was taken up in 50% TFA in CH₂Cl₂ (0.2 mL) and stirred at 20 °C. After 20 min, the volatiles were removed with a gentle stream of N₂ and residual TFA was azetroped with CH₂Cl₂ (3 × 5 mL). The residue was purified reverse-phase HPLC using a gradient of 5-40% MeCN/water with 0.07% TFA to provide 6 (34%) and 7 (65%). Characterization of the TFA salt of 6 (R = H, t_R = 23.8 min, 2.20 mg, 34%): white powder; ¹H NMR (CD₃OD, 600 MHz) δ 7.72 (br s, 2H), 7.63 (s, 1H), 7.58 (d, 1H, J = 8.4 Hz), 7.47 (br s, 1H), 7.19 (d, 1H, J = 8.4 Hz), 7.01 (s, 1H), 6.71 (br s, 2H), 6.52 (s, 1H),6.40 (d, 1H, J = 2.4 Hz), 5.95 (br s, 1H), 5.83 (br s, 1H), 5.38–5.32 (m, 4H), 4.70 (s, 1H), 4.64 (br s, 1H), 4.34 (br s, 1H), 4.11 (s, 1H), 3.98 (t, 1H, J = 5.4 Hz), 3.83 (br s, 2H), 2.89 (d, 1H, J = 15.0 Hz), 2.79 (s, 3H), 2.16 (br s, 1H); HR ESI-TOF m/z 1117.2342 (M + H⁺, C₅₀H₄₇Cl₂N₈O₁₈⁺, requires 1117.2380); $[\alpha]^{25}_{D}$ +33 (c 0.2, CH₃OH). Characterization for the TFA salt of 7 (R = tBu, t_R = 26.9 min, 4.40 mg, 65%): white powder; ¹H NMR (CD₃OD, 600 MHz) δ 7.82 (br s, 2H), 7.73 (br s, 1H), 7.63 (s,

1H), 7.57 (br s, 1H), 7.57 (br s, 1H), 7.55 (d, 1H, J = 8.4 Hz), 7.17 (d, 1H, J = 8.4 Hz), 7.04 (s, 1H), 6.67–6.64 (m, 2H), 6.44 (s, 1H), 6.40 (s, 1H), 6.06 (br s, 1H), 5.94 (br s, 1H), 5.37 (s, 1H), 5.33 (s, 1H), 5.27 (s, 1H), 4.75 (br s, 1H), 7.74 (s, 1H), 4.26 (d, 1H, J = 9.0 Hz), 4.15 (s, 1H), 4.10 (t, 1H, J = 6.6 Hz), 3.73 (d, 2H, J = 7.8 Hz), 2.95 (d, 1H, J = 15.6 Hz), 2.83 (s, 3H), 1.99 (br s, 1H), 1.2 (s, 9H); HR ESITOF m/z 1173.2975 (M + H⁺, C₅₄H₅₅Cl₂N₈O₁₈⁺, requires 1173.3006); $[\alpha]^{25}_{D}$ +49 (c 0.2, CH₃OH).

(2R,3S)-2-(tert-Butoxycarbonylamino)-3-(tert-butyldimethylsilyloxy)butanoic Acid (18)

A chilled solution of Boc-D-Thr (521 mg, 2.38 mmol) in anhydrous DMF (4.7 mL) at 0 °C was treated with *tert*-butyldimethylsilyl chloride (466 mg, 3.09 mmol) followed by imidazole (486 mg, 7.14 mmol), and the mixture was stirred for 1 h at 0 °C, and then warmed to 20 °C. After 18 h, the mixture was partitioned between aqueous 1 N HCl and Et₂O, and aqueous phase was washed Et₂O (2 × 40 mL). The combined organics layers were dried over Na₂SO₄, filtered, and the volatiles were removed in vacuo. Column chromatography (50 g SiO₂, 60% EtOAc/hexanes) provided **18** (636 mg, theoretical 648 mg, 98%) as a colorless oil: 1 H NMR (CDCl₃, 600 MHz) δ 7.71 (br s, 1H), 5.23 (d, J = 9.0 Hz, 1H), 4.44 (dd, J = 2.4, 6.6 Hz, 1H), 4.25 (dd, J = 2.4, 9.0 Hz, 1H), 1.46 (s, 9H), 1.19 (d, J = 6.0 Hz, 3H), 0.85 (s, 9H), 0.073 (s, 3H), 0.05 (s, 3H); 13 C NMR (CDCl₃, 150 MHz) δ 175.3, 156.1, 80.1, 68.7, 59.0, 28.3, 25.6, 20.0, 17.8, -4.6, -5.2; IR (neat) ν_{max} 3454 ω , 2954 ω , 1721 ω , 1694 ω , 1510 ω , 1366 ω , 1311 ω , 1253 ω , 1163 ω , 1129 ω , 1100 ω , 1040 ω , 833 ω , 773 ω cm⁻¹; HR ESI-TOF ω /z 356.1866 (M + Na⁺, C₁₅H₃₁NNaO₅Si⁺, requires 356.1864); [α]²⁵D -14 (ω 0.5, CH₂Cl₂).

(2R,3S)-2-(tert-Butoxycarbonyl(methyl)amino)-3-(tert-butyldimethylsilyloxy)butanoic Acid (20)

The title compound was prepared according to *GP1* from **18** (366 mg, 1.09 mmol), NaH (145 mg, 3.63 mmol), MeI (206 μ L, 3.32 mmol), THF (3.6 mL), and DMF (175 μ L). Column chromatography (30 g, SiO₂, 0–50% EtOAc/hexanes) provided **20** (231 mg, theoretical 381 mg, 60%) as an off-white crystalline solid (rotomer ratio in CDCl₃ A/B = 2:1): mp 87 °C (decomp.); ¹H NMR (CDCl₃, 600 MHz) δ 8.41 (br s, 2H), 4.75 (br s, 2H), 4.63–4.52 (m, 1H), 3.02 (s, 3H_B), 3.01 (s, 3H_A), 1.48 (s, 9H_A), 1.44 (s, 9H_B), 1.20 (s, 3H_A), 1.9 (s, 3H_B), 0.85 (s, 18H), 0.08 (s, 6H), 0.05 (s, 6H); ¹³C NMR (CDCl₃, 150 MHz) δ 175.4/175.3, 157.1/155.7, 80.5/80.4, 68.9/68.8, 64.1, 34.1/33.2, 28.3 (3C), 25.6 (3C), 20.7/20.4, 17.8, -4.43/-4.46/-5.26 (2C); IR (neat) ν_{max} 2956m, 2932m, 1695s, 1475w, 1369m, 1315m, 1255m, 1151s, 1069m, 940m, 835m, 776m cm⁻¹; HR ESI-TOF m/z 348.2206 (M + H⁺, C₁₆H₃₄NO₅Si⁺, requires 348.2201); $[\alpha]^{25}$ _D -20 (c 0.5, CH₂Cl₂).

1-(D-Threonyl)desleucylaglucovancomycin Trifluoroacetate (8)

The title compound was prepared from 20 (5.30 mg, 0.014 mmol), and dicyclohexylcarbodiimide (1.46 mg, 0.007 mmol) in CH₂Cl₂ (0.1 mL), followed by treatment with NaHCO₃ (1.19 mg, 0.015 mmol), desleucylaglucovancomycin (16, 4.00 mg, 0.004 mmol) in DMF (0.2 mL) according to GP3. The protected intermediate was isolated as a white solid (3.30 mg, theoretical 4.70 mg, 70%) by reversephase HPLC using a gradient of 35–95 % MeCN/water with 0.07% TFA ($t_R = 20.0$ min), and lyophilized, and taken directly to the next step without further characterization. The intermediate (3.3) mg, 0.0025 mmol) was taken up in 50% TFA in CH₂Cl₂ (0.2 mL) and stirred at 20 °C. After 10 min, the volatiles were removed with a gentle stream of N_2 and residual TFA was azetroped with CH_2Cl_2 (3 × 5 mL). The residue was purified by reverse-phase HPLC using a gradient of 5-40% MeCN/water with 0.07% TFA ($t_R = 24.4$ min) and afforded the TFA salt of 7 (3.3 mg, theoretical 4.3 mg, 76%) as white powder after lyophilization: ¹H NMR (CD₃OD, 600 MHz) δ 8.94 (d, 1H, NH, J = 6.0 Hz), 8.78 (d, 1H, NH, J = 5.4 Hz), 7.75 (br s, 1H), 7.71 (br s, 1H), 7.66 (s, 1H), 7.57 (d, 2H, J = 8.4 Hz), 7.19 (d, 1H, J =8.4 Hz), 7.04 (s, 1H), 6.67 (br s, 2H), 6.42 (dd, 2H, J = 1.8, 14.4 Hz), 6.02 (br s, 1H), 5.91 (br s, 1H), 5.35 (s, 1H), 5.33 (s, 1H), 5.27 (d, 1H, J = 1.8 Hz), 4.75–4.74 (m, 2H), 4.28 (d, 1H, J = 8.4 Hz), 4.22 (br s, 1H), 4.14 (s, 1H), 3.88 (d, 1H, J = 6.0 Hz), 2.94 (d, 1H, J = 15.6 Hz), 2.79 (s, 3H), 2.05 (br s, 1H), 1.06 (br s, 3H) (two signals buried under residual water); HR ESI-TOF m/z 1131.2484 (M + H⁺, $C_{51}H_{49}Cl_2N_8O_{18}^+$, requires 1131.2536); $[\alpha]_D^{25} + 47$ (c 0.2, CD₃OD).

(R)-2-(tert-Butoxycarbonyl(methyl)amino)-3-(3-nitrophenyl)propanoic Acid (22)

The title compound was prepared starting from commercially available Boc-D-3-nitrophenylalanine (**21**, 0.50 g, 1.62 mmol), NaH (0.214 g, 5.32 mmol), MeI (0.30 mL, 4.84 mmol), THF (5.4 mL), and DMF (0.26 mL) according to *GP1*. Column chromatography (50 g SiO₂, 10% MeOH/CH₂Cl₂) afforded **22** (423 mg, theoretical 518 mg, 82%) as a peach powder (rotomer ratio in CDCl₃ A/B = 3:4): ¹H NMR (CDCl₃, 500 MHz) δ 10.07 (br s, 2H), 8.08 (d, 4H, J = 11.0 Hz), 7.59–7.47 (m, 4H), 4.90 (dd, 1H_B, J = 5.0, 11.0 Hz), 4.70 (dd, 1H_B, J = 4.0, 10.0 Hz), 3.45–3.41 (m, 2H), 3.24–3.13 (m, 2H), 2.76 (s, 3H_A), 2.73 (s, 3H_B), 1.36 (s, 9H_B), 1.34 (s, 9H_A); ¹³C NMR (CDCl₃, 125 MHz) δ 175.3/175.2, 156.0/154.9, 148.4/148.2, 139.5/139.4, 135.3/135.2, 129.5/129.4, 123.9/123.7, 121.9/121.8, 81.8/81.0, 60.8/59.8, 34.9/34.3, 32.7/32.5, 28.1 (3C); IR (neat) ν_{max} 2977 ν_{max} 1688 ν_{m} , 1528 ν_{m} , 1480 ν_{m} , 1449 ν_{m} , 1350 ν_{m} , 1264 ν_{m} , 1144 ν_{m} , 732 ν_{m} cm⁻¹; HR ESI-TOF ν_{m} 347.1219 (M + Na⁺, C₁₅H₂₀N₂NaO₆⁺, requires 347.1213); [α] 25 D +56 (ν_{m}) 47.1219.

(R)-Methyl 2-(tert-Butoxycarbonyl(methyl)amino)-3-(3-nitrophenyl)propanoate (23)

Crushed and vacuum-oven dried KHCO₃ (260 mg, 2.74 mmol) followed by MeI (112 μ L, 1.83 mmol) was added to a solution of **22** (423 mg, 1.30 mmol) in anhydrous DMF (4.3 mL) at 0 °C. After the addition was complete, the mixture was allowed to warm to 25 °C. After 12 h, the mixture was acidified to a pH of 2–3 with aqueous 1 M NaHSO₄ and extracted with EtOAc (3 × 100 mL). The combined organic layers were washed with water (20 mL), saturated aqueous NaHCO₃ (20 mL), water (20 mL), then dried over MgSO₄, filtered, and concentrated in vacuo. Column chromatography (50 g SiO₂, 20% EtOAc/hexanes) afforded **23** (416 mg, theoretical 442 mg, 94%) as a yellow oil (rotomer ratio in CDCl₃ A/B = 1:1): ¹H NMR (CDCl₃, 500 MHz) δ 8.05 (d, 4H, J = 12.5 Hz), 7.56–7.42 (m, 4H), 4.87 (dd, 1H_A, J = 5.0, 10.5 Hz), 4.59 (dd, 1H_B, J = 4.5, 10.0 Hz), 3.74 (s, 3H_B), 3.73 (s, 3H_A), 3.38 (dd, 2H, J = 5.0, 14.0 Hz), 3.16–3.09 (m, 2H), 2.70 (s, 6H), 1.33 (s, 9H_A), 1.32 (s, 9H_B); ¹³C NMR (CDCl₃, 125 MHz) δ 171.0/170.8, 155.63/155.61, 148.3/148.2, 139.8/139.6, 135.3/135.2, 129.4/129.2, 123.9/123.7, 121.8/121.7, 80.6/80.3, 60.9/59.4, 52.3, 35.1/34.5, 32.4/32.2, 28.1 (3C); IR (neat) ν _{max} 2976 ν ₂, 1741 ν ₂, 1690 ν ₃, 1527 ν ₃, 1479 ν ₃, 1436 ν ₃, 1348 ν ₃, 1224 ν ₃, 1141 ν ₃, 734 ν ₃ cm⁻¹; HR ESI-TOF ν ₂ 361.1375 (M + Na⁺, C₁₆H₂₂N₂NaO₆⁺, requires 361.1370); [α |²⁵D +56 (ν ₂0.5, CHCl₃).

(R)-Methyl 3-(3-Aminophenyl)-2-(tert-butoxycarbonyl(methyl)amino)propanoate (20)

A vacuum-degassed and N₂ flushed solution of **23** (400 mg, 1.18 mmol) in MeOH (11.8 mL) was treated with 10% Pd/C (0.1% wt., 0.040 g), and the mixture was again vacuum-degassed and flushed with N₂ (3 ×). The mixture was then placed under an atmosphere of H₂ (balloon) at 20 °C. After 13 h, the solution was filtered through a short plug of Celite and the volatiles were removed in vacuo. Column chromatography (50 g SiO₂, 50% EtOAc/hexanes) afforded **20** (358 mg, theoretical 365 mg, 98%) as a dark brown oil (rotomer ratio in CDCl₃ A/B = 3:4): ¹H NMR (CDCl₃, 500 MHz) δ 7.06–7.04 (m, 2H), 6.60–6.48 (m, 6H), 4.90 (dd, 1H_A, J = 4.5, 9.5 Hz), 4.49 (dd, 1H_B, J = 4.0, 10.0 Hz), 3.73 (s, 3H_B), 3.71 (s, 3H_A), 3.62 (br s, 4H), 3.21–3.15 (m, 2H), 2.93–2.88 (m, 2H), 2.73 (s, 3H_B), 2.71 (s, 3H_A), 1.39 (s, 9H_A), 1.35 (s, 9H_B); ¹³C NMR (CDCl₃, 150MHz) δ 171.9/171.6, 155.8/154.9, 146.5/146.4, 138.8/138.4, 129.3/129.1, 119.1, 115.6/115.5, 113.33/113.27, 80.1/79.9, 61.6/59.3, 52.1, 35.5/34.9, 32.7/31.9, 28.2/28.1 (3C); IR (neat) n_{max} 3459w, 3366w, 1738m, 1684s, 1605m, 1435m, 1392m, 1329m, 1256m, 1221m, 1142s, 1030w, 774m cm⁻¹; HR ESI-TOF m/z 309.1807 (M + H⁺, C₁₆H₂₅N₂O₄⁺, requires 309.1809); [α]²⁵_D +62 (c 0.5, CH₂Cl₂).

(R)-2-(tert-Butoxycarbonyl(methyl)amino)-3-(3-(tert-butoxycarbonylamino)phenyl)propanoic Acid (24)

The title compound was prepared in two steps of which the first step was conducted according to GP2 from 20 (100 mg, 0.32 mmol), aqueous 0.2 N LiOH (3.24 mL, 0.648 mmol), in THF (4.62 mL) at 0 °C Column chromatography (10 g SiO₂, 0–5% MeOH/EtOAc) provided the intermediate carboxylic acid (58.4 mg, 62%) as a brown oil, which was carried to the next step without full characterization. Boc₂O (50 uL, 0.22 mmol) was added to a solution of the carboxylate acid (58.4 mg. 0.198 mmol) in 1:1 THF/water (0.5 mL) at 20 °C. After 18 h, the reaction mixture was acidified to a pH of 2–3 with aqueous 3 N HCl and the aqueous phase was extracted with EtOAc (3 \times 50 mL). The combined organic layers were dried (Na₂SO₄), filtered, and the volatiles were removed in vacuo. Column chromatography (7g SiO₂, 2–10% MeOH/CH₂Cl₂) afforded **24** (58.8 mg, theoretical 78.3 mg, 75%) as a yellow oil (rotomer ratio in CDCl₃ A/B = 1:1): 1 H NMR (CDCl₃, 600 MHz) δ 7.31–7.15 (m, 8H), 6.88 (d, 1H, J = 6.6 Hz), 6.85 (d, 1H, J = 6.6 Hz), 6.60 (s, 2H), 4.82 (d, 1H, J = 5.4 Hz), 4.63 (d, 1H, J = 7.8 Hz), 3.3–3.28 (m, 2H), 3.07 (t, 1H, J = 12.0 Hz), 2.99 (t, 1H, J = 12.0 Hz), 2.76 (s, 3H), 2.70 (s, 3H), 1.51 (s, 18H), 1.40 (s, 9H), 1,35 (s, 9H); ¹³C NMR (CDCl₃, 150 MHz) δ 175.6/175.2, 156.4/155.1, 152.8/152.6 (br), 138.52/138.48, 138.0, 129.1/129.0, 123.7/123.6, 119.0 (br), 116.9 (br), 80.7 (2C), 61.3/60.5, 35.4/34.7, 32.9/32.6, 28.3 (3C), 28.2/28.1 (3C); IR (neat) v_{max} 2980w, 1717m, 1610w, 1523w, 1489w, 1441w, 1392w, 1368w, 1264m, 1156m, 702s cm⁻¹; HR ESI-TOF m/z 395.2177 $(M + H^{+}, C_{20}H_{31}N_{2}O_{6}^{+}, requires 395.2177); [\alpha]^{25}_{D} + 47 (c 0.5, CH_{2}Cl_{2}).$

1-[(3-Amino)-D-phenylalanyl]desleucylaglucovancomycin Bis(trifluoroacetate) (9)

The title compound was prepared from **24** (5.58 mg, 0.015 mmol), and dicyclohexylcarbodiimide (1.46 mg, 0.007 mmol) in CH₂Cl₂ (100 μ L), followed by treatment with NaHCO₃ (1.19 mg, 0.015 mmol), desleucylaglucovancomycin (**16**, 4.00 mg, 0.004 mmol) in DMF (200 μ L) according to *GP3*. The protected intermediate was isolated as a white solid (2.70 mg, theoretical 4.90 mg, 55%) by reverse-phase HPLC with a gradient of 10–80 % MeCN/water with 0.07% TFA (t_R = 29.2 min) and lyophilized, and taken directly to the next step without further characterization. The intermediate (2.70 mg, 0.002 mmol) was taken up in 50% TFA in CH₂Cl₂ (0.2 mL) and stirred at 20 °C. After 15 min, the volatiles were removed with a gentle stream of N₂ and residual TFA was azetroped with CH₂Cl₂ (3 × 5 mL). The residue was purified by reverse-phase HPLC using a gradient of 5–40% MeCN/water with 0.07% TFA (t_R = 24.1 min) and afforded the bis-TFA salt of **9** (2.3 mg, theoretical 2.5 mg, 92%) as white powder after lyophilization: ¹H NMR ((CD₃)₂SO, 600 MHz) δ 9.55 (s, 1H), 9.42 (s, 1H), 9.13 (s, 1H), 9.04–9.00 (m, 2H), 8.65 (s, 1H), 8.55 (d, 3H, J = 3.6 Hz), 7.84 (s, 1H), 7.57 (d, 1H, J = 8.4 Hz), 7.45 (d, 3H, J =

8.4 Hz), 7.25 (t, 2H, J = 8.4 Hz), 7.18 (s, 1H), 7.13 (s, 1H), 7.10 (s, 1H), 7.04–6.97 (m, 3H), 6.76 (dd, 1H, J = 1.8, 8.4 Hz), 6.70 (d, 3H, J = 8.4 Hz), 6.50 (br s, 4H), 6.38 (d, 1H, J = 1.8 Hz), 6.25 (d, 1H, J = 1.8 Hz), 5.94 (br s, 1H), 5.88 (br s, 1H), 5.69 (d, 1H, J = 7.8 Hz), 5.63 (br s, 1H), 5.17 (s, 1H), 5.15 (s, 1H), 5.11 (br s, 1H), 4.88 (br s, 1H), 4.44–4.42 (m, 2H), 4.27 (br s, 2H), 4.16 (d, 1H, J = 11.4 Hz), 3.08 (dd, 1H, J = 5.4, 14.4 Hz), 2.81 (dd, 1H, J = 7.8, 13.8 Hz), 2.54 (s, 3H), 2.22 (dd, 1H, J = 7.8, 15.6 Hz); HR ESI-TOF m/z 1192.2870 (M + H⁺, C₅₆H₅₂Cl₂N₉O₁₇⁺, requires 1192.2853); $[\alpha]^{25}_{D}$ +25 (c 0.2, CH₃OH).

(R)-Methyl 2-(tert-Butoxycarbonyl(methyl)amino)-3-(3-(3-ethylureido)phenyl)propanoate (25)

Ethylisocyanate (74 μL, 0.94 mmol) was added dropwise to a stirred solution of **20** (0.15 g, 0.47 mmol) in anhydrous CH₂Cl₂ (5.8 mL). After 15 h, the volatiles were removed in vacuo. Column chromatography (18 g SiO₂, 50% EtOAc/hexanes) provided **25** (0.17 g, theoretical 0.18 mg, 97%) as a yellow oil (rotomer ratio in CDCl₃ A/B = 3:4): 1 H NMR (CDCl₃, 600 MHz) δ 7.21–7.13 (m, 8H), 6.98 (br s, 1H), 6.83–6.62 (m, 1H), 5.65 (br s, 1H), 5.32 (br s, 1H), 5.02 (dd, 1H_B, J = 4.8. 10.8 Hz), 4.50 (d, 1H_A, J = 7.2 Hz), 3.73 (s, 3H_A), 3.72 (s, 3H_B), 3.27–3.20 (m, 6H), 2.96–2.88 (m, 2H), 2,71 (s, 6H), 1.31 (s, 18H), 1.15–1.10 (m, 6H); 13 C NMR (CDCl₃, 150 MHz) δ 171.5/171.4, 156.2/156.1, 155.9/155.1, 139.4/138.8, 138.7/138.1, 129.2, 124.2/123.6, 121.5/120.7, 119.3/118.3, 80.4/80.2, 61.6/59.0, 52.23/52.20, 35.5/35.0, 34.95/34.91, 32.7/31.4, 28.2/28.1 (3C), 15.4; IR (neat) ν_{max} 3333 ν_{max} 1741 ν_{max} 1651 ν_{max} 1482 ν_{max} 1436 ν_{max} 1436 ν_{max} 1435 ν_{max} 1435, 910 ν_{max} 1482 ν_{max} 1436 ν_{max} 1

(R)-2-(tert-Butoxycarbonyl(methyl)amino)-3-(3-(3-ethylureido)phenyl)propanoic Acid (26)

An aqueous solution of 0.2 N LiOH (3.95 mL) was added to a solution of **25** (100 mg, 0.263 mmol) in THF (3.75 mL) according to GP2. Column chromatography (10 g SiO₂, 5–10% MeOH/EtOAc with 1% AcOH) provided **26** (84 mg, theoretical 96 mg, 88%) as an off-white foam (rotomer ratio in acetone-D₆ A/B = 3:4): 1 H NMR ((CD₃)₂CO, 600 MHz) δ 7.90 (br s, 1H_B), 7.88 (br s, 1H_A), 7.42 (br s, 1H), 7.35–7.32 (m, 3H), 7.15–7.12 (m, 2H), 6.83 (d, 2H, J = 7.2 Hz), 5.86 (br s, 1H_A), 5.80 (br s, 1H_B), 4.94 (dd, 1H, J = 4.8, 11.4 Hz), 4.72 (dd, 1H, J = 3.6, 10.8 Hz), 3.26–3.22 (m, 6H), 3.03–2.96 (m, 2H), 2.75 (s, 3H_A), 2.72 (s, 3H_B), 1.34 (s, 9H_A), 1.31 (s, 9H_B), 1.11–1.08 (m, 6H) (CO₂H not observed); 13 C NMR ((CD₃)₂CO, 150 MHz) δ 174.0/173.9, 157.4/157.3, 157.2/156.5, 142.6/142.4, 140.8/140.5, 130.4/130.3, 124.1/124.0, 120.9/120.7, 118.5/118.2, 81.0/80.9, 62.8/61.3, 37.2/36.3, 36.2, 33.4/33.3, 29.4/29.3 (3C), 16.9; IR (neat) ν_{max} 3348 ν_{max} 1654 ν_{max} 1654 ν_{max} 3348 ν_{max} 1654 ν_{max} 1757 $\nu_$

1-[3-(3-Ethylureido)-D-phenylalanyl]desleucylaglucovancomycin Trifluoroacetate (10)

The title compound was prepared from 26 (5.17 mg, 0.015 mmol), and dicyclohexylcarbodiimide (1.46 mg, 0.007 mmol) in CH₂Cl₂ (100 µL), followed by treatment with NaHCO₃ (1.19 mg, 0.015 mmol), desleucylaglucovancomycin (16, 4.00 mg, 0.004 mmol) in DMF (200 µL) according to GP3. The protected intermediate was isolated as a white solid (3.3 mg, theoretical 5.6 mg, 59%) by reverse-phase HPLC with a gradient of 35–95% MeCN/water with 0.07% TFA ($t_R = 19.9 \text{ min}$), and lyophilized, and was taken directly to the next step without further characterization. The intermediate (3.30 mg, 0.002 mmol) was taken up in 50% TFA in CH₂Cl₂ (0.2 mL) and stirred at 20 °C. After 10 min, the volatiles were removed with a gentle stream of N_2 and residual TFA was azetroped with CH_2Cl_2 (3 × 5 mL). The residue was purified by reverse-phase HPLC using a gradient of 10-60% MeCN/water with 0.07% TFA $(t_R = 20.6 \text{ min})$ and afforded the TFA salt of 10 (2.50 mg, theoretical 3.30 mg, 75%) as white powder after lyophilization: ¹H NMR (CD₃OD, 600 MHz) δ 7.82 (br s, 2H), 7.65 (s, 1H), 7.62 (br s, 2H), 7.56 (d, 1H, J = 8.4 Hz), 7.28 (t, 1H, J = 7.8 Hz), 7.19 (d, 1H, J = 6.6 Hz), 7.07 (s, 1H), 7.03 (d, 2H, J = 6.6Hz), 6.57 (br s, 3H), 6.40 (d, 1H, J = 1.2 Hz), 6.12 (br s, 1H), 5.93 (br s, 1H), 5.41 (s, 1H), 5.38 (s, 1H), 5.30 (d, 1H, J = 2.4 Hz), 4.80 (br s, 2H), 4.26–4.22 (m, 2H), 4.14 (s, 1H), 3.19–3.14 (m, 4H), 2.97 (d, 1H, J = 15.6 Hz), 2.88–2.83 (m, 1H), 2.64 (s, 3H), 2.01 (br s, 1H), 1.11 (t, 3H, J = 7.2 Hz); ESI-TOF HRMS m/z: 1263.3225 (M + H⁺, C₅₉H₅₇Cl₂N₁₀O₁₈⁺, requires 1263.3224); $[\alpha]^{25}_{D}$ +45 (c 0.1, CH₃OH).

(R)-Methyl 3-(3-(2,3-Bis(tert-butoxycarbonyl)guanidino)phenyl)-2-(tert-butoxycarbonyl(methyl)amino)propanoate (28)

Amine **20** (100 mg, 0.325 mmol) was added to a solution of *N*, *N*′-di-Boc-1*H*-pyrazole-1-carboxamidine (**35**, 100 mg, 0.325 mmol) in anhydrous CH₃CN (1.63 mL) and the mixture was warmed at 80 °C. After 2 h, the volatiles were removed in vacuo. Column chromatography (20 g SiO₂, 0–10 % MeOH/CH₂Cl₂) provided **28** (85.0 mg, theoretical 179 mg, 47%) as a white foam (rotomer ratio in CDCl₃ A/B = 10:11): 1 H NMR (CDCl₃, 600 MHz) δ 11.63 (s, 2H), 10.33 (d, 2H, J = 8.4 Hz), 7.57 (d, 1H_A, J = 7.8 Hz), 7.54 (d, 1H_B, J = 7.8 Hz), 7.43 (s, 1H_B), 7.37 (s, 1H_A), 7.29–7.25 (m, 2H), 6.99 (d, 1H_A, J = 7.2 Hz), 6.94 (d, 1H_B, J = 7.8 Hz), 4.89 (dd, 1H_A, J = 4.8, 10.2 Hz), 4.51 (d, 1H_B, J = 6.0 Hz), 3.76 (s, 6H), 3.30 (dd, 2H, J = 4.8, 13.8 Hz), 3.04 (dd, 2H, J = 10.8, 13.4 Hz), 2.75 (s, 6H), 1.55–1.50 (m, 36H), 1.39 (s, 18H); 13 C NMR (CDCl₃, 150 MHz) δ 171.8/171.6, 163.5, 155.8/154.9, 153.4, 153.3, 138.4/138.1, 137.0/136.8, 129.02/128.96, 125.3/125.2, 122.6/122.5, 120.39/120.36, 83.7, 80.3/80.0, 79.5, 61.6/59.5, 35.5/34.8, 33.1/32.2, 29.7/29.2, 28.2 (3C), 28.1 (3C), 28.0 (3C); IR (neat) 3270w, 2977w, 2361w, 1699m, 1634m, 1394m, 1367m, 1334m, 1300m, 1246m, 1149s, 1109m, 1057w cm⁻¹; HR ESI-TOF m/z 573.2895 (M +

 Na^{+} , $C_{27}H_{42}N_4NaO_8^{+}$, requires 573.2895); $[\alpha]^{25}D + 36$ (c 0.5, CH_2Cl_2).

(R)-3-(3-(2,3-Bis(tert-butoxycarbonyl)guanidino)phenyl)-2-(tert-butoxycarbonyl(methyl)amino)propanoic Acid (29)

An aqueous solution of 0.2 N LiOH (1.08 mL) was added to a solution of **28** (80 mg, 0.07 mmol) in THF (1.0 mL) according to GP2. Column chromatography (5 g SiO₂, 100% EtOAc) provided **29** (52 mg, theoretical 78 mg, 66%) as a clear oil (rotomer ratio in acetone-D₆ A/B = 3:4): ¹H NMR ((CD₃)₂CO, 600 MHz) δ 11.75 (br s, 2H), 10.29 (d, 2H, J = 9.6 Hz), 7.70–7.67 (m, 2H), 7.52–7.48 (m, 2H), 7.31–7.26 (m, 2H), 7.08 (d, 2H, J = 7.2 Hz), 4.90 (d, 1H_A, J = 7.8 Hz), 4.76 (d, 1H_B, J = 8.4 Hz), 3.32–3.30 (m, 2H), 3.12–3.05 (m, 2H), 2.79 (s, 3H_A), 2.74 (s, 3H_B), 1.53–1.47 (m, 36H), 1.34 (s, 18H) (CO₂H protons not observed); ¹³C NMR ((CD₃)₂CO, 150 MHz) δ 174.4, 157.4, 156.5, 155.3, 153.2, 141.3/141.1, 139.1/139.0, 130.5/130.4, 127.3/127.2, 124.5/124.4, 122.0/121.9, 83.8, 80.9, 80.7, 62.8/61.5, 37.1/36.4, 33.6, 29.5/29.4 (3C), 29.2 (3C), 29.1 (3C); IR (neat) ν_{max} 3270 ν_{max} 2977 ν_{max} 1718 ν_{max} 1696 ν_{max} 1394 ν_{max} 1368 ν_{max} 1331 ν_{max} 1299 ν_{max} 1247 ν_{max} 1109 ν_{max} 1058 ν_{max} 777 ν_{max} 1718 ν_{max} 1696 ν_{max} 1394 ν_{max} 1368 ν_{max} 131 ν_{max} 1299 ν_{max} 1247 ν_{max} 1109 ν_{max} 1058 ν_{max} 777 ν_{max} 1718 ν_{max} 1696 ν_{max} 1394 ν_{max} 1368 ν_{max} 131 ν_{max} 1299 ν_{max} 1247 ν_{max} 1109 ν_{max} 1058 ν_{max} 1718 ν_{max} 1299 ν_{max} 1247 ν_{max} 1109 ν_{max} 1058 ν_{max} 1718 ν_{max} 1299 ν_{max} 1247 ν_{max} 1199 ν_{max} 1058 ν_{max} 171 ν_{max} 1299 ν_{max} 1247 ν_{max} 1299 ν_{max} 129

1-[(3-Guanudyl)-D-phenylalanyl]desleucylaglucovancomycin Bis(trifluoroacetate) (11)

The title compound was prepared starting from 29 (7.60)mg, $0.015 \, \text{mmol}$), dicyclohexylcarbodiimide (1.46 mg, 0.007 mmol) in CH₂Cl₂ (100 µL), followed by treatment with NaHCO₃ (1.19 mg, 0.015 mmol), desleucylaglucovancomycin (16, 4.00 mg, 0.004 mmol) in DMF (200 μL) according to GP3. The protected intermediate was isolated as a white solid (2.70 mg, theoretical 5.40 mg, 50%) by reverse-phase HPLC with a gradient of 35–95 % MeCN/water with 0.07% TFA ($t_R =$ 19.7 min), and lyophilized, and was taken directly to the next step without further characterization. The residue was taken up in 50% TFA in CH₂Cl₂ (0.2 mL) and stirred at 20 °C. After 15 min, the volatiles were removed with a gentle stream of N₂ and residual TFA was azetroped with CH₂Cl₂ (3 × 5 mL). Purification by reverse-phase HPLC using a gradient of 5–40% MeCN/water with 0.07% TFA (t_R = 24.2 min) and afforded the bis-TFA salt of 11 (2.70 mg, theoretical 5.10 mg, 53%) as a white powder: ¹H NMR (CD₃OD, 600 MHz) δ 8.99 (d, 1H, NH, J = 4.8 Hz), 8.84 (d, 1H, NH, J = 5.4 Hz), 8.81 (br s, 1H), 7.82 (br s, 1H), 7.78 (s, 1H), 7.63 (br s, 1H), 7.56 (s, 1H), 7.47 (d, 1H, J = 7.8 Hz), 7.35 (t, 1H, J =7.2 Hz), 7.21–7.12 (m, 5H), 6.92 (br s, 1H), 6.86 (br s, 1H), 6.72 (d, 1H, J = 8.4 Hz), 6.44 (d, 1H, J = 8.4 Hz), 6.45 (d, 1H, J = 8.4 Hz), 6.46 (d, 1H, J = 8.4 Hz), 6.46 (d, 1H, J = 8.4 Hz), 6.47 (d, 1H, J = 8.4 Hz), 6.48 (d, 1H, J = 8.4 Hz), 6.49 (d, 1H, J = 8.4 Hz), 6.49 (d, 1H, J = 8.4 Hz), 6.40 (d, 1H, J = 8.4 Hz), 6.40 (d, 1H, J = 8.4 Hz), 6.41 (d, 1H, J = 8.4 Hz), 6.42 (d, 1H, J = 8.4 Hz), 6.44 (d, 1H, J = 8.4 Hz), 6.45 (d, 1H, J = 8.4 Hz), 6.44 (d, 1H, J = 8.4 Hz), 6.45 (d, 1H, J = 8.4 2.4 Hz), 6.42 (d, 1H, J = 1.8 Hz), 6.08 (br s, 1H), 5.97 (br s, 1H), 5.45 (s, 1H), 5.31 (s, 1H), 5.16 (s, 1H), 4.80 (d, 1H, J = 6.0 Hz), 4.73 (s, 1H), 4.30-4.18 (m, 3H), 3.23-3.21 (m, 1H), 2.84 (d, 1H, J = 16.2 Hz), 2.77 (s, 3H), 1.84 (br s, 1H); ESI-TOF HRMS m/z 1234.3011 (M + H⁺, C₅₇H₅₄Cl₂N₁₁O₁₇⁺, requires 1234.3071); $\left[\alpha\right]^{25}_{D}$ +22 (c 0.5, CH₃OH).

Analogues modified at the N-terminus for covalent attachment to Ac₂-L-Lys-D-Ala-D-Lac (3)

(R)-2-(tert-Butoxycarbonyl(methyl)amino)-3-(3-(2-(tert-butoxycarbonyl)hydrazinyl)phenyl)propanoic Acid (30)

A solution of oxaziridine 31^{15} (0.234 g, 0.811 mmol) in anhydrous CH₂Cl₂ (2.35 mL) was added dropwise to a solution of 20 (0.250 g, 0.811 mmol) in CH₂Cl₂ (2.35 mL) at 20 °C. After 18 h, the volatiles were removed in vacuo. Column chromatography (15 g SiO₂, 50% Et₂O/hexanes) provided the crude hydrazine 32 (80 mg, 23%) as a pale yellow foam. The residue (70 mg, 0.16 mmol) was taken up in THF (2.35 mL) at 0 °C and treated with a solution of aqueous 0.2 N LiOH (2.4 mL, 0.48 mmol) and was allowed to stir for 1.75 h at 0 °C according to *GP2*. Column chromatography (7 g SiO₂, 5–10% MeOH/CH₂Cl₂ with 0.1% CH₃CO₂H) afforded 30 (43 mg, 66 mg theoretical, 65%) as a pale yellow foam (rotomer ratio A/B = 6:5): ¹H NMR (CDCl₃, 500 MHz) δ 7.15 (t, 2H, J = 7.8 Hz), 6.74–6.66 (m, 6H), 6.50 (br s, 2H), 5.77 (br s, 4H), 4.72 (br s, 1H_B), 4.52 (br s, 1H_A), 3.24 (br s, 2H), 3.11–2.95 (m, 2H), 2.75 (s, 3H_B), 2.68 (s, 3H_A), 1.46 (s, 18H), 1.42 (s, 9H), 1.37 (s, 9H); ¹³C NMR (CDCl₃, 150 MHz) δ 175.4/175.1, 156.4, 155.0, 148.54/148.47, 138.7/138.3, 129.4/129.2, 121.4, 113.6, 111.4, 81.44/81.42, 80.69/80.64, 61.6/60.9, 35.5/34.8, 33.2/32.9, 28.4 (6C); IR (neat) ν_{max} 3309 ν_{max} 3309 ν_{max} (br), 2977 ν_{max} 2931 ν_{max} 1679 ν_{max} 1610 ν_{max} 1392 ν_{max} 1368 ν_{max} 1368 ν_{max} 1368 ν_{max} 1575 ν_{max} 1570F ν_{max} 410.2287 (M + H⁺, C₂₀H₃₂N₃O₆⁺, requires 410.2286); [α]²⁵ ν_{max} 156 (ν_{max} 1570F ν_{max} 2410.2287 (M + H⁺, C₂₀H₃₂N₃O₆⁺, requires 410.2286); [α]²⁵ ν_{max} 156 (ν_{max} 1570F ν_{max} 2410.2287 (M + H⁺, C₂₀H₃₂N₃O₆⁺, requires 410.2286); [α]²⁵ ν_{max} 158 (ν_{max} 159 (ν_{max} 159 (ν_{max} 159 (ν_{max} 159 (ν_{max} 150 (ν_{\text

1-[(3-Hydrazino)-D-phenylalanyl]desleucylaglucovancomycin Bis(trifluoroacetate) (13)

The title compound was prepared starting from **30** (11.7 mg, 0.029 mmol), and dicyclohexyl carbodiimide (2.95 mg, 0.014 mmol) in CH_2Cl_2 (205 μ L), followed by treatment with NaHCO₃ (2.41 mg, 0.029 mmol), desleucylaglucovancomycin (**16**, 11.0 mg, 0.009 mmol) in DMF (0.514 mL) according to *GP3*. The protected intermediate was isolated as a white solid (9.80 mg, theoretical 12.6 mg, 77%) by reverse-phase HPLC with a gradient of 10–80% MeCN/water with 0.07% TFA (t_R = 28.5 min), and lyophilized, and was taken directly to the next step without further characterization. The intermediate (2.50 mg, 0.002 mmol) was taken up in 10% TFA in CH_2Cl_2 (0.1 mL) and stirred at 0 °C. After 30 min, the volatiles were removed with a gentle stream of N_2 and residual TFA was azetroped with CH_2Cl_2 (3 × 5 mL). The residue was purified by reverse-phase HPLC using a gradient of 5–40%

MeCN/water with 0.07% TFA (t_R = 23.7 min) and afforded the bis-TFA salt of **13** (1.6 mg, theoretical 2.5 mg, 64%) as an off-white powder: ¹H NMR (CD₃OD, 600 MHz) δ 8.98 (d, 1H, NH, J = 3.0 Hz), 8.88 (br s, 1H, NH), 7.76 (br s, 2H), 7.66 (br s, 1H), 7.57–7.54 (m, 2H), 7.31 (t, 1H, J = 7.8 Hz), 7.21 (d, 1H, J = 7.8 Hz), 7.10 (s, 1H), 7.03 (d, 1H, J = 7.2 Hz), 6.84 (dd, 1H, J = 1.8, 8.4 Hz), 6.76–6.70 (m, 3H), 6.47 (s, 1H), 6.44 (d, 1H, J = 1.8 Hz), 6.01 (br s, 1H), 5.87 (br s, 1H), 5.38 (s, 1H), 5.36 (s, 1H), 5.28 (d, 1H, J = 2.4 Hz), 4.79–4.77 (m, 1H), 4.33 (br s, 1H), 4.27 (d, 1H, J = 8.4 Hz), 4.18 (br s, 1H), 3.22 (dd, 1H, J = 6.0, 13.8 Hz), 2.95 (d, 1H, J = 15.6 Hz), 2.70 (s, 3H), 2.08 (br s, 1H) (three signals buried); ESI-TOF HRMS m/z 1207.2907 (M + H⁺, C₅₆H₅₃Cl₂N₁₀O₁₇⁺, requires 1207.2962); [α]²⁵_D +18 (c 0.1, CH₃OH).

(R)-2-(tert-Butoxycarbonyl(methyl)amino)-3-(3-cyanophenyl)propanoic Acid (34)

The title compound was prepared from Boc-D-3-cyanophenylalanine (**33**, 1.00 g, 3.44 mmol) with NaH (454 mg, 11.4 mmol), iodomethane (643 µL, 10.3 mmol), and anhydrous THF (11.5 mL), and DMF (546 µL) according to *GP1*. Column chromatography (10 g SiO₂, 0–0.2% AcOH/EtOAc) provided **34** (1.01 g, theoretical 1.05 g, 97%) as a colorless opaque oil (rotomer ratio in CDCl₃ A/B = 8:5): ¹H NMR (CDCl₃, 600 MHz) δ 7.55–7.50 (m, 6H), 7.43–7.41 (m, 2H), 4.80 (dd, 1H, J = 3.6, 9.0 Hz), 4.64 (d, 1H, J = 7.2 Hz), 3.37–3.34 (m, 2H), 3.16 (t, 1H, J = 12.0 Hz), 3.08 (t, 1H, J = 12.0 Hz), 2.77 (s, 3H_B), 2.71 (s, 3H_A), 1.41 (s, 9H_A), 1.36 (s, 9H_B) (CO₂H not observed); ¹³C NMR (CDCl₃, 150 MHz) δ 174.9/174.7, 156.2/154.7, 139.0/138.8, 133.6/133.5, 132.5, 130.6, 129.5/129.3, 118.7/118.6, 112.7/112.6, 81.2, 60.8/60.3, 34.9/34.3, 33.1/32.5, 28.2 (3C); IR (neat) v_{max} 2976w, 2230w, 1684s, 1481m, 1447m, 1391m, 1367m, 1322m, 1252m, 1142s, 1074m cm⁻¹; ESI-TOF HRMS m/z 327.1312 (M + Na⁺, C₁₆H₂₀N₂NaO₄⁺, requires 327.1315); $[\alpha]^{25}_{\text{D}}$ +65 (c 0.5, CH₂Cl₂).

(R)-Methyl 2-(tert-Butoxycarbonyl(methyl)amino)-3-(3-cyanophenyl)propanoate (35)

A solution of **34** (600 mg, 1.97 mmol) in anhydrous DMF (6.6 mL) at 0 °C was treated with oven-dried KHCO₃ (374 mg, 3.94 mmol) followed by iodomethane (171 μL, 2.76 mmol). After 1 h at 0 °C, the mixture was gently warmed to ambient temperature. After 18 h, the reaction was quenched with water (10 mL), and the mixture was extracted with EtOAc (100 mL), the organic was discarded. The aqueous phase was acidified to pH of 2–3 with aqueous 0.2 N NaHSO₄ and extracted with EtOAc (3 × 200 mL) and the combined organic layers were dried over Na₂SO₄, filtered, and concentrated. Column chromatography (75 g SiO₂, 20–60% hexanes/EtOAc) provided **35** (617 mg, theoretical 626 mg, 99%) as a white solid (rotomer ratio in CDCl₃ A/B = 6:5): mp 83–84 °C; ¹H NMR (CDCl₃, 600 MHz) δ 7.54–7.39 (m, 6H), 7.41–7.39 (m, 2H), 4.90 (dd, 1H_A, J = 5.4, 10.2 Hz), 4.55 (dd, 1H_B, J = 3.6, 9.6 Hz), 3.77 (s, 3H_B), 3.75 (s, 3H_A), 3.36–3.32 (m, 2H), 3.08–3.03 (m, 2H), 2.73 (s, 3H_B), 2.71 (s, 3H_A), 1.38 (s, 9H_A), 1.35 (s, 9H_B); ¹³C NMR (CDCl₃, 150 MHz) δ 172.2/171.9, 155.7/154.7, 139.3/139.1, 133.7/133.6, 132.5, 130.5/130.4, 129.3/129.2, 118.8/118.6, 112.7/112.4, 80.7/80.5, 61.1, 59.3, 35.1/34.6, 32.5/32.0, 28.2 (3C); IR (neat) ν_{max} 2976 ν_{max} 2976 ν_{max} 2930 ν_{max} 1741 ν_{max} 1741 ν_{max} 2976 ν_{max} 2930 ν_{max} 1741 ν_{max} 174

1367*m*, 1322*m*, 1223*m*, 1141*s*, 734*s* cm⁻¹; ESI-TOF HRMS m/z 341.1476 (M + Na⁺, C₁₇H₂₂N₂NaO₄⁺, requires 341.1472); [α]²⁵_D +68 (c 1.0, CH₂Cl₂).

(R)-Methyl 3-(3-(Aminomethyl)phenyl)-2-(tert-butoxycarbonyl(methyl)amino)propanoate (36)

An aqueous solution of Raney-Ni (cat.) was added to an oxygen degassed solution of **35** (300 mg, 0.940 mmol) in MeOH (9.4 mL) and aqueous ammonium hydroxide (1.4 mL). The mixture was vigorously stirred and placed under an atmosphere of H_2 gas (balloon). After 18 h, the solids were removed by filtration over Celite, and washed with MeOH. The filtrate was concentrated in vacuo. The residue was taken up in water (10 mL) and extracted with EtOAc (3 × 200 mL). The combined organics were dried over Na_2SO_4 , filtered, and the volatiles removed in vacuo providing **36** (293 mg, theoretical 303 mg, 97%) as a deep yellow oil (rotomer ratio in CDCl₃ A/B = 10:11): 1H NMR (CD₂Cl₂, 600 MHz) δ 7.26–7.05 (m, 8H), 4.89 (dd, 1H_A, J = 4.5, 10.0 Hz), 4.58 (dd, 1H_B, J = 4.0, 10.5 Hz), 3.84 (br s, 4H), 3.73 (s, 3H_B), 3.71 (s, 3H_A), 3.26 (d, 2H, J = 14.5 Hz), 2.99 (dd, 2H, J = 11.0, 14.5 Hz), 2.68 (s, 6H), 1.69 (br s, 4H), 1.35 (s, 9H_A), 1.30 (s, 9H_B); 13 C NMR (CDCl₃, 150 MHz) δ 173.0/172.8, 157.6/156.9, 141.0/140.5, 139.8/139.6, 130.0/129.9, 129.7, 129.57/129.55, 127.3, 81.9/81.5, 63.2/61.5, 52.84/52.77, 45.9/45.8, 36.3/35.7, 33.7/33.1, 28.6/28.5 (3C); IR (neat) v_{max} 2975w, 1740w, 1688s, 1608w, 1480w, 1437w, 1391w, 1324w, 1222w, 1140s, 773w, 733w, 701w cm⁻¹; HR ESI-TOF w/z 323.1965 (M + H⁺, C₁₇H₂₇N₂O₄⁺, requires 323.1965); [α]²⁵p +56 (c 1.0, CH₂Cl₂).

(R)-Methyl 2-(tert-Butoxycarbonyl(methyl)amino)-3-(3-((tert-butoxycarbonylamino)methyl)phenyl)propanoate (37)

A solution of di-*tert*-butyl dicarbonate (30 μL, 0.13 mmol) in CHCl₃ (250 μL) was added to a solution of **36** (41 mg, 0.13 mmol) in CHCl₃ (520 μL) and the solution was stirred at 20 °C. After 24 h, the solution was diluted with CHCl₃ (5 mL) and washed with 0.2 N NaHSO₄, saturated aqueous NaHCO₃, water, and the combined organic layers were dried over Na₂SO₄, filtered and concentrated. Column chromatography (10 g SiO₂, 30–100% hexanes/EtOAc) provided **37** (rotomer ratio in CDCl₃ A/B = 10:11) as a pale yellow oil (47 mg, theoretical 55 mg, 86%): ¹H NMR (CDCl₃, 500 MHz) δ 7.25–7.23 (m, 2H), 7.15–7.06 (m, 6H), 4.93 (dd, 1H_A, J = 5.5, 10.5 Hz), 4.82 (br s, 2H), 4.52 (dd, 1H_B, J = 4.0, 10.0 Hz), 4.28 (br s, 4H), 3.74 (s, 3H_B), 3.73 (s, 3H_A), 3.31–3.24 (m, 2H), 2.99 (dd, 2H, J = 11.0, 14.5 Hz), 2.71 (s, 3H_B), 2.69 (s, 3H_A), 1.45 (s, 18H), 1.36 (s, 9H_A), 1.33 (s, 9H_B); ¹³C NMR (CDCl₃, 125 MHz) δ 171.8/171.5, 155.8 (2C), 139.2/138.9, 138.1/137.8, 128.8/128.7 (2C), 128.1/128.0, 125.8, 80.3/80.0, 79.4, 61.5/59.4, 52.2, 44.6, 35.5/34.9, 32.6/31.9, 28.4 (3C), 28.2 (3C); IR (film) v_{max} 3366w, 2975w, 2361w, 1742w, 1691s, 1512w, 1450w, 1392w, 1366w, 1328w, 1248m, 1164s, 1046w, 776w cm⁻¹; HRESI-TOF m/z 423.2496 (M + H⁺, C₂₂H₃₅N₂O₆⁺, requires 423.2490); [α]²⁵_D +46 (c 1.0, CH₂Cl₂).

(R)-2-(tert-Butoxycarbonyl(methyl)amino)-3-(3-((tert-butoxycarbonylamino)methyl)phenyl)propanoic Acid (38)

The title compound was prepared from **37** (58.0 mg, 0.136 mmol), 0.2 N aqueous LiOH (1.36 ml), and THF (1.94 mL) according to GP2. The mixture was stirred for 3 h at 0 °C. Column chromatography (5 g SiO₂, 50% EtOAc/hex) afforded **38** (rotomer ratio in acetone-D₆ A/B = 10:12) as a white foam (49.8 mg, theoretical 55.5 mg, 88%): ¹H NMR ((CD₃)₂CO, 600 MHz) δ 7.25–7.15 (m, 10H), 6.40 (br s, 1H), 6.38 (br s, 1H), 4.91 (dd, 1H_A, J = 3.6, 10.2 Hz,), 4.71 (d, 1H_B, J = 10.2 Hz), 4.25 (br s, 4H), 3.30–3.26 (m, 2H), 3.10–3.03 (m, 2H), 2.72 (s, 3H_A), 2.70 (s, 3H_B), 1.42 (s, 18H), 1.36 (s, 9H_A), 1.32 (s, 9H_B); ¹³C NMR ((CD₃)₂CO, 150 MHz) δ 173.6/173.5, 157.8/157.3, 156.4, 142.2/142.0, 140.3/140.1, 130.2/130.1, 129.9/129.8, 129.4/129.3, 127.12/127.09, 81.0/80.8, 79.7, 62.8/61.3, 45.8, 37.0/36.4, 33.6/33.5, 29.6 (3C), 29.5/29.4 (3C); IR (film) ν_{max} 2976 ν_{max} 1686 ν_{max} 1515 ν_{max} 1480 ν_{max} 1392 ν_{max} 1366 ν_{max} 1366 ν_{max} 151 ν_{max} 162 ν_{max} 151 ν_{max} 1686 ν_{max} 170 $\nu_{\text{$

1-[(3-Aminomethyl)-D-phenylalanyl]desleucylaglucovancomycin Bis(trifluoroacetate) (12)

The title compound was prepared starting from 38 (8.65)mg, 0.021 mmol), and dicyclohexylcarbodiimide (2.18 mg, 0.014 mmol), in CH₂Cl₂ (0.150 mL), followed by treatment with NaHCO₃ (1.78 mg, 0.021 mmol), desleucylaglucovancomycin (16, 10.0 mg, 0.009 mmol) in DMF (0.30 mL) according to GP3. The protected intermediate was isolated as a white solid (9.80 mg, theoretical 12.6 mg, 77%) by reverse-phase HPLC with a gradient of 35–95% MeCN/water with 0.07% TFA (t_R = 13.9 min), and lyophilized, and was taken directly to the next step without further characterization. The intermediate (9.80 mg, 0.007 mmol) was taken up in 10% TFA in CH₂Cl₂ (0.2 mL) and stirred at 0 °C. After 30 min, the volatiles were removed with a gentle stream of N₂ and residual TFA was azetroped with CH₂Cl₂ (3 \times 5 mL). The residue was purified by reverse-phase HPLC using a gradient of 5–40% MeCN/water with 0.07% TFA ($t_R = 23.3$ min) and afforded the bis-TFA salt of 12 (7.8 mg, theoretical 12.6 mg, 62%): ¹H NMR (CD₃OD, 600 MHz) δ 9.04 (br s, 1H, NH), 8.91 (d, 1H, NH, J = 5.4 Hz) 8.72 (br s, 1H, NH), 7.77 (br s, 1H), 7.70 (s, 1H), 7.66 (s, 1H), 7.53 (d, 1H, J = 7.8 Hz), 7.40–7.31 (m, 4H), 7.20 (t, 2H, J = 7.8 Hz), 7.12 (s, 1H), 6.81 (br s, 1H), 6.72 (d, 1H, J = 8.4 Hz), 6.46 (s, 1H), 6.44 (d, 1H, J = 2.4 Hz), 6.12 (br s, 1H), 5.92 (br s, 1H), 5.41 (s, 1H), 5.34 (s, 1H), 5.21 (d, 1H, J = 2.4 Hz), 4.84 (d, 1H, J = 6.0 Hz), 4.77 (d, 1H, J = 4.8 Hz) 4.30 (t, 1H, J = 7.8 Hz), 4.25 (s, 1H), 4.23 (s, 1H), 4.10 (s, 2H), 3.33-3.27 (m, 1H), 3.16-3.12 (m, 1H), 2.84 (d, 1H, J = 16.2 Hz), 2.75 (s, 3H), 1.98 (br s, 1H) (one signal buried below residual solvent); ESI-TOF HRMS m/z 1206.2988 (M + H⁺, C₅₇H₅₄Cl₂N₉O₁₇⁺,

requires 1206.3009); $[\alpha]^{25}_D + 33$ (c 0.2, CH₃OH).

(R)-2-(tert-Butoxycarbonyl(methyl)amino)-3-(3-((2-(tert-butoxycarbonyl)hydrazinyl)methyl)phenyl)propanoic Acid (40)

A solution of oxaziridine 31^{15} (67.0 mg, 0.232 mmol) in anhydrous toluene (0.8 mL) was added dropwise over a 15 min period to a solution of 36 (75.0 mg, 0.232 mmol) in anhydrous toluene (0.8 mL) at 0 °C. After stirring for a total of 30 min at 0 °C, the volatiles were removed in vacuo. Column chromatography (15 g SiO₂, 50-60% EtOAc/hexanes) afforded 39 (58 mg, theoretical 100 mg, 58%) as a yellow oil. The residue (58 mg, 0.133 mmol) was taken up in THF (1.9 mL) and treated with aqueous 0.2 N LiOH (2.0 mL, 0.44 mmol) at 0 °C, according to GP2. Column chromatography (7 g, SiO₂ 50–100% EtOAc/hexanes) provided 40 (35.0 mg, theoretical 56.3 mg, 62%) as a white foam (rotomer ratio in acetone-D₆ A/B = 3:4): ¹H NMR ((CD₃)₂CO, 600 MHz) δ 7.31 (s, 2H), 7.27–7.14 (m, 8H), 4.93 (dd, 1H_A, J = 4.8, 11.4 Hz), 4.73 (dd, 1H_B, J = 4.2, 11.4 Hz), 3.97–3.91 (m, 4H), 3.31–3.24 (m, 2H), 3.12–3.04 (m, 2H), 2.71 (s, 3H_B), 2.71 (s, 3H_A), 2.08 (d, 2H, J = 0.6 Hz), 2.07–2.06 (m, 2H), 1.41 (s, 18H), 1.34 (s, 9H_A), 1.29 (s, 9H_B); ¹³C NMR ((CD₃)₂CO, 150 MHz) δ 173.7/173.5, 158.5, 157.2/156.6, 140.6/140.4, 140.1/139.8, 131.1, 130.0/129.9, 129.6/129.5, 128.6/128.5, 81.0/80.8, 80.6/80.5, 61.5/61.3, 56.7/56.5, 36.9/36.3, 33.5, 29.5 (3C), 29.4/29.3 (3C); IR (film) ν_{max} 1687s, 1479m, 1450m, 1391m, 1366s, 1251m, 1246s, 1061w, 863w, 769m, 735m cm⁻¹; ESI-TOF HRMS m/z 424.2446 (M + H⁺, C₂₁H₃₄N₃O₆⁺, requires 424.2442); $[\alpha]^{25}_{D}$ +33 (c 0.5, CH₂Cl₂).

(R)-3-(3-((1,2-Bis(tert-butoxycarbonyl)hydrazinyl)methyl)phenyl)-2-(tert-butoxycarbonyl(methyl)amino)propanoic Acid (41)

NaHCO₃ (0.027 g, 0.318 mmol) and di-*tert*-butyl dicarbonate (0.026 mL, 0.117 mmol) were added to a solution of **40** (0.045 g, 0.106 mmol) in 1:1 THF/H₂O (0.265 mL) at 0 °C. The mixture was slowly warmed to room temperature and stirred for 18 h. The solution was then quenched with the addition of water (2 mL) and then acidified to a pH of 2 with aqueous 0.2 N NaHSO₄. The mixture was extracted with EtOAc (6 × 5 mL). The combined organic layers were dried over Na₂SO₄, filtered, and concentrated in vacuo. Column chromatography (5 g SiO₂ 50–100% EtOAc/hexanes) provided **41** (48.0 mg, theoretical 55.5 mg, 87%) as a pale yellow oil (rotomer ratio in CD₃OD A/B = 3:1): ¹H NMR (CD₃OD, 500 MHz) δ 7.28–7.14 (m, 8H), 4.60 (dd, 2H, J = 3.5, 11.0 Hz), 4.55 (br s, 2H), 3.27 (dd, 4H, J = 4.5, 14.5 Hz), 3.05 (dd, 2H, J = 11.5, 13.5 Hz,), 2.71 (s, 3H_B), 2.68 (s, 3H_A), 1.53–1.30 (m, 54H); ¹³C NMR ((CD₃)₂CO, 150 MHz) δ 173.6/173.4, 157.5, 157.3/156.8, 140.4/140.2, 139.8/139.6, 130.7, 130.5/130.4, 129.8/129.6, 128.1/127.9, 82.3/81.8, 81.5/81.4, 81.2/80.9, 62.9, 56.0/55.9, 54.3/54.3, 36.9/36.4, 33.8/33.6, 29.5 (3C), 29.4 (6C); IR (film) ν_{max} 1679s, 1479m, 1452m, 1392s, 1367s, 1252m, 1150s, 1053w, 1024w, 858w, 761m, 736m cm⁻¹; ESI-TOF HRMS m/z 524.2961 (M + H⁺, C₂₆H₄₂N₃O₈⁺, requires 524.2966); $[\alpha]^{25}_{D}$ +42 (c 0.5, CH₂Cl₂).

1-[(3-Methylhydrazino)-D-phenylalanyl|desleucylaglucovancomycin Bis(trifluoroacetate) (15)

The title compound was prepared from 41 (11.1 mg, 0.021 mmol), and dicyclohexylcarbodiimide (2.18 mg, 0.011 mmol) in CH₂Cl₂ (0.15 mL), followed by treatment with NaHCO₃ (1.78 mg, 0.021 mmol), desleucylaglucovancomycin (16, 10.0 mg, 0.009 mmol) in DMF (0.3 mL) according to GP3. The protected intermediate was isolated as a white solid (7.50 mg, theoretical 13.8 mg, 55%) by reversephase HPLC with a gradient of 35–95% MeCN/water with 0.07% TFA ($t_R = 23.0 \text{ min}$), and lyophilized, and was taken directly to the next step without further characterization. The intermediate (3.0 mg, 0.002 mmol) was taken up in 40% TFA in CH₂Cl₂ (0.2 mL) at 0 °C and slowly allowed to room temperature. After 45 min, the volatiles were removed with a gentle stream of N₂ and residual TFA was azetroped with CH₂Cl₂ (3 x 5 mL). The residue was purified by reverse-phase HPLC using a gradient of 5–40% MeCN/water with 0.07% TFA ($t_R = 23.6$ min) and afforded the bis-TFA salt of 15 (2.60 mg, theoretical 2.75 mg, 94%): ¹H NMR (CD₃OD, 600 MHz) δ 8.99 (br s, 1H, NH), 8.23 (br s, 1H, NH), 7.76 (br s, 1H) 7.71 (s, 1H), 7.62 (s, 1H), 7.54–7.50 (m, 2H), 7.35 (d, 1H, J = 7.2 Hz), 7.32 (s, 2H), 7.19 (d, 1H, J = 7.2 Hz), 7.32 (s, 2H), 7.19 (d, 1H, J = 7.2 Hz), 7.35 (d, 1H, J = 7.2 Hz), 7.32 (s, 2H), 7.19 (d, 1H, J = 7.2 Hz), 7.35 (d, 1H, J = 7.= 7.2 Hz), 7.14 (br s, 1H), 7.10 (s, 1H), 6.86 (br s, 1H), 6.73 (d, 1H, J = 6.0 Hz), 6.46 (s, 1H), 6.44 (s, 1H), 6.05 (br s, 1H), 5.88 (br s, 1H), 5.41 (s, 1H), 5.34 (s, 1H), 5.20 (s, 1H), 4.84–4.81 (m, 2H), 4.73 (br s, 1H), 4.33-4.23 (m, 3H), 4.15-4.10 (m, 2H), 2.87 (d, 1H, J = 15.0 Hz), 2.77 (s, 3H), 2.01 (br s, 1H) (two signals buried below residual solvent/water peak); ESI-TOF HRMS m/z 1221.3107 (M + H⁺, $C_{57}H_{55}Cl_2N_{10}O_{17}^+$, requires 1221.3118); $[\alpha]_D^{25} + 17$ (c 0.2, CH₃OH).

(R)-4-(Benzyloxy)-2-(tert-butoxycarbonyl(methyl)amino)butanoic Acid (43)

The title compound was prepared from Boc-D-*O*-benzylhomoserine (**42**, 1.00 g, 3.23 mmol), NaH (426 mg, 10.6 mmol), methyl iodide (0.603 mL, 9.69 mmol) in anhydrous THF (10.7 mL), and DMF (0.514 mL) according to *GP1*. Column chromatography (100 g SiO₂, 0–0.2 % AcOH/EtOAc) afforded **43** (1.00 g, theoretical 1.04 g, 96%) as a white solid (rotomer ratio in acetone-D₆ A/B = 1:1): 1 H NMR (CDCl₃, 600 MHz) δ 7.36–7.28 (m, 10H), 4.70 (dd, 1H, J = 4.8, 8.4 Hz), 4.51 (s, 4H), 4.48 (dd, 1H, J = 7.8, 9.0 Hz), 3.60–3.56 (m, 2H), 3.52–3.46 (m, 2H), 2.89 (s, 3H_A), 2.84 (s, 3H_B), 2.36–2.30 (m, 2H), 2.10–2.01 (m, 2H), 1.46 (s, 9H_A), 1.41 (s, 9H_B) (CO₂H not observed); 13 C NMR (CDCl₃, 150 MHz) δ 177.0/175.9, 156.5/155.4, 137.9, 128.4 (2C), 127.7 (2C), 127.6, 80.7/80.6, 73.2/73.1, 66.8/66.3, 57.3/57.0, 33.3/32.8, 29.8/29.0, 28.3/28.3 (3C); IR (neat) ν_{max} 1738m, 1696s, 1480m, 1453m, 1392m, 1367m, 1151s, 1103m cm⁻¹; ESI-TOF HRMS m/z 346.1625 (M + Na⁺, C₁₇H₂₅NNaO₅⁺, requires 346.1625); $\left[\alpha\right]^{25}_{\text{D}}$ +22 (c 1.0, CH₂Cl₂).

(R)-2-(tert-Butoxycarbonyl(methyl)amino)-4-(2-(tert-butoxycarbonyl)hydrazono)butanoic Acid (44)

An O₂ degassed solution of 43 (500 mg, 1.54 mmol) in EtOAc (15.4 mL) with 10% Pd/C (100 mg) was placed under an atmosphere of H₂ gas (balloon) and the mixture was vigorously stirred at room temperature. After 18 h, the solution was filtered over Celite. The filtrate was concentrated in vacuo to afford the primary alcohol as a mixture with lactone (358 mg, theoretical 359 mg, 99%). The residue (300.0 mg, 1.28 mmol) was dissolved in anhydrous CH₂Cl₂ (12.8 mL) and Dess-Martin periodinane (600 mg, 1.41 mmol) was added and the mixture was vigorously stirred for 2 h at room temperature. The volatiles were removed in vacuo and the residue was diluted in CH₂Cl₂ (2 mL). The solid precipitate was removed by filtration, and the filtrate was concentrated in vacuo. The crude aldehyde was dissolved in anhydrous THF (1.35 mL) and tert-butyl carbazate (170 mg, 1.28 mmol) in anhydrous THF (1.35 mL) was added. After 16 h, the volatiles were removed in vacuo and the residue was partitioned between water (15 mL) and EtOAc (30 mL). The aqueous phase was acidified to a pH of 2 with aqueous 0.2 N NaHSO₄, and extracted with EtOAc (15 mL). The organic phase were dried over Na₂SO₄, filtered, and concentrated. Column chromatography (15 g SiO₂, 70–100% EtOAc/hexanes) provided 44 (210 mg, theoretical 441 mg, 48%) as a white foam (rotomer ratio in acetone-D₆ A/B = 1:1): ${}^{1}H$ NMR ((CD₃)₂CO, 600 MHz) δ 9.53 (br s, 2H), 7.44 (br s, 1H_A), 7.38 (br s, 1H_B), 4.86 (dd, 1H_A), J = 4.8, 9.6 Hz), 4.63 (dd, 1H_B, J = 4.2, 9.0 Hz), 2.89 (s, 3H_A), 2.86 (s, 3H_B), 2.78–2.69 (m, 4H), 1.44 (s, 18H), 1.42 (s, 9H_A), 1.41 (s, 9H_B) (CO₂H not observed); 13 C NMR ((CD₃)₂CO, 150 MHz) δ 173.3/173.2, 157.5/156.6, 154.2 (br), 145.1 (br), 81.3/81.1, 81.0, 59.7/58.3, 34.6, 33.9/33.3, 29.5 (3C), 29.4 (3C); IR (neat) v_{max} 1678s (br), 1529w, 1480w, 1453w, 1391m, 1367m, 1328w, 1247m, 1148s, 1045w, 1017w, 859w, 770w, 734s cm⁻¹; HR ESI-TOF m/z 368.1797 (M + Na⁺, C₁₅H₂₇N₃NaO₆⁺, requires 368.1792); $\left[\alpha\right]^{25}_{D} + 14$ (c 0.5, CH₂Cl₂).

(R)-2-(tert-Butoxycarbonyl(methyl)amino)-4-(2-(tert-butoxycarbonyl)hydrazinyl)butanoic Acid (45)

An O₂ degassed solution of **44** (104 mg, 0.301 mmol) in methanol (3.0 mL) was treated with *tert*-butylcarbazate (79.6 mg, 0.602 mmol). The solution was degassed and backfilled with N₂, and 5% Pt/C (50% wt., 52 mg) was added. The mixture was subsequently degassed and backfilled with N₂ then placed under an atmosphere of H₂ (balloon) and vigorously stirred. After 18 h, the solids were removed by filtration over Celite and the filtrate was concentrated in vacuo. Column chromatography (5 g SiO₂, 70–100% EtOAc/hexanes) provided **45** (79.8 mg, theoretical 104 mg, 76%) as a clear colorless oil (rotomer ratio in (CD₃)₂CO A/B = 1:1): ¹H NMR ((CD₃)₂CO, 500 MHz) δ 4.81 (dd, 1H_A, J = 4.5, 10.0 Hz), 4.55 (dd, 1H_B J = 4.5, 10.0 Hz), 2.93–2.78 (m, 12H), 2.14–2.05 (m, 6H), 1.94–1.87 (m, 2H), 1.45–1.42 (m, 36H) (CO₂H not observed); ¹³C NMR (CD₃OD, 150 MHz) δ 175.0/174.9, 158.9, 158.0/157.6, 81.9/81.5, 80.9, 59.6/57.8, 33.6, 32.1, 28.7/28.6 (3C), 28.5/27.9 (3C), 20.8; IR (neat) ν_{max} 3293w (br),

1680s, 1478m, 1452m, 1391m, 1367m, 1250m, 1146s, 865m, 773m cm⁻¹; HR ESI-TOF m/z 348.2131 (M + H⁺, C₁₅H₃₀N₃O₆⁺, requires 348.2129); $\left[\alpha\right]^{25}$ _D +59 (c 0.5, CH₃OH).

(R)-4-(1,2-Bis(tert-butoxycarbonyl)hydrazinyl)-2-(tert-butoxycarbonyl(methyl)amino)butanoic Acid (46)

Di-*tert*-butyl dicarbonate (58.0 μ L, 0.253 mmol) was added dropwise to a solution of **45** (79.8 mg, 0.229 mmol) in 1:1 THF/H₂O (0.573 mL) at 0 °C. The solution was warmed to room temperature and stirred for 16 h. The mixture was partitioned between water and EtOAc and the aqueous was washed with EtOAc (6 × 5 mL). The combined organic layers were dried over Na₂SO₄, filtered and concentrated. Column chromatography (5 g SiO₂, 30–50% EtOAc/hexanes) provided **46** (49.2 mg, theoretical yield 102 mg, 48%) as a white foam (rotomer ratio in acetone-D₆ A/B = 1:1): mp 148–149 °C; ¹H NMR ((CD₃)₂CO, 600 MHz) δ 8.14–7.51 (br m, 2H), 4.67 (d, 1H_A, J = 7.2 Hz), 4.38 (d, 1H_B, J = 9.0 Hz), 3.61–3.30 (m, 4H), 2.87 (s, 3H_B), 2.85 (s, 3H_A), 2.29 (br s, 2H), 2.03 (br s, 2H), 1.46 (s, 18H), 1.45 (s, 18H) 1.42 (s, 18H); ¹³C NMR ((CD₃)₂CO, 150 MHz) δ 174.0, 157.7, 157.2/157.0, 156.9/156.8/156.8/156.7/156.6, 82.1/81.7/81.6, 81.5, 81.11/81.07, 59.5/57.8, 50.1/50.0, 48.4/48.0, 34.2/32.9, 29.6 (3C), 29.5 (3C), 29.4 (3C); IR (neat) ν_{max} 3294 ν_{max} 3294 ν_{max} 1453 ν_{max} 1392 ν_{max} 1367 ν_{max} 1150 ν_{max} 1150

1-[(R)-4-(Hydrazinyl-2-methylamino)butanoyl]desleucylaglucovancomycin Bis(trifluoroacetate) Salt (14)

The title compound was prepared from **46** (9.50 mg, 0.021 mmol), and dicyclohexylcarbodiimide (2.19 mg, 0.011 mmol) in CH₂Cl₂ (0.151 mL), followed by treatment with NaHCO₃ (1.78 mg, 0.021 mmol), desleucylaglucovancomycin (**16**, 10.0 mg, 0.009 mmol) in DMF (0.3 mL) according to *GP3*. The protected intermediate was isolated as a white solid (10.4 mg, theoretical 12.8 mg, 81%) by reverse-phase HPLC with a gradient of 35–75% MeCN/water with 0.07% TFA (t_R = 18.0 min), and lyophilization, and was taken directly to the next step without further characterization. The intermediate (5.00 mg, 0.004 mmol) was taken up in 40% TFA in CH₂Cl₂ (0.280 mL) and stirred at 0 °C for 5 min then gently warmed to room temperature. After 45 min, the volatiles were removed with a slowly stream of N₂ and residual TFA was azetroped with CH₂Cl₂ (3 × 5 mL). The residue was purified by reverse-phase HPLC using a gradient of 5–40% MeCN/water with 0.07% TFA (t_R = 23.4 min) and afforded the bis-TFA salt of **14** (2.90 mg, theoretical 4.80 mg, 60%): ¹H NMR (CD₃OD, 600 MHz) δ 9.01 (d, 1H, NH, J = 4.2 Hz), 8.79 (br s, 1H, NH), 7.67–7.57 (m, 4H), 7.38 (br s, 1H), 7.24 (d, 1H, J =

8.4 Hz), 7.05 (s, 1H), 6.86 (br s, 1H), 6.87 (d, 1H, J = 8.4 Hz), 6.46 (s, 1H), 4.32 (s, 1H), 5.90 (br s, 1H), 5.81 (br s, 1H), 5.38–5.34 (m, 4H), 4.74 (s, 1H), 4.60 (br s, 1H), 4.46 (br s, 1H), 4.15 (s, 1H), 4.10 (t, 1H, J = 6.3 Hz), 3.10–3.08 (m, 1H), 2.97 (d, 1H, J = 15.6 Hz), 2.81 (s, 3H), 2.27 (br s, 1H), 2.18 (br s, 2H) (one signal buried under residual solvent/water peak); ESI-TOF HRMS m/z 1145.2782 (M + H⁺, $C_{51}H_{51}Cl_2N_{10}O_{17}^+$, requires 1145.2805); $[\alpha]^{25}_D$ +30 (c 0.2, CH_3OH).

Table 2SI. Purity Analysis 6–15.^a

Compound	Gradient (Solvent A/B) ^b	Percent purity
6	5-40% A/B(30 min)	94%
7	5-40% A/B(30 min)	94%
8	5-40% A/B(30 min)	94%
9	5-40% A/B(30 min)	93%
10	5-40% A/B(30 min)	96%
11	5-40% A/B(30 min)	99%
12	5-40% A/B(30 min)	96%
13	5-40% A/B(30 min)	90%
14	5-40% A/B(30 min)	93%
15	5-40% A/B(30 min)	86%

^a Purity of each compound was determined on a preparative reverse-phase WatersTM Nova-Pac[®] HR C18 6 μm, 60 Å column (25 x 100 mm) using a WatersTM 600 HPLC with a WatersTM Photodiode Array Detector. A gradient of acetonitrile/water (containing 0.07% TFA) at flow rate of 5 mL min⁻¹ was used for all samples with detection at 220, 254, 280 nm. ^b solvent A = acetonitrile, solvent B = water with 0.07% TFA.

Supporting Information References

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