

SUPPLEMENTARY INFORMATION

Photochemical Degradation of Marbofloxacin and Enrofloxacin in Natural Waters

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Figure S1. Representative variation of the solar radiation intensity during the day, both in summer and

in winter. Figure S2. Comparison of photoproducts from ENR under solar and artificial light

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(9 pages)

Reagents and Materials.

MAR and ENR were supplied by Fluka (Sigma-Aldrich), acetonitrile (ACN) by VWR, H₃PO₄ (85% w/w) by Carlo Erba and ultra-pure water from a Millipore Milli-Q system. Humic acid sodium salt (MW=100,000-150,000, Aldrich) was used to quantify aquatic humic acids (HAs, see 31). FQs stock solutions of 300 µg mL⁻¹ in methanol 0.1% v/v NaOH 1 M were prepared under red light and stored at 4 °C for <3 months. Working solutions of 30 µg mL⁻¹ in 25 mM H₃PO₄ were stored at 4 °C and renewed weekly.

CaSO₄ (99%, Sigma-Aldrich), NaCl (100.1%, J.T. Baker), MgNO₃ hexahydrate (97%, Sigma-Aldrich), K₂HPO₄ (≥ 99.0%, Sigma-Aldrich) and KH₂PO₄ (99.5%, Merck) salts were used. Ultra-pure HCl acid (37% w/w) and NaOH 0.1 M solution prepared from NaOH anhydrous pellets (97%, Carlo Erba) were employed for pH correction.

Details of Analytical Determination.

The HPLC system consisted of a pump Series 200 (Perkin Elmer) equipped with vacuum degasser, programmable fluorescence detector (FD) and diode array detector (UV) Series 200 (Perkin Elmer). The FD excitation/emission wavelengths selected were 297/507 nm for MAR and 280/450 nm for ENR. After an equilibration period of 5 min, 50 µL of each sample were injected into a 250 × 4.6 mm, 5 µm Ascentis RP-Amide (Supelco) coupled with a similar guard-column. The mobile phase was 25 mM H₃PO₄-ACN (85:15) at a flow rate of 1 mL min⁻¹.

Preparative HPLC experiments were performed with the same HPLC system on a 250 x 10 mm, 5 µm Inertsil ODS-2 (GL Sciences Inc.) preceded by a similar guard-column. Mobile phase was H₂O (pH adjusted to 2.5 with HCl)-ACN (90:10) at a flow rate of 4 mL min⁻¹.

LC-MS analysis was performed by using an Agilent 1100 HPLC with a Gemini C18 (250 x 4.6 mm, 5 µm) column, maintained at 30 °C. A gradient was used for the mobile phase (solvent A: formic acid

0.5% v/v in ultra-pure water; solvent B: ACN) as follows: 15% B until 10 min, 20 % B from 10 to 12 min and 0% B until 1 min, 60% B from 1 to 50 min, for ENR and MAR, respectively. The flow rate was 1.2 mL min⁻¹ and the injection volume was 5 µL. The MS-system consisted of a linear trap Thermo LXQ.

A DX 500 Dionex Ion Chromatograph equipped with a GP40 gradient pump, CD20 conductivity detector and anion self-regenerating suppressor (ASRS 400, 4 mm) has been used for the determination of anions content in tap, ditch and river water. 70 µL of each sample were injected into a 250 x 4 mm IonPac AS23 coupled with a AG23 50 x 4 mm guard-column. The eluent was 0.8 mM NaHCO₃-4.5 mM Na₂CO₃ at a flow rate of 1 mL min⁻¹.

A Perkin Elmer ICP-OES Optima 3300 DV was used for calcium and magnesium determination, following the operating conditions suggested by the manufacturer.

The pH was monitored with a combined Orion glass electrode 9102 SC, standardized in H⁺ activity. ¹H-NMR, ¹³C-NMR and ¹³C-DEPT spectra were acquired on a Bruker Avance 300 MHz spectrometer and the chemical shifts are reported relative to TMS.

Table S3. Ions Concentrations Determined in Tap and River Ticino

Ion	Concentration (mg L ⁻¹)	
	Tap water	River Ticino water
Calcium	35.0	37.0
Magnesium	10.0	7.6
Chloride	4.8	10.5
Phosphate	< 0.2	< 0.2
Nitrate	0.6	8.5
Sulphate	4.4	33.2

Table S1. Mass Spectroscopic Data of the Photoproducts of ENR

Fragment	HPLC/ESI-MS/MS											
	comp. D		comp. C		comp. A		comp. B		comp. E		ENR	
	m/e	int. ^a	m/e	int.	m/e	int.	m/e	int.	m/e	int.	m/e	int.
[M+1] ⁺	358.2	10	316.3	80	342.3	15	334.3	25	374.2	100	360.3	10
[M+1-HF] ⁺	-	-	-	-	-	-	314.3	10	-	-	-	-
[M+1-H ₂ O] ⁺	340.3	5	298.3	20	324.3	5	-	-	356.3	70	-	-
[M+1-CO ₂] ⁺	314.3	100	-	-	298.3	100	-	-	-	-	316.3	100
[M+1-C ₂ H ₇ N] ⁺	-	-	271.2	25	-	-	289.2	10	-	-	-	-
[M+1-C ₄ H ₉ N] ⁺	-	-	245.2	100	-	-	263.2	100	-	-	-	-
[M+1-C ₅ H ₉ NO ₂] ⁺	-	-	-	-	-	-	219.2	15	-	-	-	-

^a Relative percent.

Table S4. ^1H -NMR Signals of ENR Photodegradation Product D in DMSO-d6

Signal	^1H - NMR		Signal	^1H - NMR	
	comp. D	comp. D		comp. D	comp. D
H_A	n.p.	3	H_G	n.p.	1
	δ [ppm]	1.2		δ [ppm]	8.6
	M	m		m	s
$\text{H}_\text{B},$ $\text{H}_\text{C,C'}$	n.p.	6	H_H	n.p.	1
	δ [ppm]	3.1-3.2		δ [ppm]	3.8
	M	m		m	m
$\text{H}_\text{D,D'}$	n.p.	4	$\text{H}_\text{I}, \text{H}_\text{J}$	n.p.	4
	δ [ppm]	3.6		δ [ppm]	1.3
	M	bs		m	m
H_E	n.p.	1	H_K	n.p.	1
	δ [ppm]	7.7		δ [ppm]	10.8
	M	s		m	s
H_F	n.p.	1	H_L	n.p.	1
	δ [ppm]	7.5		δ [ppm]	10.5
	M	s		m	bs

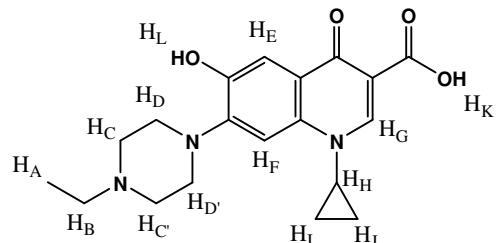


Table S5. ^{13}C -NMR Signals of ENR Photodegradation Product D in DMSO-d6

Signal	$^{13}\text{C-NMR}$ comp. D		Signal	$^{13}\text{C-NMR}$ comp. D		
	C	c.t.		C	c.t.	
C ₁	c.t.	C	C ₁₀	c.t.	C	
		δ [ppm]	135.9		δ [ppm]	176.1
C ₂	c.t.	C	C ₁₃	c.t.	C	
		δ [ppm]	148.9		δ [ppm]	35.7
C ₃	c.t.	CH	C ₁₄₋₁₅	c.t.	2xCH ₂	
		δ [ppm]	105.7		δ [ppm]	7.5
C ₄	c.t.	C	C ₁₇₋₂₁	c.t.	2xCH ₂	
		δ [ppm]	144.8		δ [ppm]	50.7
C ₅	c.t.	C	C ₁₈₋₂₀	c.t.	2xCH ₂	
		δ [ppm]	120.2		δ [ppm]	50.2
C ₆	c.t.	CH	C ₂₂	c.t.	CH ₂	
		δ [ppm]	108.6		δ [ppm]	46.0
C ₈	c.t.	CH	C ₂₃	c.t.	CH ₃	
		δ [ppm]	146.0		δ [ppm]	8.9
C ₉	c.t.	C	C ₂₄	c.t.	C	
		δ [ppm]	106.2		δ [ppm]	166.4

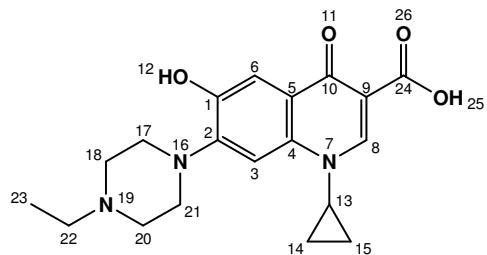


Table S2. Mass Spectroscopic Data of the Photoproducts of MAR

Fragment	HPLC/ESI-MS/MS					
	comp. A		comp. B		MAR	
	m/e	int. ^a	m/e	int.	m/e	int.
[M+1] ⁺	641.5	1	322.3	5	363.1	100
[M+1-H ₂ O] ⁺	623.4	100	304.3	75		
[M+1-CO ₂] ⁺	-	-	278.3	100		
[M+1-C ₄ H ₇ NO ₂] ⁺	-	-	221.2			

^a Relative percent.

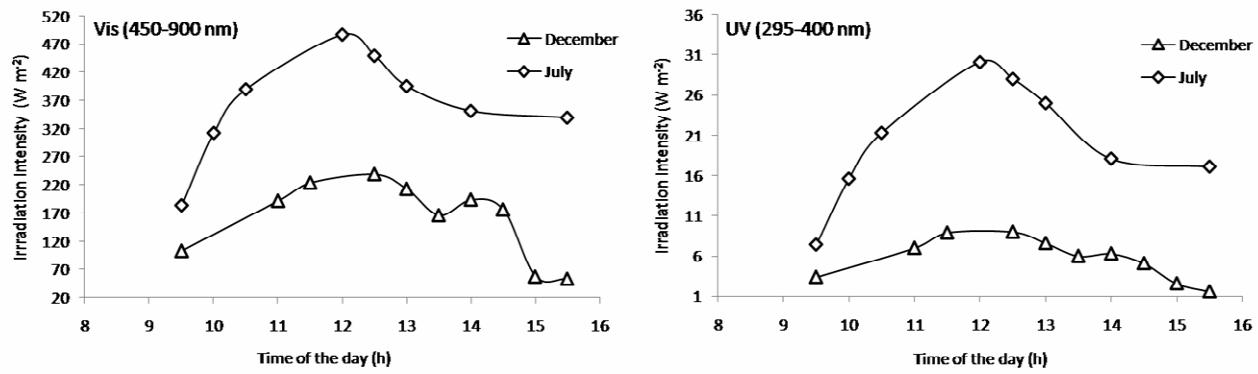


FIGURE S1. Irradiation intensity (W m^{-2}) measured in two different seasons under natural sunlight conditions.

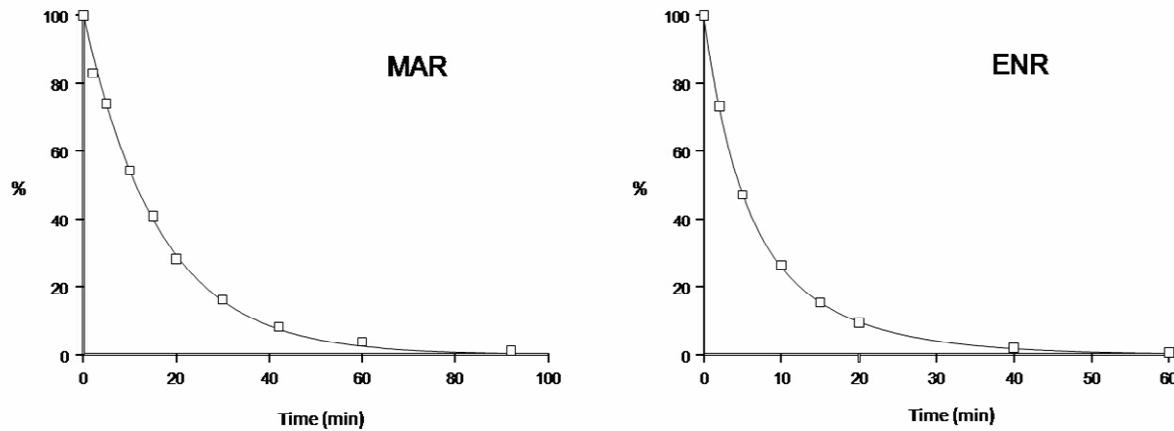


FIGURE S3. Photodegradation profiles obtained by solar-simulated irradiation of MAR and ENR in tap water samples (500 mL, pH 7.9) enriched with $50 \mu\text{g L}^{-1}$ of either FQs.

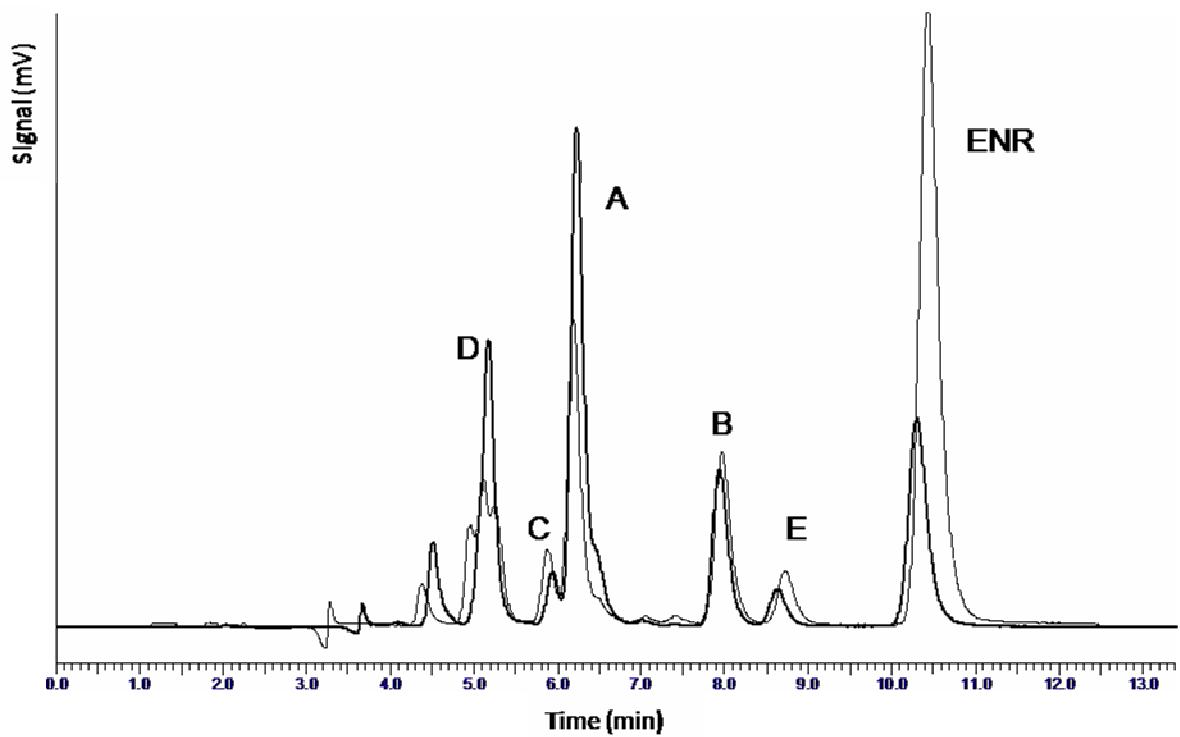


FIGURE S2. FD chromatograms obtained by irradiation of a tap water sample enriched with $50 \mu\text{g L}^{-1}$ of ENR under natural solar light for 15 min (bold line) and under mercury lamp light (315 nm, 200 W) for 10 min (thin line).