Supporting Information:

Pd-Catalyzed Asymmetric Hydrogenation of Unprotected Indoles Activated by Brønsted Acids

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1. General and Materials

General: All reactions were carried out under an atmosphere of nitrogen using standard Schlenk techniques, unless otherwise noted. ¹H NMR and ¹³C NMR spectra were recorded on Bruker DRX-400 spectrometers. The chemical shifts for ¹H NMR were recorded in ppm downfield from tetramethylsilane (TMS) with the solvent resonance as the internal standard. The chemical shifts for ¹³C NMR were recorded in ppm downfield using the central peak of deuterochloroform (77.23 ppm) as the internal standard. Coupling constants (*J*) are reported in Hz and refer to apparent peak multiplications. TLC analysis was performed using glass-backed plates coated with 0.2 mm silica. Quantitative analysis was performed by ¹H NMR on Bruker DRX 400 instrument. Flash column chromatography was performed on silica gel (200-300 mesh). Enantiomeric excess was determined by HPLC analysis, using chiral column described below in detail. Optical rotations were measured with JASCO P-1010 polarimeter. The configuration was determined by comparison of rotation sign with the literature data or by analogue.

Materials: Commercially available reagents were used throughout without further purification other than those detailed below. Acetone was dried with anhydrous CaSO₄ and distilled over KMnO₄. The solvents for asymmetric hydrogenation reaction were purchased without further purification.

2. Typical Procedure for the Synthesis of Indoles

Indoles **1b-f**^{1a,1b} and **1o-p**^{1c,1d} were synthesized according to the literature procedure. 2-Benzyl substituted indoles **1g-k** were prepared following modified known methods. ^{1e,1f} Procedure for the synthesis of unknown compounds **1h-k** were depicted as follows.

To a solution of indole (20 mmol, 2.343 g) in ethanol (20 mL) at room temperature, KOH pellets were added (25 mmol, 1.403 g), and the mixture was stirred until total solubilization. The ethanol was completely removed in vacuum and acetone (20 mL) added followed by the benzyl bromide (20 mmol). A precipitate was formed instantly. The solid was filtered and the solution concentrated in vacuum to give *N*-benzylindole which was purified by flash chromatography.

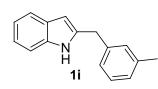
The *N*-benzylindole (4.2 mmol) was added to polyphosphoric acid (50 mL), which was prewarmed to the desired reaction temperature and vigorously stirred. Stirring and heating was continued until the starting material was consumed as indicated by TLC. Then the dark reaction mixture was cooled in an ice bath and mated carefully with ice water (40 mL). Extraction with EtOAc (3×30 mL) and washing the combined organic layers with saturated aqueous sodium bicarbonate solution and brine (30 mL each) was followed by drying over sodium sulfate. The solvent was removed under reduced pressure, and the residue was purified by flash chromatography to give **1h-k**.

2-(2'-Methylbenzyl)indole (1h). Yellow solid, mp = 72-75 °C; 1 H NMR (400 MHz, CDCl₃) δ

N H 1h 2.26 (s, 3H), 4.07 (s, 2H), 6.23 (s, 1H), 7.03-7.11 (m, 2H), 7.16-7.21 (m, 5H), 7.51 (d, J = 7.3 Hz, 1H), 7.64 (br, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 19.6, 32.8, 101.1, 110.6, 119.9, 120.1, 121.3, 126.5, 127.3, 128.9, 129.7, 130.8, 136.2, 136.7, 137.1, 137.4; HRMS Calculated for

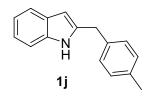
 $C_{16}H_{14}N$ [M-H]⁻ 220.1126, found 220.1117.

2-(3'-Methylbenzyl)indole (1i). Yellow solid, mp = 69-70 °C; 1 H NMR (400 MHz, CDCl₃) δ



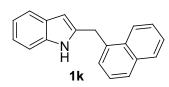
2.32 (s, 3H), 4.08 (s, 2H), 6.32 (s, 1H), 7.05-7.12 (m, 5H), 7.19-7.23 (m, 2H), 7.54 (d, J = 7.4 Hz, 1H), 7.73 (br, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 21.6, 34.8, 101.2, 110.7, 119.9, 120.2, 121.4, 126.1, 127.7, 128.8, 129.8, 136.4, 138.1, 138.6; HRMS Calculated for C₁₆H₁₄N [M-H] 220.1126, found 220.1134.

2-(4'-Methylbenzyl)indole (1j). Yellow solid, mp = 91-93 °C; 1 H NMR (400 MHz, CDCl₃) δ



2.32 (s, 3H), 4.05 (s, 2H), 6.29 (s, 1H), 7.03-7.13 (m, 6H), 7.19 (d, J = 7.9 Hz, 1H), 7.52 (d, J = 7.4 Hz, 1H), 7.67 (br, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 21.2, 34.5, 101.1, 110.7, 119.9, 120.2, 121.4, 128.9, 129.6, 135.6, 136.4, 136.5, 138.3; HRMS Calculated for C₁₆H₁₄N [M-H]⁻ 220.1126, found 220.1130.

2-(1'-Naphthylmethyl)indole (1k). Yellow solid, mp = 126-128 °C; ¹H NMR (400 MHz, CDCl₃)



δ 4.58 (s, 2H), 6.39 (s, 1H), 7.09-7.11 (m, 2H), 7.18-7.19 (m, 1H), 7.39-7.56 (m, 5H), 7.72 (br, 1H), 7.85 (d, J = 8.1 Hz, 1H), 7.92 (d, J = 7.4 Hz, 1H), 8.09 (d, J = 8.6 Hz, 1H); 13 C NMR (100 MHz, CDCl₃) δ 32.5, 101.0, 110.7, 119.9, 120.1, 121.4, 124.1, 125.8, 126.1, 126.6, 127.3, 128.0, 128.9, 132.2, 134.2, 134.4, 136.2,

137.7; HRMS Calculated for C₁₉H₁₄N [M-H]² 256.1126, found 256.1129.

3. The Effect of Acids and Ligands on the Reactivity and Enantioselectivity

The effect of various Brønsted acids on the reactivity and enantioselectivity was achieved for the asymmetric hydrogenation of simple 2-methylindole **1a** (Table 1, entries 1-10). It was found that this reaction was strongly Brønsted acid-dependent, strong acids gave full conversion and moderate enantioselectivity whereas weak acids showed poor reactivity and enantioselectivity (entries 3-6 *vs* entries 8-9). Without the addition of acid, the reaction did not occur (entry 1). L-CSA gave full conversion and the highest enantioselectivity (entry 6, 71% ee). In contrast, D-CSA with opposite configuration gave product with somewhat lower ee and 44% conversion (entry 7, 66% ee). Subequivalent L-CSA resulted in deteriorated enantioselectivity and activity. Therefore, L-CSA was selected as the activator for further study.

Table 1. The Effect of Brønsted Acids on the Reactivity and Enantioselectivity^a

Pd(OCOCF₃) / (R)-SegPhos

Acid (1 equiv.), TFE

H₂ (700 psi), RT

entry acid^b convn.
$$(\%)^c$$
 ee $(\%)^d$

entry	acid^b	convn. (%) ^c	ee (%) ^d
1 ^e	no	<5	-
2	TfOH	61	54 (R)
3	PhSO ₃ H	>95	62 (R)
4	$TsOH \cdot H_2O$	>95	69 (R)
5	TFA	>95	8 (R)
6	L-CSA	>95	71 (R)
7	D-CSA	44	66 (R)
8	PhCO ₂ H	<5	-
9	Salicylic acid	35	4 (R)
10^f	L-CSA	94	63 (R)

^a Conditions: 0.25 mmol **1a**, Pd(OCOCF₃)₂ (2 mol %), (*R*)-SegPhos (2.4 mol %), Acid (1.0 equiv.), 3 mL TFE, 24 h, RT. ^b TfOH: Triflic acid; PhSO₃H: Benzenesulfonic acid; TsOH·H₂O: *p*-Toluenesulfonic acid monohydrate; TFA: trifluoroacetic acid; L-CSA: L-(-)-Camphorsulfonic acid; D-CSA: D-(+)-Camphorsulfonic acid. ^c Determined by ¹H-NMR. ^d Determined by HPLC. ^e with no acid. ^f with 0.9 equiv. L-CSA.

Next, the effect of chiral ligands on the reactivity and enantioselectivity was examined for the asymmetric hydrogenation of simple 2-methylindole **1a**. Axially chiral bisphosphine ligands were effective, full conversions and high to excellent enantioselectivities were obtained (Table 2, entry 1-6, 84-91% ee). Ferrocene-based bisphosphine ligand (JosiPhos-type) displayed moderate activity and excellent enantioselectivity (entry 7, 80% ee). Me-DuPhos showed no activity for this transformation (entry 8). Subsequently, higher temperature was applied for the activity. At 50 °C, the catalyst loading could be reduced to 0.5% but with lower enantioselectivity (entry 9, 84% ee *vs* 91% ee).

Table 2. The Effect of Chiral Ligands on the Reactivity and Enantioselectivity^a

entry	ligand	convn. $(\%)^b$	ee (%) ^c
1	(R)-SegPhos	>95	85 (R)
2	(R)-SynPhos	>95	86 (R)
3	(R)-MeOBiPhep	>95	84 (R)
4	(R)-C4-TunePhos	>95	84 (R)
5	(R)-BINAP	>95	85 (R)
6	(<i>R</i>)-H8-BINAP	>95	91 (R)
7	(R)- (S) -PPF-PCy ₂ -JosiPhos	49	80 (R)
8	(R,R)-Me-DuPhos	<5	-
9^d	(R)-H8-BINAP	>95	84 (R)
	Ph ₂ OPPh ₂ MeOPh ₂ PPh ₂ MeO	PPh ₂ PPh ₂	PPh ₂
(<i>R</i>)-SegPho	s (<i>R</i>)-SynPhos (<i>R</i>)-N	MeO-BiPhep (R)	-C4-TunePhos
	PPh ₂ PPh ₂ Ph ₂ P	PCy ₂ Fe H .C ₂ H ₅ OH	P
(<i>R</i>)-BINAP	(<i>R</i>)-H8-BINAP (<i>R</i>)-(<i>S</i>)-PP	F-PCy ₂ -JosiPhos (R,I	R)-Me-DuPhos

 a Conditions: 0.25 mmol **1a**, Pd(OCOCF₃)₂ (2 mol %), Ligand (2.4 mol %), L-CSA (1.0 equiv.), 3 mL TFE, 24 h, RT. b Determined by 1 H-NMR. c Determined by HPLC. d With oil bath of 50 o C and 0.5 mol % catalyst.

4. Typical Procedure for Pd-catalyzed Asymmetric Hydrogenation of Unprotected Indoles

$$R^{1} \stackrel{||}{\underset{H}{||}} R^{2} \xrightarrow{R^{2}} R^{3} \xrightarrow{Pd(OCOCF_{3})_{2} / (R)-H8-BINAP} R^{1} \stackrel{||}{\underset{H}{||}} R^{2}$$

$$R^{1} \stackrel{||}{\underset{H}{||}} R^{3} \xrightarrow{R^{2}} R^{2}$$

(*R*)-H8-BINAP (3.8 mg, 0.006 mmol) and Pd(OCOCF₃)₂ (1.7 mg, 0.005 mmol) were placed in a dried Schlenk tube under nitrogen atmosphere, and degassed anhydrous acetone was added. The mixture was stirred at rt for 1 h, then solvent was removed under vacuum to give the catalyst. In a glovebox, L-CSA (0.25 mmol) and substrate 1 (0.25 mmol) were stirred in 1 mL solvent (DCM and TFE were mixed in ratio of 1:1 before use) at room temperature for 5 min. Subsequently, the above catalyst together with 2 mL solvent was added to the reaction mixture. The hydrogenation was performed at room temperature under H₂ (700 psi) in a stainless steel autoclave for 24 h. After carefully releasing the hydrogen, the resulting mixture was concentrated under vacuum and dissolved in saturated aqueous NaHCO₃ (5 mL). After stirring for 10 min, the mixture was extracted with CH₂Cl₂ (3×5 mL) and dried over Na₂SO₄. After purified by silica gel chromatography using petroleum ether/EtOAc (10/1) as eluent, the enantiomeric excess of the products were determined by HPLC with chiral columns (OD-H, OJ-H or IC).

Racemates of $\mathbf{2}$ were prepared by the hydrogenation of the indoles catalyzed by $Pd(OCOCF_3)_2/(+/-)$ -SegPhos in TFE or reduced by refluxing in EtOH with Sn/HCl.²

(+)-(*R*)-2-Methylindoline (2a).³ 88% yield, 91% ee, $[\alpha]^{RT}_{D} = +6.96$ (c 0.63, benzene), {Lit. $[\alpha]^{RT}_{D} = -12.2$ (c 2.60, benzene) for (*S*)-enantiomer}³; ¹H NMR (400 MHz, CDCl₃) δ 1.30 (d, J = 6.2 Hz, 3H), 2.64 (dd, J = 15.4, 7.8 Hz, 1H), 3.15 (dd, J = 15.4, 4.4 Hz, 1H), 3.76 (br, 1H), 3.90-4.00 (m, 1H), 6.61 (d, J = 7.8 Hz, 1H), 6.69 (t, J = 7.4 Hz, 1H), 7.01 (t, J = 7.6 Hz, 1H), 7.08 (d, J = 7.3 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 22.5, 38.0, 55.4, 109.4, 118.7, 124.9, 127.4, 129.1, 151.1; HPLC (OD-H, elute: Hexanes/*i*-PrOH = 97/3, detector: 254 nm, flow rate: 0.8 mL/min), (*R*) $t_1 = 9.3$ min, (*S*) $t_2 = 10.1$ min.

(+)-(*R*)-2-Butylindoline (2b).⁴ 82% yield, 93% ee, $[\alpha]^{RT}_{D} = +12.6$ (c 1.1, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 0.95 (t, J = 6.6 Hz, 3H), 1.36-1.40 (m, 4H), 1.63 (q, J = 6.2 Hz, 2H), 2.69 (dd, J = 15.4, 8.4 Hz, 1H), 3.14 (dd, J = 15.4, 8.6 Hz, 1H), 3.81-3.89 (m, 2H), 6.62 (d, J = 7.7 Hz, 1H), 6.70 (t, J = 7.3 Hz, 1H), 7.02 (t, J = 7.6 Hz, 1H), 7.09 (d, J = 7.2 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 14.3, 23.0, 29.0, 36.3, 36.7, 60.3, 109.3, 118.6, 124.8, 127.4, 129.1, 151.2; HPLC (OD-H, elute: Hexanes/*i*-PrOH = 99/1, detector: 254 nm, flow rate: 1.0 mL/min), t_1 = 6.5 min, t_2 = 7.9 min.

(+)-(*R*)-2-Pentylindoline (2c). 89% yield, 92% *ee*, $[\alpha]^{RT}_{D} = +15.5$ (*c* 1.40, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 0.93 (t, J = 6.4 Hz, 3H), 1.35-1.43 (m, 6H), 1.59-1.64 (m, 2H) 2.69 (dd, J = 15.4, 8.5 Hz, 1H), 3.14 (dd, J = 15.4, 8.6 Hz, 1H), 3.83-3.89 (m, 2H), 6.61 (d, J = 7.7 Hz, 1H), 6.70 (t, J = 15.4)

7.4 Hz, 1H), 7.02 (t, J = 7.6 Hz, 1H), 7.09 (d, J = 7.2 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 14.2, 22.8, 26.5, 32.1, 36.4, 37.0, 60.3, 109.2, 118.6, 124.8, 127.4, 129.1, 151.2; HPLC (OD-H, elute: Hexanes/*i*-PrOH = 99/1, detector: 254 nm, flow rate: 1.0 mL/min), $t_1 = 6.2$ min, $t_2 = 7.6$ min; HRMS Calculated for $C_{13}H_{19}N$ [M]⁺ 189.1517, found 189.1524.

(+)-(*R*)-2-Cyclohexanylindoline (2d).⁴ White solid, mp = 61-63 °C, 90% yield, 95% ee, $[\alpha]^{RT}_{D} = +1.82$ (c 1.50, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 0.97-1.04 (m, 2H), 1.20-1.31 (m, 3H), 1.44-1.47 (m, 1H), 1.70-1.90 (m, 5H), 2.75 (dd, J = 15.4, 9.8 Hz, 1H), 3.07 (dd, J = 15.5, 8.7 Hz, 1H), 3.57 (q, J = 8.5 Hz, 1H), 3.80 (br, 1H), 6.61 (d, J = 7.7 Hz, 1H), 6.69 (t, J = 7.4 Hz, 1H), 7.01 (t, J = 7.6 Hz, 1H), 7.08 (d, J = 7.2 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 26.1, 26.2, 26.7, 29.8, 30.3, 34.3, 44.1, 65.8, 109.0, 118.5, 124.7, 127.3, 129.3, 151.4; HPLC (OD-H, elute: Hexanes/*i*-PrOH = 97/3, detector: 254 nm, flow rate: 0.8 mL/min), $t_1 = 8.0$ min, $t_2 = 11.9$ min.

(+)-(*R*)-2-Cyclopentylindoline (2e). White solid, mp = 54-57 °C, 85% yield, 95% ee, $[\alpha]^{RT}_{D}$ = +17.1 (c 1.20, CHCl₃); 1 H NMR (400 MHz, CDCl₃) δ 1.24-1.29 (m, 2H), 1.58-1.68 (m, 4H), 1.70-1.88 (m, 2H), 1.99-2.06 (m, 1H), 2.75 (dd, J = 15.5, 8.9 Hz, 1H), 3.11 (dd, J = 15.5, 8.6 Hz, 1H), 3.66 (q, J = 8.7 Hz, 1H), 3.82 (br, 1H), 6.61 (d, J = 7.8 Hz, 1H), 6.69 (t, J = 7.4 Hz, 1H), 7.02 (t, J = 7.6 Hz, 1H), 7.08 (d, J = 7.2 Hz, 1H); 13 C NMR (100 MHz, CDCl₃) δ 25.6, 25.7, 29.7, 30.4, 35.5, 46.3, 65.4, 109.1, 118.5, 124.8, 127.3, 129.3, 151.3; HPLC (OD-H, elute: Hexanes/*i*-PrOH = 97/3, detector: 254 nm, flow rate: 0.8 mL/min), t_1 = 8.6 min, t_2 = 12.2 min; HRMS Calculated for $C_{13}H_{17}N$ [M]⁺ 187.1361, found 187.1368.

(+)-(*R*)-2-Phenethylindoline (2f).⁵ 89% yield, 93% *ee*, $[\alpha]^{\text{RT}}_{\text{D}} = +12.7$ (*c* 1.70, CHCl₃), {Lit. $[\alpha]^{23}_{\text{D}} = +12.0$ (*c* 0.17, CHCl₃)}⁵; ¹H NMR (400 MHz, CDCl₃) δ 1.89-1.96 (m, 2H), 2.67-2.74 (m, 3H), 3.13 (dd, J = 15.4, 8.6 Hz, 1H), 3.80-3.88 (m, 2H), 6.57 (d, J = 7.8 Hz, 1H), 6.67 (t, J = 7.4 Hz, 1H), 6.99 (t, J = 7.6 Hz, 1H), 7.05(d, J = 7.2 Hz, 1H), 7.16-7.20 (m, 3H), 7.26-7.30 (m, 2H); ¹³C NMR (CDCl₃, 125 MHz): δ 33.1, 36.3, 38.6, 59.7, 109.3, 118.7, 124.8, 126.1, 127.4, 128.5, 128.6, 128.9, 141.9, 151.1; HPLC (OD-H, elute: Hexanes/*i*-PrOH = 90/10, detector: 254 nm, flow rate: 0.8 mL/min), $t_1 = 11.5$ min, $t_2 = 14.0$ min.

(+)-(*R*)-2-Benzylindoline (2g).⁴ 99% yield, 95% ee, $[\alpha]^{RT}_{D} = +80.2$ (c 1.00, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 2.74-2.91 (m, 3H), 3.12 (dd, J = 15.5, 8.4 Hz, 1H), 3.79 (br, 1H), 4.01-4.09 (m, 1H), 6.54 (d, J = 7.7 Hz, 1H), 6.68 (t, J = 7.4 Hz, 1H), 6.99 (t, J = 7.6 Hz, 1H), 7.07 (d, J = 7.2 Hz, 1H), 7.19-7.30 (m, 3H), 7.31-7.34 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 36.1, 42.9, 61.2, 109.3, 118.7, 125.0, 126.6, 127.5, 128.5, 128.8, 129.3, 139.3, 150.7; HPLC (OD-H, elute: Hexanes/*i*-PrOH = 99/1, detector: 254 nm, flow rate: 1.0 mL/min), t_1 = 10.7 min, t_2 = 12.0 min.

(+)-(*R*)-2-(2'-Methylbenzyl)indoline (2h). 84% yield, 94% *ee*, $[\alpha]^{\text{RT}}_{\text{D}} = +74.8$ (*c* 1.50, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 2.35 (s, 3H), 2.82-2.91 (m, 3H), 3.77 (dd, J = 15.5, 8.5 Hz, 1H), 3.80 (br, 1H), 4.08-4.13 (m, 1H), 6.59 (d, J = 7.7 Hz, 1H), 6.71 (t, J = 7.3 Hz, 1H), 7.03 (t, J = 7.6

Hz, 1H), 7.11 (d, J = 7.2 Hz, 1H), 7.13-7.19 (m, 4H); ¹³C NMR (100 MHz, CDCl₃) δ 19.8, 36.3, 39.9, 59.8, 109.3, 118.7, 125.0, 126.3, 126.8, 127.6, 128.6, 129.9, 130.7, 136.7, 137.4, 150.8; HPLC (OD-H, elute: Hexanes/*i*-PrOH = 99/1, detector: 254 nm, flow rate: 1.0 mL/min), $t_1 = 9.8$ min, $t_2 = 10.8$ min; HRMS Calculated for $C_{16}H_{17}N$

[M]⁺ 223.1361, found 223.1368.

(+)-(*R*)-2-(3'-Methylbenzyl)indoline (2i). 95% yield, 94% ee, [α]^{RT}_D = +75.4 (c 1.60, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 2.38 (s, 3H), 2.78-2.91 (m, 3H), 3.16 (dd, J = 15.4, 8.4 Hz, 1H), 3.80 (br, 1H), 4.07-4.11 (m, 1H), 6.59 (d, J = 7.7 Hz, 1H), 6.69 (t, J = 7.3 Hz, 1H), 7.00-7.11 (m, 5H), 7.24 (t, J = 7.7 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 21.6, 36.1, 42.8, 61.2, 109.3, 118.6, 118.7, 125.0, 126.3, 127.4, 127.5, 128.7, 130.1, 138.4, 139.2, 150.7; HPLC (OD-H, elute: Hexanes/*i*-PrOH = 99/1, detector: 254 nm, flow rate: 1.0 mL/min), t_1 = 8.5 min, t_2 = 9.3 min; HRMS Calculated for $C_{16}H_{17}N$ [M]⁺ 223.1361, found 223.1369.

(+)-(*R*)-2-(4'-Methylbenzyl)indoline (2j). 82% yield, 93% *ee*, $[\alpha]^{RT}_{D} = +75.5$ (*c* 1.50, CHCl₃); ^{1}H NMR (400 MHz, CDCl₃) δ 2.36 (s, 3H), 2.77-2.90 (m, 3H), 3.14 (dd, J = 15.5, 8.4 Hz, 1H), 3.80 (br, 1H), 4.04-4.08 (m, 1H), 6.57 (d, J = 7.5 Hz, 1H), 6.70 (t, J = 7.3 Hz, 1H), 7.02 (t, J = 7.5 Hz, 1H), 7.08-7.17 (m, 5H); ^{13}C NMR (100 MHz, CDCl₃) δ 21.3, 36.1, 42.4, 61.3, 109.3, 118.7, 125.0, 127.5, 128.6, 129.2, 129.5, 129.6, 136.2, 150.8; HPLC (OD-H, elute: Hexanes / *i*-PrOH = 99/1, detector: 254 nm, flow rate: 1.0 mL/min), t₁ = 8.5 min, t₂ = 9.3 min; HRMS Calculated for C₁₆H₁₇N [M]⁺ 223.1361, found 223.1364.

(+)-(*R*)-2-(1'-Naphthylmethyl)indoline (2k). 78% yield, 96% ee, $[\alpha]^{RT}_{D} = +43.5$ (c 1.20, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 2.93 (dd, J = 15.5, 6.5 Hz, 1H), 3.18-3.39 (m, 3H), 3.80 (br, 1H), 4.25-4.29 (m, 1H), 6.57 (d, J = 7.7 Hz, 1H), 6.73 (t, J = 7.7 Hz, 1H), 7.04 (t, J = 7.6 Hz, 1H), 7.14 (d, J = 7.2 Hz, 1H), 7.39 (d, J = 6.9 Hz, 1H), 7.44-7.54 (m, 3H), 7.80 (d, J = 8.2 Hz, 1H), 7.91 (d, J = 8.1 Hz, 1H), 8.07 (d, J = 7.6 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 36.4, 39.7, 60.3, 109.4, 118.7, 124.0, 125.0, 125.7, 125.9, 126.2, 127.3, 127.5, 127.6, 128.5, 129.0, 132.3, 134.2, 135.3, 150.6; HPLC (OD-H, elute: Hexanes/*i*-PrOH = 90/10, detector: 254 nm, flow rate: 0.8 mL/min), t_1 = 13.3 min, t_2 = 16.6 min; HRMS Calculated for $C_{19}H_{17}N$ [M]⁺ 259.1361, found 259.1368.

(+)-(*R*)-5-Fluoro-2-methylindoline (2l).⁶ 84% yield, 88% ee, $[\alpha]^{RT}_{D} = +7.56$ (c 0.80, CHCl₃), {Lit. $[\alpha]^{RT}_{D} = -10.1$ (c 0.50, CHCl₃) for 99% ee of (*S*)-enantiomer }⁶; ¹H NMR (400 MHz, CDCl₃) δ 1.28 (d, J = 6.2 Hz, 3H), 2.62 (dd, J = 15.6, 7.8 Hz, 1H), 3.11 (dd, J = 15.7, 8.4 Hz, 1H), 3.60 (br, 1H), 3.96-4.00 (m, 1H), 6.47-6.51(m, 1H), 6.66-6.70 (m, 1H), 6.78-6.81 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 22.4, 38.2, 56.1, 109.5, 112.3, 113.3, 130.0, 147.1, 158.4; HPLC (OD-H, elute: Hexanes/*i*-PrOH = 99/1, detector: 254 nm, flow rate: 1.0 mL/min), t_1 = 6.5 min, t_2 = 9.1 min.

(+)-(*R*)-2,5-Dimethylindoline (2m). 81% yield, 84% ee, [α]^{RT}_D = +12.4 (c 1.10, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 1.30 (d, J = 6.2 Hz, 3H), 2.27 (s, 3H), 2.62 (dd, J = 15.4, 7.8 Hz, 1H), 3.12 (dd, J = 15.4, 8.4 Hz, 1H), 3.60 (br, 1H), 3.93-4.01 (m, 1H), 6.54 (d, J = 7.8 Hz, 1H), 6.84 (d, J = 7.8 Hz, 1H), 6.93 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 21.0, 22.4, 38.0, 55.6, 109.4,

125.7, 127.7, 128.1, 129.5, 148.7; HPLC (OD-H, elute: Hexanes/i-PrOH

= 99/1, detector: 254 nm, flow rate: 1.0 mL/min), t_1 = 7.2 min, t_2 = 9.2 min.

- (+)-(2*R*,3*R*)-*cis*-5,6,7,8,8a,9-Hexahydro-4bH-carbazole (2n).⁵ 91% yield, 91% *ee*, $[\alpha]^{\text{RT}}_{\text{D}} = +23.4$ (*c* 1.20, CHCl₃), {Lit. $[\alpha]^{23}_{\text{D}} = -20.0$ (*c* 0.25, CHCl₃) (84% ee)}⁵; ¹H NMR (400 MHz, CDCl₃) δ 1.36-1.44 (m, 3H), 1.55-1.60 (m, 2H), 1.61-1.74 (m, 1H), 1.76-1.81 (m, 2H), 3.12 (q, J = 6.6 Hz, 1H), 3.70 (br, 1H), 3.74 (q, J = 6.4 Hz, 1H), 6.69 (d, J = 7.7 Hz, 1H), 6.76 (t, J = 7.4 Hz, 1H), 7.04 (t, J = 7.6 Hz, 1H), 7.10 (d, J = 7.2 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 21.9, 22.7, 27.1, 29.4, 41.1, 59.8, 110.3, 119.0, 123.3, 127.2, 133.7, 150.9; HPLC (IC, elute: Hexanes/*i*-PrOH = 99/1, detector: 254 nm, flow rate: 1.0 mL/min), $t_1 = 5.0$ min, $t_2 = 7.5$ min.
- (+)-(2*R*,3*R*)-*cis*-1-methyl-5,6,7,8,8a,9-hexahydro-4bH-carbazole (2o). 8 83% yield, 96% *ee*, $[\alpha]^{\text{RT}}_{D} = +17.9 \ (c\ 0.90, \text{CHCl}_3); ^1\text{H NMR (400 MHz, CDCl}_3) \ \delta \ 1.37-1.43 \ (m, 3H), 1.55-1.58 \ (m, 2H), 1.60-1.70 \ (m, 1H), 1.75-1.79 \ (m, 2H), 2.15 \ (m, 3H), 3.13 \ (q, J = 6.6 \ Hz, 1H), 3.15 \ (br, 1H), 3.75 \ (q, J = 6.7 \ Hz, 1H), 6.70 \ (t, J = 7.4 \ Hz, 1H), 6.88 \ (d, J = 7.5 \ Hz, 1H), 6.96 \ (d, J = 7.2 \ Hz, 1H); <math>^{13}\text{C NMR}$ (100 MHz, CDCl₃) $\delta \ 17.0, 22.0, 22.7, 27.2, 29.7, 41.4, 59.7, 119.1, 119.6, 120.8, 128.2, 132.9, 149.5; HPLC (IC, elute: Hexanes/$ *i* $-PrOH = 99/1, detector: 254 nm, flow rate: 1.0 mL/min), <math>t_1 = 4.4 \ \text{min}, t_2 = 4.9 \ \text{min}.$
- (+)-(2*R*,3*R*)-*cis*-5,5a,6,7,8,9,10,10a-Octahydrocyclohepta[b]indole (2p).⁵ White solid, mp = 61-64 °C, 96% yield, 90% *ee*, [α]^{RT}_D = +32.6 (*c* 1.20, CHCl₃), {Lit. [α]^{RT}_D = -30.0 (*c* 0.52, CHCl₃) (86% ee)}⁵; ¹H NMR (400 MHz, CDCl₃) δ 1.33-1.43 (m, 3H), 1.70-1.91 (m, 6H), 1.94-2.00 (m, 1H), 3.48 (td, *J* = 10.6, 3.2 Hz, 1H), 3.64 (br, 1H), 4.01-4.08 (m, 1H), 6.56 (d, *J* = 7.7 Hz, 1H), 6.70 (t, *J* = 7.4 Hz, 1H), 6.99-7.04 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 26.2, 28.9, 31.6, 33.8, 46.9, 63.6, 108.6, 118.3, 124.3, 127.5, 133.7, 150.4; HPLC (IC, elute: Hexanes/*i*-PrOH = 99/1, detector: 254 nm, flow rate: 1.0 mL/min), t_1 = 5.6 min, t_2 = 7.1 min.
- (+)-(2*R*,3*R*)-*cis*-2,3-Dimethylindoline (2q). 84% yield, 92% *ee*, [α]^{RT}_D = +26.6 (*c* 0.83, CHCl₃), {Lit. [α]²⁶_D = +19.3 (*c* 0.30, CHCl₃)} 9; ¹H NMR (400 MHz, CDCl₃) δ 1.14 (d, J = 6.6 Hz, 3H), 1.19 (d, J = 7.2 Hz, 3H), 3.23-3.29 (m, 1H), 3.65 (br, 1H), 3.91-3.97 (m, 1H), 6.62 (d, J = 7.7 Hz, 1H), 6.73 (t, J = 7.4 Hz, 1H), 7.00-7.08 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 13.8, 16.5, 39.6, 58.6, 109.5, 118.9, 124.0, 127.4, 134.5, 150.3; HPLC (OJ-H, elute: Hexanes/*i*-PrOH = 99/1, detector: 254 nm, flow rate: 1.0 mL/min), t_1 = 16.7 min, t_2 = 20.6 min.

5. Mechanism Study

To elucidate the mechanism of Pd-catalyzed asymmetric hydrogenation of unprotected indoles in the presence of Brønsted acid, ¹H-NMR and two isotopic labeling experiments were carried out.

¹**H-NMR Study:** TsOH·H₂O (48 mg, 0.25 mmol) was added to 2-methylindole (33 mg, 0.25 mmol) solution in CDCl₃ and stirred for 5 min at room temperature.

¹H-NMR analysis of the resulted yellow solution, it was showed that the signal of the 2-methyl and hydrogen at the 3-position become broad peaks and shifted to downfield, and the N-H peak and olefinic hydrogen at the 3-position of 2-methylindole disappeared rapidly (**Figure 1**). These results indicated that fast reversible process of protonation and deprotonation existed.

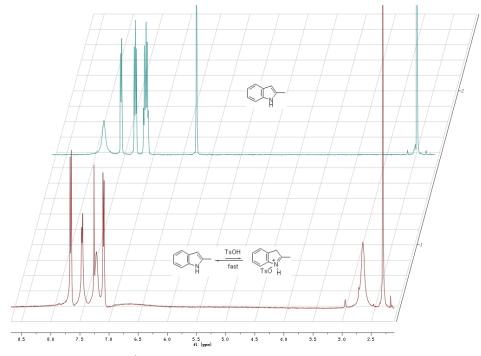


Figure 1. ¹H-NMR Study of TsOH and 2-Methylindole in CDCl₃

Asymmetric Hydrogenation of 2-Methylindole with the Deuterated L-CSA in d³-TFE: Deuterated L-CSA was obtained by stirring d³-TFE (1000 mg) and L-CSA (58 mg, 0.25 mmol) at room temperature in glove box for 0.5 h for sufficient Hydrogen/Deuterium exchange. Subsequently, the hydrogenation was carried out.

¹H-NMR analysis of the crude hydrogenated product showed that two deuterium atoms were incorporated to the 3-position (both with 94% incorporation) of hydrogenation product 2-methylindoline **2a**, which suggested that a rapid reversible process of protonation and

deprotonation existed (**Figure 2**), and the equilibrium was faster than hydrogenation. Thus, two deuterium atoms was imported to the 3-position of the 2-methylindoline before hydrogenation occurred.

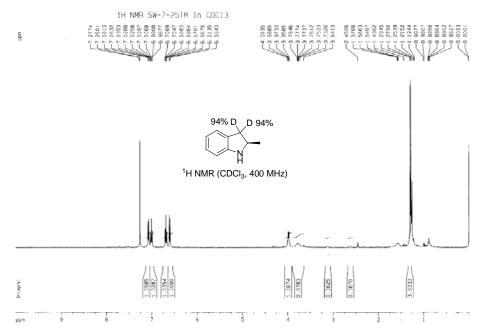


Figure 2. ¹H-NMR of Product 1a with d³-TFE and L-CSA

Asymmetric Hydrogenation of 2-Methylindole with D₂: 2-Methylindole **1a** was hydrogenated in D_2 (116 psi) with the Pd(OCOCF₃)₂/(R)-BINAP/TFE condition.

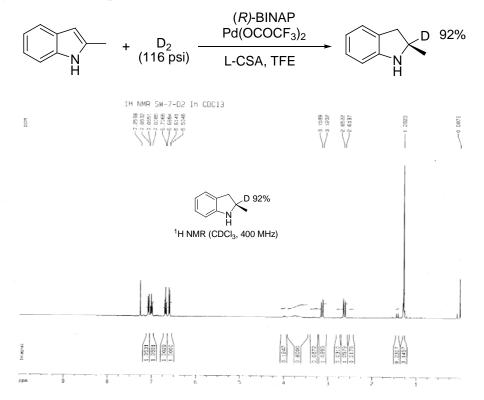


Figure 3. ¹H-NMR of Product 1a with D₂

2-deuterio-2-Methylindoline with 92% incorporation was obtained, deuterium at 3-position was not observed (**Figure 3**). These results confirmed that the simple unprotected indole can be activated by a Brønsted acid to form iminium *in situ*, which was then hydrogenated by the Pd-catalyst.

$$\begin{array}{c|c} R' \\ R \\ R \\ R \\ R \\ R \\ R' \\ R \\ R' \\ R'$$

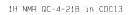
Figure 4. Mechanism of Hydrogenation of 2,3-Disubstituted Indoles

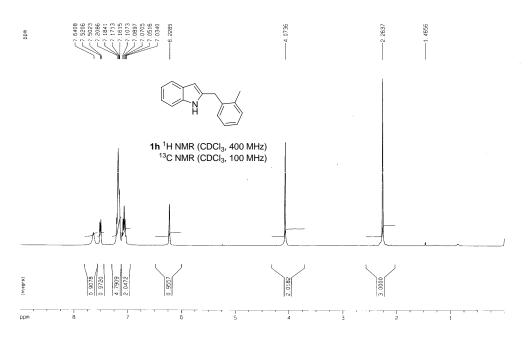
For the Pd catalyzed asymmetric hydrogenation of 2,3-disubstituted indole activated by a Brønsted acid, the mechanism was slightly different from that of 2-substituted indoles. The hydrogenation of an intermediate iminium salt of 2-substituted indole is the enantioselectivity-controlled step, while enantio-selectivity-controlled step of 2,3-disubstituted indole is the protonation of carbon-carbon double bond and the hydrogenation of iminium salt, which is in fact a dynamic kinetic resolution process (**Figure 4**). To obtain high ee, it should meet the equation of $k_1 >> k_2$. The above mechanism study indicated that rate of protonation k_1 is faster than rate of hydrogenation k_2 ($k_1 >> k_2$).

6. References

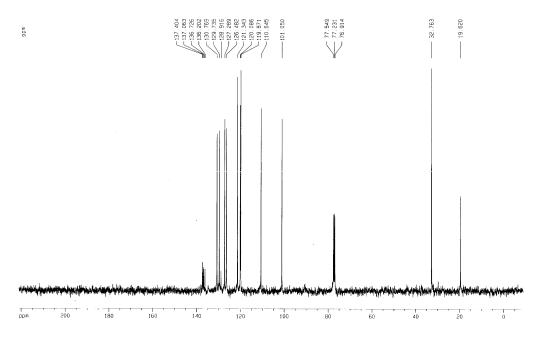
- (a) Saito, T.; Nihei, H.; Otani, T.; Suyama, T.; Furukawa, N.; Saito, M. Chem. Commun.
 2008, 172. (b) Okuma, K.; Seto, J.; Sakaguchi, K.; Ozaki, S.; Nagahora, N.; Shioji, K.
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 Tetrahedron 1998, 54, 13915. (f) Wiedenau, P.; Blechert, S. Synth. Commun. 1997, 27, 2033.
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7. Copy of NMR and HRMS Spectra





13C NMR QC-4-21B In CDC13



Page 1

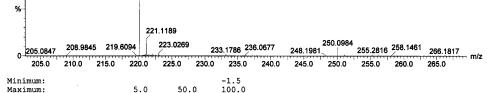
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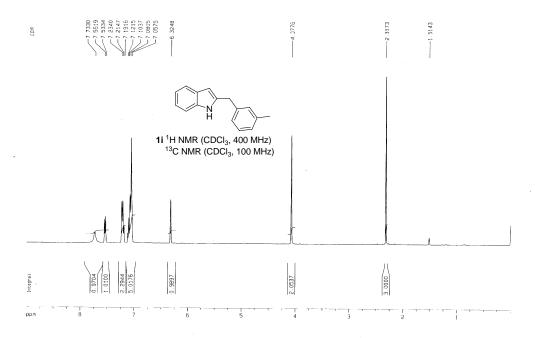
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Elements Used:
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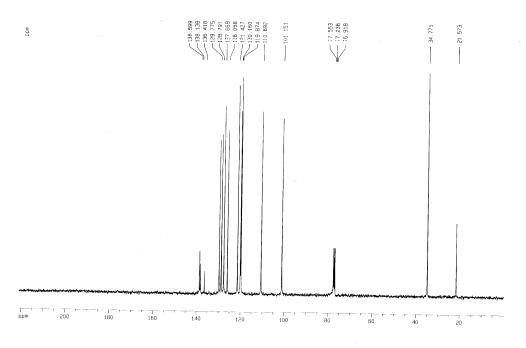
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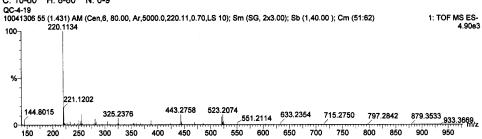
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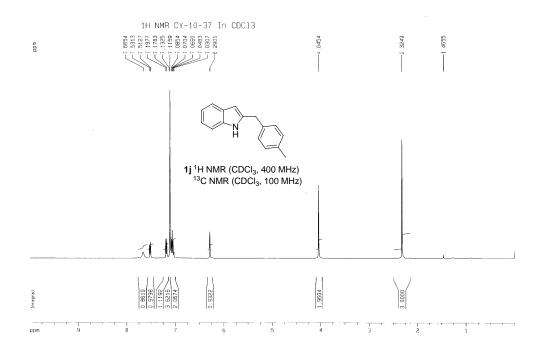
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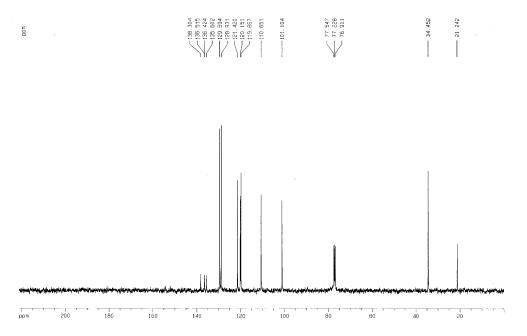
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20 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass)
Elements Used:
C: 10-60 H: 8-80 N: 0-9



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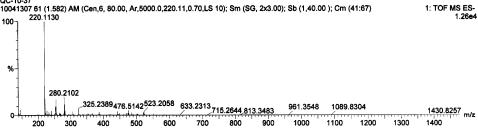
13C NMR CY-10~37 in CDC13



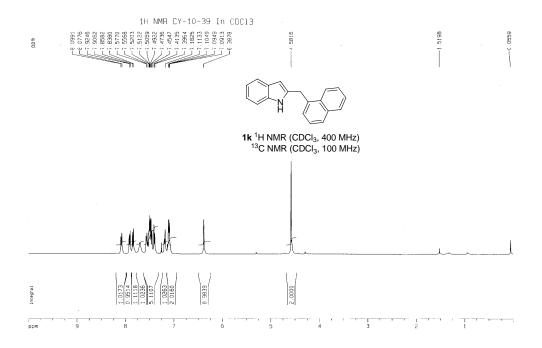
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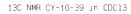
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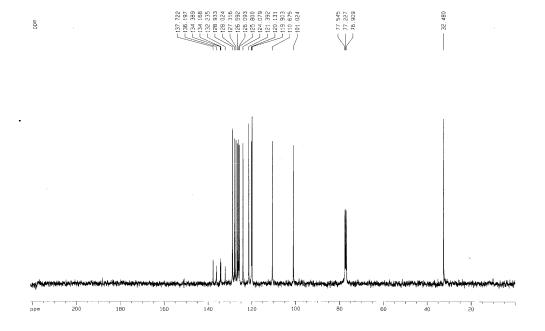
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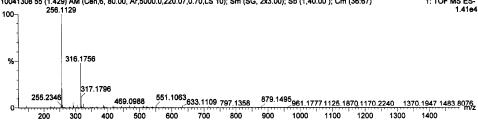


Page 1

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QC-10-39
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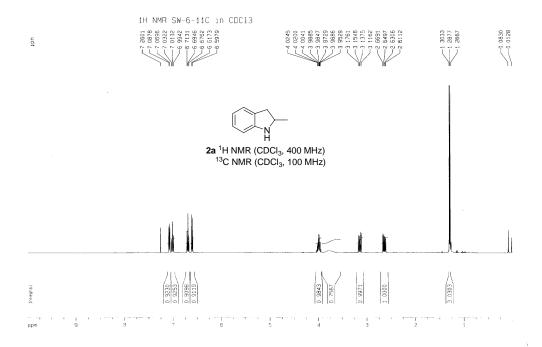
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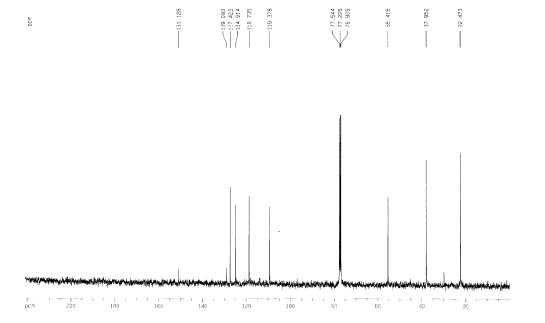
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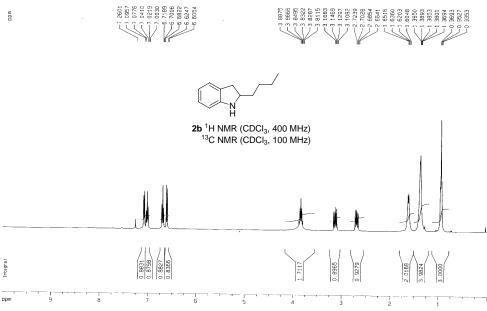
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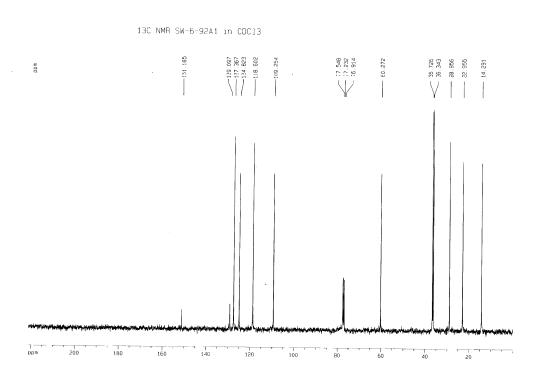
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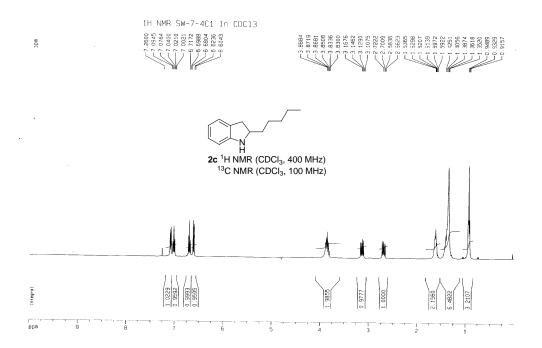


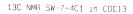


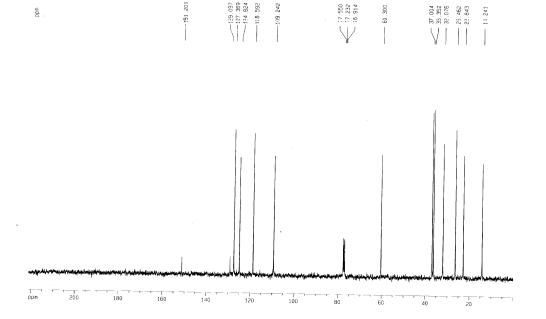








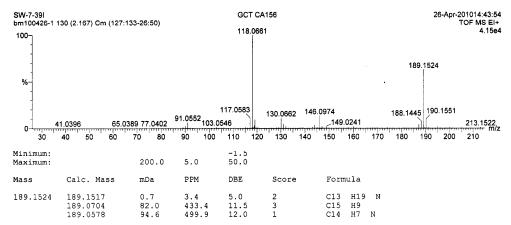


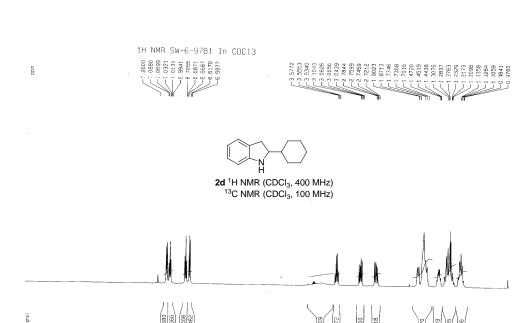


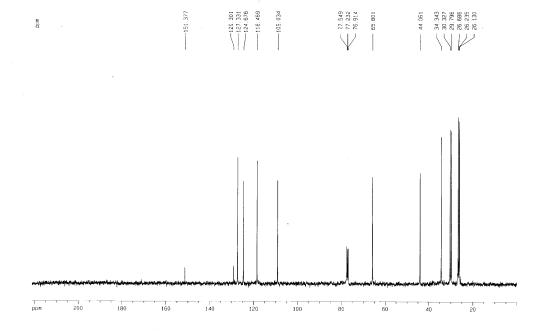
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Monoisotopic Mass, Odd and Even Electron Ions 3 formula(e) evaluated with 3 results within limits (up to 50 closest results for each mass)







13C NMR SW-6-97B1 in CDC13

1H NMR SW-7-32H In CDC13 2601 0375 0375 0375 0185 9996 7118 6934 6750 6750 Edd **2e** ¹H NMR (CDCl₃, 400 MHz) ¹³C NMR (CDCl₃, 100 MHz) Integral 13C NMR SW-7-32H in CDC13 mdd

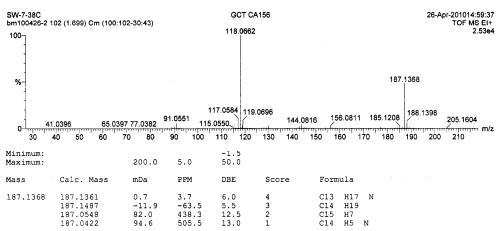
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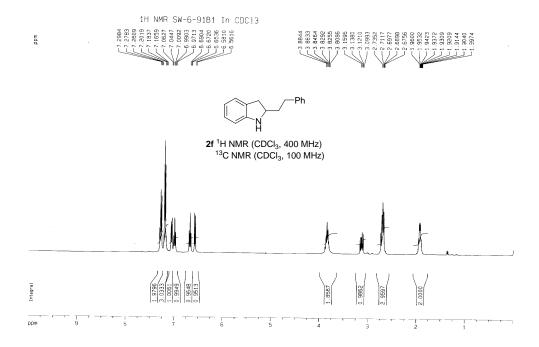
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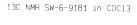
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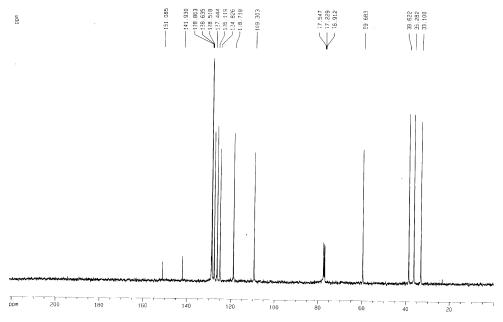
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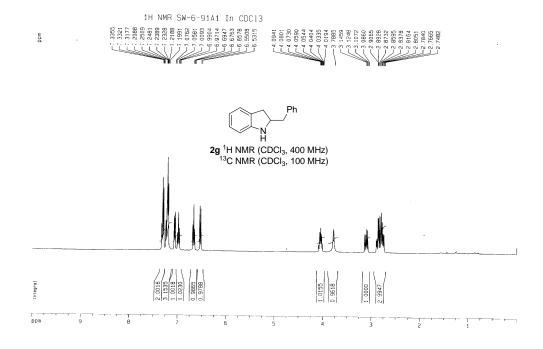
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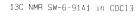


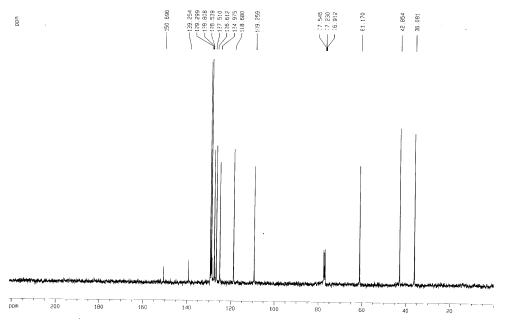


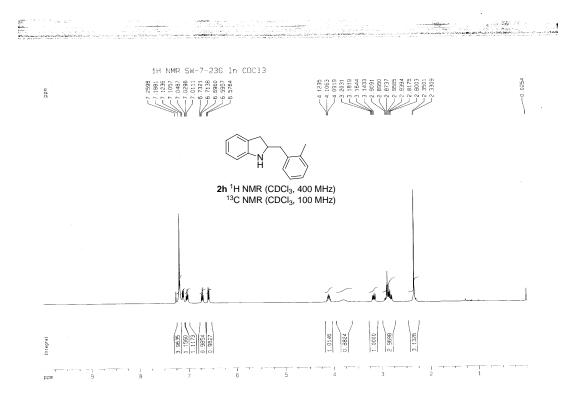


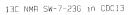


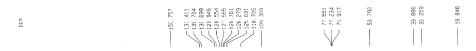


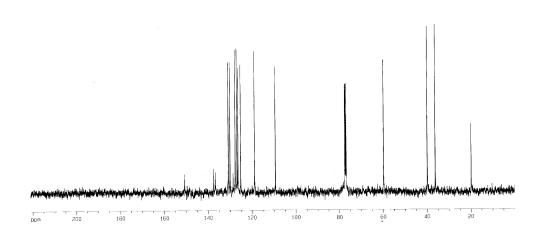












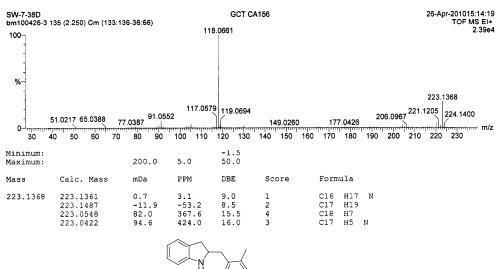
Page 1

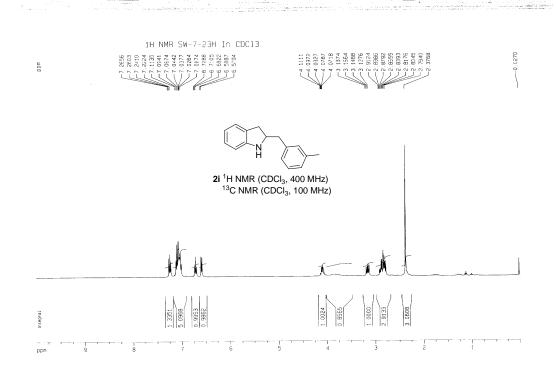
Single Mass Analysis

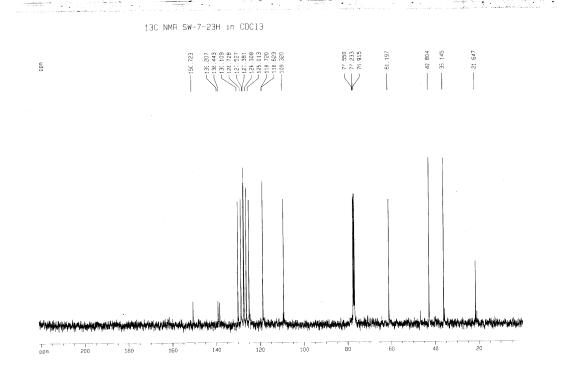
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Monoisotopic Mass, Odd and Even Electron Ions 4 formula(e) evaluated with 4 results within limits (up to 50 closest results for each mass)

2h HRMS





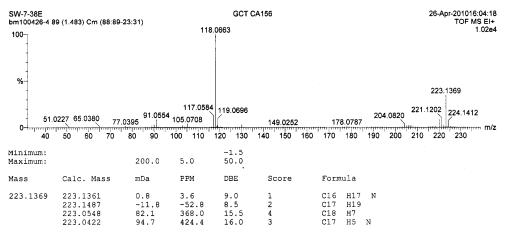


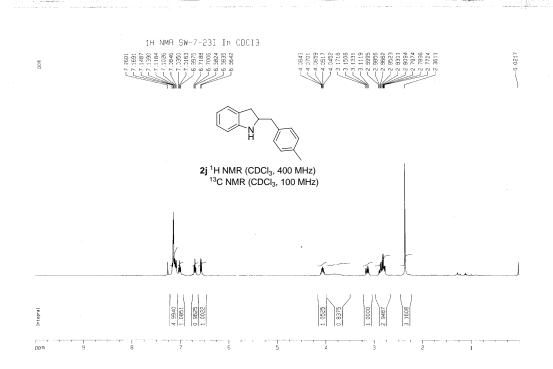
Page 1

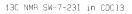
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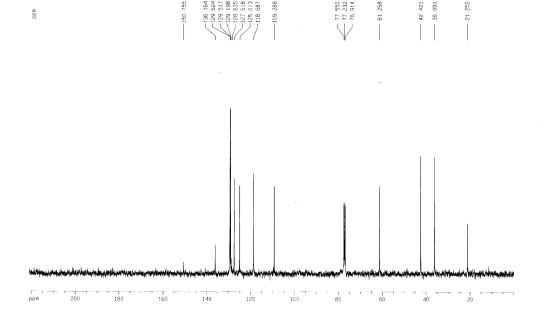
Tolerance = 200.0 mDa / DBE: min = -1.5, max = 50.0 Isotope cluster parameters: Separation = 1.0 Abundance = 1.0%

Monoisotopic Mass, Odd and Even Electron Ions 4 formula(e) evaluated with 4 results within limits (up to 50 closest results for each mass)







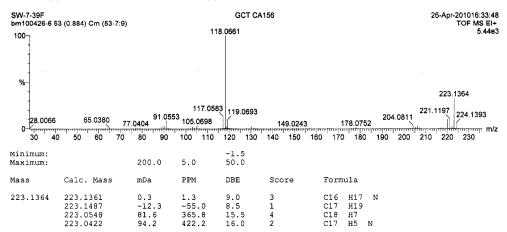


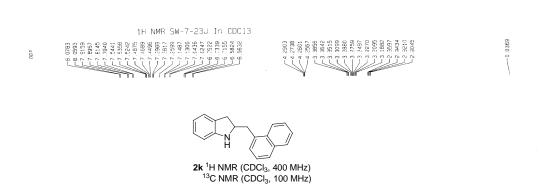
Page 1

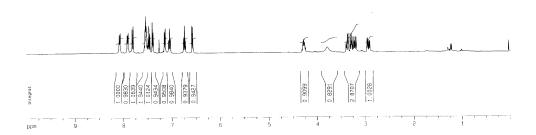
Single Mass Analysis

Tolerance = 200.0 mDa / DBE: min = -1.5, max = 50.0 Isotope cluster parameters: Separation = 1.0 Abundance = 1.0%

Monoisotopic Mass, Odd and Even Electron Ions 4 formula(e) evaluated with 4 results within limits (up to 50 closest results for each mass)



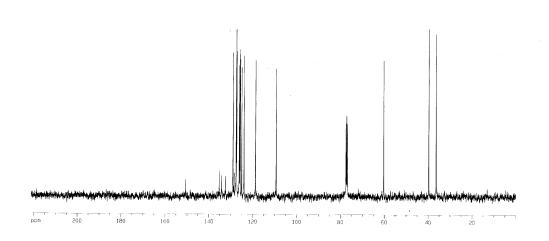




13C NMR SW-7-23J in CDC13

mdd

150 658 113 218 122 113 318 318 113 318 318 114 318 318 116 323 116 328 117 545 118 728 119 328 119 328 119 328

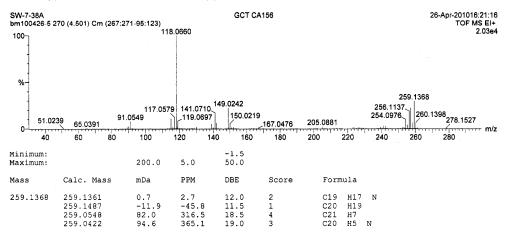


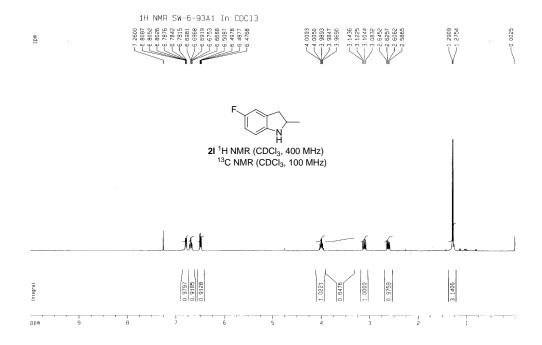
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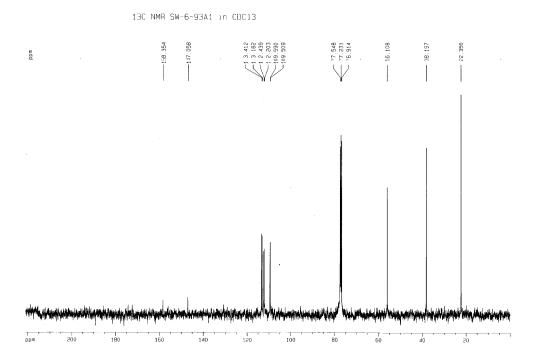
Single Mass Analysis

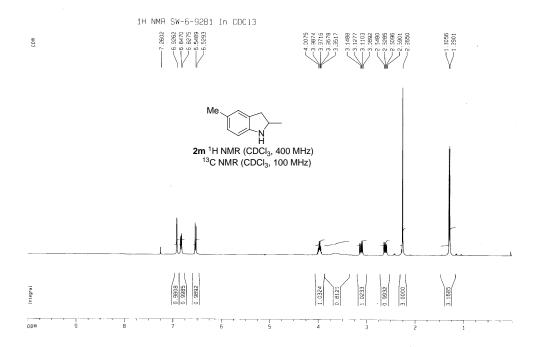
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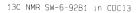
Monoisotopic Mass, Odd and Even Electron Ions 4 formula(e) evaluated with 4 results within limits (up to 50 closest results for each mass)

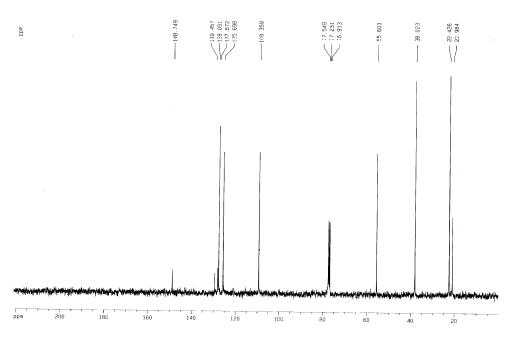


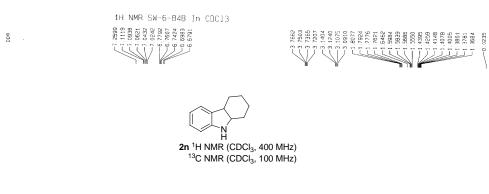


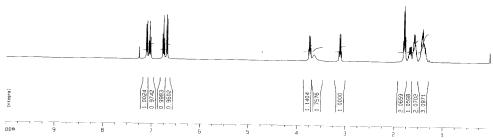




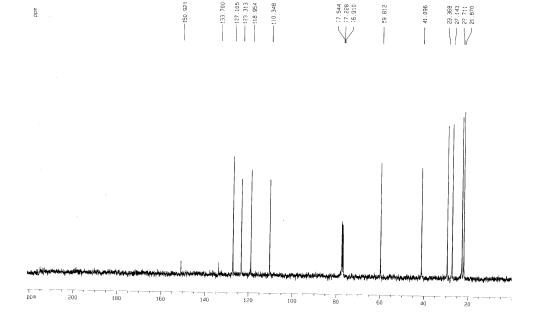


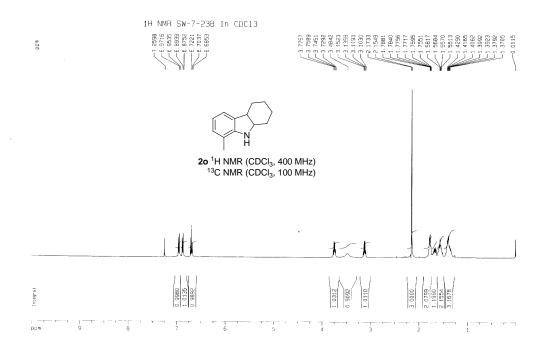


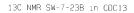




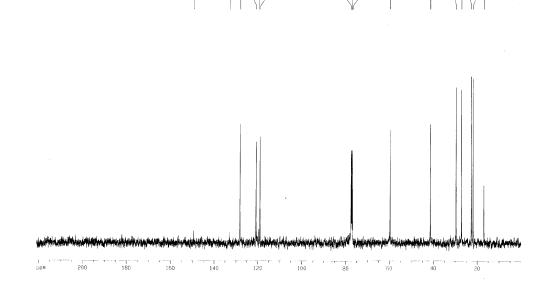
13C NMR SW-6-84B in CDC13

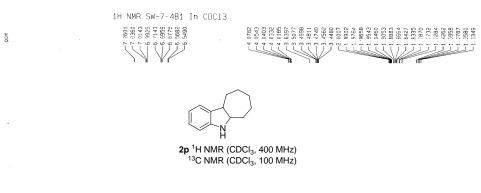


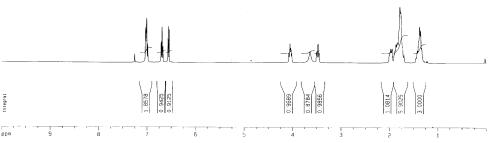




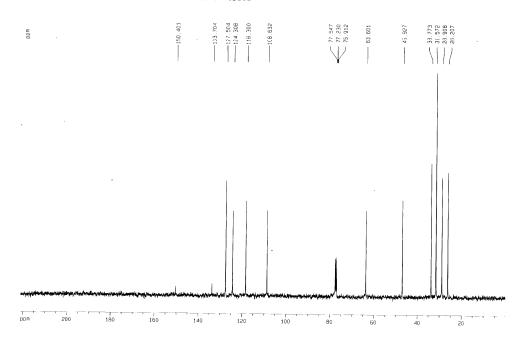
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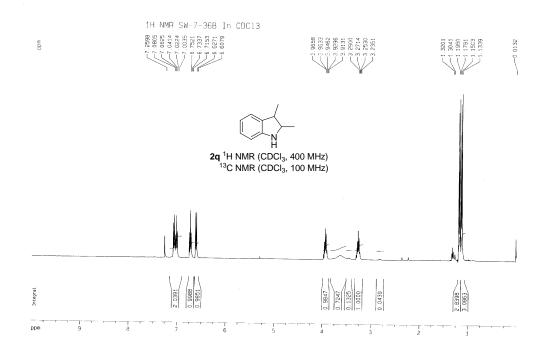


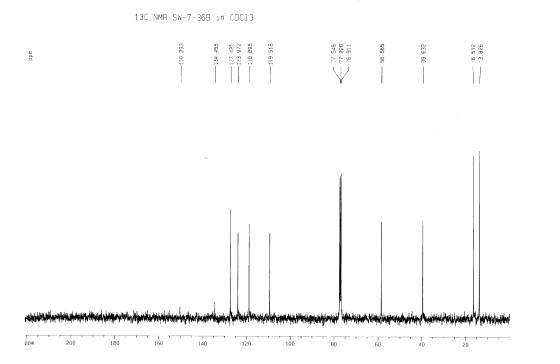




13C NMR SW-7-4B1 in CDC13







8. Copy of HPLC for Racemic and Chiral Compounds

Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008620.D OD-H, H/i-PrOH =97/3, 0.8 mL/min, 30 oC, 254 nm

Sample Name: SW-6-11C

OD-H, H/i-PrOH =97/3, 0.8 mL/min, 30 oC, 254 nm _____

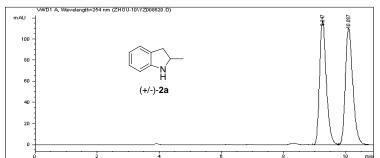
Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008619.D

Sample Name: SW-7-35A

_____ Injection Date : 4/24/2010 10:44:17 PM Sample Name : SW-6-11C Acg. Operator : Acg. Method : C:\HPCHEM\1\METHODS\SW Location : Vial 1

C:\HPCHEM\1\METHODS\SW.M Last changed : 4/24/2010 10:48:55 PM (modified after loading) Analysis Method : C:\HPCHEM\1\METHODS\SW.M Last changed : 4/24/2010 10:57:12 PM

(modified after loading)



Area Percent Report

Sigmal 1.0000 1.0000 Sorted By Dilution

Signal 1: VWD1 A, Wavelength=254 nm

1 9.247 VV 2 10.087 VB

Totals : 3624.63062 227.78693

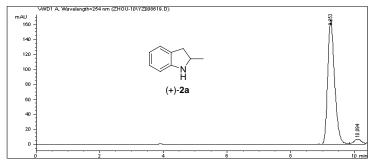
Results obtained with enhanced integrator! *** End of Report ***



Location : Vial 1

Acg. Operator Acg. Method C:\HPCHEM\1\METHODS\SW.M Last changed : 4/24/2010 10:26:52 PM (modified after loading)
Analysis Method : C:\HPCHEM\l\METHODS\SW.M Last changed : 4/24/2010 10:48:52 PM

(modified after loading)



_____ Area Percent Report

Sorted By Signal Multiplier Dilution

Signal 1: VWD1 A, Wavelength=254 nm

Peak RetTime Type Width Area Height Area 1 9.253 VV 0.2311 2520.55015 2 10.094 VV 0.2576 118.02444 6.95730 4.4729

Totals : 2638.67459 172.90644

Results obtained with enhanced integrator! *** End of Report ***

Instrument 1 4/24/2010 10:57:15 PM Page 1 of 1 Instrument 1 4/24/2010 10:48:55 PM

Page 1 of 1

Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008651.D OD-H, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm

: SW-6-92A1

Analysis Method : C:\HPCHEM\1\METHODS\SW.M Last changed : 4/26/2010 12:02:22 AM

Injection Date : 4/25/2010 11:51:49 PM

Sample Name

Acg. Operator Acg. Method

Last changed

Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008650.D Sample Name: SW-6-92Al OD-H, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm

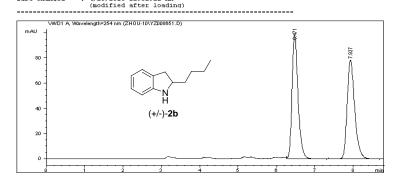
> Injection Date : 4/25/2010 11:29:01 PM

Sample Name : SW-7-39H

Location : Vial 1

Acg. Operator Acg. Method C:\HPCHEM\1\METHODS\SW.M 4/25/2010 11:24:00 PM (modified after loading) C:\HPCHEM\l\METHODS\SW.M 4/26/2010 12:02:25 AM Last changed

Analysis Method : Last changed : (modified after loading)



Location : Vial 1

-----Area Percent Report

C:\HPCHEM\1\METHODS\SW.M

: 4/25/2010 11:48:44 PM (modified after loading)

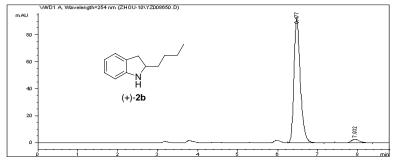
Sorted Bv Multiplier Sigmal 1.0000 1.0000

Dilution

Signal 1: VWD1 A, Wavelength=254 nm

2140.94299 176.64323

Results obtained with enhanced integrator!



______ Area Percent Report

Sorted Bv Multiplier Signal 1.0000 1.0000 Dilution

Signal 1: VWD1 A, Wavelength=254 nm

Totals : 1051.08424 94.67502

Results obtained with enhanced integrator!

Instrument 1 4/26/2010 12:02:25 AM

Page 1 of 1

Instrument 1 4/26/2010 12:02:48 AM

Page 1 of 1

Sample Name: SW-7-39H

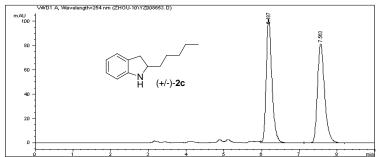
Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008653.D OD-H, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm

Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008652.D OD-H, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm Sample Name: SW-7-4C1

Sample Name: SW-7-39I

: 4/26/2010 12:21:39 AM (modified after loading) Last changed Analysis Method : C:\HPCHEM\1\METHODS\SW.M Last changed : 4/26/2010 7:01:15 PM

(modified after loading)



_____ Area Percent Report

Sorted Bv Multiplier Signal 1.0000 1.0000

Dilution

Signal 1: VWD1 A, Wavelength=254 nm

 Peak RetTime Type
 Width [min]
 Area [max]
 Height [max]
 Area [max]
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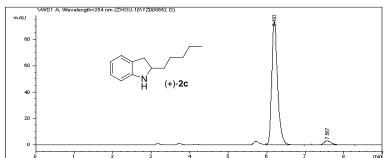
Totals : 2138.49084 183.43617

Results obtained with enhanced integrator! *** End of Report ***

Injection Date : 4/26/2010 12:07:51 AM Sample Name : SW-7-39I Location : Vial 1

Sample Name Acc. Operator Acq. Method C:\HPCHEM\1\METHODS\SW.M 4/26/2010 12:02:48 AM (modified after loading) Last changed Analysis Method : Last changed : C:\HPCHEM\1\METHODS\SW.M 4/26/2010 7:00:57 PM

(modified after loading)



_____ Area Percent Report

Sorted Bv Multiplier Signal 1.0000 1.0000

Signal 1: VWD1 A, Wavelength=254 nm

Dilution

Totals : 1027.14674 97.03624

Results obtained with enhanced integrator!

*** End of Report ***

Instrument 1 4/26/2010 7:03:19 PM

Page 1 of 1

Instrument 1 4/26/2010 7:01:15 PM

Page 1 of 1

Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008634.D OD-H, H/i-PrOH =97/3, 0.8 mL/min, 30 oC, 254 nm

Analysis Method : C:\HPCHEM\1\METHODS\SW.M Last changed : 4/25/2010 5:52:49 PM

Injection Date : 4/25/2010 4:21:55 PM Sample Name : SW-6--97B1

Sample Name Acc. Operator Acq. Method

Last changed

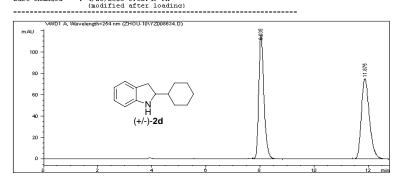
Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008638.D OD-H, H/i-PrOH =97/3, 0.8 mL/min, 30 oC, 254 nm

_____ Location : Vial 1

Injection Date : 4/25/2010 5:31:21 PM Sample Name : SW-7-38B Acc. Operator : C:\hPCHEM\l\METHODS\SU

: C:\HPCHEM\1\METHODS\SW.M : 4/25/2010 4:59:47 PM (modified after loading) Last changed Analysis Method : C:\HPCHEM\l\METHODS\SW.M Last changed : 4/25/2010 5:56:18 PM

(modified after loading)



Location : Vial 1

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: C:\HPCHEM\1\METHODS\SW.M

: 4/25/2010 4:17:14 PM (modified after loading)

Sorted Bv Multiplier Signal 1.0000 1.0000 Dilution

Signal 1: VWD1 A, Wavelength=254 nm

Totals : 3091.84155 191.25714

Results obtained with enhanced integrator! *** End of Report ***

VWD1 A, Wavelength=254 nm (ZHOU-10\YZ008638.D) mAU 160 120 100 80 (+)-2d60 20

_____ Area Percent Report ned labano napozo

Sorted Bv Multiplier Signal 1.0000 1.0000 Dilution

Signal 1: VWD1 A, Wavelength=254 nm

Totals : 2486.60168 183.25887

Results obtained with enhanced integrator! *** End of Report ***

Instrument 1 4/25/2010 5:52:52 PM

Page 1 of 1

Sample Name: SW-6--97B1

Instrument 1 4/25/2010 5:56:39 PM

Page 1 of 1

Sample Name: SW-7-38B

Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008639.D OD-H, H/i-PrOH =97/3, 0.8 mL/min, 30 oC, 254 nm

Analysis Method : C:\HPCHEM\1\METHODS\5W.M Last changed : 4/25/2010 6:37:51 PM

Acc. Method

Last changed

Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008640.D OD-H, H/i-PrOH =97/3, 0.8 mL/min, 30 oC, 254 nm Sample Name: SW-7-23E

Sample Name Acc. Operator Acq. Method

: C:\HPCHEM\1\METHODS\SW.M : 4/25/2010 6:22:18 PM (modified after loading) Last changed

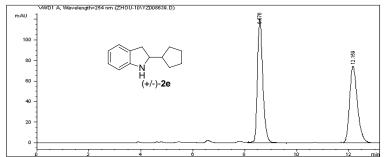
Analysis Method : C:\HPCHEM\1\METHODS\SW.M Last changed : 4/25/2010 6:37:54 PM (modified after loading)

(modified after loading)

Injection Date : 4/25/2010 6:00:35 PM Sample Name : SW-7-23E Location : Vial 1 Acc. Oberator :

C:\HPCHEM\1\METHODS\SW.M

: 4/25/2010 5:56:39 PM (modified after loading)



-----Area Percent Report

Sorted Bv Multiplier Signal 1.0000 1.0000

Signal 1: VWD1 A, Wavelength=254 nm

Dilution

Totals : 3263.08997 194.33997

Results obtained with enhanced integrator! *** End of Report ***

VWD1 A, Wavelength=254 nm (ZHOU-10\YZ008640.D) mAU 200 175 150 125 -100 75 -25

Sample Name: SW-7-38C

Page 1 of 1

_____ Area Percent Report

Sorted Bv Multiplier Signal 1.0000 1.0000 Dilution

Signal 1: VWD1 A, Wavelength=254 nm

 Peak RetTime Type
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 2 (2.21)

Totals : 3002.65131 208.17336

Results obtained with enhanced integrator! *** End of Report ***

Instrument 1 4/25/2010 6:37:54 PM Page 1 of 1 Instrument 1 4/25/2010 6:38:07 PM

S47

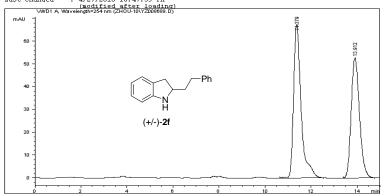
Sample Name: SW-7-11C Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008686.D Sample Name: SW-7-40R

OD-H, H/i-PrOH =90/10, 0.8 mL/min, 30 oC, 254 nm

______ Injection Date : 4/27/2010 10:19:23 PM Sample Name : SW-7-11C Location : Vial 1

Acq. Operator Acq. Method : C:\HPCHEM\1\METHODS\DEF_LC.M : 4/27/2010 10:14:52 PM Last changed

(modified after loading)
Analysis Method : C:\HPCHEM\l\METHODS\DEF LC.M Last changed : 4/27/2010 10:47:39 PM



..... Area Percent Report

ALCA TELEGIO NEDELO

Sorted By Signal 1.0000 Multiplier Dilution 1.0000

Signal 1: VWD1 A, Wavelength=254 nm

Totals : 2856.10120 120.12859

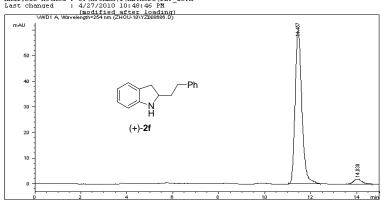
Results obtained with enhanced integrator! _____ *** End of Report ***

OD-H, H/i-PrOH =90/10, 0.8 mL/min, 30 oC, 254 nm

______ Injection Date : 4/27/2010 6:24:08 PM Sample Name : SW-7-40R Location : Vial 1

Acq. Operator Acq. Method C:\HPCHEM\1\METHODS\DEF_LC.M

Last changed : 4/27/2010 5:49:27 PM (modified after loading)
Analysis Method : C:\HPCHEM\l\METHODS\DEF_LC.M



Area Percent Report

Sorted By Signal Multiplier 1.0000 Dilution 1.0000

Signal 1: VWD1 A, Wavelength=254 nm

 Peak RetTime Type
 Width [min]
 Area [mAU]
 Height [min]
 Area [mAU]
 Area [mAU]</th

Totals : 1278.47350 61.91430

Results obtained with enhanced integrator! *** End of Report ***

Instrument 1 4/27/2010 10:47:41 PM Page 1 of 1 Instrument 1 4/27/2010 10:48:48 PM Page 1 of 1 Sample Name: SW-7-11B Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008534.D Sample Name: SW-7-39G

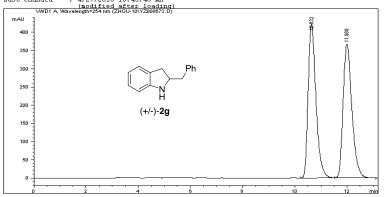
Page 1 of 1

OD-H, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm

_____ Injection Date : 4/27/2010 10:23:57 AM Sample Name : SW-7-11B Location : Vial 1

Acq. Operator Acq. Method : C:\HPCHEM\1\METHODS\DEF_LC.M : 4/27/2010 10:01:51 AM Last changed

(modified after loading)
Analysis Method: C:\HPCHEM\l\METHODS\DEF_LC.M Last changed : 4/27/2010 10:45:46 AM



..... Area Percent Report

ALCG TELECHO MEDILO

Sorted By Sigmal Multiplier Dilution 1.0000

Signal 1: VWD1 A, Wavelength=254 nm

Totals : 1.73676e4 790.05811

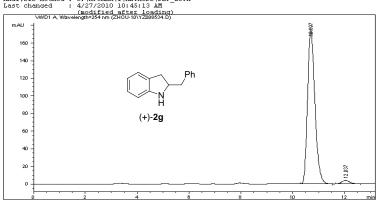
Results obtained with enhanced integrator! ______ *** End of Report ***

OD-H, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm

______ Injection Date : 4/19/2010 12:36:34 AM Sample Name : SW-7-39G Location : Vial 1

Acq. Operator Acq. Method : C:\HPCHEM\1\METHODS\SW.M

: 4/19/2010 12:32:49 AM Last changed (modified after loading)
Analysis Method: C:\HPCHEM\l\METHODS\DEF LC.M



Area Percent Report ALCA TELECHO NEPOLO

1.0000

Sorted By Signal Multiplier 1.0000

Signal 1: VWD1 A, Wavelength=254 nm

Dilution

Totals : 3504.99538 178.22451

Results obtained with enhanced integrator! ______ *** End of Report ***

Instrument 1 4/27/2010 10:45:48 AM Page 1 of 1 Instrument 1 4/27/2010 10:45:17 AM Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008665.D OD-H, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm

Injection Date : 4/26/2010 10:57:05 PM Sample Name : SW-7-23G

Analysis Method : C:\HPCHEM\1\METHOD\$\5W.M Last changed : 4/26/2010 11:31:59 PM

Sample Name Acc. Operator Acq. Method

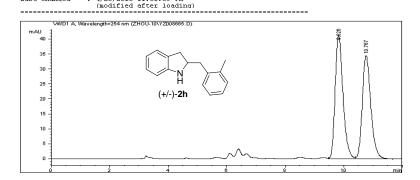
Last changed

Sample Name: SW-7-23G

Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008666.D OD-H, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm

Injection Date : 4/26/2010 11:12:42 PM Sample Name : SW-7-38D Location : Vial 1

(modified after loading)



Location : Vial 1

Area Percent Report

: C:\HPCHEM\1\METHODS\SW.M

: 4/26/2010 10:52:04 PM (modified after loading)

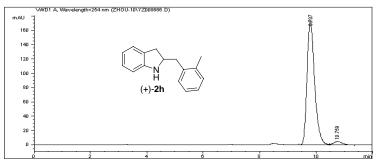
Sorted Bv : Signal Multiplier : 1.0000 Dilution : 1.0000

Signal 1: VWD1 A, Wavelength=254 nm

Totals: 1466.91119 75.74163

Results obtained with enhanced integrator!

*** End of Report ***



Area Percent Report

 Sorted By
 : Signal

 Multiplier
 : 1.0000

 Dilution
 : 1.0000

Signal 1: VWD1 A, Wavelength=254 nm

Totals: 3330.81712 177.89236

Results obtained with enhanced integrator!

**** End of Report ***

Sample Name: SW-7-38D

Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008663.D OD-H, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm

Injection Date : 4/26/2010 10:24:02 PM Sample Name : 5W-7-24H Acc. Operator : C:\hPCHEM\1\METHODS\5W.

Analysis Method : C:\HPCHEM\1\METHOD\$\5\W.M Last changed : 4/26/2010 10:51:31 PM

Last changed

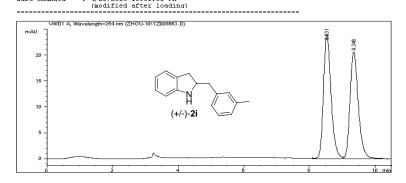
Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008664.D OD-H, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm Sample Name: SW-7-24H

> _____ Location : Vial 1

Injection Date : 4/26/2010 10:39:30 PM Sample Name : SW-7-38E Acg. Operator : Acg. Method : C:\hPCHEN\1\METHODS\SW : C:\HPCHEM\1\METHODS\SW.M

Last changed : 4/26/2010 10:35:52 PM (modified after loading) Analysis Method : C:\HPCHEM\l\METHODS\SW.M Last changed : 4/26/2010 10:52:02 PM

(modified after loading)



Location : Vial 1

-----Area Percent Report Inter resource repose

.....

C:\HPCHEM\1\METHODS\SW.M

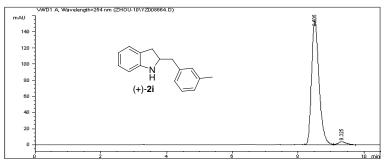
: 4/26/2010 10:20:45 PM (modified after loading)

Sorted Bv Multiplier Signal 1.0000 1.0000 Dilution

Signal 1: VWD1 A, Wavelength=254 nm

Totals : 747.26825 44.53579

Results obtained with enhanced integrator! *** End of Report ***



_____ Area Percent Report

Sorted Bv Multiplier Signal 1.0000 1.0000 Dilution

Signal 1: VWD1 A, Wavelength=254 nm

 Peak RetTime Type
 Width [min]
 Area [Height]
 Harea [Height]
 Area [Height]
 Area

Totals : 2480.20841 157.06700

Results obtained with enhanced integrator! *** End of Report ***

Instrument 1 4/26/2010 10:51:43 PM

Page 1 of 1

Instrument 1 4/26/2010 10:52:04 PM

Page 1 of 1

Sample Name: SW-7-38E

Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008667.D OD-H, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm

Injection Date : 4/26/2010 11:37:13 PM

: SW-7-24I

Analysis Method : C:\HPCHEM\1\METHOD\$\S\U.M Last changed : 4/26/2010 11:52:55 PM

Sample Name

Last changed

Acg. Operator Acg. Method

Sample Name: SW-7-24I

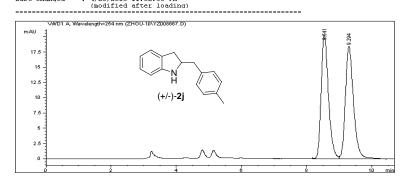
Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008661.D OD-H, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm

-----Location : Vial 1

Injection Date : 4/26/2010 9:45:55 PM Sample Name : SW-7--39F Acg. Operator : C:\HPCHEM\1\METHODS\SI

C:\HPCHEM\1\METHODS\SW.M 4/26/2010 9:20:58 PM (modified after loading) Last changed Analysis Method : C:\HPCHEM\l\METHODS\SW.M Last changed : 4/26/2010 11:53:13 PM

(modified after loading)



Location : Vial 1

______ Area Percent Report

: C:\HPCHEM\1\METHODS\SW.M

: 4/26/2010 11:32:57 PM (modified after loading)

Sorted Bv Multiplier Signal 1.0000 1.0000 Dilution

Signal 1: VWD1 A, Wavelength=254 nm

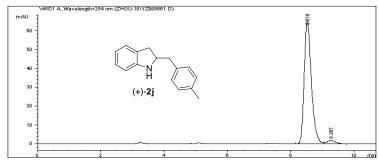
 Peak RetTime Type
 Width fmin
 Area fmin
 Height fmin
 Area fmin

 1
 8.541 VV
 0.2384 315.90903
 20.29078
 49.6497

 2
 9.294 VV
 0.2654 320.36688
 18.43404
 50.3503

Totals : 636.27591 38.72482

Results obtained with enhanced integrator! *** End of Report ***



_____ Area Percent Report

Sorted Bv Multiplier Signal 1.0000 1.0000 Dilution

Signal 1: VWD1 A, Wavelength=254 nm

Totals : 1064.28774 67.52117

Results obtained with enhanced integrator!

*** End of Report ***

Instrument 1 4/26/2010 11:52:59 PM

Page 1 of 1

Instrument 1 4/26/2010 11:53:16 PM

Page 1 of 1

Sample Name: SW-7--39F

Sample Name: SW-7-24J

Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008678.D

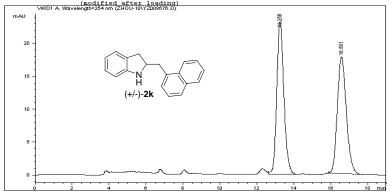
Sample Name: SW-7-38A

OD-H, H/i-PrOH =90/10, 0.8 mL/min, 30 oC, 254 nm

_____ Injection Date : 4/27/2010 11:55:30 AM Sample Name : SW-7-24J Location : Vial 1

Acq. Operator Acq. Method

: C:\HPCHEM\1\METHODS\DEF_LC.M



..... Area Percent Report

Sorted By Signal Multiplier Dilution 1.0000

Signal 1: VWD1 A, Wavelength=254 nm

Totals : 1223.57715 41.00914

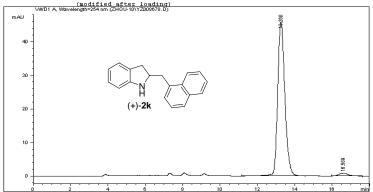
Results obtained with enhanced integrator! ______

*** End of Report ***

OD-H, H/i-PrOH =90/10, 0.8 mL/min, 30 oC, 254 nm

Injection Date : 4/27/2010 12:55:54 PM Sample Name : SW-7-38A

Acq. Operator Acq. Method : C:\HPCHEM\1\METHODS\DEF_LC.M : 4/27/2010 12:31:40 PM Last changed



Location : Vial 1

Area Percent Report

Sorted By Signal Multiplier Dilution

Signal 1: VWD1 A, Wavelength=254 nm

 Peak RetTime
 Type
 Width [min]
 Area [mak]
 Height Area [min]
 Area [mak]
 1 %

 1 13.260 BB
 0.4101 1221.06433
 45.79693
 97.8249

 2 16.589 BB
 0.5158 27.15018
 7.95293e-1
 2.1751

1248.21452 46.59022 Totals :

Results obtained with enhanced integrator!

*** End of Report ***

Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008647.D OD-H, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm

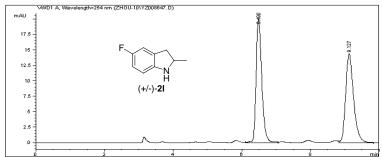
Sample Name: SW-6-93Al

Sample Name: SW-7-39J

Injection Date : 4/25/2010 10:01:32 PM Sample Name : SW-6-93Al Acc. Operator : Location : Vial 1

C:\HPCHEM\1\METHODS\SW.M Acc. Method : 4/25/2010 9:32:32 PM (modified after loading) Last changed Analysis Method : C:\HPCHEM\1\METHOD\$\5\W.M Last changed : 4/25/2010 11:23:43 PM

(modified after loading)



_____ Area Percent Report

Sorted Bv Multiplier Signal 1.0000 1.0000

Signal 1: VWD1 A, Wavelength=254 nm

Dilution

 Peak RetTime Type
 Width | Area | Fall | 1
 Height | Area |

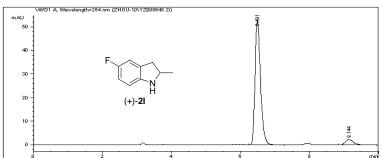
Totals : 456.76411 34.27350

Results obtained with enhanced integrator! *** End of Report *** Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008648.D OD-H, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm

Sample Name Acc. Operator Acq. Method C:\HPCHEM\1\METHODS\SW.M : 4/25/2010 10:58:32 PM (modified after loading) Last changed

Analysis Method : C:\HPCHEM\1\METHODS\SW.M Last changed : 4/25/2010 11:23:19 PM

(modified after loading)



_____ Area Percent Report

Sorted Bv Multiplier Signal 1.0000 1.0000 Dilution

Signal 1: VWD1 A, Wavelength=254 nm

Peak RetTime Type Width Area Height Area

Totals : 614.31222 54.75600

Results obtained with enhanced integrator! *** End of Report ***

Instrument 1 4/25/2010 11:24:00 PM

Page 1 of 1 Instrument 1 4/25/2010 11:23:42 PM Page 1 of 1

Injection Date : 4/27/2010 5:36:59 PM Sample Name : SW-6-92B1

Acq. Operator Acq. Method

Last changed

OD-H, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm

C:\HPCHEM\1\METHODS\DEF_LC.M

: 4/27/2010 4:44:38 PM

Sample Name: SW-6-92B1

mAU

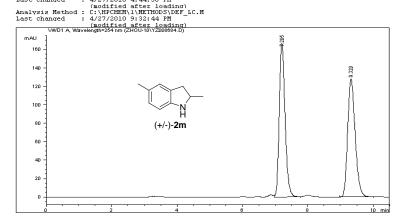
Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008581.D OD-H, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm

4/21/2010 11:44:04 PM (modified after loading) Last changed

Analysis Method : C:\HPCHEM\l\METHODS\SW.M Last changed : 4/23/2010 9:15:24 PM

/W/D1 A, Wavelength=254 nm (ZHOU-10\YZ008581.D)

(modified after loading)



Location : Vial 1

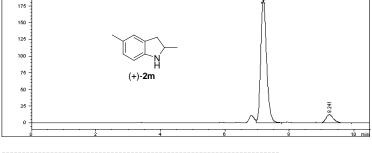
..... Area Percent Report

Sorted By Signal Multiplier Dilution

Signal 1: VWD1 A, Wavelength=254 nm

Totals : 4063.10010 294.67008

Results obtained with enhanced integrator! ______ *** End of Report ***



_____ Area Percent Report ------

Sorted Bv Multiplier Signal 1.0000 1.0000 Dilution

Signal 1: VWD1 A, Wavelength=254 nm

 Peak RetTime Type
 Width [min]
 Area [maid]
 Height Area
 Area
 Indicated
 I

Totals : 2495.37077 197.76667

Results obtained with enhanced integrator! *** End of Report *** Sample Name: SW-7-40P

Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008625.D IC, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm

Injection Date : 4/25/2010 1:54:48 PM Sample Name : SW-7-11D Acg. Operator : C:\hPCHEM\l\METHODS\S

Analysis Method : C:\HPCHEM\1\METHOD\$\S\W.M Last changed : 4/25/2010 2:25:58 PM

Last changed

20

Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008626.D IC, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm

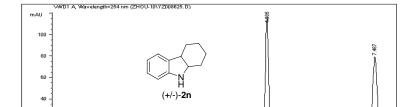
Sample Name: SW-7-11D

.....

Injection Date : 4/25/2010 2:10:28 PM Sample Name : SW-7-39K Acg. Operator : C:\HPCHEM\1\METHODS\SU

: C:\HPCHEM\1\METHODS\SW.M 4/25/2010 2:05:24 PM (modified after loading) Analysis Method : C:\HPCHEM\l\METHODS\SW.M Last changed : 4/25/2010 2:25:33 PM

(modified after loading)



Location : Vial 1

_____ Area Percent Report

(modified after loading)

: C:\HPCHEM\1\METHODS\SW.M

: 4/25/2010 1:30:54 PM (modified after loading)

Sorted Bv Multiplier Signal 1.0000 1.0000 Dilution

Signal 1: VWD1 A, Wavelength=254 nm

Totals : 1254.40228 194.71261

Results obtained with enhanced integrator! *** End of Report ***

WWD1 A, Wavelength=254 nm (ZHOU-10\YZ008626.D) mAU 140 120 100 80 60 20

Location : Vial 1

_____ Area Percent Report mad Tarotho Naporo

Sorted Bv Multiplier Signal 1.0000 1.0000 Dilution

Signal 1: VWD1 A, Wavelength=254 nm

 Peak RetTime Type
 Width [min]
 Area [midt]
 Height Area
 Area [midt]
 Area [midt]
 4.874
 Area [midt]
 4.974
 Area [midt]
 4.974

Totals : 962.24188 174.14795

Results obtained with enhanced integrator! *** End of Report ***

Instrument 1 4/25/2010 2:26:01 PM Page 1 of 1 Instrument 1 4/25/2010 2:25:36 PM Page 1 of 1

Sample Name: SW-7-39K

Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008669.D IC, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm

Analysis Method : C:\HPCHEM\1\METHOD\$\5\W.M Last changed : 4/27/2010 12:48:01 AM

Sample Name Acc. Operator Acq. Method

Last changed

Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008544.D IC, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm Sample Name: SW-7-23B

Last changed

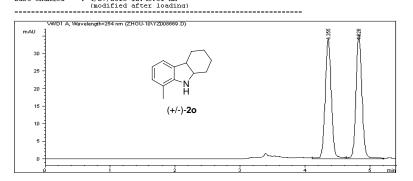
_____ Location : Vial 1

Injection Date : 4/19/2010 3:50:58 PM Sample Name : SW-7-39L Acg. Operator : C:\HPCHEN\1\METHODS\S

C:\HPCHEM\1\METHODS\SW.M : 4/19/2010 3:47:09 PM (modified after loading)

Analysis Method : C:\HPCHEM\1\METHODS\5W.M Last changed : 4/27/2010 12:49:41 AM

(modified after loading)



_____ Area Percent Report

: C:\HPCHEM\1\METHODS\SW.M

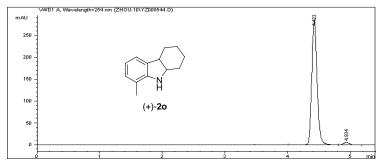
: 4/26/2010 11:58:54 PM (modified after loading)

Sorted Bv Multiplier Signal 1.0000 1.0000 Dilution

Signal 1: VWD1 A, Wavelength=254 nm

Totals : 453.57089 69.33657

Results obtained with enhanced integrator! *** End of Report ***



______ Area Percent Report

Sorted Bv Multiplier Signal 1.0000 1.0000 Dilution

Signal 1: VWD1 A, Wavelength=254 nm

 Peak RetTime Type
 Width [min]
 Area [midt]
 Height Area

 # [min]
 [min]
 *s [mad]
 *s [mad]

 1
 4.423 VV
 0.0936 1703.24229
 282.84137 97.7914

 2
 4.934 VV
 0.0992 38.46766
 5.91249 2.2086

Totals : 1741.71094 288.75386

Results obtained with enhanced integrator! *** End of Report ***

Instrument 1 4/27/2010 12:49:24 AM

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Instrument 1 4/27/2010 12:49:44 AM

Page 1 of 1

Sample Name: SW-7-39L

Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008627.D IC, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm

Injection Date : 4/25/2010 2:33:22 PM Sample Name : SW-7-4B1 Acc. Operator :

Analysis Method : C:\HPCHEM\1\METHOD\$\5\W.M Last changed : 4/25/2010 2:55:24 PM

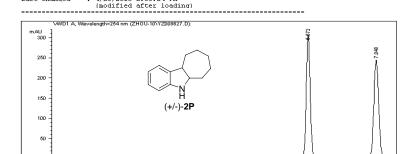
Acc. Method

Last changed

Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008628.D IC, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm Sample Name: SW-7-4B1

: 4/25/2010 2:43:54 PM (modified after loading) Last changed Analysis Method : C:\HPCHEM\1\METHODS\SW.M Last changed : 4/25/2010 2:55:21 PM

(modified after loading)



Location : Vial 1

_____ Area Percent Report ------

C:\HPCHEM\1\METHODS\SW.M

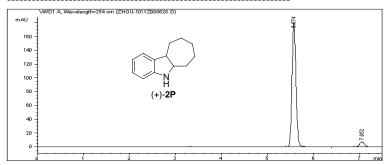
: 4/25/2010 2:26:02 PM (modified after loading)

Sorted Bv Multiplier Signal 1.0000 1.0000 Dilution

Signal 1: VWD1 A, Wavelength=254 nm

Totals : 3658.27271 550.92253

Results obtained with enhanced integrator! *** End of Report ***



_____ Area Percent Report med letotho hapoto

Sorted Bv Multiplier Signal 1.0000 1.0000

Signal 1: VWD1 A, Wavelength=254 nm

Dilution

Totals : 1122.40987 186.90912

Results obtained with enhanced integrator! *** End of Report ***

Instrument 1 4/25/2010 2:55:42 PM

Page 1 of 1

Instrument 1 4/25/2010 2:55:23 PM

Page 1 of 1

Sample Name: SW-7-40M

Sample Name: SW-7-40N+-Data File C:\HPCHEM\1\DATA\ZHOU-10\YZ008691.D Sample Name: SW-7-40N

Page 1 of 1

OJ-H, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm

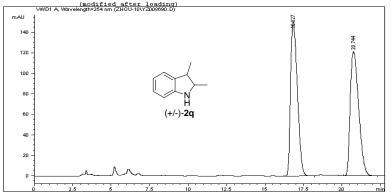
_____ Injection Date : 4/27/2010 11:10:18 PM Sample Name : SW-7-40N+-Location : Vial 1

Acq. Operator Acq. Method

C:\HPCHEM\1\METHODS\DEF_LC.M Analysis Method: (-Y427/2010 11:32:29 PM | modified after loading)

Analysis Method: (-YHFCHEMYINETHODS)VDFF_LC.M
Last changed: 4/27/2010 11:38:59 PM | modified after loading)

WWD1 A Wavelength-254 nm (ZHOU-10Y/ZD06600.D)



..... Area Percent Report

Sorted By Signal Multiplier Dilution 1.0000

Signal 1: VWD1 A, Wavelength=254 nm

Totals : 9239.93311 270.98679

Results obtained with enhanced integrator! ______ *** End of Report ***

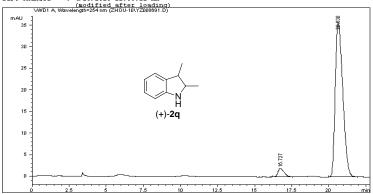
OJ-H, H/i-PrOH =99/1, 1.0 mL/min, 30 oC, 254 nm

______ Injection Date : 4/27/2010 11:36:01 PM Sample Name : SW-7-40N Location : Vial 1

Acq. Operator Acq. Method

C:\HPCHEM\1\METHODS\DEF_LC.M Last changed : 4/27/2010 11:39:02 PM

Analysis Method: C:\HFCHEM\;\METHOS\DEF_LC.M Last changed : C:\HFCHEM\;\METHOS\DEF_LC.M Last changed : 4/28/2010 12:00:22 AM \text{(aodified after loading)} \text{\text{WD1A \text{\text{WaveIndex}}}



Area Percent Report

Sorted By Signal Multiplier 1.0000 Dilution

Signal 1: VWD1 A, Wavelength=254 nm

 Peak RetTime Type
 Width [min]
 Area [max]
 Height [max]
 Area [max]
 Area [max]
 1
 Area [max]
 Area [

Totals : 1316.34644 37.50347

Results obtained with enhanced integrator! _____ *** End of Report ***

Instrument 1 4/27/2010 11:39:02 PM Page 1 of 1 Instrument 1 4/28/2010 12:00:24 AM