

Enzymatic Dynamic Kinetic Resolution of (\pm)-*cis*-*N*-(Alkoxycarbonyl)cyclopentane-1,2-diamines based on Spontaneous Racemization

F. Javier Quijada, Vicente Gotor, and Francisca Rebolledo**

Departamento de Química Orgánica e Inorgánica, and Instituto Universitario de Biotecnología de Asturias, Universidad de Oviedo, 33071-Oviedo, Spain

vgs@uniovi.es frv@uniovi.es

SUPPORTING INFORMATION (page 1 of 39 pages)

Table of Contents

- General Spectroscopic and Experimental Data (S2).
- Experimental Procedures. Spectroscopic Data:
 - *tert*-Butyl (\pm)-*trans*-(2-hydroxycyclopentyl)carbamate, (\pm)-**2a** (S2).
 - General procedure for the synthesis of carbamates (\pm)-**2b-d** (S3).
 - Alkyl (\pm)-*cis*-(2-azidocyclopentyl)carbamates, (\pm)-**3a-d**. General Procedure (S3).
 - *tert*-Butyl (\pm)-*cis*-(2-aminocyclopentyl)carbamate, (\pm)-**5a** (S5).
 - General procedure for the Staudinger reduction of (\pm)-*cis*-(2-azidocyclopentyl)carbamates **3b-d** (S5).
 - Enzymatic kinetic resolution of *tert*-butyl (\pm)-*cis*-(2-aminocyclopentyl)carbamate, (\pm)-**5a** (S6).
 - Enzymatic dynamic kinetic resolution of alkyl (\pm)-*cis*-(2-aminocyclopentyl)carbamates (\pm)-**5a-d** (S6).
 - Determination of the enantiomeric excesses (S8).
 - Monitorization of the enzymatic DKRs of alkyl (\pm)-*cis*-(2-aminocyclopentyl)carbamates (\pm)-**5b-d** (S8)
- Copy of ^1H and ^{13}C NMR Spectra of:

2c (S10, S11), **2d** (S12, S13), **3b** (S14, S15), **3c** (S16, S17), **3d** (S18, S19), **5a** (S20, S21), **5b** (S22, S23), **5c** (S24, S25), **5d** (S26, S27), **6a** (S28, S29), **6b** (S30, S31), **6c** (S32, S33), **6d** (S34, S35).

- Copy of Chiral HPLC Chromatograms of:

(±)-**6a** (S36), (1*S*,2*R*)-**6a** (S36), (1*R*,2*S*)-**6a** (S36); (±)-**6b** (S37), (1*S*,2*R*)-**6b** (S37), (±)-**6c** (S38), (1*S*,2*R*)-**6c** (S38), (±)-**6d** (S39), (1*S*,2*R*)-**6d** (S39).

General Spectroscopic and Experimental Data

Lipase B from *Candida antarctica* (CAL-B, available immobilized on polyacrylamide as Novozyme 435, 7300 PLU/g) was supplied by Novo Nordisk Co. For the enzymatic reactions, ethyl acetate of spectrophotometric grade (stored with 4 Å molecular sieves), other anhydrous solvents and (±)-1-phenylethyl acetate were used. Thin-layer chromatography was performed on precoated TLC plates of Merck silica gel 60F₂₅₄, using a potassium permanganate solution as developing reagent. Merck silica gel 60 (particle size, 40 - 63 µm) was used for column chromatography. Optical rotations were measured at the sodium D line at 20 °C. Mass spectra (*m/z*) were recorded in ElectronSpray Ionisation (ESI) and Electron Impact (EI). ¹H NMR and proton-decoupled ¹³C NMR spectra (CDCl₃ solutions) were recorded using AC-300 or DPX-300 (¹H, 300.13 MHz and ¹³C, 75.5 MHz) and AV-400 MHz (¹H, 400.13 MHz and ¹³C, 100.63 MHz) spectrometers using the δ scale (ppm) for chemical shifts; calibration was made on the CDCl₃ (¹³C, 76.95 ppm) or the residual CHCl₃ (¹H, 7.26 ppm).

Experimental Procedures

tert-Butyl (±)-trans-(2-hydroxycyclopentyl)carbamate, (±)-2a. (±)-*trans*-2-Aminocyclopentanol hydrochloride (12.0 mmol) was dissolved in methanol (40 mL) and Na₂CO₃ (12.0 mmol) and di-*tert*-butyl dicarbonate (14.0 mmol) were added to the solution. After stirring 24 h at room temperature, solvent was evaporated and the residue extracted with CH₂Cl₂. The usual work-up of the organic phase gave pure (±)-**2a** with 83% yield. M.p.: 105.6-106.8 °C (Lit.¹ m.p.: 103 °C). Spectroscopic data are in good agreement with those previously published.¹

General procedure for the synthesis of carbamates (±)-2b-d.

¹ Page, M. F. Z.; Jalisatgi, S. S.; Maderna, A.; Hawthorne, M. F. *Synthesis* **2008**, 555-563.

To a solution of 2-aminocyclopentanol hydrochloride (7.3 mmol) in water (15 mL), sodium carbonate (14.6 mmol) and the corresponding alkyl chloroformate (8.7 mmol) were added. After 24 h stirring at room temperature, the reaction mixture was extracted with CH₂Cl₂ (3 × 15 mL). The organic layers were combined, washed with brine (20 mL), dried with Na₂SO₄, and evaporated under reduced pressure to give the corresponding pure product, except in the case of (±)-**2b**.

Benzyl (±)-*trans*-(2-hydroxycyclopentyl)carbamate, (±)-2b. The solid crude material was repeatedly washed with warm hexane to remove benzylic alcohol. Thus, pure product was isolated as a white solid. Yield: 94%; m.p.: 59.1-60.4 °C (Lit.² m.p.: 57-59 °C). Spectroscopic data were in good agreement with the literature.²

Allyl (±)-*trans*-(2-hydroxycyclopentyl)carbamate, (±)-2c. Yield: 97%; colourless oil; ¹H NMR (300 MHz): δ (ppm) = 1.38 (m, 1H), 1.54-1.82 (m, 3H), 1.90-2.16 (m, 2H), 3.58-3.80 [m + br s, 2H, CH-NH + OH (3.75)], 4.00 (q, 1H, ³J = 6.3 Hz, CH-OH), 4.55 (d, 2H, ³J = 5.5 Hz, CH₂-O), 4.93 (br s, 1H, NH), 5.21 (dd, 1H, ³J_{cis} = 10.4 Hz, ¹J = 1.0 Hz, =CHH), 5.30 (dd, 1H, ³J_{trans} = 17.2 Hz, ¹J = 1.0 Hz, =CHH), 5.90 (m, 1H, HC=); ¹³C NMR (100.6 MHz): δ (ppm) = 20.4 (CH₂), 29.7 (CH₂), 31.6 (CH₂), 59.9 (CH), 65.4 (CH₂), 78.2 (CH), 117.4 (CH₂), 132.4 (CH), 156.9 (C=O); MS (ESI), *m/z* (%) = 186 (100) [M + H]⁺; elemental analysis (%) calcd. for C₉H₁₅NO₃: C, 58.36; H, 8.16; N, 7.56; found: C, 58.15; H, 8.34; N, 7.49.

Ethyl (±)-*trans*-(2-hydroxycyclopentyl)carbamate, (±)-2d. Yield: 92%; colourless oil; ¹H NMR (300 MHz): δ (ppm) = 1.23 (t, 3H, ³J = 7.1 Hz, CH₃), 1.28-1.45 (m, 1H), 1.57-1.83 (m, 3H), 1.89-2.17 (m, 2H), 3.66 (m, 1H, CH-N), 3.88 (br s, 1H, OH), 3.98 (q, 1H, ³J = 6.3 Hz, CH-O), 4.10 (q, 2H, ³J = 7.1 Hz, CH₂-O), 4.87 (br s, 1H, NH); ¹³C NMR (100.6 MHz): δ (ppm) = 14.3 (CH₃), 20.5 (CH₂), 29.9 (CH₂), 31.8 (CH₂), 60.0 (CH), 60.8 (CH₂), 78.6 (CH), 157.4 (C=O); MS (ESI), *m/z* (%) = 174 (100) [M + H]⁺, 258 (48) [2M – NHCOOEt]⁺; elemental analysis (%) calcd. for C₈H₁₅NO₃: C, 55.47; H, 8.73; N, 8.09; found: C, 55.21; H, 8.45; N, 8.38.

Alkyl (±)-*cis*-(2-azidocyclopentyl)carbamates. General Procedure.

Triethylamine (30.0 mmol) and the corresponding alkyl (±)-*trans*-(2-hydroxycyclopentyl)carbamate (±)-**2a-d** (10.0 mmol) were dissolved in CH₂Cl₂ (12 mL). The solution was cooled at 0 °C, and methanesulfonyl chloride (16.0 mmol) was added, under a nitrogen atmosphere, during 10 min. After 3 h at 0 °C (for **2a**) or 50 min at rt (for **2b-d**), the solvent was eliminated under reduced pressure. The crude

² Maestro, A.; Astorga, C.; Gotor, V. *Tetrahedron: Asymmetry* **1997**, 8, 3153-3159.

was dissolved in CH₂Cl₂ and the resulting organic solution successively washed with 1 M aq. NaHSO₄, water and brine. After dried with Na₂SO₄, solvent was evaporated under reduced pressure yielding the corresponding (±)-*trans*-mesylate, which was used without purification in the following reaction. Thus, a mixture of the crude mesylate and NaN₃ (30.0 mmol) was dissolved in anhydrous DMF (17 mL) and the solution was heated at 50 °C for 24 h. After this time, CH₂Cl₂ was added and the organic solution repeatedly washed with water. The organic layer was dried with Na₂SO₄, the solvents were eliminated, and the resulting crude was purified by flash chromatography (hexane-ethyl acetate 6:1) to yield the pure product.

***tert*-Butyl (±)-*cis*-(2-azidocyclopentyl)carbamate, (±)-3a.** Yield: 54%; m.p.: 75.3-76.4 °C. Spectroscopic data are in good agreement with those previously published for optically active **3a**.³

Benzyl (±)-*cis*-(2-azidocyclopentyl)carbamate, (±)-3b. Yield: 69%; colourless oil; ¹H NMR (300 MHz): δ (ppm) = 1.45 (m, 1H), 1.63 (m, 1H), 1.72-2.06 (m, 4H), 4.03 (m, 2H, H-1 and H-2), 5.03 (br s, 1H, NH), 5.11 (AB system, 2H, ¹J = 12.3 Hz, CH₂-O), 7.36 (m, 5H); ¹³C NMR (75.5 MHz): δ (ppm) = 19.7 (CH₂), 28.6 (CH₂), 28.8 (CH₂), 54.9 (CH), 64.0 (CH), 66.6 (CH₂), 127.89 (CH), 127.92 (CH), 128.3 (CH), 136.2 (C), 155.7 (C=O); MS (ESI), *m/z* (%) = 283 (92) [M + Na]⁺, 233 (45) 102 (100); elemental analysis calcd. for C₁₃H₁₆N₄O₂: C, 59.99; H, 6.20; N, 21.52; found: C, 60.27; H, 6.03; N, 21.80.

Allyl (±)-*cis*-(2-azidocyclopentyl)carbamate, (±)-3c. Yield: 67%; colourless oil; ¹H NMR (400 MHz): δ (ppm) = 1.45 (m, 1H), 1.56-1.64 (m, 1H), 1.72-2.11 (m, 4H), 4.01 (br s, 2H, H-1 and H-2), 4.56 (d, 2H, ³J = 5.4 Hz, CH₂-O), 5.00 (br s, 1H, NH), 5.20 (dd, 1H, ³J_{cis} = 10.4 Hz, ¹J = 1.4 Hz, =CHH), 5.30 (dd, 1H, ³J_{trans} = 17.2 Hz, ¹J = 1.4 Hz, =CHH), 5.91 (m, 1H, HC=); ¹³C NMR (100.6 MHz): δ (ppm) = 19.7 (CH₂), 28.5 (CH₂), 28.8 (CH₂), 54.9 (CH), 64.0 (CH₂), 65.4 (CH), 117.4 (CH₂), 132.6 (CH), 155.6 (C=O); MS (ESI), *m/z* (%) = 211 (60) [M + H]⁺, 183 (100); elemental analysis calcd. for C₉H₁₄N₄O₂: C, 51.42; H, 6.71; N, 26.65; found: C, 51.26; H, 6.97; N, 26.43.

Ethyl (±)-*cis*-(2-azidocyclopentyl)carbamate, (±)-3d. Yield: 68%; colourless oil; ¹H NMR (400 MHz): δ (ppm) = 1.21 (t, 3H, ³J = 7.1 Hz), 1.36-1.48 (m, 1H), 1.52-1.64 (m, 1H), 1.72-1.96 (m, 4H), 3.98 (m, 2H), 4.08 (q, 2H, ³J = 7.1 Hz), 4.99 (br s, 1H, NH); ¹³C NMR (100.6 MHz): δ (ppm) = 14.4 (CH₃), 19.8 (CH₂), 28.7 (CH₂), 28.9 (CH₂), 54.9 (CH), 60.8 (CH₂), 64.1 (CH), 156.0 (C=O); MS (ESI), *m/z* (%) = 199 (58) [M + H]⁺, 171 (100); elemental analysis calcd. for C₈H₁₄N₄O₂: C, 48.47; H, 7.12; N, 28.26; found: C, 48.25; H, 7.00; N, 28.39.

³ Govindaraju, T.; Kumar, V. A.; Ganesh, K. N. *J. Org. Chem.* **2004**, *69*, 5725-5734.

***tert*-Butyl (±)-*cis*-(2-aminocyclopentyl)carbamate, (±)-5a.** A suspension of (±)-3a (2.20 mmol) and Pd-C (10%, 230 mg) in deoxygenated methanol (30 mL) was stirred for 23 h under a hydrogen atmosphere. The reaction mixture was filtered through Celite[®], and the filtrate was evaporated to yield a crude, which was purified by flash chromatography (ethyl acetate and ethyl acetate:methanol 4:1 were successively used as eluents). Yield: 79%; colourless oil; ¹H NMR (400 MHz): δ (ppm) = 1.30-1.55 (m + s, 12H. Singlet to 1.39 corresponds to Bu^t), 1.69 (m, 1H), 1.89 (m, 2H), 2.23 (br s, 2H, NH₂), 3.29 (q, 1H, ³*J* = 5.7 Hz, H-2), 3.76 (br m, 1H, H-1), 5.14 (br s, 1H, NH); ¹³C NMR (100.6 MHz): δ (ppm) = 20.2 (CH₂), 28.2 (3×CH₃), 29.7 (CH₂), 32.4 (CH₂), 53.0 (CH), 54.6 (CH), 78.9 (C), 155.8 (C=O); MS (ESI), *m/z* (%) = 201 (100) [M + H]⁺; elemental analysis calcd. for C₁₀H₂₀N₂O₂: C, 59.97; H, 10.07; N, 13.99; found: C, 60.23; H, 10.33; N, 13.81.

General procedure for the Staudinger reduction of (±)-*cis*-(2-azidocyclopentyl)carbamates 3b-d.

Triphenylphosphine (4.32 g, 16.5 mmol) was added to a solution of the corresponding azidocarbamate (±)-3b-d (13.8 mmol) in THF-H₂O (10:1, 55 mL). After stirring at room temperature during 72h, solvents were eliminated under reduced pressure and the resulting crude was purified by flash chromatography (a gradient of ethyl acetate to ethyl acetate-methanol was used as eluent).

Benzyl (±)-*cis*-(2-aminocyclopentyl)carbamate, (±)-5b. Yield: 89%; white solid; m.p.: 61.6-63.0 °C; ¹H NMR (300 MHz): δ (ppm) = 1.30-1.62 [m + s, 5H. Singlet corresponds to NH₂ (1.45)], 1.74 (m, 1H), 1.84-2.02 (m, 2H), 3.34 (m, 1H, H-2), 3.84 (m, 1H, H-1), 5.09 (s, 2H, CH₂-O), 5.38 (br d, 1H, ³*J* = 6.5 Hz, NH), 7.20-7.40 (m, 5H); ¹³C NMR (75 MHz): δ (ppm) = 19.6 (CH₂), 28.8 (CH₂), 31.8 (CH₂), 52.5 (CH), 54.6 (CH), 65.6 (CH₂), 127.2 (CH), 127.3 (CH), 127.7 (CH), 136.0 (C), 155.7 (C=O); MS (ESI), *m/z* (%) = 235 (100) [M + H]⁺, 257 (40) [M + Na]⁺, 469 (60) [2M + H]⁺; elemental analysis calcd. for C₁₃H₁₈N₂O₂: C, 66.64; H, 7.74; N, 11.96; found: C, 66.39; H, 7.98; N, 12.15.

Allyl (±)-*cis*-(2-aminocyclopentyl)carbamate, (±)-5c. Yield: 84%; colourless oil; ¹H NMR (400 MHz): δ (ppm) = 1.39 (m, 1H), 1.45-1.59 (m, 4H, NH₂ + 2H), 1.64-1.78 (m, 1H), 1.84-1.98 (m, 2H), 3.35 (q, 1H, ³*J* = 5.6 Hz, H-2), 3.81 (br m, 1H, H-1), 4.55 (br s, 2H, CH₂-O), 5.19 (d, 1H, ³*J*_{cis} = 10.3 Hz, =CHH), 5.29 (dd, 1H, ³*J*_{trans} = 17.2 Hz, ²*J* = 1.3 Hz, =CHH), 5.35 (br s, 1H, NH), 5.91 (ddt, 1H, ³*J*_{trans} = 17.2 Hz, ³*J*_{cis} = 10.3 Hz, ³*J* = 5.6 Hz, HC=); ¹³C NMR (100.6 MHz): δ (ppm) = 19.6 (CH₂), 28.8 (CH₂), 31.7 (CH₂), 52.4 (CH), 54.4 (CH), 64.4 (CH₂), 116.4 (CH₂), 132.3 (CH), 155.5 (C=O); MS (ESI), *m/z* (%) = 185 (100) [M + H]⁺; elemental analysis calcd. for C₉H₁₆N₂O₂: C, 58.67; H, 8.75; N, 15.21; found: 58.39; H, 8.97; N, 15.45.

Ethyl (±)-*cis*-(2-aminocyclopentyl)carbamate, (±)-5d. Yield: 79%; colourless oil; ¹H NMR (400 MHz): δ (ppm) = 1.22 (t, 3H, ³*J* = 7.1 Hz), 1.37 (m, 1H), 1.56 (m, 2H), 1.62-1.80 [br s + m, 3H, br s corresponds to NH₂ (1.66)], 1.94 (m, 2H), 3.33 (q, 1H, ³*J* = 5.6 Hz, H-2), 3.81 (br m, 1H, H-1), 4.08 (q, 2H, ³*J* = 7.1 Hz), 5.25 (br d, 1H, NH, ³*J* = 6.1 Hz); ¹³C NMR (100.6 MHz): δ (ppm) = 14.2 (CH₃), 20.0 (CH₂), 29.4 (CH₂), 32.4 (CH₂), 52.8 (CH), 54.8 (CH), 60.1 (CH₂), 156.2 (C=O); MS (ESI), *m/z* (%) = 173 (100) [M + H]⁺; elemental analysis calcd. for C₈H₁₆N₂O₂: C, 55.79; H, 9.36; N, 16.27; found: C, 55.68; H, 9.59; N, 16.08.

Enzymatic kinetic resolution of *tert*-butyl (±)-(2-aminocyclopentyl)carbamate, (±)-5a.

tert-Butyl methyl ether (5.0 mL) and (±)-1-phenylethyl acetate (3.0 mmol) were added under nitrogen atmosphere to a mixture of racemic amino carbamate (±)-5a (200 mg, 1.0 mmol), CAL-B (100 mg) and 4 Å molecular sieves (25 mg). The suspension was shaken at 28 °C and 200 rpm during 6 days. The solvent was removed under vacuum, CH₂Cl₂ was added to the suspension and the enzyme filtered. After, the enzyme was successively washed with CH₂Cl₂ and aq. 3N NaOH. Both layers were separated and the aqueous phase was extracted with CH₂Cl₂ (2 × 15 mL). All the organic layers were combined, dried with Na₂SO₄ and concentrated in vacuo to give a residue. Flash chromatography (hexane:ethyl acetate 1:1, ethyl acetate and ethyl acetate:methanol 4:1 were successively used as eluents) of the residue yielded pure enantioenriched compounds 5a and 6a.

***tert*-Butyl (1*R*,2*S*)-(2-aminocyclopentyl)carbamate, (1*R*,2*S*)-5a.** Yield: 41%. [α]_D²⁰ = +7.8, ee = 97%.

***tert*-Butyl (1*S*,2*R*)-[(2-acetylamino)cyclopentyl]carbamate, (1*S*,2*R*)-6a.** Yield: 45%; white solid; m.p.: 118.9-120.2 °C; [α]_D²⁰ = +11.4 (*c* 1.0 in CHCl₃), ee = 98%; ¹H NMR (300 MHz): δ (ppm) = 1.40-1.85 [m + s, 13H. Singlet corresponds to Bu^t (1.44)], 1.90-2.20 [m + s, 5H. Singlet corresponds to CH₃ (1.96)], 3.94 (quintet, 1H, ³*J* = 6.2 Hz, H-1), 4.13 (br m, 1H, H-2), 4.76 (br s, 1H, NH), 6.11 (br s, 1H, NH); ¹³C NMR (100.6 MHz): δ (ppm) = 20.0 (CH₂), 23.1 (CH₃), 28.2 (3×CH₃), 29.4 (CH₂), 29.8 (CH₂), 52.9 (CH), 53.4 (CH), 79.2 (C), 156.2 (C=O), 170.4 (C=O, Ac); MS (ESI), *m/z* (%) = 243 (45) [M + H]⁺, 485 (100) [2M + Na]⁺; HRMS (EI) calcd. for C₁₀H₁₇NO₂ [M - (CH₂=C=O) - (NH₃)]^{•+}: 183.1259; found: 183.1255. Elemental analysis calcd. for C₁₂H₂₂N₂O₃: C, 59.48; H, 9.15; N, 11.56; found: C, 59.63; H, 9.02; N, 11.79.

Enzymatic dynamic kinetic resolution of alkyl (±)-(2-aminocyclopentyl)carbamates (±)-5a-d.

To a mixture of racemic amino carbamate (±)-5a-d (1.2 mmol) and CAL-B (120 mg) under a nitrogen atmosphere, a mixture of anhydrous TBME-Et₃N (10:1; 7.2 mL) and (±)-1-phenylethyl acetate (575 μ L,

3.6 mmol) were added. The suspension was circularly shaken at 50 °C and 200 rpm during 9 days. After, the enzyme was filtered and washed with methanol. Once solvents were eliminated, the crude was submitted to flash chromatography (a gradient of hexane-ethyl acetate 10:1 to ethyl acetate was used as eluent) to yield pure the corresponding acetamide.

Benzyl (1*S*,2*R*)-[(2-acetylamino)cyclopentyl]carbamate, (1*S*,2*R*)-6b. Yield: 94%; white solid; m.p.: 143.3-144.7 °C; $[\alpha]_{\text{D}}^{20} = +61.2$ (*c* 1.0 in CHCl₃), ee = 96%; ¹H NMR (300 MHz): δ (ppm) = 1.42-1.75 (m, 4H), 1.91 (s, 3H), 2.07 (m, 2H), 4.02 (quintet, 1H, ³*J* = 5.8 Hz), 4.18 (quintet, 1H, ³*J* = 5.8 Hz), 5.02 (br s, 1H, NH), 5.10 (s, 2H, CH₂-O), 5.98 (br s, 1H, NH), 7.35 (m, 5H, Ph); ¹³C NMR (75.5 MHz): δ (ppm) = 20.1 (CH₂), 23.1 (CH₃), 29.4 (CH₂), 29.9 (CH₂), 53.0 (CH), 54.1 (CH), 66.7 (CH₂), 128.0 (CH), 128.4 (CH), 136.3 (C), 156.6 (C=O), 170.6 (C=O, Ac); MS (ESI), *m/z* (%) = 277 (100) [M + H]⁺, 553 (38) [2M + H]⁺; HRMS (EI) calcd. for C₁₅H₂₀N₂O₃ (M^{•+}): 276.1474; found: 276.1479.

Allyl (1*S*,2*R*)-[(2-acetylamino)cyclopentyl]carbamate, (1*S*,2*R*)-6c. Yield: 85%; white solid; m.p.: 107.2-108.5 °C; $[\alpha]_{\text{D}}^{20} = +20.4$ (*c* 1.0 in CHCl₃), ee = 97%; ¹H NMR (300 MHz): δ (ppm) = 1.42-1.78 (m, 4H), 1.90-2.20 [m + s, 5H. Singlet corresponds to CH₃ (2.00)], 4.01 (br s, 1H, H-1), 4.19 (br m, 1H, H-2), 4.56 (d, 2H, ³*J* = 5.5 Hz), 5.02 (br s, 1H, NH), 5.22 (dd, 1H, ³*J*_{cis} = 10.4 Hz, ²*J* = 1.4 Hz, =CHH), 5.32 (dd, 1H, ³*J*_{trans} = 17.2 Hz, ²*J* = 1.4 Hz, =CHH), 5.92 (m, 1H, =CH), 6.13 (br s, 1H, NH); ¹³C NMR (75.5 MHz): δ (ppm) = 20.1 (CH₂), 23.1 (CH₃), 29.4 (CH₂), 30.0 (CH₂), 53.0 (CH), 54.0 (CH), 65.6 (CH₂), 117.7 (CH₂), 132.7 (CH), 156.5 (C=O), 170.7 (C=O, Ac); MS (ESI), *m/z* (%) = 249 (80) [M + Na]⁺, 475 (100) [2M + Na]⁺; HRMS (EI) calcd. for C₉H₁₃NO₂ [M – (CH₂=C=O) – (NH₃)]^{•+}: 167.0946; found: 155.0949. Elemental analysis calcd. for C₁₁H₁₈N₂O₃: C, 58.39; H, 8.02; N, 12.38; found: C, 58.12; H, 8.31; N, 12.09.

Ethyl (1*S*,2*R*)-[(2-acetylamino)cyclopentyl]carbamate, (1*S*,2*R*)-6d. Yield: 72%; white solid; m.p.: 125.6-127.3 °C; $[\alpha]_{\text{D}}^{20} = +19.9$ (*c* 1.0 in CHCl₃), ee = 95%; ¹H NMR (300 MHz): δ (ppm) = 1.24 (t, 3H, ³*J* = 7.1 Hz), 1.42-1.80 (m, 3H), 1.90-2.18 [m + s, 5H. Singlet corresponds to CH₃ (1.97)], 3.99 (quintet, 1H, ³*J* = 6.1 Hz), 4.10-4.30 [q + m, 3H. Quartet corresponds to CH₂-O (4.11 ppm, ³*J* = 7.2 Hz)], 5.03 (br s, 1H, NH), 6.10 (br s, 1H, NH); ¹³C NMR (75.5 MHz): δ (ppm) = 14.5 (CH₃), 20.0 (CH₂), 23.1 (CH₃), 29.5 (CH₂), 30.0 (CH₂), 52.8 (CH), 53.8 (CH), 60.7 (CH₂), 156.9 (C=O), 170.7 (C=O, Ac); MS (ESI), *m/z* (%) = 237 (100) [M + Na]⁺; HRMS (EI) calcd. for C₈H₁₃NO₂ [M – (CH₂=C=O) – (NH₃)]^{•+}: 155.0946; found: 155.0951. Elemental analysis calcd. for C₁₀H₁₈N₂O₃: C, 56.06; H, 8.47; N, 13.07; found: C, 56.29; H, 8.53; N, 12.89.

Determination of the enantiomeric excesses.

The ee for each optically active compound involved in the enzymatic reactions was determined by chiral HPLC using Chiralpak IA (25 cm × 4.6 mm i.d.) or Chiralpak AS (25 cm × 4.6 mm i.d.) columns.

The products isolated from the enzymatic reactions, that is, acetamides, were directly analyzed:

For *tert*-butyl (±)-2-[(acetylamino)cyclopentyl]carbamate: Chiralpak AS, hexane/ethanol 93:7, 0.8 mL/min, 20 °C; t_R = 10.0 (1*S*,2*R*) and 11.4 (1*R*,2*S*) min; R_S = 1.3.

For benzyl (±)-2-[(acetylamino)cyclopentyl]carbamate: Chiralpak AS, hexane/ethanol 90:10, 0.8 mL/min, 20 °C; t_R = 13.9 (1*S*,2*R*) and 18.7 (1*R*,2*S*) min; R_S = 2.3. At 30 °C; t_R = 13.0 (1*S*,2*R*) and 16.4 (1*R*,2*S*) min; R_S = 2.9.

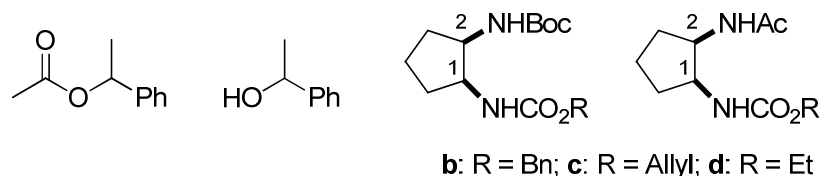
For allyl (±)-2-[(acetylamino)cyclopentyl]carbamate: Chiralpak IA, hexane/ethanol 94:6, 1.0 mL/min, 25 °C; t_R = 14.3 (1*S*,2*R*) and 15.9 (1*R*,2*S*) min; R_S = 1.5.

For ethyl (±)-2-[(acetylamino)cyclopentyl]carbamate: Chiralpak IA, hexane/ethanol 96:4, 0.9 mL/min, 25 °C; t_R = 24.3 (1*S*,2*R*) and 26.3 (1*R*,2*S*) min; R_S = 1.3.

In order to analyze the ee of the remaining substrate isolated in the reaction of amino-Boc **5a**, it was previously transformed into the acetamide (acetyl chloride, Et₃N, CH₂Cl₂). For the other remaining amino carbamates, see below.

Monitorization of the enzymatic DKRs of alkyl (±)-*cis*-2-(aminocyclopentyl)carbamates (±)-**5b-d**:

Aliquots of 50 µL were taken periodically and treated with di-*tert*-butyl dicarbonate (Boc₂O). After 5 min, organic solvents were eliminated obtaining a crude, which consisted of a mixture of four compounds: 1-phenylethyl acetate, 1-phenylethanol, biscarbamate (alkyl *tert*-butyl cyclopentane-1,2-diylbiscarbamate proceeding from the reaction of the remaining amino carbamate with Boc₂O) and acetamide (see below).



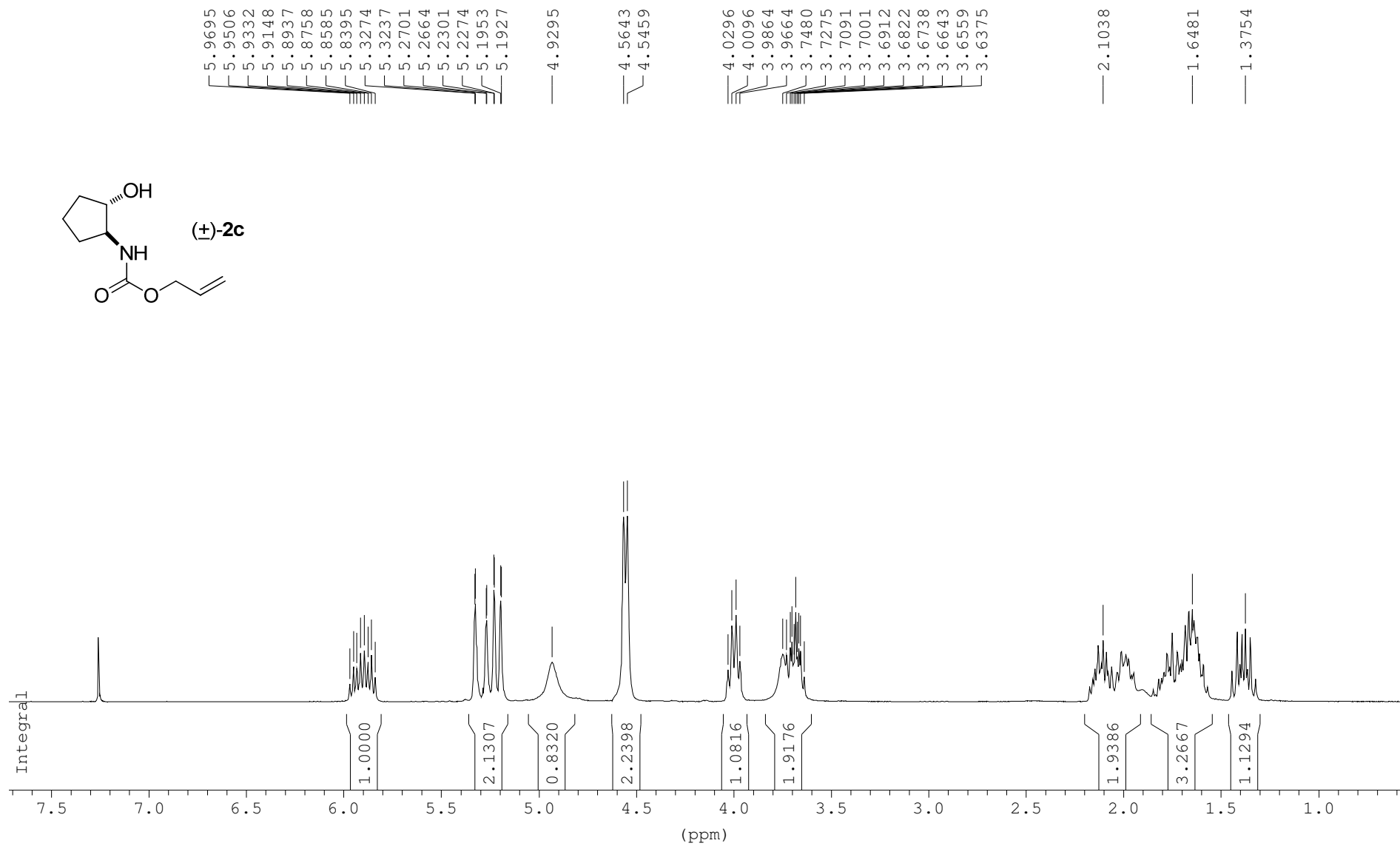
These crudes were analyzed by chiral HPLC and conditions and reaction times were as follows:

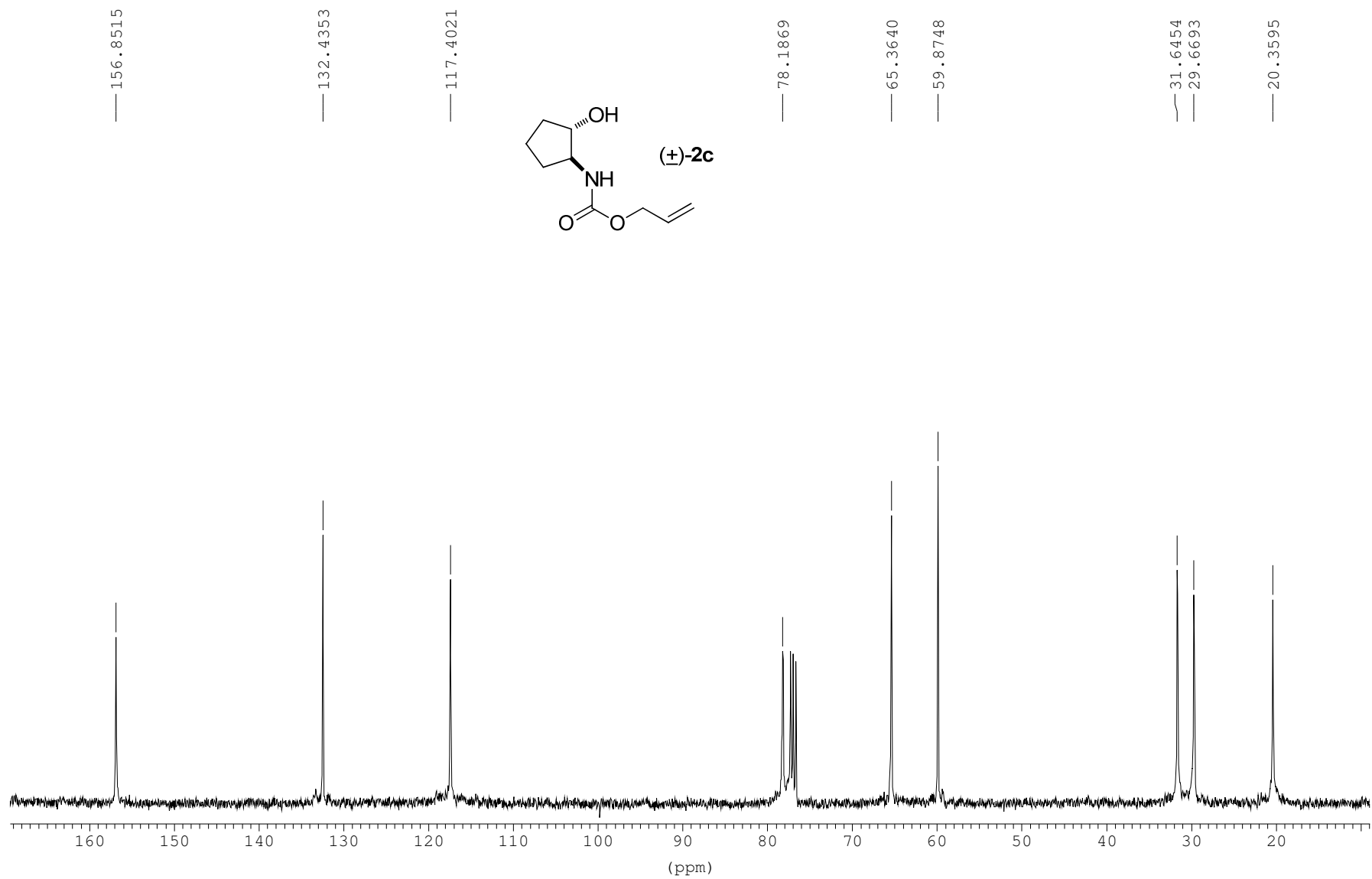
Reactions using (±)-5b** as substrate:** Chiralpak IA, hexane/ethanol 96:4 (7 min) and then, 85:15, 30 °C, 0.8 mL/min. (±)-1-Phenylethyl acetate: t_R = 4.9 (*R*) and 5.2 (*S*) min, R_S = 1.4. (±)-1-Phenylethanol: t_R =

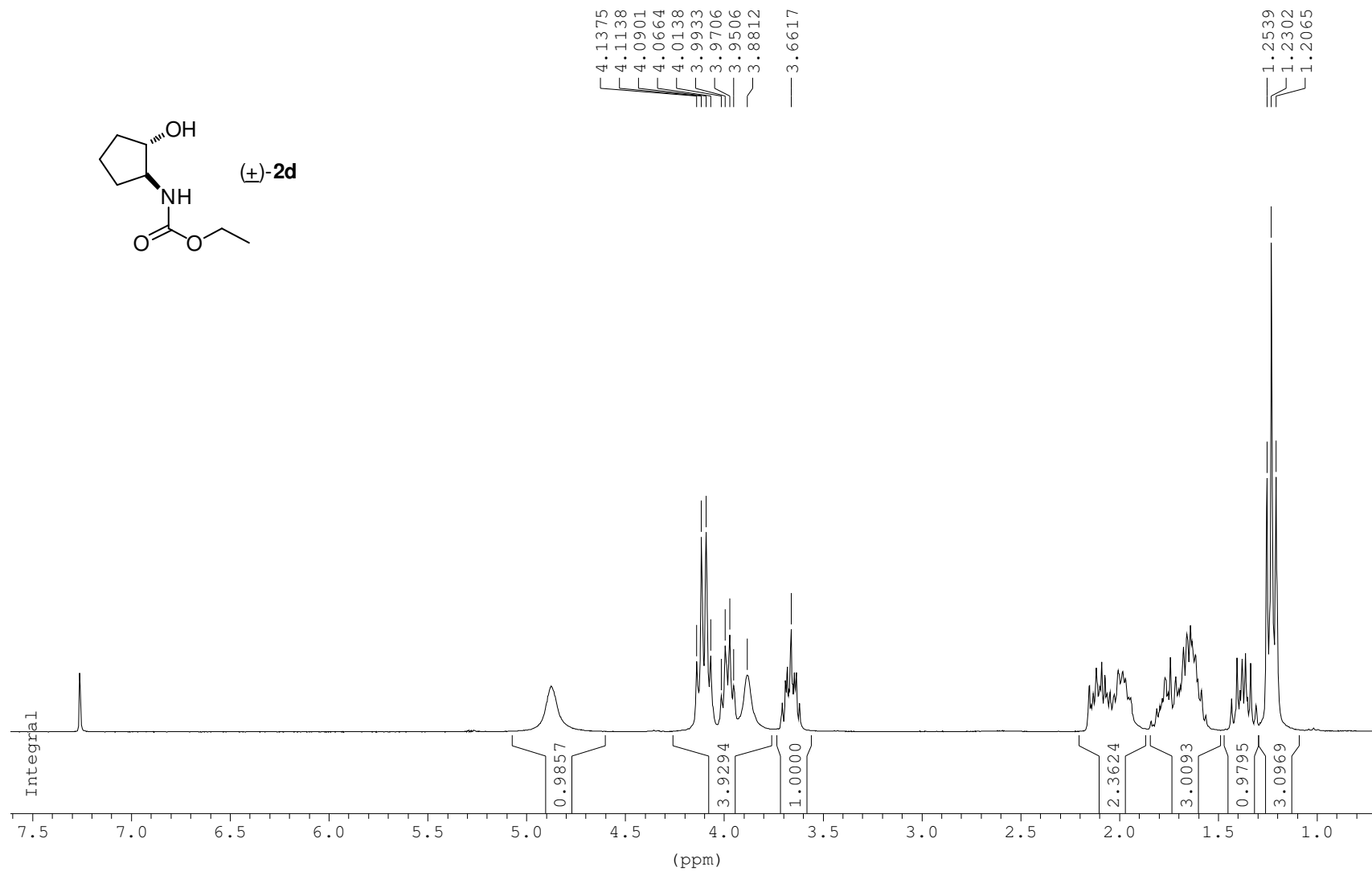
9.2 (*R*) and 9.7 (*S*) min; $R_S = 1.5$. (±)-Biscarmate: $t_R = 13.6$ (1*S*,2*R*) and 18.8 (1*R*,2*S*) min; $R_S = 11.3$. (±)-Acetamide: $t_R = 16.4$ (1*S*,2*R*) and 17.0 (1*R*,2*S*) min; $R_S = 1.5$.

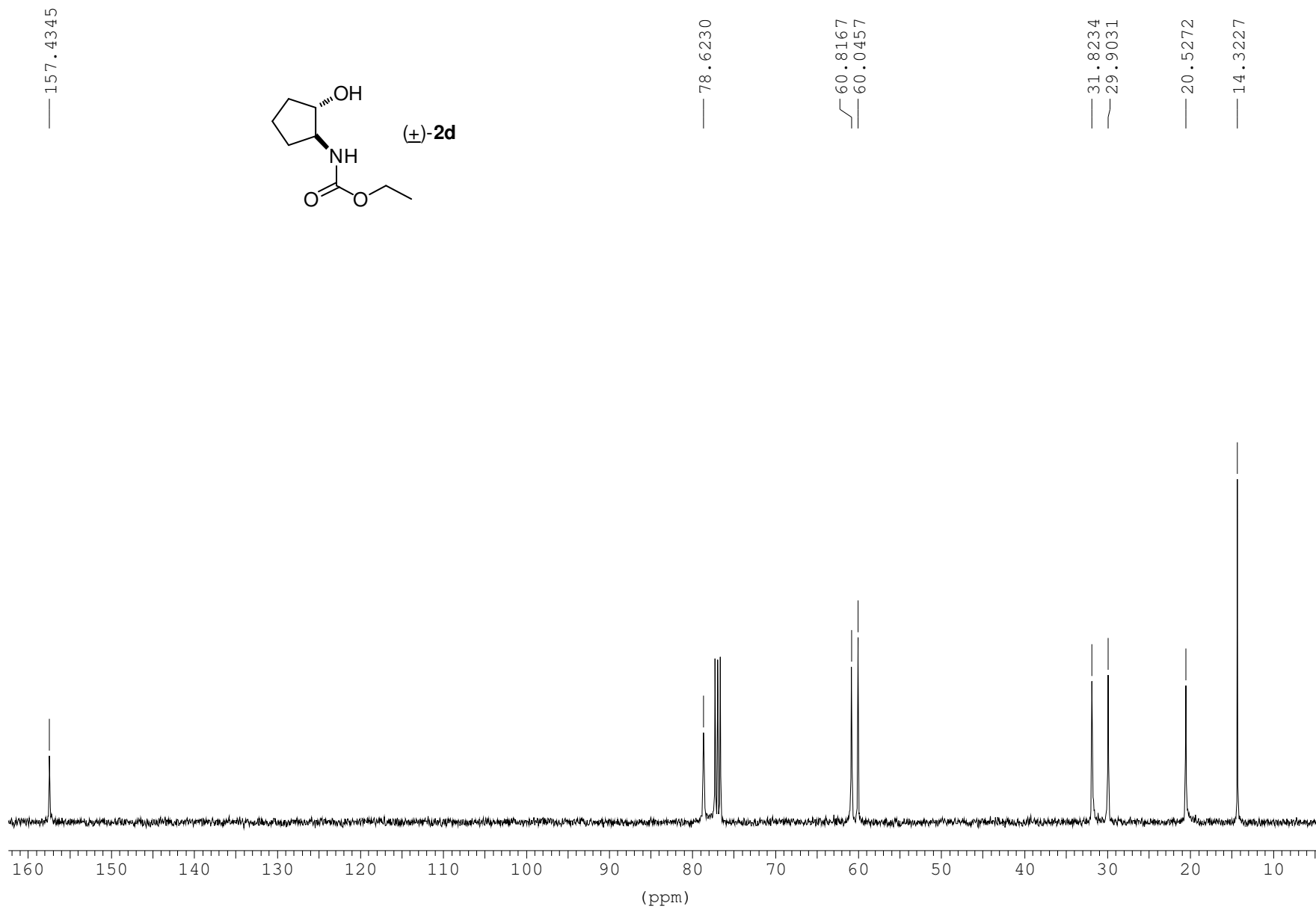
Reaction using (±)-5c as substrate: Chiralpak IA, hexane/ethanol 94:6, 20 °C, 0.8 mL/min. (±)-1-Phenylethyl acetate: $t_R = 5.0$ (*R*) and 5.3 (*S*) min. (±)-1-Phenylethanol: $t_R = 8.0$ (*R*) and 8.4 (*S*) min. (±)-Biscarmate: $t_R = 11.2$ (1*S*,2*R*) and 14.1 (1*R*,2*S*) min; $R_S = 4.9$. (±)-Acetamide: $t_R = 20.0$ (1*S*,2*R*) and 22.6 (1*R*,2*S*) min; $R_S = 1.8$.

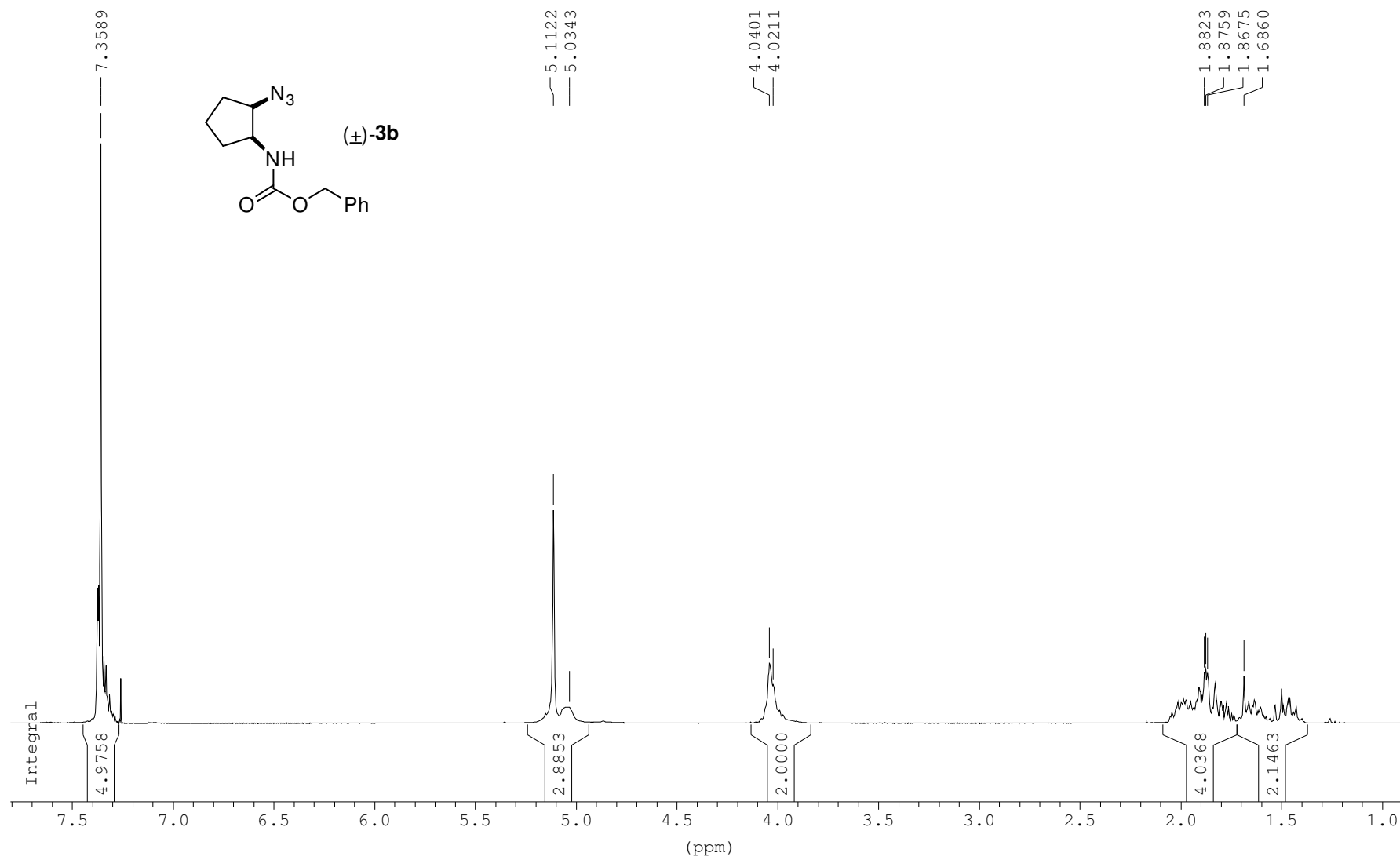
Reaction using (±)-5d as substrate: Chiralpak IA, hexane/ethanol 96:4, 20 °C, 0.8 mL/min. (±)-1-Phenylethyl acetate: $t_R = 5.2$ (*R*) and 5.5 (*S*) min. (±)-1-Phenylethanol: $t_R = 9.7$ (*R*) and 10.3 (*S*) min. (±)-Biscarmate: $t_R = 14.3$ (1*S*,2*R*) and 16.5 (1*R*,2*S*) min; $R_S = 3.4$. (±)-Acetamide: $t_R = 32.3$ (1*S*,2*R*) and 36.3 (1*R*,2*S*) min; $R_S = 1.7$.

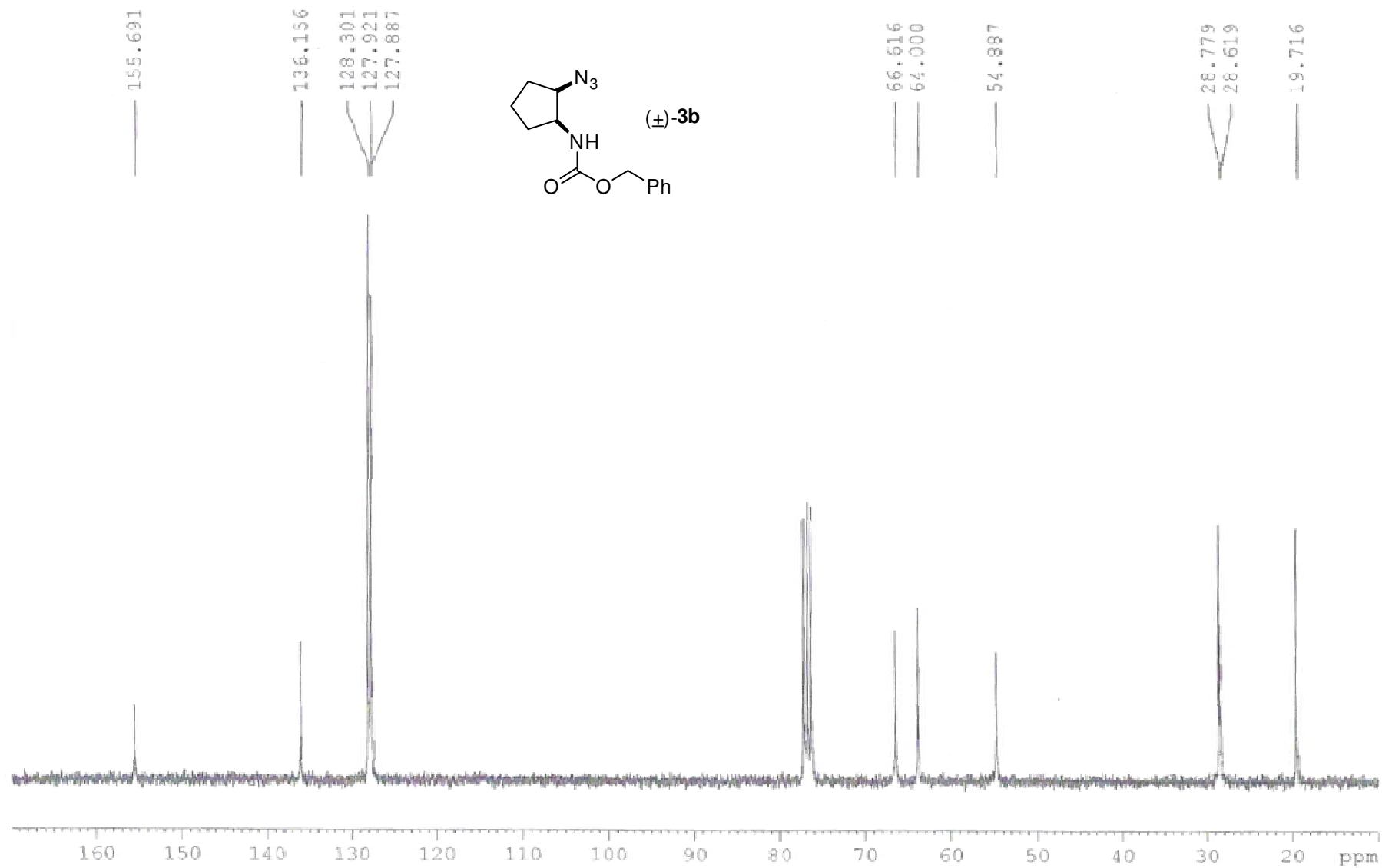


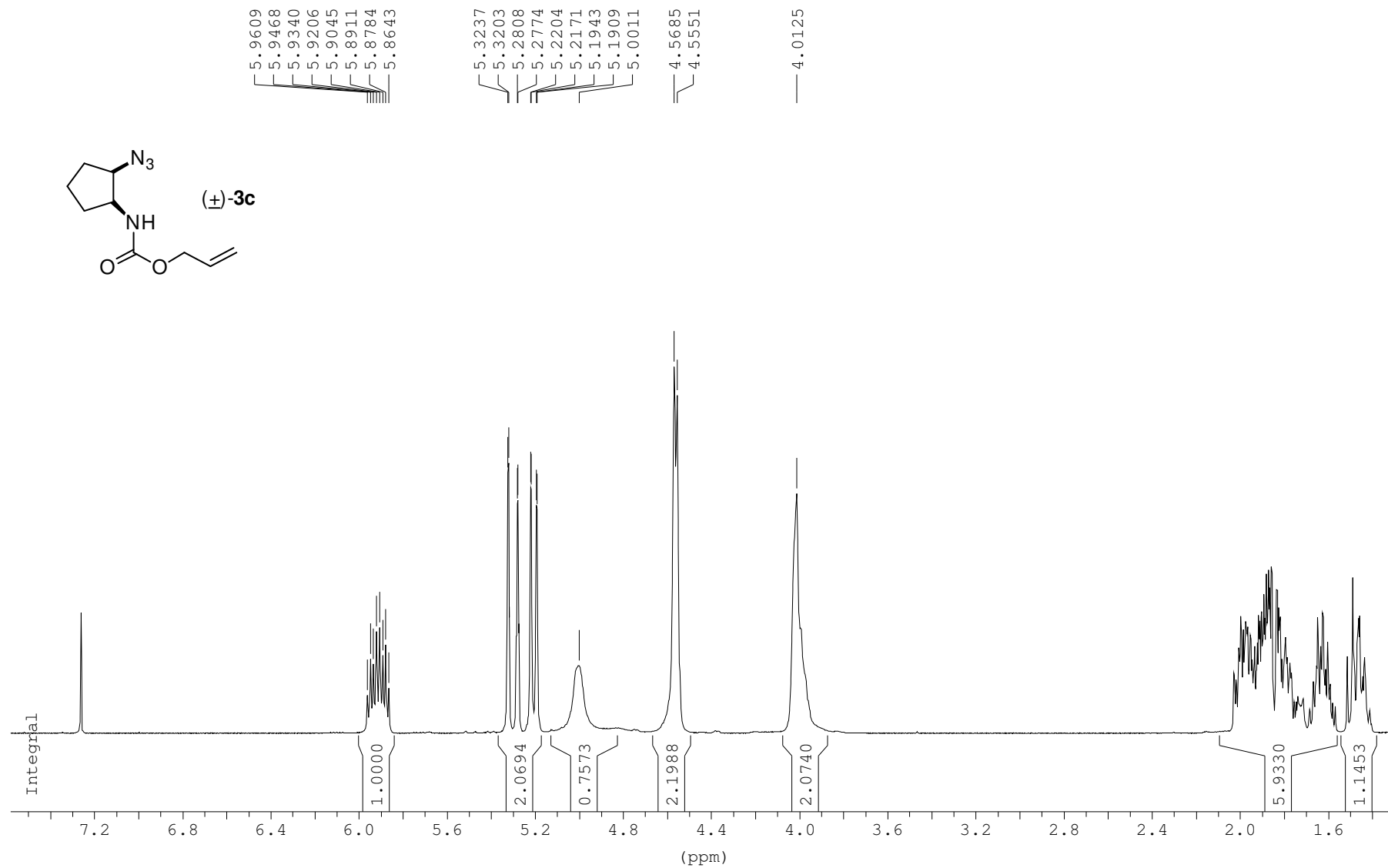


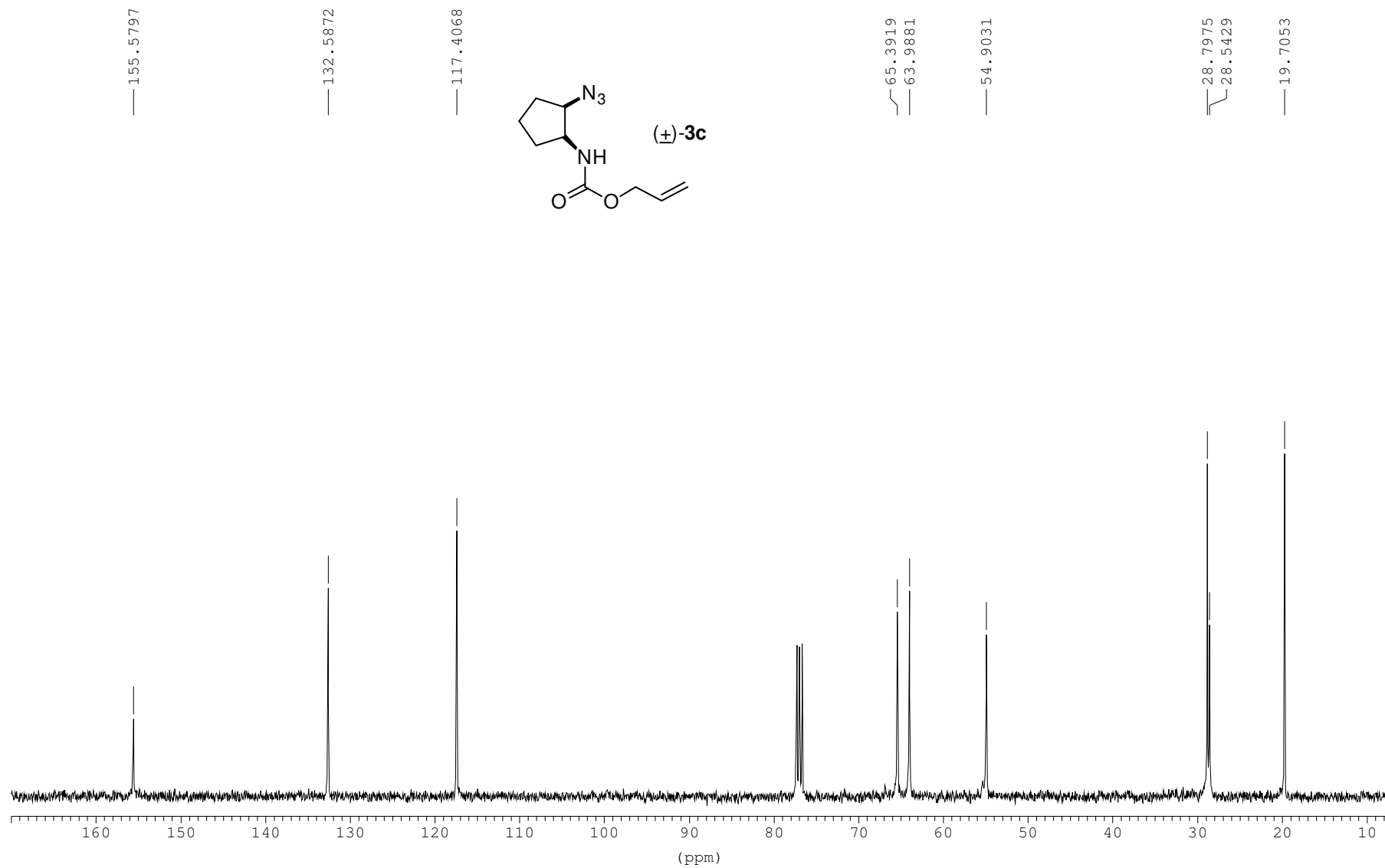


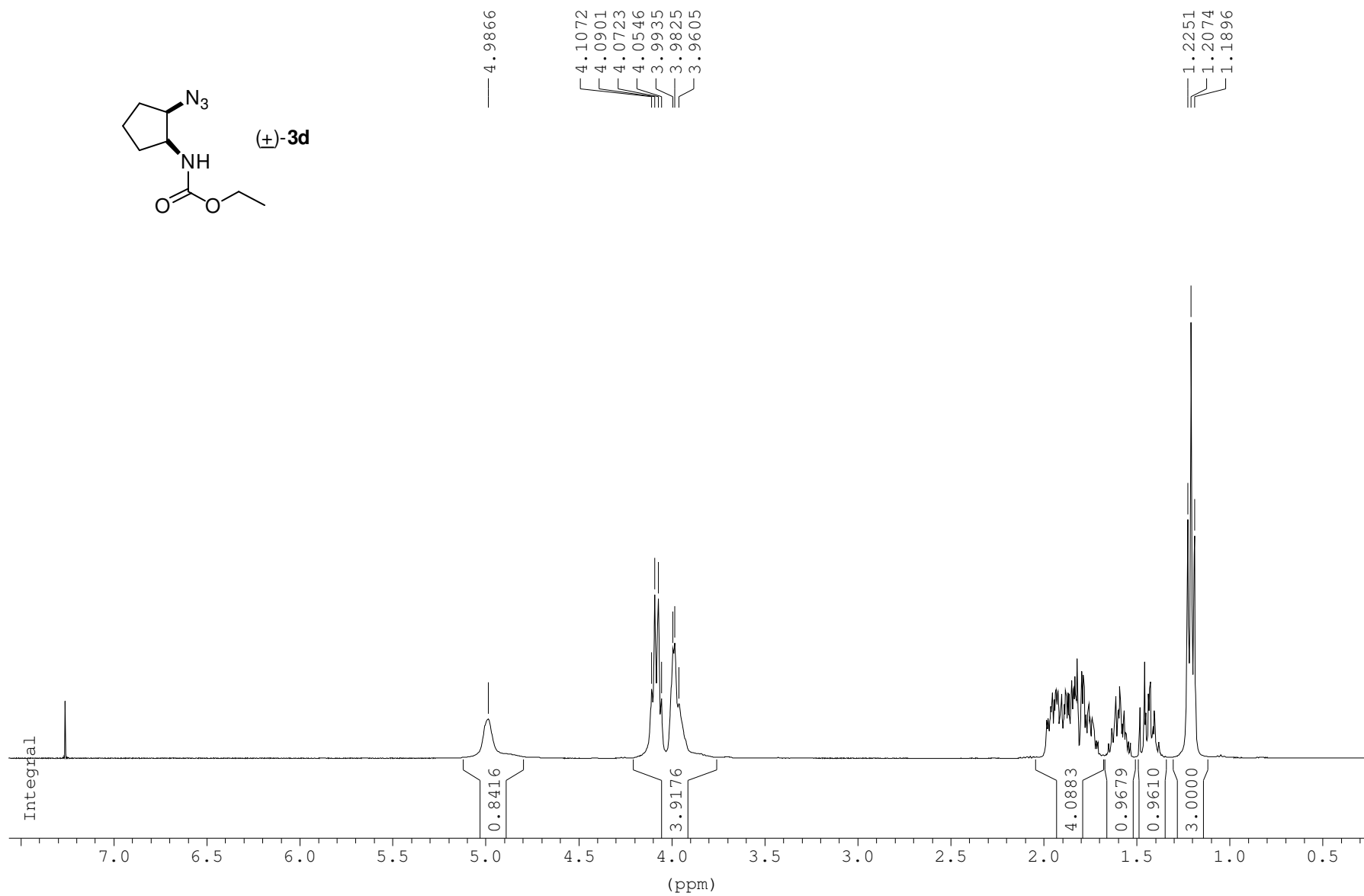
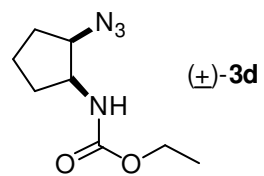


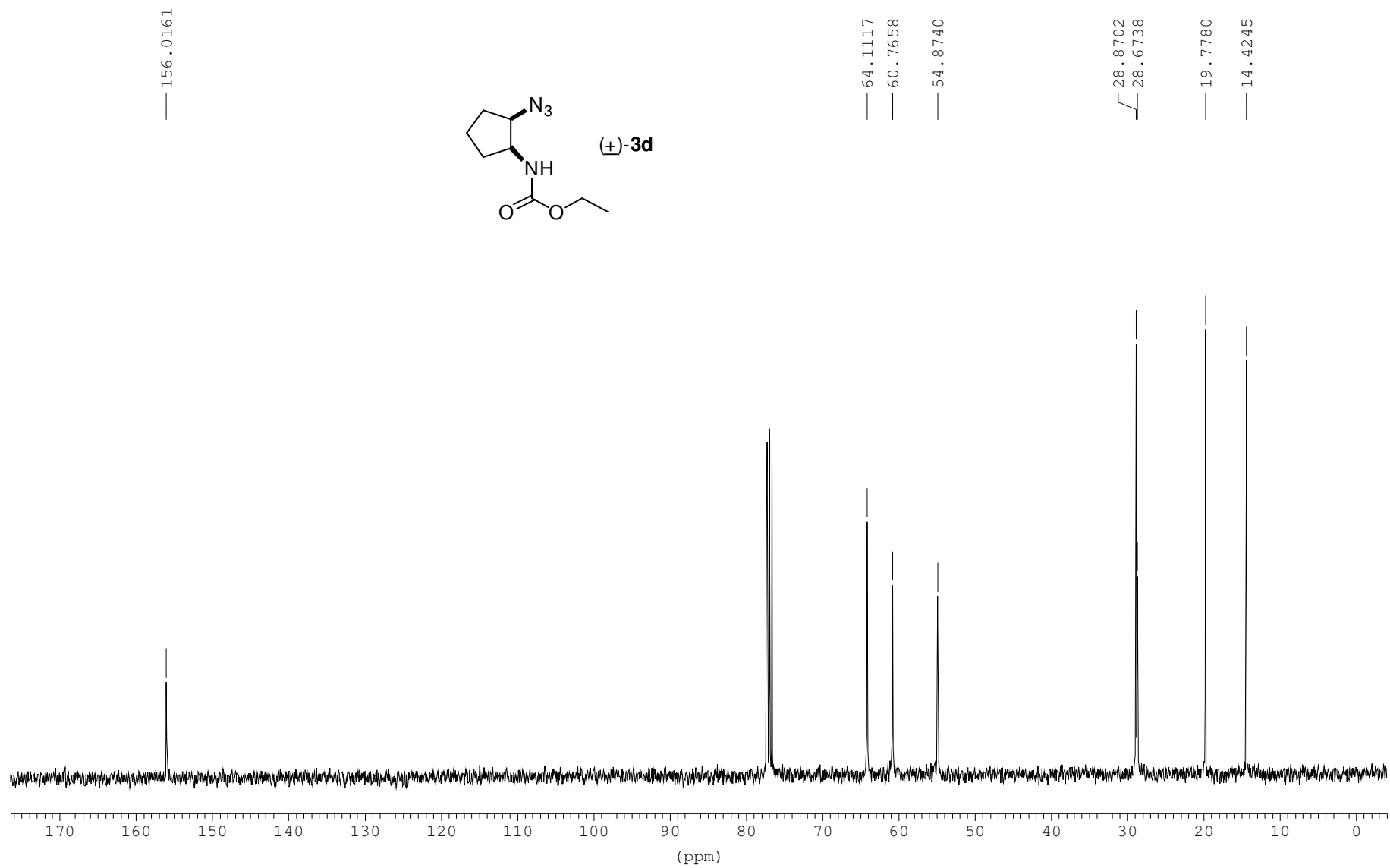


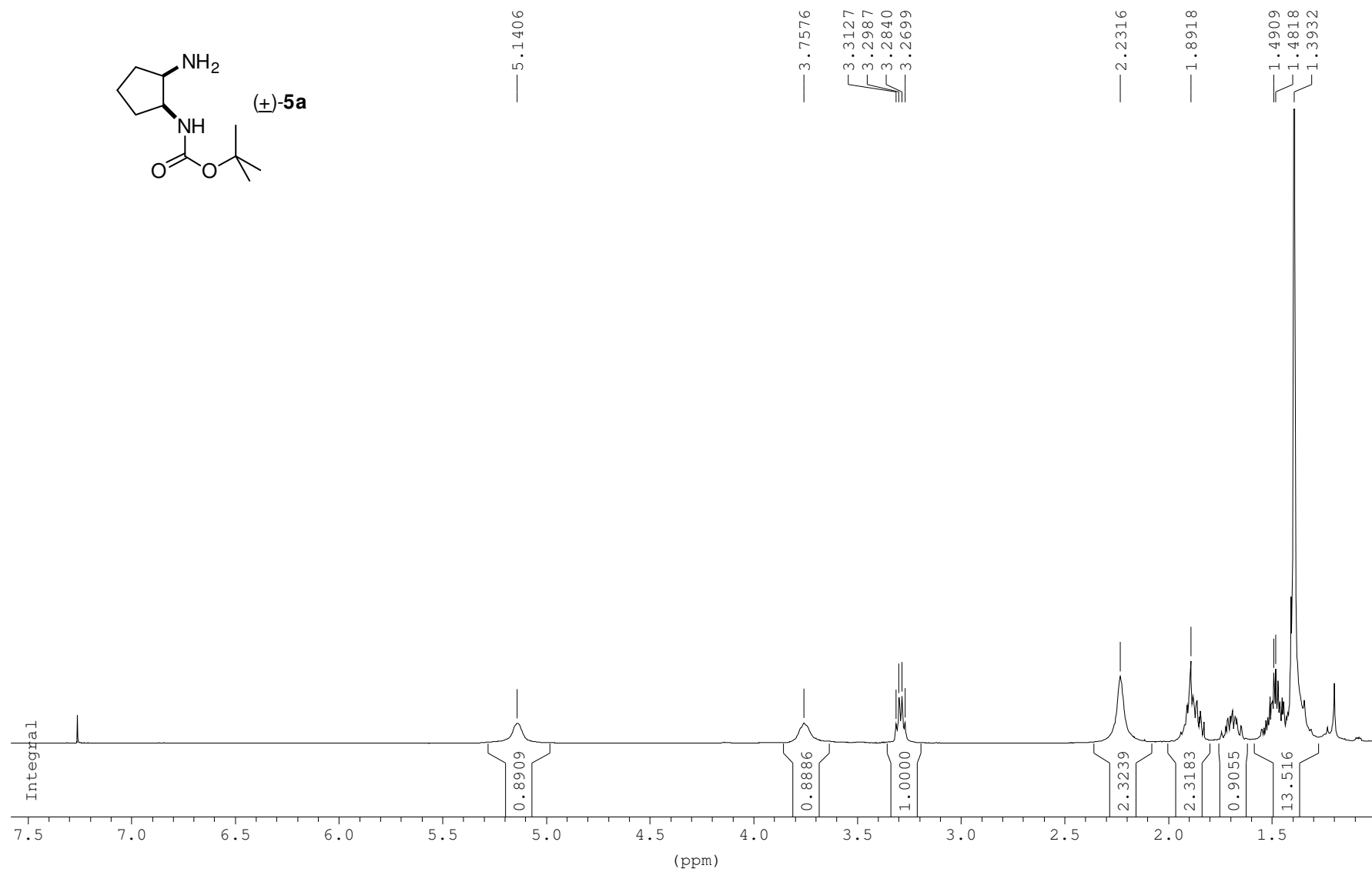


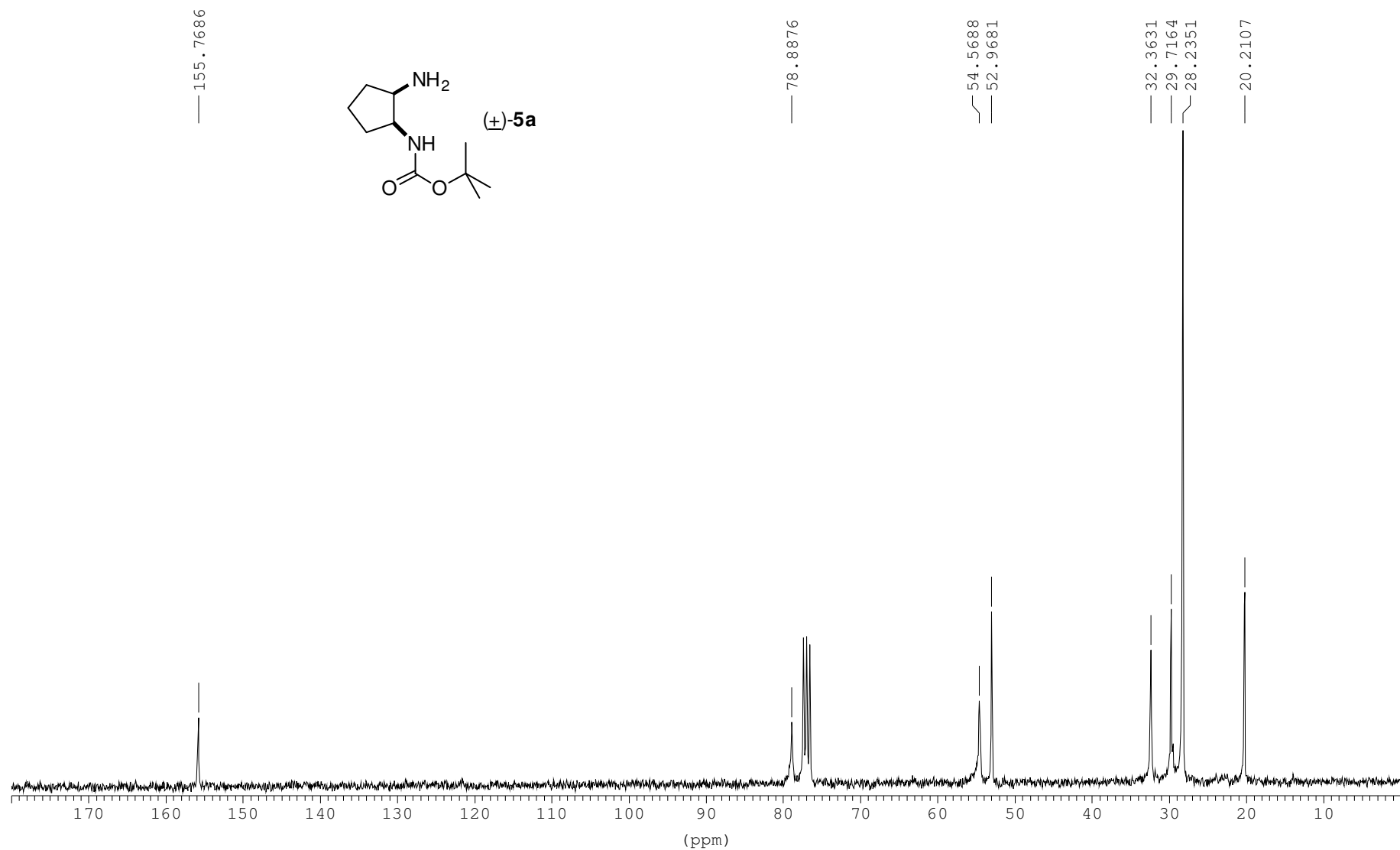


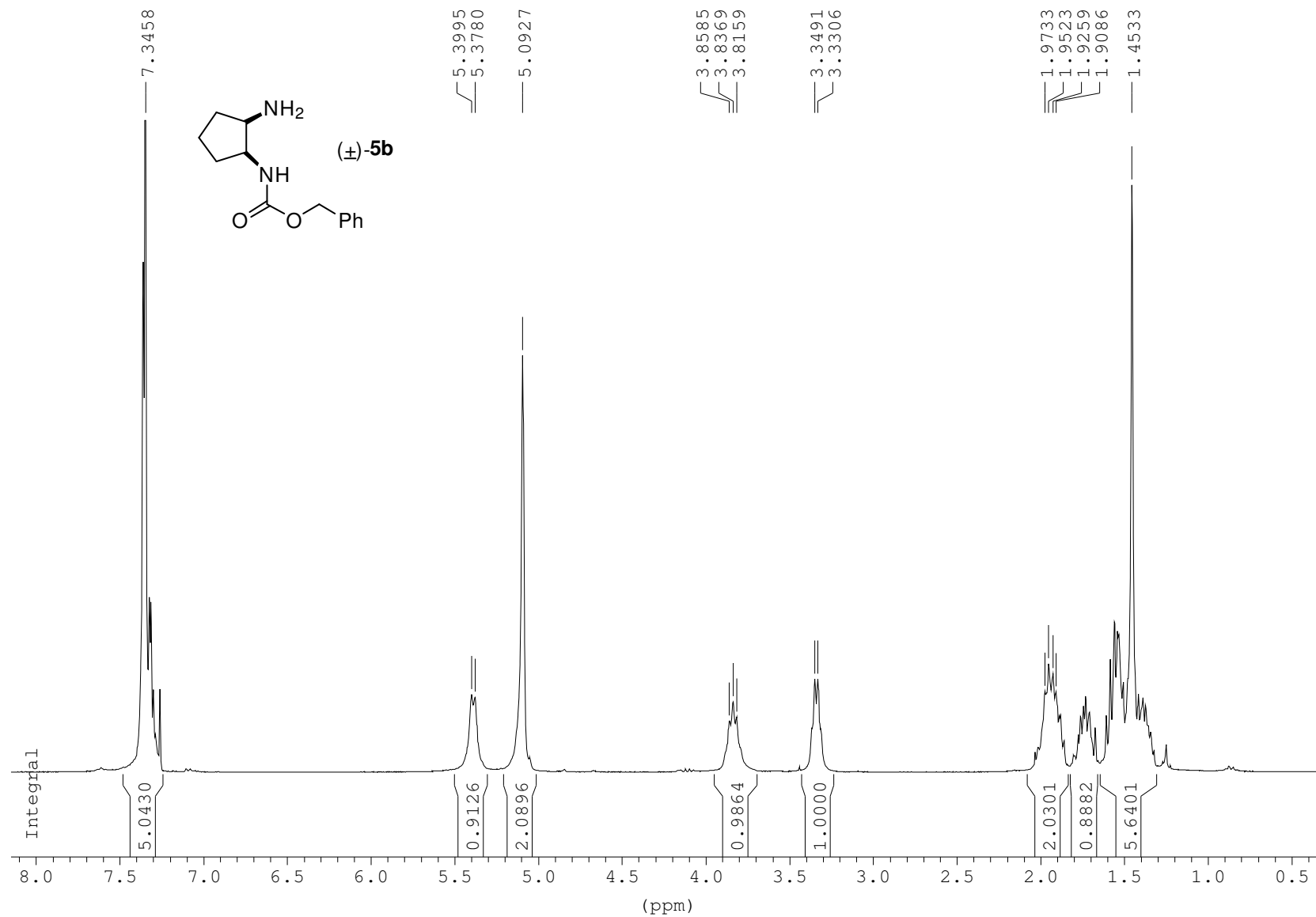


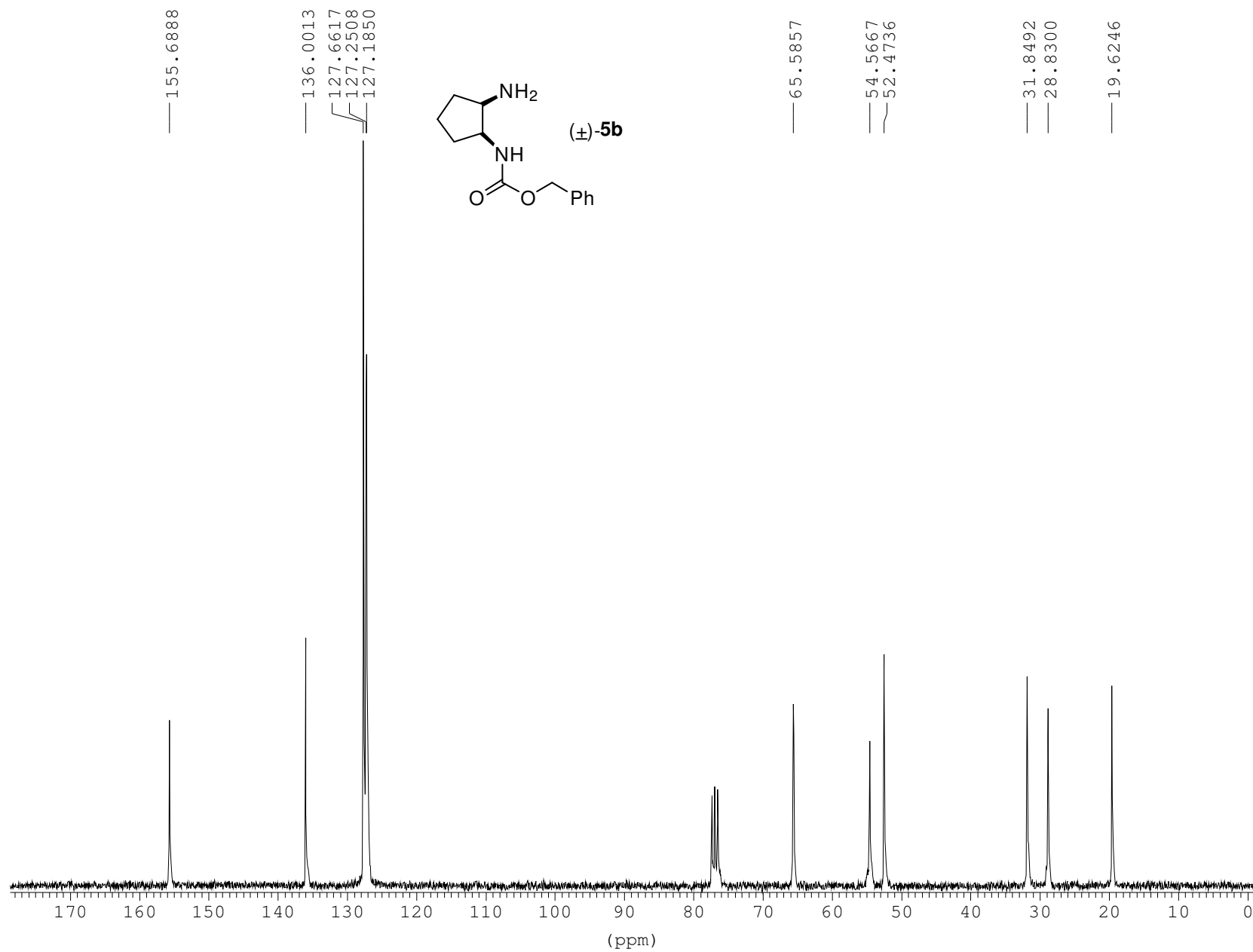


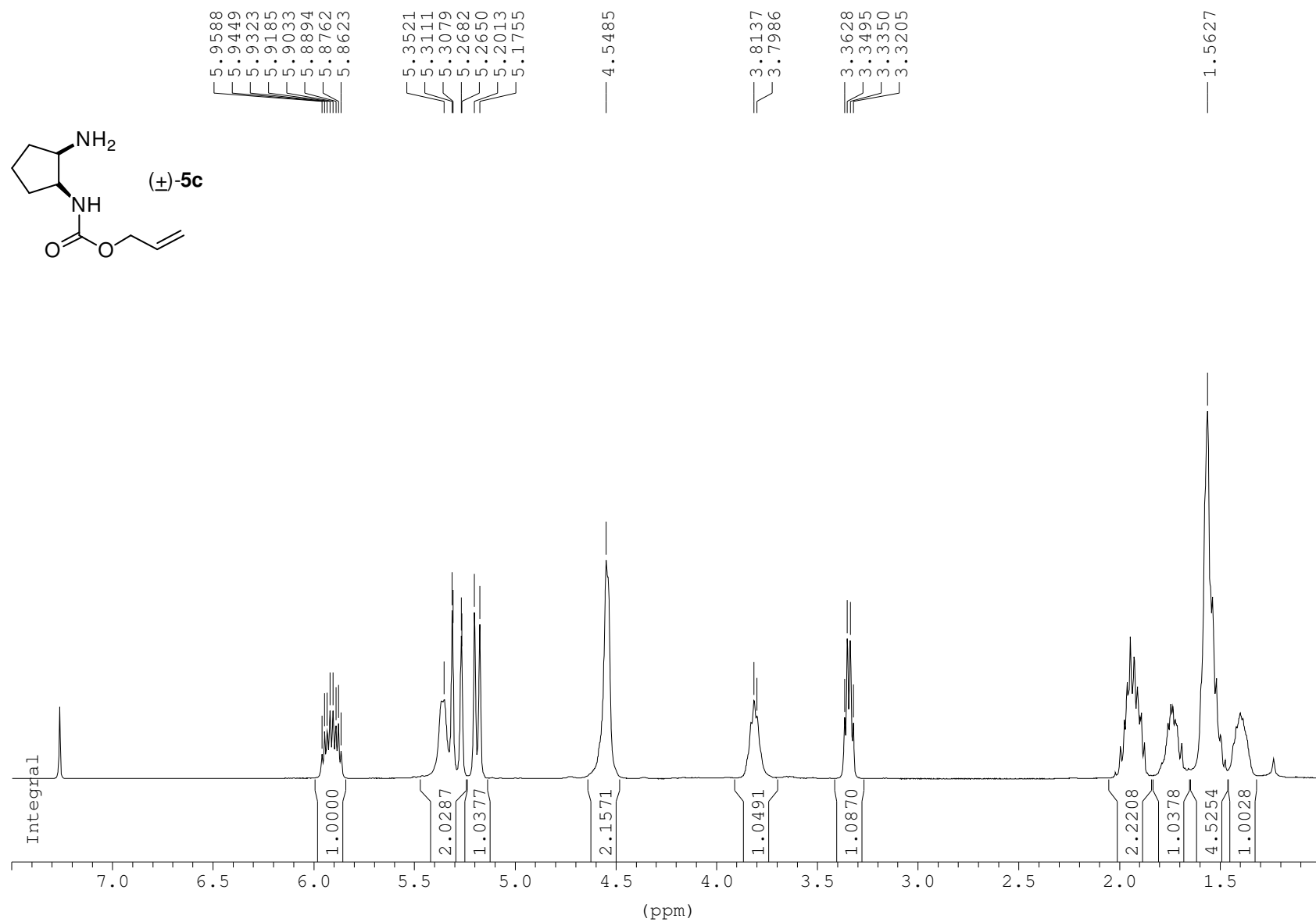


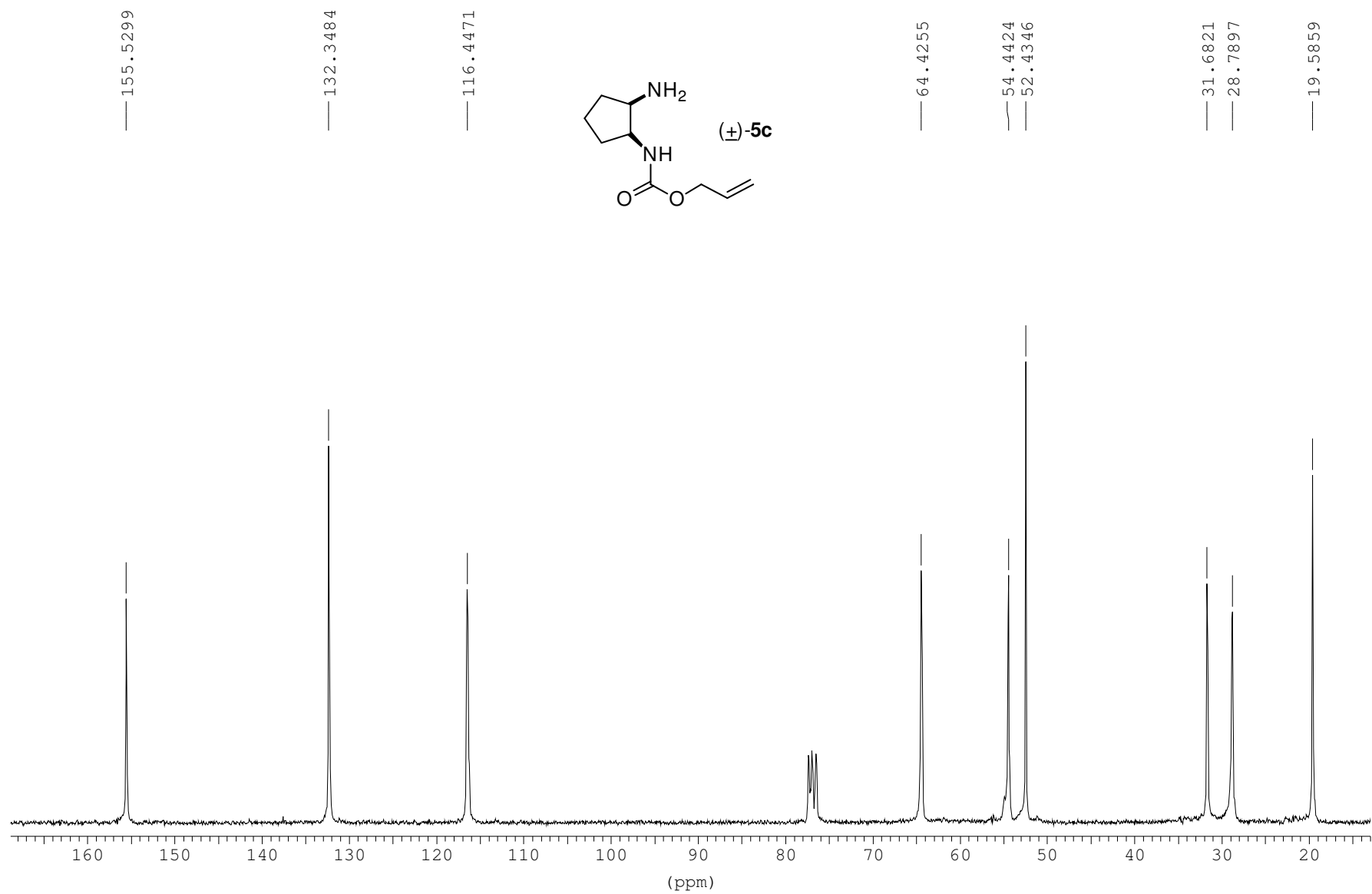


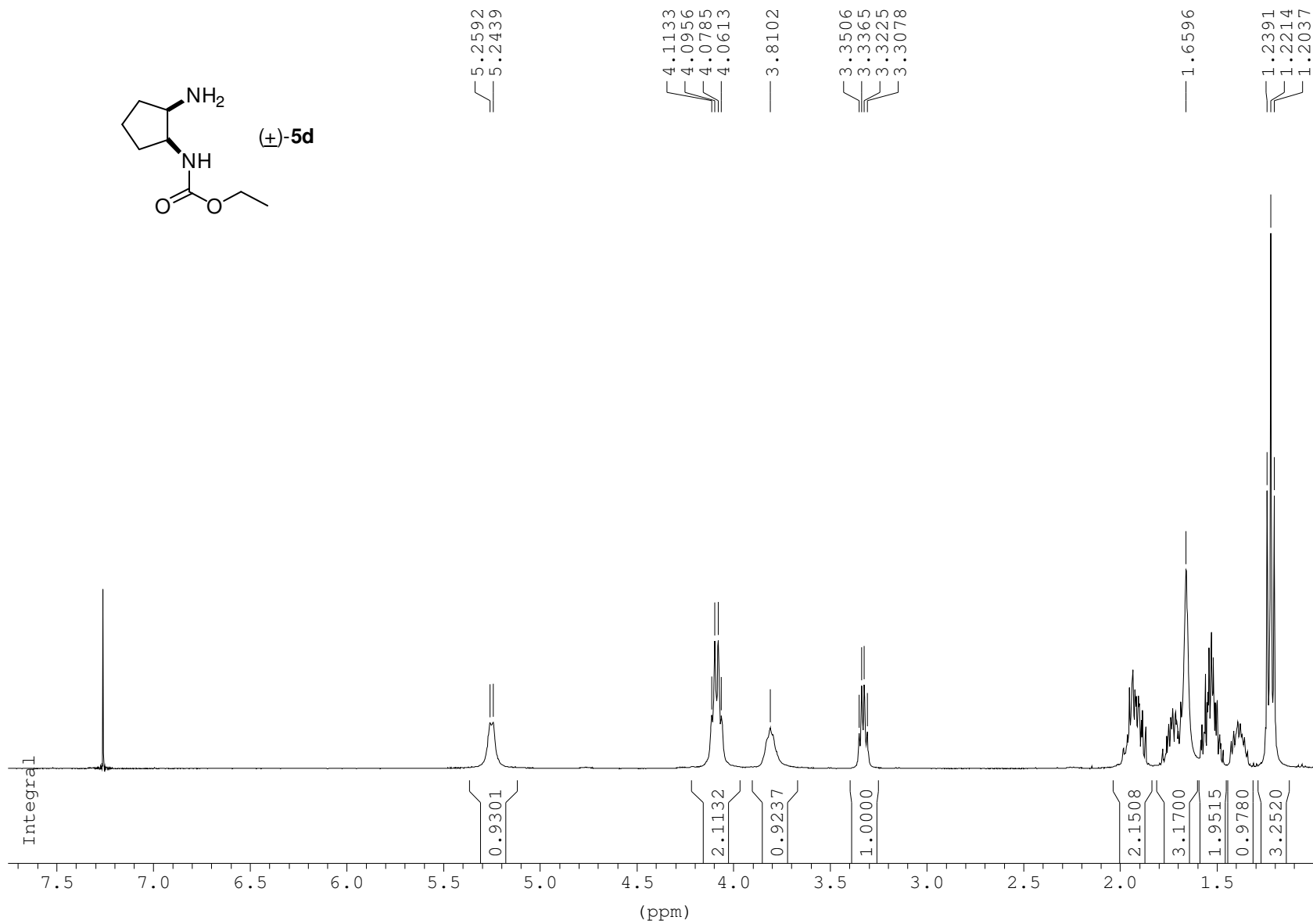


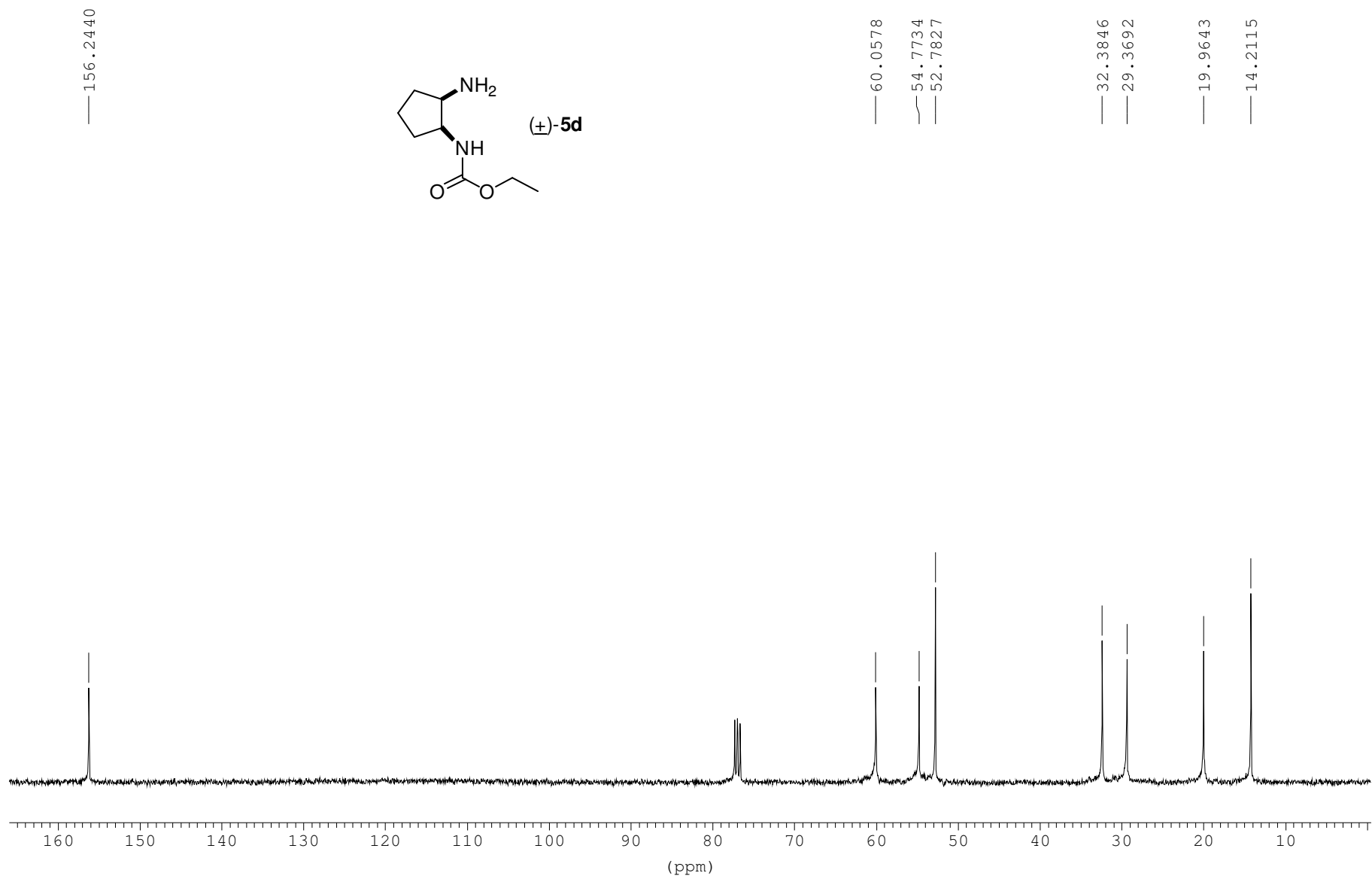


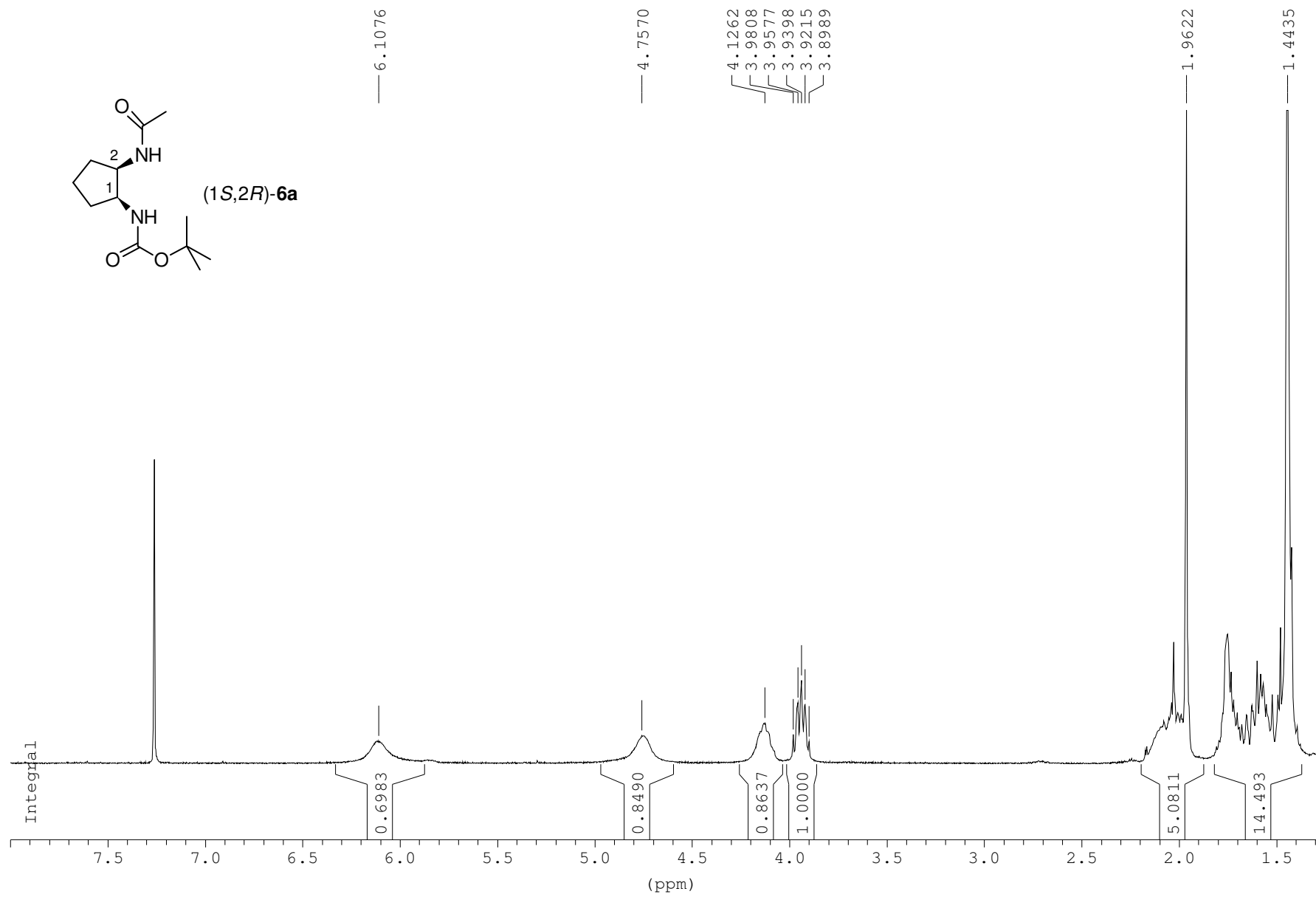


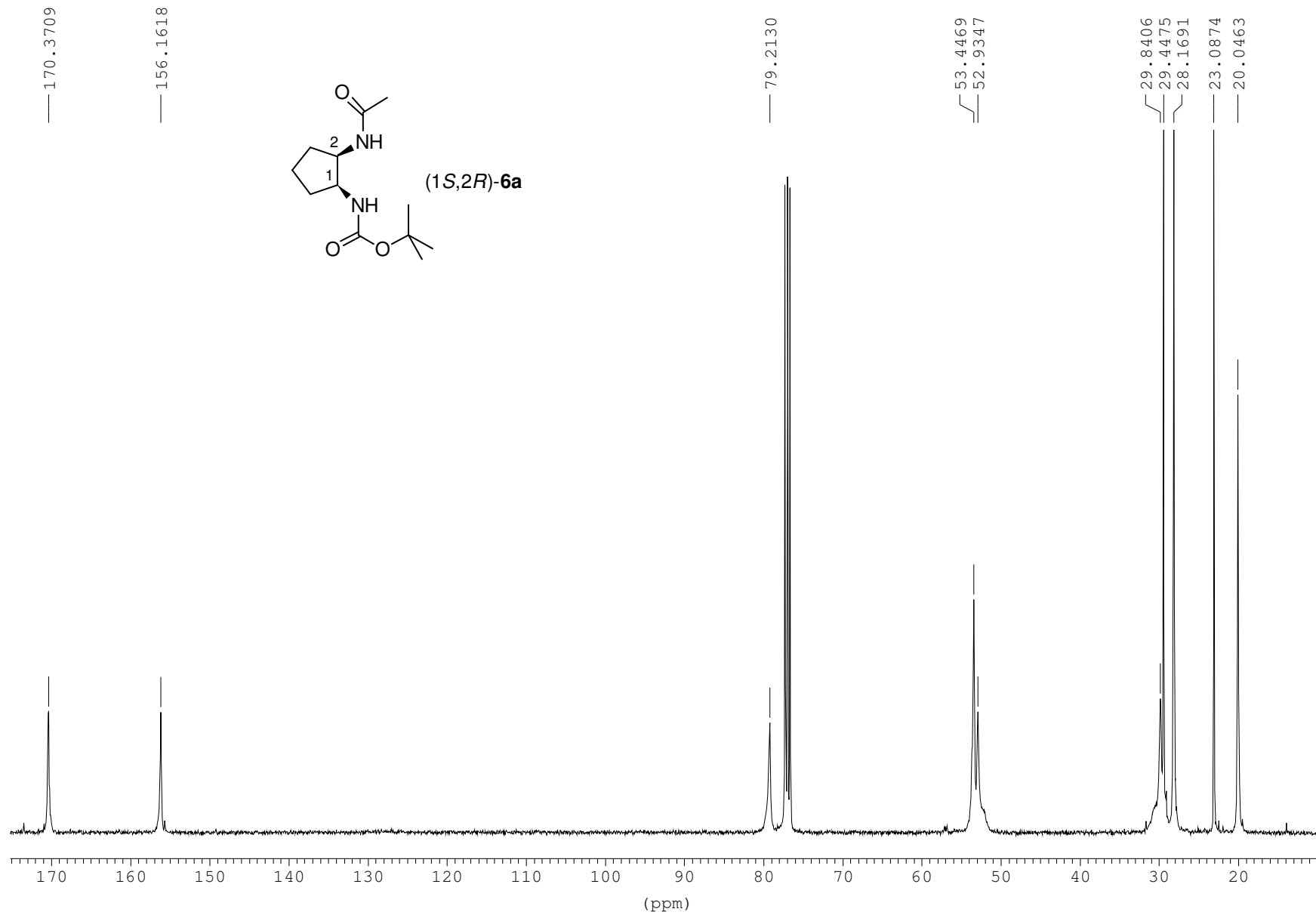


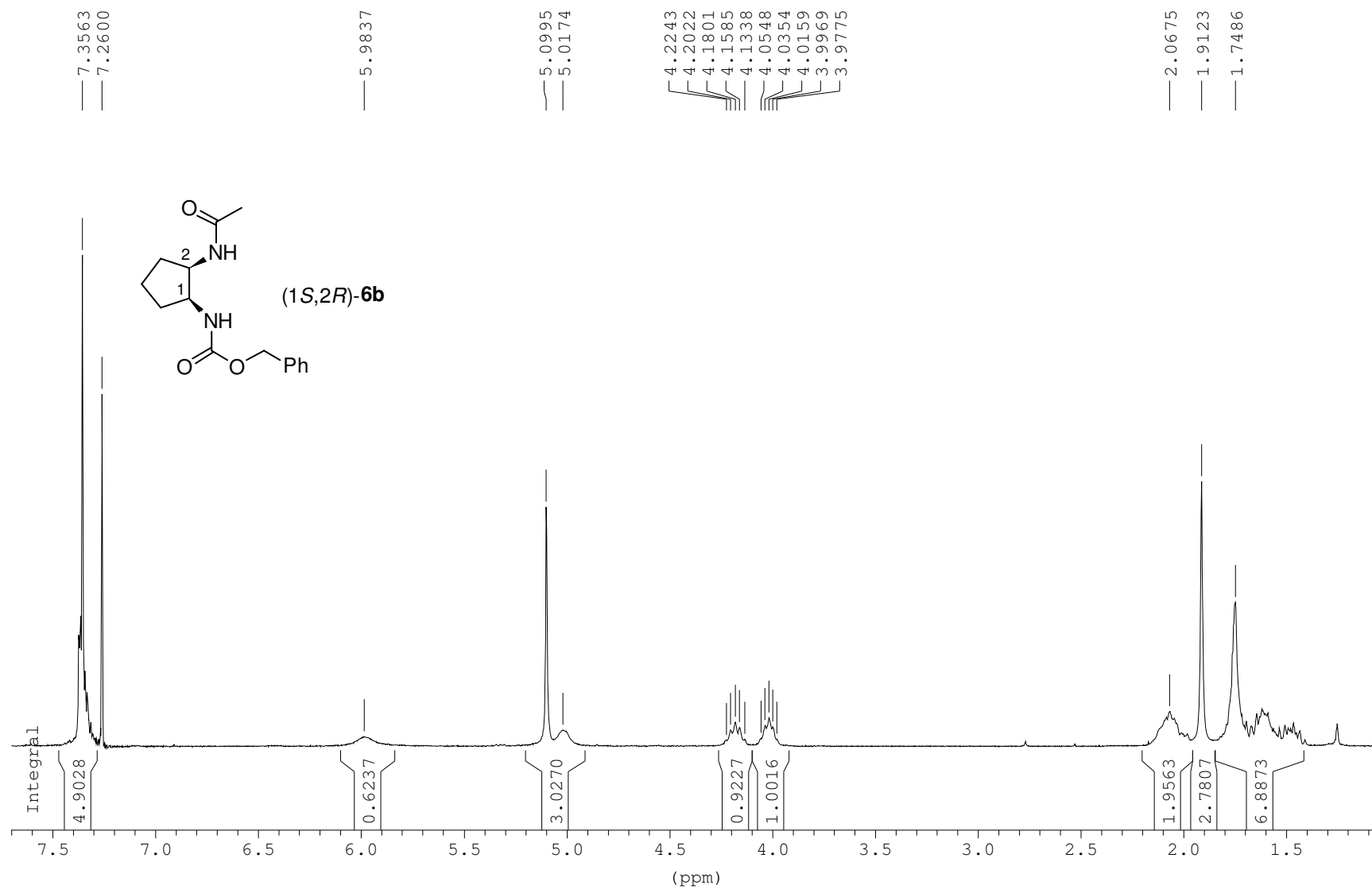


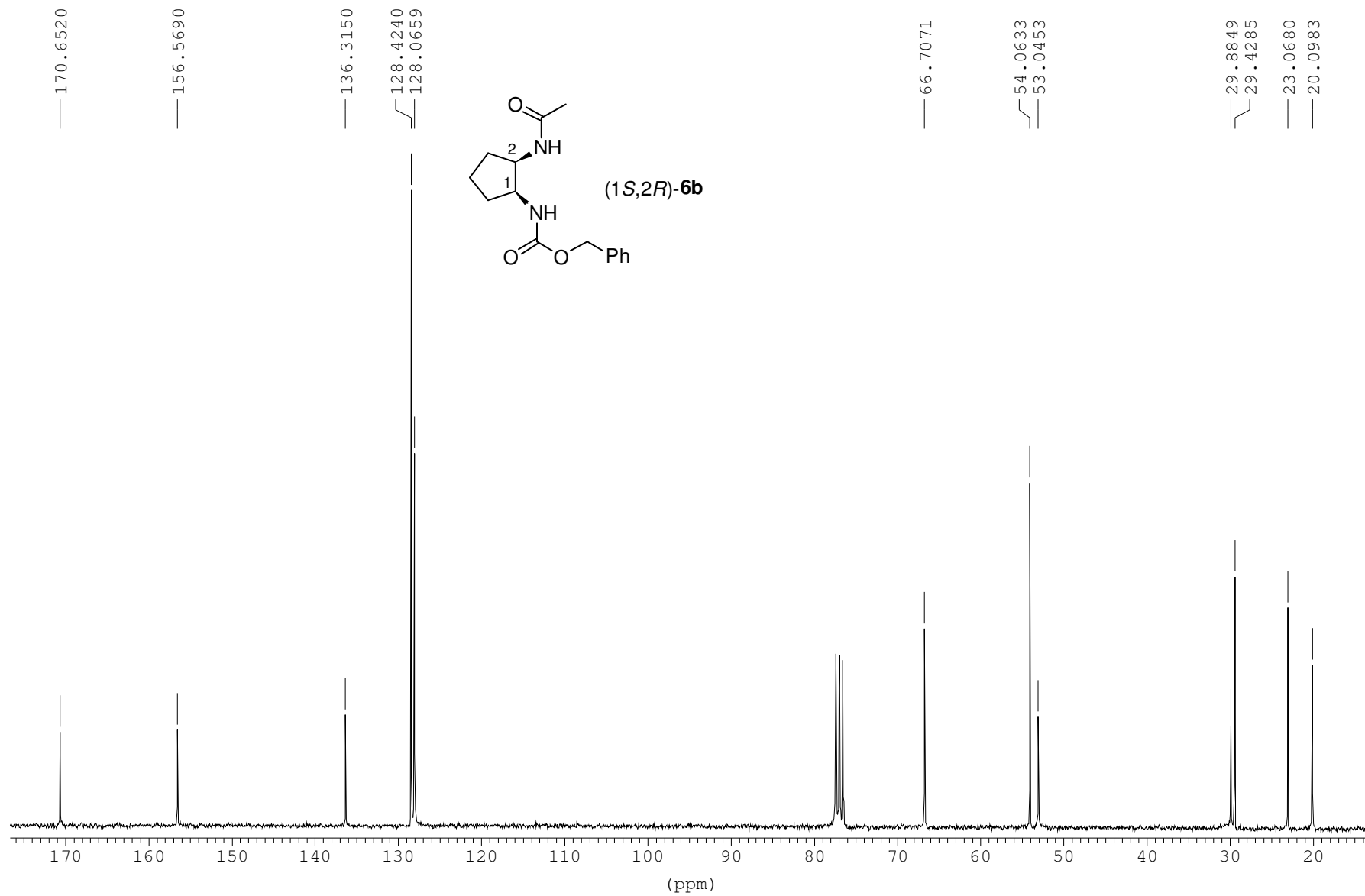


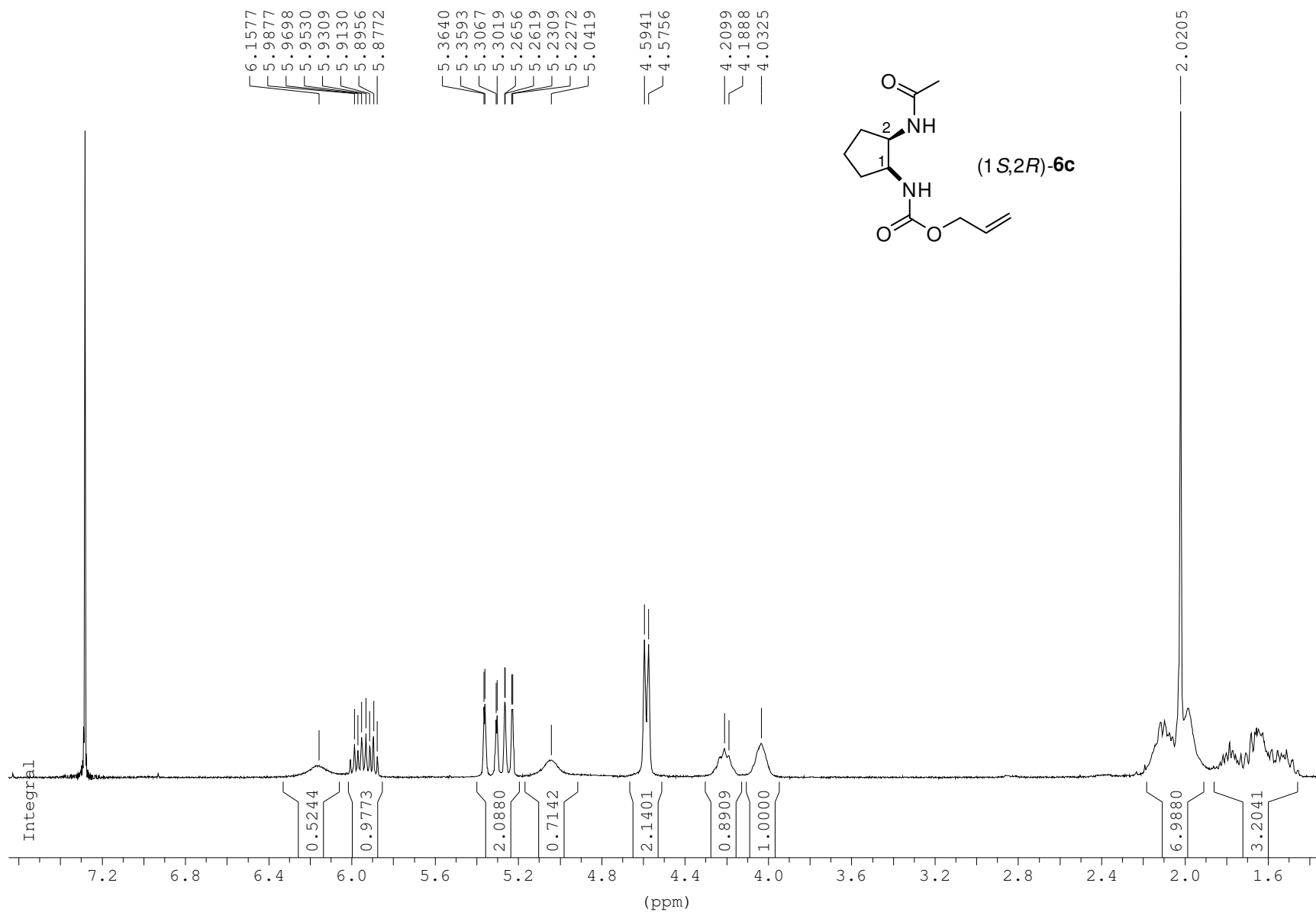


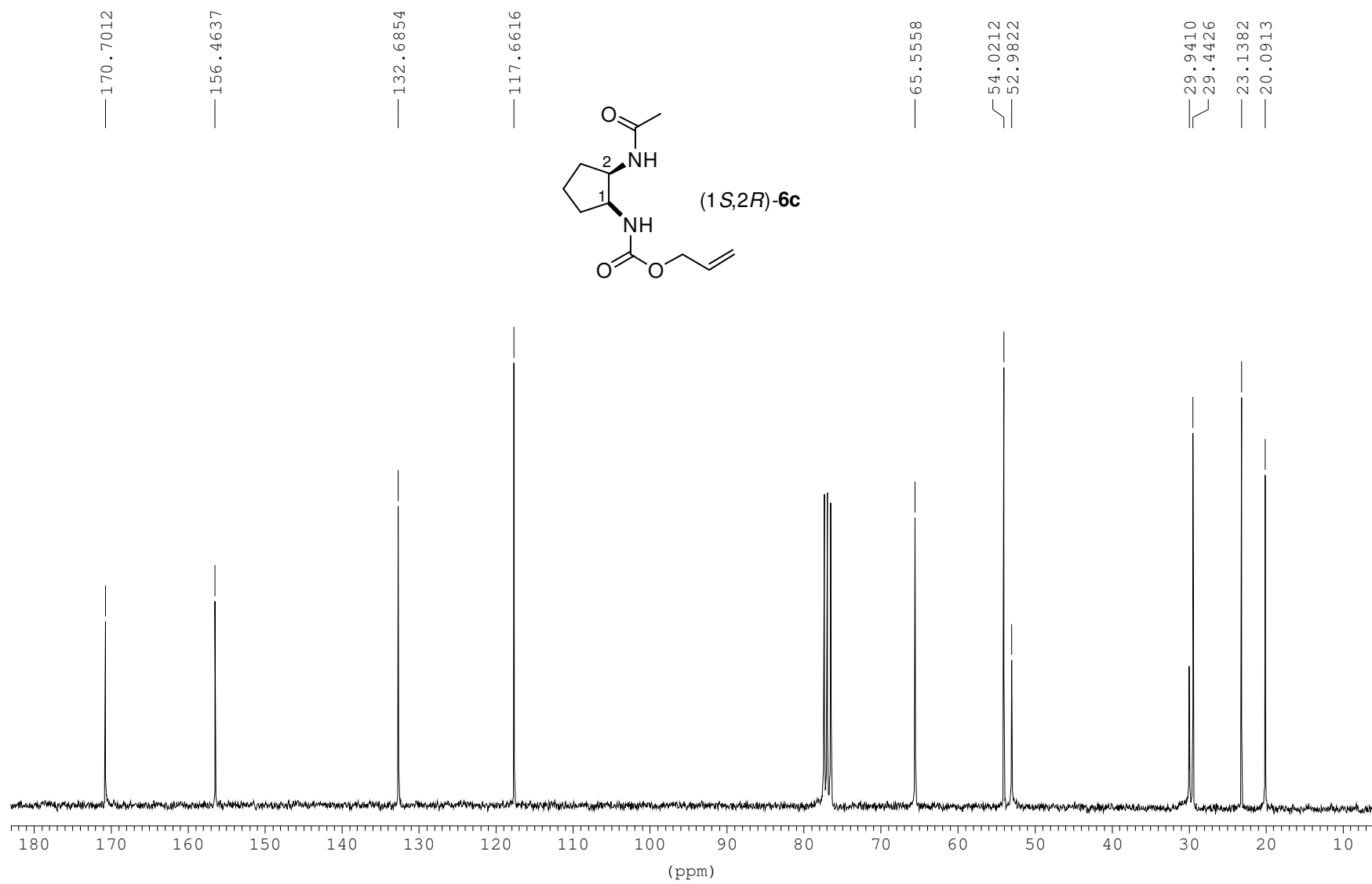


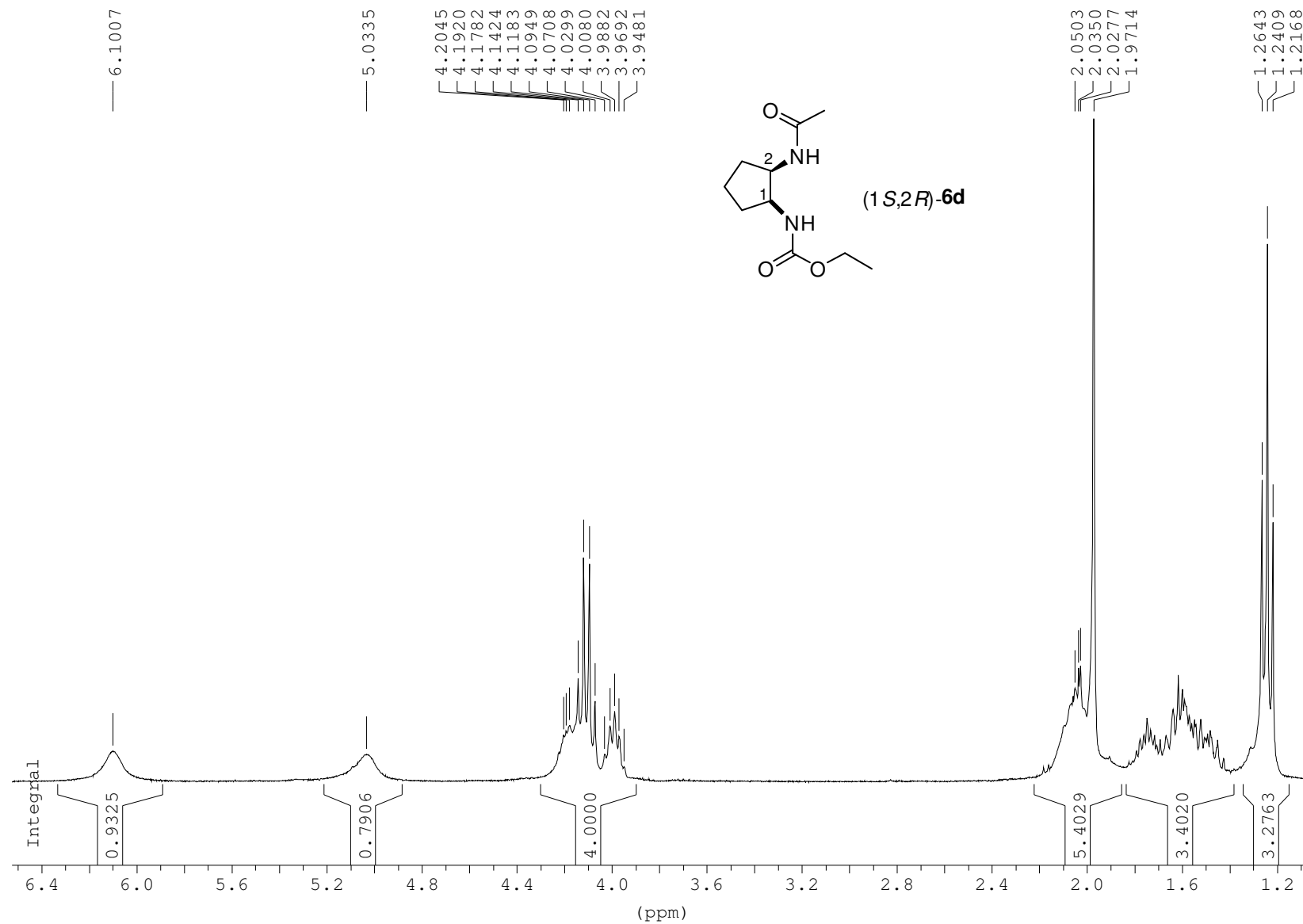


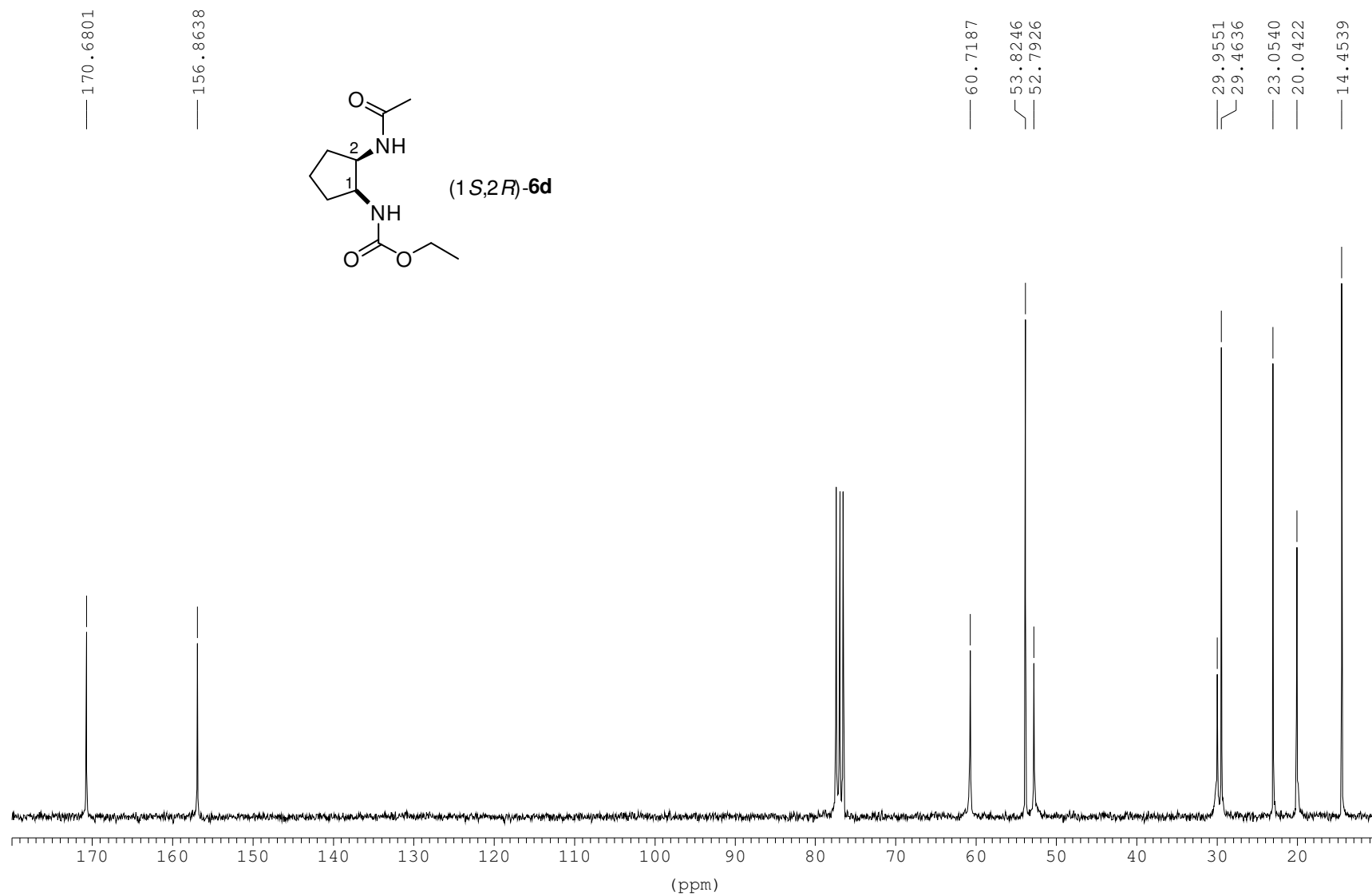




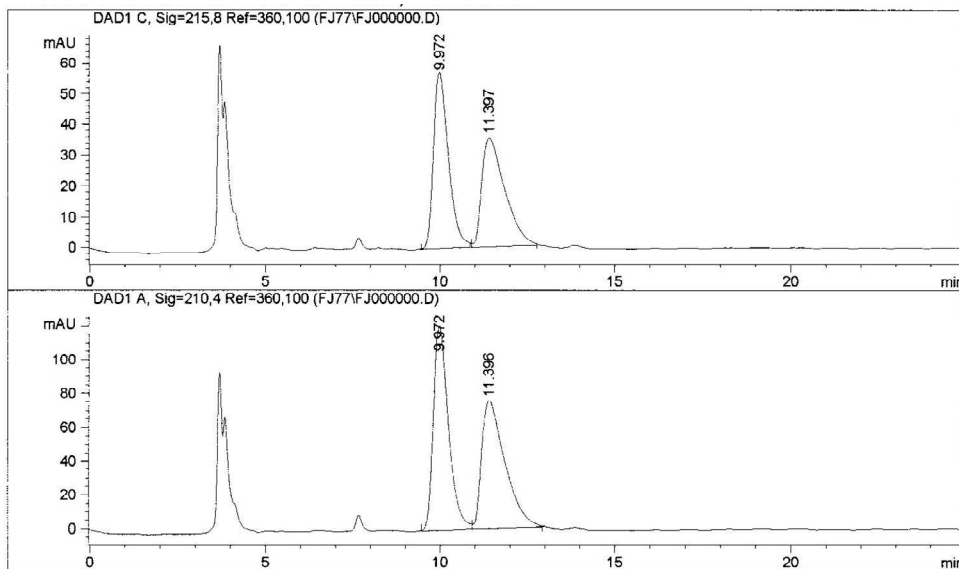
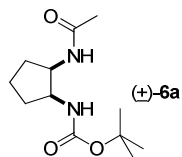




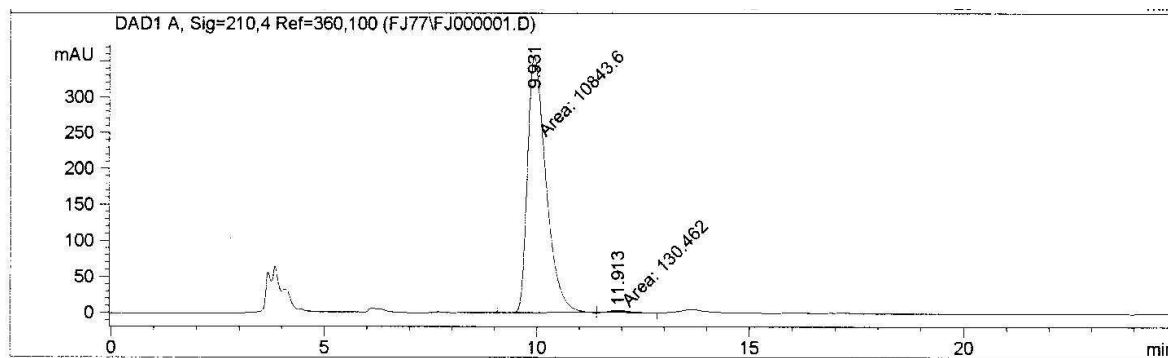




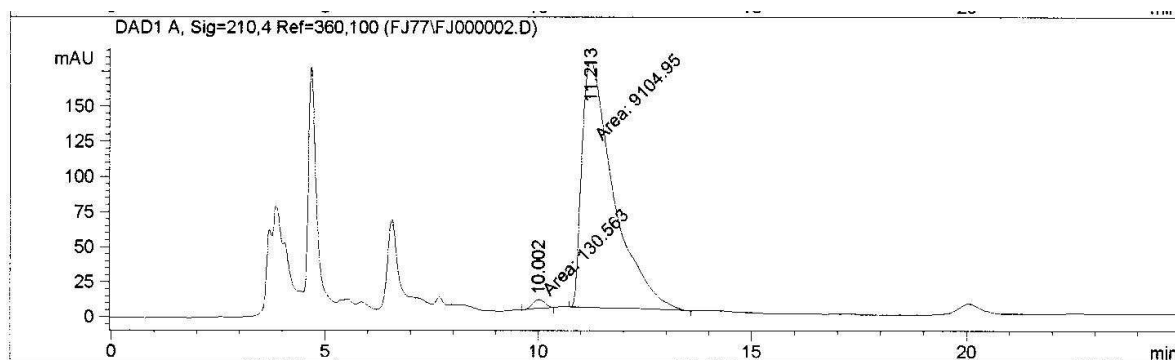
=====

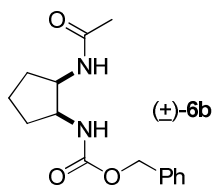


Product isolated in the KR of (±)-5a: (1*S*,2*R*)-6a (ee = 98%)

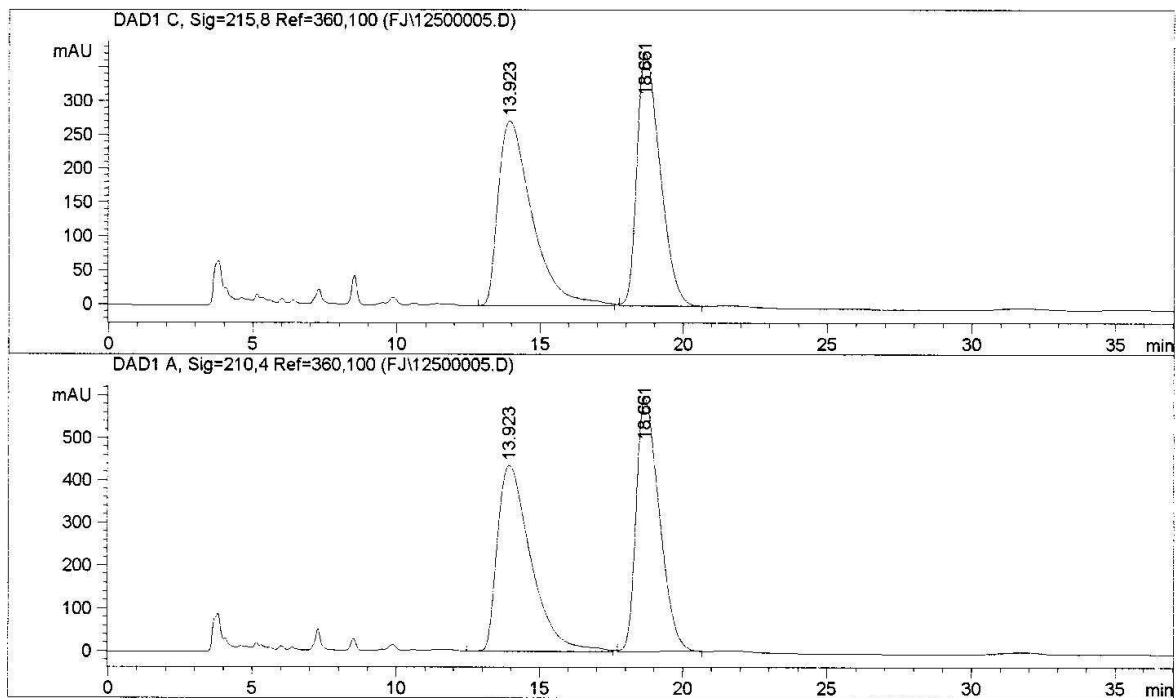


Substrate isolated in the KR of (±)-5a: (1*R*,2*S*)-5a, which is transformed into (1*R*,2*S*)-6a (ee = 97%).

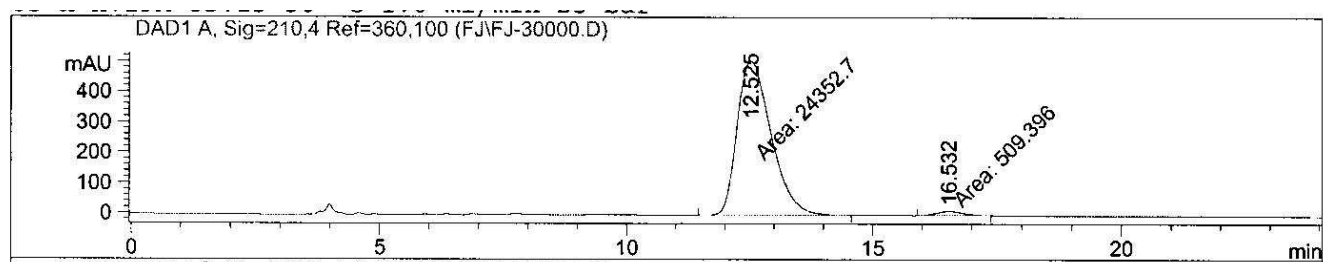




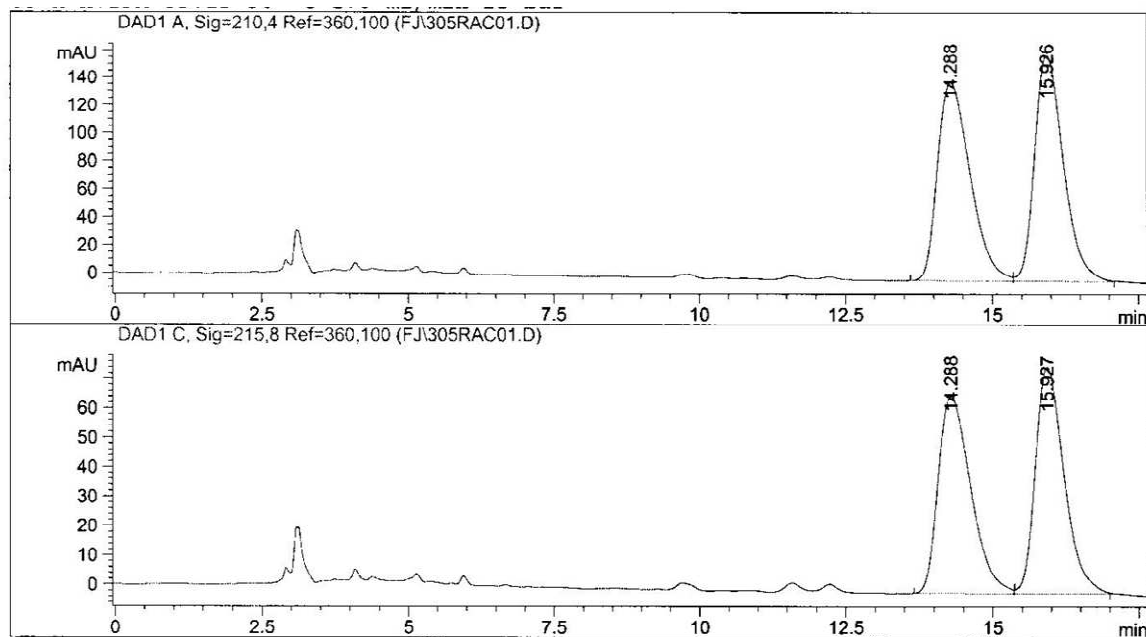
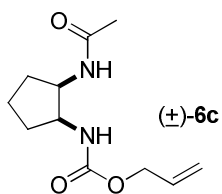
AS-APH H:EtOH 90:10 0.8mL/min 20 °C 13 bar



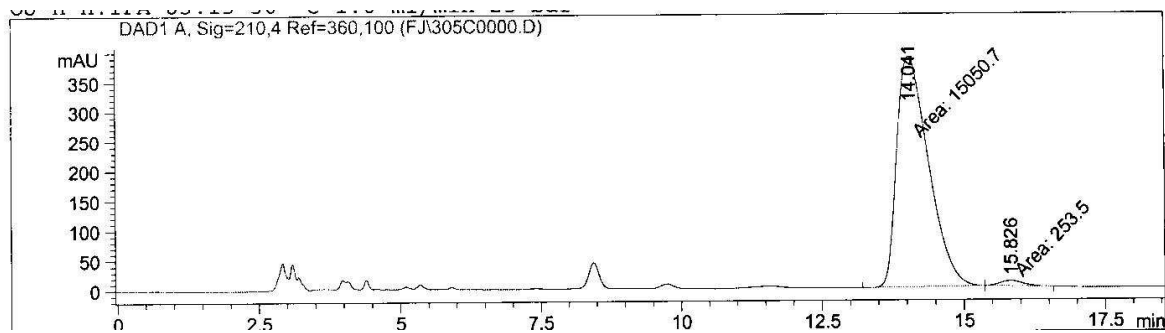
Product isolated in the DKR of (±)-**5b**: (1*S*,2*R*)-**6b** (ee = 96%). Analysis was carried out at 30 °C.



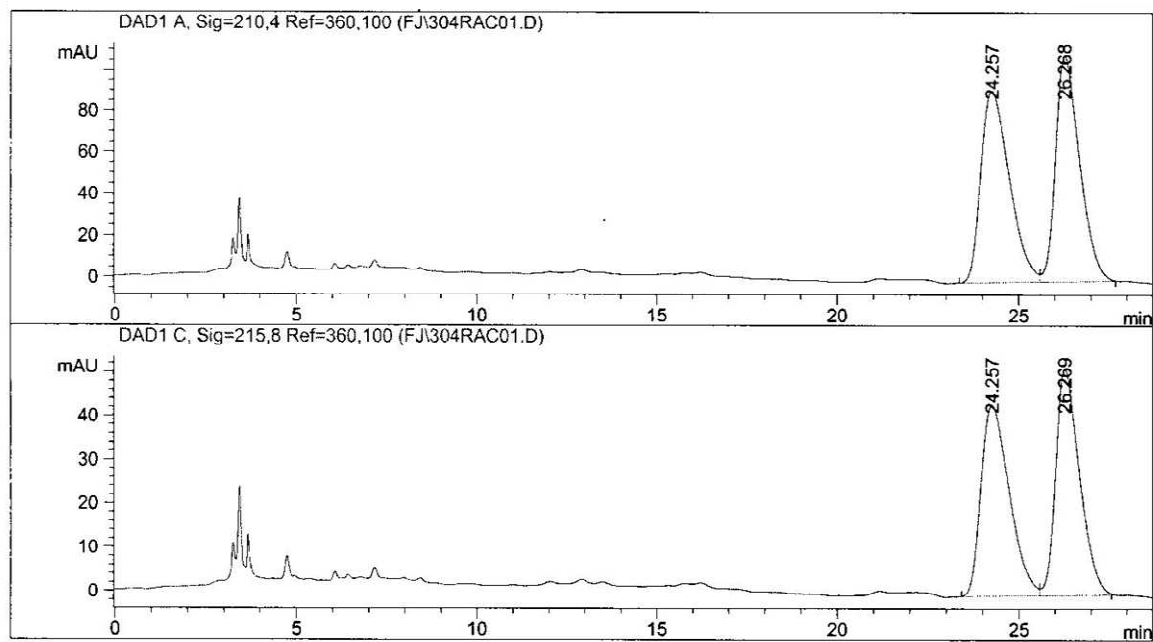
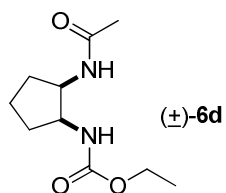
FJ-305rac
Chiralpak IA H:EtOH 94:6 1.0 ml/min 25 °C 29 BAR



Product isolated in the DKR of (±)-**5c**: (1*S*,2*R*)-**6c** (ee = 97%).



FJ-304rac
Chiralpak IA H:EtOH 96:4 0.9 ml/min 25 °C 25 BAR



Product isolated in the DKR of (±)-5d: (1*S*,2*R*)-6d (ee = 95%).

