Ligand-Accelerated C–H Activation Reactions: Evidence for a Switch of Mechanism

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SUPPORTING INFORMATION

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General Information: Unless otherwise noted, all materials were used as received from commercial sources without further purification. The phenylacetic acid substrates and olefin coupling partners were purchased from Acros, Sigma-Aldrich, TCI and Alfa-Aesar and were used as received. 2-(Trifluoromethyl)phenylacetic acid (1) was purchased from TCI; samples of 1 from other commercial sources were found to give irreproducible results. 1,4-Benzoquinone (BQ) was sublimed prior to used. Freshly distilled methyl vinyl ketone was used in the synthesis of 1g. Commercially available organic acid ligands were purchased from Acros, Sigma Aldrich, and Alfa Aesar. In the optimization studies 5a was used as a ligand; its synthesis is described herein. Commercially available amino acid ligands were purchased from Bachem, EMD, or Novabiochem. L4 was prepared according to a method developed by Burgess. L6 was prepared according to a literature procedure.² All others were prepared following literature precedent.^{3,4} Palladium acetate and potassium hydrogen carbonate were purchased from Sigma-Aldrich and Fisher, respectively, and were used without further purification. All reactions were run on hot plates with oil baths calibrated to an external thermometer. Prior to beginning an experiment, the hot plate was turned on, and the oil bath was allowed to equilibrate to the desired temperature for 30 minutes. Infrared spectra were recorded on a Perkin Elmer FT-IR Spectrometer. ¹H and ¹³C NMR spectra were recorded on Varian Mercury (300 MHz and 75 MHz, respectively), Varian Inova (400 MHz and 100 MHz, respectively) and Bruker DRX (500 MHz and 125 MHz, respectively) instruments internally referenced to SiMe₄ or chloroform signals. The following abbreviations (or combinations thereof) were used to explain multiplicities: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, and a = apparent. High resolution mass spectra were recorded at the Center for Mass Spectrometry. The Scripps Research Institute.

Experimental:

Initial Rate Measurements:

General procedure for determining the initial reaction rate for C–H olefination of 1 (or 2) under different conditions: Four different reaction conditions were examined: (1) without Boc-Val-OH, without BQ, (2) without Boc-Val-OH, with BQ, (3) with Boc-Val-OH, without BQ, (4) with Boc-Val-OH, with BO. To establish the initial rate under each of the conditions, four parallel reactions were set up simultaneously. Four 50 mL Schlenk-type sealed tube (with a Teflon high pressure valve and side arm) were obtained, each equipped with a magnetic stir bar. Each tube was charged with 1 (or 2) (0.5 mmol), Pd(OAc)₂ (5.6 mg, 0.025 mmol), KHCO₃ (100.1 mg, 1.0 mmol), BQ (2.7 mg, 0.025 mmol) (when used), Boc-Val-OH (10.9 mg, 0.05 mmol) (when used), ethyl acrylate (106 µL, 1.0 mmol), and t-AmylOH (2.5 mL). The reaction tubes were capped, then evacuated briefly under high vacuum and charged with O₂ (1 atm, balloon) (×3). The reaction mixtures were stirred at room temperature for 5 min, then at 90 °C for the appropriate time. At regular intervals (every 5 minutes, or every 30 minutes), one of the reactions would be removed from the hot plate and cooled to 0 °C in an ice bath. A 2.0 N HCl solution (5 mL) and diethyl ether (10 mL) were then added. A small aliquot of the organic phase was taken, concentrated in vacuo, and analyzed by ¹H NMR. The conversion was determined by integration of the benzylic methylene proton signals, which appear as singlets (approximately 3.87 ppm for 1, 4.01 ppm for 1a, 3.67 ppm for 2, and 3.84 ppm for 2a). For each condition, this process was repeated three times. The resulting data was plotted, and linear regression

established the initial rate. Representative data for the determination of one initial rate are shown in Figure S1.

26.2

43.1

42.4

43.7

0.0

1.0

7.5

8.1

4.3

34.0

45.9

	0.12						
	0.1 -					•	
	0.08				•		
[1a] (M)	0.06			• /	•		
	0.04			/•			
	0.02	$y = 0.0047x - 0.004$ $R^2 = 0.928$					
	0		5	10	15	20	25
		,	Ŭ	Time		20	20

Figure \$1: Representative initial rate data for C-H olefination of 1 with Boc-Val-OH and without BQ.

Synthesis of New Ligands:

Entry^a

1

2

3

4

5

15

20

33.3

50.8

$$\begin{array}{c} \text{Me} \\ \text{H}_2\text{N} \\ \text{CO}_2\text{H} \end{array} + \underbrace{\begin{array}{c} \text{O} \\ \text{X} \\ \text{CI} \\ \text{(1.5 equiv.)} \end{array}}_{\text{(1.5 equiv.)}} \underbrace{\begin{array}{c} \text{NaOH} \\ \text{H}_2\text{O/1,4-dioxane} \\ \text{0 °C to RT} \end{array}}_{\text{NaOH}} \underbrace{\begin{array}{c} \text{Me} \\ \text{NaOH} \\ \text$$

Scheme S1: General procedure for *N*-protection for L-valine.

General procedure for the synthesis of mono-*N*-protected L-valine ligands:^{3,4} A 500 mL round bottom flask equipped with a magnetic stir bar was charged with distilled H_2O (100 mL) and NaOH (100 mmol, 4.0 g). The resulting solution was cooled to 0 °C in an ice bath. L-valine (35 mmol, 4.1 g) was added, and the solution was stirred until it was homogeneous. The flask was equipped with an addition funnel. The corresponding carbonyl chloride (45.5 mmol) in 1,4-dioxane (40 mL) was added dropwise. The reaction mixture was then allowed to stir at room temperature overnight. The following morning, the solution was extracted with Et_2O (3 × 50 mL), and the organic layers were discarded. The aqueous layer was again cooled to 0 °C in an ice bath, and concentrated HCl was added dropwise until the pH had reached 2 (as observed by pH paper). The aqueous solution was extracted with Et_2O (3 × 100 mL). The organic layers were combined, dried over anhydrous Na_2SO_4 , filtered, and concentrated *in vacuo* to give the crude product. The pure product was obtained following recrystallization from Et_2O /hexanes or column chromatography using 15:1 DCM:MeOH as the solvent system. For a general depiction of this procedure, see Scheme S1.

Characterization of New Ligands:

for $C_{16}H_{25}NO_3$ [M+H]⁺ 280.1907, found 280.1915.

N-Carbamoyl-protected amino acids are known to exist as *syn*- and *anti*-rotamers,⁵ with the latter being more thermodynamically stable than the former. In the ¹H and ¹³C NMR spectra of MeO₂C-Val-OH, EtO₂C-Val-OH, *i*-BuO₂C-Val-OH, Cbz-Val-OH, and Men-Val-OH, we detected both rotamers, generally observing <10% of the *syn*-rotamers in solution at room temperature. For simplicity, the ¹H and ¹³C NMR spectral data described below correspond exclusively to the *anti*-rotamers.

(neat) v 3376, 2964, 2875, 1725, 1625, 1518, 1486, 1403, 1370, 1198, 1141, 917, 840 cm⁻¹; **HRMS** (ESI-TOF) m/z Calcd for $C_{10}H_{19}NO_3$ [M+H]⁺ 202.1438, found 202.1447.

(S)-3-methyl-2-adamantanecarbonylamidobutanoic acid (Ada-Val-OH): The title compound was obtained as a white solid (3.7 g, 75% yield). ¹H NMR (400 MHz, CDCl₃)
$$\delta$$
 6.11 (br, 1 H), 4.51 (dd, J_1 = 8.0 Hz, J_2 = 5.2 Hz, 1 H), 2.30–2.22 (m, 1 H), 2.06 (s, 3 H), 1.90–1.86 (m, 6 H), 1.77–1.69 (m, 6 H), 0.98 (d, J = 6.8 Hz, 3 H), 0.96 (d, J = 6.8 Hz, 3 H); ¹³C NMR (100 MHz, CDCl₃) δ 174.96, 56.96, 40.83, 39.12, 36.41, 30.71, 28.02, 19.07, 17.77; IR (neat) v 3375, 2962, 2906, 2852, 1728, 1624, 1520, 1301, 1207, 1147, 1110, 984 cm⁻¹; HRMS (ESI-TOF) m/z Calcd

MeO₂C-Val-OH): The title compound was obtained as a white solid (4.1 g, 67% yield). ¹H NMR (400 MHz, CDCl₃) δ 5.17 (br, 1 H), 4.33 (dd, $J_I = 8.8$ Hz, $J_2 = 4.4$ Hz, 1 H), 3.70 (s, 3 H), 2.28–2.20 (m, 1 H), 1.01 (d, J = 6.8 Hz, 3 H), 0.94 (d, J = 6.8 Hz, 3 H); ¹³C NMR (100 MHz, CDCl₃) δ 176.97, 157.08, 58.78, 52.50, 30.96, 18.98, 17.29; **IR** (neat) ν 3324, 2966, 1695, 1526, 1466, 1373, 1304, 1214, 1098, 1041, 1021, 840, 778 cm⁻¹; **HRMS** (ESITOF) m/z Calcd for C₇H₁₃NO₄ [M+H]⁺ 176.0917, found 176.0918.

(S)-2-(ethoxycarbonylamino)-3-methylbutanoic acid (EtO₂C-Val-OH): The title compound was obtained as a colorless oil (3.9 g, 59% yield). ¹H NMR (400 MHz, CDCl₃) δ 5.13 (br, 1 H), 4.33 (dd, $J_I = 8.8$ Hz, $J_2 = 4.8$ Hz, 1 H), 4.15 (q, $J_1 = 6.8$ Hz, 3 H), 0.94 (d, $J_1 = 6.8$ Hz, 3 H), 1.26 (t, $J_1 = 6.8$ Hz, 3 H), 1.01 (d, $J_1 = 6.8$ Hz, 3 H), 0.94 (d, $J_1 = 6.8$ Hz, 3 H); ¹³C NMR (100 MHz, CDCl₃) δ 176.81, 156.69, 61.36, 58.68, 30.98, 18.99, 17.31, 14.45; IR (neat) v 3323, 2967, 2936, 1693, 1523, 1419, 1382, 1343, 1303, 1216, 1098, 1030, 871, 778 cm⁻¹; HRMS (ESI-TOF) m/z Calcd for $C_8H_{15}NO_4$ [M+H]⁺ 190.1074, found 190.1073.

Me Ne CO₂H (S)-2-(isobutoxycarbonylamino)-3-methylbutanoic acid (i-BuO₂C-Val-OH): The title compound was obtained as a colorless oil (7.4 g, 97% yield). ¹H NMR (400 MHz, CDCl₃) δ 5.16 (br, 1 H), 4.33 (dd, J_I = 8.8 Hz, J_2 = 4.4 Hz, 1 H), 3.86 (d, J = 6.8 Hz, 2 H), 2.26–2.18 (m, 1 H), 1.97–1.87 (m, 1 H), 1.01 (d, J = 6.8 Hz, 3 H), 0.94 (d, J = 6.8 Hz, 3 H), 0.92 (d, J = 6.8 Hz, 6 H); ¹³C NMR (100 MHz, CDCl₃) δ 176.81, 156.83, 71.47, 58.71, 31.03, 27.93, 18.96, 17.34; IR (neat) v 3323, 2964, 2876, 1705, 1516, 1468, 1421, 1217, 1095, 1036, 777 cm⁻¹; HRMS (ESI-TOF) m/z Calcd for C₁₀H₁₉NO₄ [M+H]⁺ 218.1387, found 218.1380.

Me (S)-2-(benzyloxycarbonylamino)-3-methylbutanoic acid (Cbz-Val-OH): The title compound was obtained as a colorless oil (8.4 mg, 96% yield). He (NMR (400 MHz, CDCl₃) δ 7.37–7.26 (m, 5 H), 5.25 (br, 1 H), 5.12 (s, 2 H), 4.36 (dd, J_I = 9.2 Hz, J_2 = 4.8 Hz, 1 H), 2.28–2.20 (m, 1 H), 1.01 (d, 6.8 Hz, 3 H), 0.93 (d, J = 6.8 Hz, 3 H); 13 C NMR (100 MHz, CDCl₃) δ 176.99, 156.35, 136.05, 128.52, 128.23, 128.14, 67.20, 58.81, 31.02, 18.98, 17.32; IR (neat) v 3321, 2966, 2936, 1705, 1514, 1454, 1413, 1346, 1302, 1214, 1094, 1026, 909, 731 cm⁻¹; HRMS (ESI-TOF) m/z Calcd for $C_{13}H_{17}NO_4$ [M+H]⁺ 252.1230, found 252.1233.

(S)-2-(((1R,2S,5R)-2-isopropyl-5-methylcyclohexyloxy)carbonylamino)-3-methylbutanoic acid (Men-Val-OH): The title compound was obtained as a white solid (6.0 g, 57% yield). ¹H NMR (400 MHz, CDCl₃) δ 5.10 (br, 1 H), 4.58–4.54 (m, 1 H), 4.33 (dd, J_I = 8.8 Hz, J_Z = 4.4 Hz, 1 H), 2.26–2.18 (m, 1 H), 2.02 (ad, J = 12.0 Hz, 1 H), 1.98–1.86 (m, 1 H), 1.66 (ad, J = 12.0 Hz, 2 H), 1.53–1.41 (m, 1 H), 1.31 (at, J = 12.0 Hz, 1 H), 1.09–0.99 (m, 2 H), 1.01 (d, J = 6.8 Hz, 3 H), 0.93 (d, J = 6.8 Hz, 3 H), 0.90–0.83 (m, 7 H), 0.76 (d, J = 6.8 Hz, 3

H); ¹³C **NMR** (100 MHz, CDCl₃) δ 177.22, 156.41, 75.16, 58.71, 47.33, 41.27, 34.25, 31.35, 31.07, 26.11, 23.45, 22.02, 20.77, 19.04, 17.37, 16.34; **IR** (neat) v 3319, 2958, 2929, 2871, 1713, 1665, 1513, 1456, 1414, 1305, 1224, 1094, 1027, 983, 909 cm⁻¹; **HRMS** (ESI-TOF) m/z Calcd for $C_{16}H_{29}NO_4$ [M+H]⁺ 300.2169, found 300.2167.

Optimization Studies:

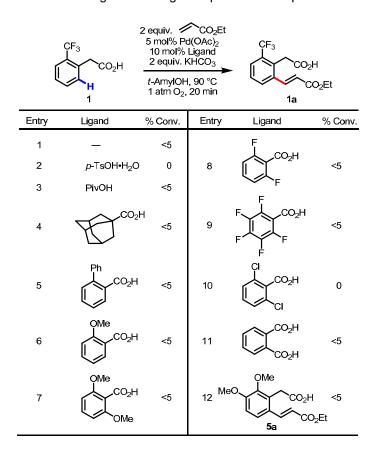
Ligand optimization for Pd(II)-catalyzed olefination with 2-(Trifluoromethyl)phenylacetic acid (1): A 50 mL Schlenk-type sealed tube (with a Teflon high pressure valve and side arm) equipped with a magnetic stir bar was charged with 1 (102.1 mg, 0.5 mmol), Pd(OAc)₂ (5.6 mg, 0.025 mmol), KHCO₃ (100.1 mg, 1.0 mmol), ligand (0.05 mmol), ethyl acrylate (106 µL, 1.0 mmol), and t-AmylOH (2.5 mL). The reaction tube was capped, then evacuated briefly under high vacuum and charged with O_2 (1 atm, balloon) (×3). The reaction mixture was stirred at room temperature for 5 min, then at 90 °C for the appropriate time. The reaction vessel was removed from the oil bath and immediately cooled to 0 °C in an ice bath. A 2.0 N HCl solution (5 mL) and diethyl ether (10 mL) were added. A small aliquot of the organic phase was taken, concentrated in vacuo, and analyzed by ¹H NMR. The conversion was determined by integration of the benzylic methylene proton signals, which appear as singlets (approximately 3.87 ppm for 1 and 4.01 ppm for 1a). When indicated, the reactions were performed in triplicates, and the values shown represent the average result from the three experiments. In our initial efforts, we measured the conversion after 2 h (Table S1), but we observed quantitative conversion for many ligands. We then adjusted our assay and examined the conversion after 20 min (Tables S2 and S3).

Table S1: Initial results from the amino acid ligand optimization experiments.^a

ĺ	CF ₃ CO ₂ H	2 equiv. 5 mol% Po 10 mol% 2 equiv. k t-AmylOH 1 atm O	Lìgand (HCO ₃), 90 °C	CF ₃ CO ₂ H CO ₂ Et		
Entry	Ligand	% Conv.	Entry	Ligand	% Conv.	
1		7 ^b	8	Boc-Ser-OH	16	
2	Boc-Val-OH	98 (96)	9	Ac-Val-OH	>99	
3	Boc-Ile-OH	98	10	Ac-IIe-OH	>99 (96)	
4	Boc-Leu-OH	92	11	Ac-Leu-OH	>99	
5	Boc-t-Leu-OH	98	12	Formyl-Ile-OH	36	
6	Boc-Ala-OH	88	13	Formyl-Leu-OH	37	
7	Boc-Phe-OH	96	14	Men-Leu-OH ^c	71	

^a The conversion was determined by ¹H NMR analysis of the crude reaction mixture. Isolated yield is given in parentheses. ^b Average of three trials. ^c Men = (–)-Menthyl(O_2C).

Table S2: Results from the organic acid ligand optimization experiments.^a



^a The conversion was determined by ¹H NMR analysis of the crude reaction mixture.

Table S3: Results from the amino acid ligand optimization experiments.^a

Ac-Ala-OH

Ac-Gly-OH

Ac-lle-OH

Ac-Phe-OH

L6

71

51 72^b

60

31

16

31

33

35

14

15

16

BocHN'

olefination 2-Additional optimization for Pd(II)-catalyzed with (Trifluoromethyl)phenylacetic acid (1): The effects of ligand loading (Table S4), olefin loading (Table S5), and reaction temperature (Tables S6–S10) were explored using Ac-Ile-OH as the ligand. A 50 mL Schlenk-type sealed tube (with a Teflon high pressure valve and side arm) equipped with a magnetic stir bar was charged with 1 (102.1 mg, 0.5 mmol), Pd(OAc)₂ (5.6 mg, 0.025 mmol), KHCO₃ (100.1 mg, 1.0 mmol), Ac-Ile-OH, ethyl acrylate, and t-AmylOH (2.5 mL). The reaction tube was capped, then evacuated briefly under high vacuum and charged with O₂ (1 atm, balloon) (×3). The reaction mixture was stirred at room temperature for 5 min, then at the appropriate temperature for the indicated time. The reaction vessel was then cooled to 0 °C in an ice bath. A 2.0 N HCl solution (5 mL) and diethyl ether (10 mL) were added. A small aliquot

^a The conversion was determined by ¹H NMR analysis of the crude reaction mixture. ^b Average of three trials. ^c Ada = Adamantyl(OC). ^d Men = (–)-Menthyl(O₂C).

of the organic phase was taken, concentrated *in vacuo*, and analyzed by ¹H NMR. The conversion was determined by integration of the benzylic methylene proton signals, which appear as singlets (approximately 3.87 ppm for **1** and 4.01 ppm for **1a**). When indicated, the reactions were performed in triplicates, and the values shown represent the average result from the three experiments.

Table S4: Optimization of ligand loading.^a

Table \$5: Optimization of olefin loading.^a

^a The conversion was determined by ¹H NMR analysis of the crude reaction mixture. Isolated yield is given in parenthesis. ^b Average of three trials.

^a The conversion was determined by ¹H NMR analysis of the crude reaction mixture. Isolated yield is given in parenthesis. ^b Average of three trials.

Table S6: Conversion versus time at 50 °C.

Table S7: Conversion versus time at 70 °C.

^a The conversion was determined by ¹H NMR analysis of the crude reaction mixture.

^a The conversion was determined by ¹H NMR analysis of the crude reaction mixture.

Table S8: Conversion versus time at 90 °C.

Table S9: Conversion versus time at 110 °C.^a

 $[^]a$ The conversion was determined by $^1{\rm H}$ NMR analysis of the crude reaction mixture. Isolated yield is given in parenthesis. b Average of three trials.

^a The conversion was determined by ¹H NMR analysis of the crude reaction mixture.

Table \$10: Conversion versus time at 130 °C.

General Procedure:

Scheme S2: General procedure for C-H olefination of phenylacetic acids.

General procedure for Pd(II)-catalyzed *ortho*-C-H olefination of phenylacetic acids 1–10: A 50 mL Schlenk-type sealed tube (with a Teflon high pressure valve and side arm) equipped with a magnetic stir bar was charged with the phenylacetic acid starting material (1–10) (0.5 mmol), Pd(OAc)₂ (5.6 mg, 0.025 mmol), KHCO₃ (100.1 mg, 1.0 mmol), ligand (0.05 mmol), the olefin coupling partner (1.0 mmol), and *t*-AmylOH (2.5 mL). The reaction tube was capped, then evacuated briefly under high vacuum and charged with O₂ (1 atm, balloon) (×3). The reaction mixture was stirred at room temperature for 5 min, then at 90 °C for 2 h (longer, when noted). The reaction vessel was then cooled to 0 °C in an ice bath. A 2.0 N HCl solution (5 mL) was then added, and the mixture was extracted with EtOAc (3 × 20 mL). The organic layers were combined, dried over anhydrous Na₂SO₄, filtered, and concentrated *in vacuo*. The resulting residue was purified by silica gel flash column chromatography using 3:1 hexanes:EtOAc (with 3% HOAc) as the eluent. For a general depiction of this procedure, see Scheme S2.

Characterization of New C-H Olefinated Products:

Our group has previously reported analytical data for 1a, 5a, 6a, and 8a'. In this same publication, 7a, 9a, and 10a were also synthesized but were isolated as their methyl esters following treatment of the crude product mixture with CH_2N_2 . This was done to simplify the separation of the product from residual unreacted starting material. Because the reactions reported here generally gave quantitative yields, the pure products could consistently be isolated as free acids. We have included the analytical data for free acids 7a, 9a, and 10a.

^a The conversion was determined by ¹H NMR analysis of the crude reaction mixture.

$$CO_2H$$

(E)-2-(2-(3-ethoxy-3-oxoprop-1-enyl)-6-methylphenyl)acetic acid (2a): The title compound was obtained as a white solid (122 mg, 98% yield). ¹H NMR (400 MHz, CDCl₃) δ 8.00 (d, J = 15.6 Hz, 1 H), 7.42, (dd, $J_1 = 6.8$ Hz, $J_2 = 2.0$ Hz, 1 H), 7.24-7.18 (m, 2 H), 6.33 (d, J = 15.6 Hz, 1 H), 4.26 (q, J = 7.2 Hz, 2 H), 3.84 (s, 2 H), 2.34 (s, 3 H), 1.33 (t, J = 7.2 Hz, 3 H); ¹³C NMR (100 MHz,

 $CDCl_3$) δ 176.45, 166.83, 142.47, 138.03, 134.75, 131.96, 131.50, 127.69, 124.92, 121.10, 60.61, 34.67, 20.21, 14.26; **IR** (neat) v 2981, 2935, 1704 (br), 1631, 1446, 1368, 1313, 1269, 1237, 1179, 1162 (br), 1093, 1035, 979, 788 cm⁻¹; **HRMS** (ESI-TOF) m/z Calcd for $C_{14}H_{17}O_4$ [M+H]⁺ 249.1121, found 249.1124.

$$F_3C \underbrace{CO_2H}_{CO_2Et}$$

(E)-2-(2-(3-ethoxy-3-oxoprop-1-enyl)-5-(trifluoromethyl)phenyl)acetic acid (3a): The title compound was obtained as a white solid (147 mg, 97% yield). ¹**H NMR** (400 MHz, CDCl₃) δ 7.91 (d, J = 15.6 Hz, 1 H), 7.69 (d, J = 15.6 Hz, 8.0 Hz, 1 H), 7.58 (d, J = 8.0 Hz, 1 H), 7.55 (s, 1 H), 6.43 (d, J = 15.6 Hz, 1 H), 4.28 (q, J = 7.2 Hz, 2 H), 3.87 (s, 2 H), 1.34 (t, J = 7.2 Hz, 3 H); ¹³C NMR (100 MHz,

CDCl₃) δ 174.95, 166.29, 140.01, 137.74, 133.40, 131.69 (q, $J_{C-F} = 32.7$ Hz), 128.02 (q, $J_{C-F} = 32.7$ Hz) 3.7 Hz), 127.47, 124.90 (q, $J_{C-F} = 3.7$ Hz), 123.60 (q, $J_{C-F} = 270.7$ Hz), 122.24, 60.98, 38.23, 14.22; **IR** (neat) v 2987, 2932, 1715 (br), 1638, 1422, 1335, 1283, 1222, 1167 (br), 1127, 1080, 1035, 979 cm⁻¹; **HRMS** (ESI-TOF) m/z Calcd for $C_{14}H_{14}F_{3}O_{4}$ [M+H]⁺ 303.0839, found 303.0844.

(E)-2-(2-(3-ethoxy-3-oxoprop-1-enyl)-5-methylphenyl)acetic acid (4a): The title compound was obtained as a colorless oil (103 mg, 99% yield). ¹H **NMR** (400 MHz, CDCl₃) δ 7.90 (d, J = 15.6 Hz, 1 H), 7.51 (d, J = 8.0 Hz, 1 H), 7.12 (d, J = 8.0 Hz, 1 H), 7.09 (s, 1 H), 6.34 (d, J = 15.6 Hz, 1 H), 4.25 (q, J = 7.2 Hz, 2 H), 3.77 (s, 2 H), 2.35 (s, 3 H), 1.32 (t, J = 7.2 Hz, 3 H); ¹³C NMR (100 MHz,

CDCl₃) δ 176.44, 167.01, 141.31, 140.54, 132.79, 131.87, 131.14, 128.92, 126.83, 119.53, 60.56, 38.31, 21.26, 14.27; **IR** (neat) v 2982, 2927, 1708 (br), 1632, 1610, 1368, 1315, 1283, 1179 (br), 1159, 1095, 1035, 979, 816 cm⁻¹; **HRMS** (ESI-TOF) m/z Calcd for $C_{14}H_{17}O_4$ [M+H]⁺ 249.1121, found 249.1118.

$$CI \qquad CO_2H \qquad CO_2Et$$

(E)-2-(2,4-dichloro-6-(3-ethoxy-3-oxoprop-1-enyl)phenyl)acetic acid (7a): The title compound was obtained as a white solid (150 mg, 99% yield). ¹H **NMR** (400 MHz, CDCl₃) δ 7.79 (d, J = 15.6 Hz, 1 H), 7.45 (s, 2 H), 6.35 (d, J= 15.6 Hz, 1 H), 4.27 (q, J = 7.2 Hz, 2 H), 3.69 (s, 2 H), 1.33 (t, J = 7.2 Hz, 3 H); ¹³C NMR (100 MHz, CDCl₃) δ 174.83, 166.03, 139.96, 137.75, 136.67,

134.17, 130.21, 129.49, 125.72, 123.73, 61.05, 34.83, 14.21; **IR** (neat) v 2982, 2930, 1708 (br), 1637, 1584, 1553, 1446, 1398, 1368, 1310, 1274, 1237, 1178 (br), 1153, 1031, 972, 911, 859 cm⁻¹; **HRMS** (ESI-TOF) m/z Calcd for $C_{13}H_{13}Cl_2O_4$ [M+H]⁺ 303.0190, [(M+2)+H]⁺ 305.0161, $[(M+4)+H]^+$ 307.0132, found 303.0188, 305.0160, 307.0140 $[M+H]^+$: $[(M+2)+H]^+$ $[(M+4)+H]^{+}=9:6:1.$

(E)-2-(2-(3-ethoxy-3-oxoprop-1-enyl)-6-nitrophenyl)acetic acid (8a): The title compound was obtained as a white solid (98 mg, 70% yield). ¹H NMR (400 MHz, CDCl₃) δ 8.01 (d, J = 7.6 Hz, 1 H), 7.92 (d, J = 15.6 Hz, 1 H), 7.78 (d, J = 7.6 Hz, 1 H), 7.48 (t, J = 7.6 Hz, 1 H), 6.38 (d, J = 15.6 Hz, 1 H), 4.29 (q, J = 7.2 Hz, 2 H), 4.08 (s, 2 H), 1.35 (t, J = 7.2 Hz, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 174.40, 165.93, 150.32, 140.10, 137.86, 132.00, 128.61, 127.26, 125.83, 124.68, 61.15 34.36, 14.22; IR (neat) v 3089, 2985, 1710 (br), 1638, 1607, 1529, 1351, 1317, 1185 (br), 1032, 975, 894, 808, 744 cm⁻¹; HRMS (ESI-TOF) m/z Calcd for $C_{13}H_{14}NO_6$ [M+H]⁺ 280.0816, found 280.0822.

(E)-2-(5-benzoyl-2-(3-ethoxy-3-oxoprop-1-enyl)phenyl)propanoic acid (9a): The title compound was obtained as a viscous yellow oil (188 mg, 98% yield). ¹H NMR (400 MHz, CDCl₃) δ 8.08 (d, J = 15.6 Hz, 1 H), 7.83–7.45 (m, 8 H), 6.44 (d, J = 15.6 Hz, 1 H), 4.29 (q, J = 7.2 Hz, 2 H), 4.20 (q, J = 7.2 Hz, 1 H), 1.55 (d, J = 7.2 Hz, 3 H), 1.35 (t, J = 6.8 Hz, 3 H); ¹³C NMR (100 MHz, CDCl₃) δ 195.66, 178.57, 166.28, 140.56, 139.19, 138.64, 137.38, 137.06, 132.74, 130.05, 129.33, 129.18, 128.37, 127.23, 123.40, 60.91, 40.92, 18.02, 14.27; IR (neat) v 2982, 2938, 1708 (br), 1657, 1635, 1598, 1447, 1410, 1368, 1314, 1272, 1217, 1178 (br), 1031, 960, 843, 754, 721 cm⁻¹; HRMS (ESI-TOF) m/z Calcd for $C_{21}H_{21}O_5$ [M+H]⁺ 353.1383, found 353.1388.

(E)-2-(3-(3-ethoxy-3-oxoprop-1-enyl)-6-methoxynaphthalen-2-yl)propanoic acid (10a): The title compound was obtained as a viscous yellow oil (153 mg, 93% yield). ¹H NMR (400 MHz, CDCl₃) δ 8.17 (d, J = 15.6 Hz, 1 H), 7.92 (s, 1 H), 7.72 (s, 1 H), 7.67 (d, J = 8.8 Hz, 1 H), 7.14 (d, J = 9.2 Hz, 1 H), 7.09 (s, 1 H), 6.46 (d, J = 15.6 Hz, 1 H), 4.29 (q, J = 7.2 Hz, 2 H), 4.21 (q, J = 6.8 Hz, 1 H), 3.92 (s, 3 H), 1.59 (d, J = 6.8 Hz, 3 H), 1.35 (t, J = 7.2, 3 H); ¹³C NMR (100 MHz, CDCl₃) δ 179.98, 166.70, 158.08, 142.39, 133.98, 133.42, 132.44, 129.53, 129.14, 126.11, 125.92, 121.53, 120.18, 105.56, 60.66, 55.30, 41.03, 18.31, 14.28; IR (neat) v 2980, 2934, 1704 (br), 1634, 1600, 1499, 1465, 1397, 1368, 1301, 1266, 1207, 1166 (br), 1029, 975, 896, 866, 811 cm⁻¹; HRMS (ESI-TOF) m/z Calcd for $C_{19}H_{21}O_{5}$ [M+H]⁺ 329.1383, found 329.1378.

(E)-2-(2-(3-tert-butoxy-3-oxoprop-1-enyl)-6-(trifluoromethyl)phenyl)acetic acid (1b): The title compound was obtained as a white solid (163 mg, 99% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.81 (d, J = 15.6 Hz, 1 H), 7.73 (d, J = 7.6 Hz, 1 H), 7.70 (d, J = 7.6 Hz, 1 H), 7.43 (t, J = 7.6 Hz, 1 H), 6.31 (d, J = 15.6 Hz, 1 H), 4.01 (s, 2 H), 1.53 (s, 9 H); ¹³C NMR (100 MHz, CDCl₃) δ 175.21, 165.63, 139.64, 137.37, 130.88, 130.61, 130.16 (q, J_{C-F} = 29.5 Hz), 127.88, 127.06 (q, J_{C-F} = 5.6 Hz), 124.66, 124.02 (q, J_{C-F} = 272.6 Hz), 81.28, 34.21, 28.07; **IR** (neat) v 2981, 2935, 1710 (br), 1637, 1454,

1324 (br), 1149, 1119, 1099, 981, 961, 842, 806, 752 cm⁻¹; **HRMS** (ESI-TOF) m/z Calcd for $C_{16}H_{17}F_3O_4Na$ [M+Na]⁺ 353.0971, found 353.0977.

(E)-2-(2-(3-(benzyloxy)-3-oxoprop-1-enyl)-6-(trifluoromethyl)phenyl)acetic acid (1c): The title compound was obtained as a white solid (165 mg, 90% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.95 (d, J = 16.0 Hz, 1 H), 7.74 (d, J = 7.6, 1 H), 7.72 (d, J = 8.0, 1 H), 7.46–7.34 (m, 6 H), 6.43 (d, J = 15.6 Hz, 1 H), 5.25 (s, 2 H), 4.01 (s, 2 H); ¹³C NMR (100 MHz, CDCl₃) δ 175.29, 166.06, 141.15, 137.06, 135.65, 131.00, 130.63, 130.25 (q, J = 29.6 Hz), 128.58, 128.44, 128.32, 128.27, 127.38 (q, J = 5.7 Hz), 123.96 (q, J = 272.3 Hz), 122.77, 66.71, 34.18; **IR** (neat) v 3034, 2962, 1702 (br), 1455, 1422, 1372, 1315, 1275, 1239, 1163 (br), 1110, 1004, 975, 963, 804, 748 cm⁻¹; **HRMS** (ESI-TOF) m/z Calcd for C₁₉H₁₆F₃O₄ [M+H]⁺ 365.0995, found 365.0997.

(E)-2-(2-styryl-6-(trifluoromethyl)phenyl)acetic acid (1d): The title compound was obtained as a white solid (153 mg, 99% yield). 1 H NMR (400 MHz, CDCl₃) δ 7.72 (d, J = 7.6, 1 H), 7.56 (d, J = 7.6, 1 H), 7.44–7.17 (m, 7 H) 6.92 (d, J = 16.0 Hz, 1 H), 3.95 (s, 2 H); 13 C NMR (100 MHz, CDCl₃) δ 176.53, 140.21, 136.61, 134.05, 130.29, 129.79 (q, $J_{C-F} = 29.3$ Hz), 129.40, 128.78, 128.35, 127.81, 126.82, 125.18 (q, $J_{C-F} = 5.7$ Hz), 124.71, 124.27 (q, $J_{C-F} = 272.8$ Hz), 34.58; IR (neat) v 3026, 1708 (br), 1495, 1455, 1412, 1315 (br), 1155, 1114, 1098, 957, 798, 749 cm $^{-1}$; HRMS (ESI-TOF) m/z Calcd for $C_{17}H_{14}F_3O_2$ [M+H] $^+$ 307.0940, found 307.0941.

(E)-2-(2-(4-chlorostyryl)-6-(trifluoromethyl)phenyl)acetic acid (1e): The title compound was obtained as a white solid (111 mg, 65% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.76 (d, J = 8.0 Hz, 1 H), 7.64 (d, J = 8.0 Hz, 1 H), 7.45–7.32 (m, 5 H), 7.22 (d, J = 16.0 Hz, 1 H), 6.93 (d, J = 16.0 Hz, 1 H), 4.00 (s, 2 H); ¹³C NMR (100 MHz, (CD₃)₂CO) δ 171.51, 140.93, 136.90, 134.07, 134.07, 1332.07, 132.59, 132.19, 131.02, 130.15 (q, J_{C-F} = 28.9 Hz), 129.62, 129.27, 128.49, 126.70, 125.88 (q, J_{C-F} = 5.9 Hz), 125.61 (q, J_{C-F} = 272.0 Hz), 34.94; IR (neat) v 3034, 2962, 1702 (br), 1634, 1455, 1422, 1372, 1315, 1275, 1239, 1163 (br), 1099, 1004, 975, 963, 804, 748 cm⁻¹; HRMS (ESI-TOF) m/z Calcd for $C_{17}H_{13}ClF_3O_2$ [M+H]⁺ 341.0551, [(M+2)+H]⁺ 343.0527, found 341.0556, 343.0527 [M+H]⁺: [(M+2)+H]⁺ = 3:1.

2-(2-(hex-2-enyl)-6-(trifluoromethyl)phenyl)acetic acid (1f): The title compound was obtained as an inseparable mixture of the E- and Z-stereoisomers (89 mg, 62% yield). By ^{1}H NMR, the E:Z ratio was determined to be approximately 4:1. The ^{1}H NMR data reported here represent only the major isomer. Following hydrogenation, 1f was converted to 1f', and a complete set of analytical data were collected (included below). ^{1}H NMR (400 MHz, CDCl₃) δ 7.55 (d, J = 8.0 Hz, 1 H), 7.42 (d, J = 7.6 Hz, 1 H), 7.34 (t, J = 8.0 Hz, 1 H), 5.53–5.39 (m, 2 H), 3.93 (s, 2 H), 3.36 (d, J = 6.0 Hz, 2 H), 1.98 (aq, J = 7.2 Hz, 2 H), 1.44–1.32 (m, 2 H), 0.88 (t, J = 7.2 Hz, 3 H).

2-(2-hexyl-6-(trifluoromethyl)phenyl)acetic acid (1f'): To a 25 mL round bottom flask equipped with a stir bar were added 1f (72.5 mg, 0.25 mmol), MeOH (10 mL), and Pd/C (10 wt%) (2.7 mg, 0.0025 mmol). The flask was evacuated under vacuum, then capped under H₂ (1 atm, balloon). After 6 h, the solution was filtered through a pad of Celite. The title compound was obtained as a colorless oil (71 mg.

99% yield). ¹H NMR (300 MHz, CDCl₃) δ 7.51 (d, J = 7.5 Hz, 1 H), 7.40 (d, J = 7.5 Hz, 1 H), 7.31 (t, J = 7.5 Hz, 1 H), 3.89 (s, 2 H), 2.60 (t, J = 7.8 Hz, 2 H), 1.62–1.51 (m, 2 H), 1.40–1.24 (m, 6 H), 0.89 (t, J = 6.6 Hz, 3 H); ¹³C **NMR** (100 MHz, CDCl₃) δ 177.09, 144.06, 132.96, 129.87, 129.54 (q, $J_{C-F} = 28.9$ Hz), 127.36, 124.48 (q, $J_{C-F} = 272.5$ hz), 123.76 (q, $J_{C-F} = 5.7$ Hz), 34.10, 32.89, 31.64, 30.55, 29.25, 22.55, 14.02; **IR** (neat) v 2929, 2856, 1713 (br), 1462, 1412, 1316, 1157, 1116 (br), 1094, 954, 803 cm⁻¹; **HRMS** (ESI-TOF) m/z Calcd for $C_{15}H_{20}F_3O_2$ [M+H]⁺ 289.1410, found 289.1409.

(E)-2-(2-(3-oxobut-1-enyl)-6-(trifluoromethyl)phenyl)acetic acid (1g): The title compound was obtained as an amorphous off-white solid (123 mg, 90% yield). ¹H **NMR** (400 MHz, CDCl₃) δ 7.76–7.72 (m, 3 H), 7.46 (t, J = 8.0 Hz, 1 H), 6.64 (d, J= 16.0 Hz, 1 H), 4.01 (s, 2 H), 2.38 (s, 3 H); 13 C NMR (100 MHz, (CD₃)₂CO) δ 197.90, 171.27, 139.72, 138.66, 133.64, 132.04, 131.75, 130.40 (q, $J_{C-F} = 29.2$

Hz), 128.80, 127.88 (q, $J_{C-F} = 5.9$ Hz), 125.39 (q, $J_{C-F} = 272.0$ Hz), 34.84, 27.72; **IR** (neat) v 2923, 1746 (br), 1719 (br), 1604, 1467, 1321, 1245, 1155, 1115 (br), 1050, 986, 800 cm⁻¹; **HRMS** (ESI-TOF) m/z Calcd for $C_{13}H_{12}F_3O_3$ $[M+H]^+$ 273.0733, found 273.0740.

(E)-2-(2-(3-oxopent-1-enyl)-6-(trifluoromethyl)phenyl)acetic acid (1h): The title compound was obtained as an amorphous off-white solid (138 mg, 97% yield). ¹**H NMR** (400 MHz, CDCl₃) δ 7.78 (d, J = 16.0 Hz, 1 H), 7.76–7.71 (m, 2 H), 7.45 (t, J = 8.0 Hz, 1 H), 6.66 (d, J = 16.0 Hz, 1 H), 4.01 (s, 2 H), 2.69 (q, J =7.2 Hz, 2 H), 1.16 (t, J = 7.2 Hz, 3 H); ¹³C NMR (100 MHz, (CDCl₃) δ 200.59, 175.18, 138.03, 137.40, 131.16, 130.59, 130.33, 130.27 (q, $J_{C-F} = 29.5 \text{ Hz}$), 128.01, 127.34 (q,

 $J_{C-F} = 5.7 \text{ Hz}$), 123.97 (q, $J_{C-F} = 273.0 \text{ Hz}$), 34.53, 34.25, 7.93; **IR** (neat) v 2982, 2943, 1716, 1671, 1614, 1454, 1319, 1188, 1159, 1119, 1100, 1042, 973, 808 cm⁻¹; **HRMS** (ESI-TOF) m/z Calcd for $C_{14}H_{14}F_3O_3 [M+H]^+ 287.0890$, found 287.0902.

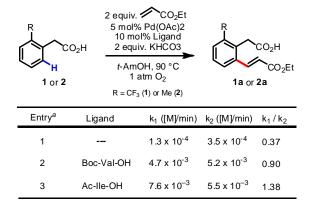
Competition Experiments:

Procedure for intermolecular competition experiments between compounds 1 and 2: A 50 mL Schlenk-type sealed tube (with a Teflon high pressure valve and side arm) equipped with a magnetic stir bar was charged with 1 (51.1 mg, 0.25 mmol), 2 (37.6 mg, 0.25 mmol), Pd(OAc)₂ (5.6 mg, 0.025 mmol), KHCO₃ (100.1 mg, 1.0 mmol), ligand (0.05 mmol), ethyl acrylate (106 μL, 1.0 mmol), and t-AmylOH (2.5 mL). The reaction tube was capped, then evacuated briefly under high vacuum and charged with O₂ (1 atm, balloon) (×3). The reaction mixture was stirred at room temperature for 5 min, then at 90 °C for the appropriate time. The reaction vessel was then cooled to 0 °C in an ice bath. A 2.0 N HCl solution (5 mL) and diethyl ether (10 mL) were added. A small aliquot of the organic phase was taken, concentrated in vacuo, and analyzed by ¹H NMR. The conversion was determined by integration of the benzylic methylene proton signals, which appear as singlets (approximately 3.87 ppm for **1**, 4.01 ppm for **1a**, 3.67 ppm for **2**, and 3.84 ppm for **2a**). The results are shown in Table S11.

Table S11: Competition experiments between 1 and 2.a

Procedure for initial rate studies for single-component reactions of 1 and 2: The procedure for reactions run with and without Boc-Val-OH is described on page S-2. An identical protocol was followed to determine the initial rate in the presence of Ac-Ile-OH (8.7 mg, 0.05 mmol), with reactions stopped at 5 min, 7.5 min, 10 min, and 15 min. The reactions were repeated three times, and determination of the initial rate was performed using linear regression. The overall results are shown in Table S12.

Table S12: Results from the single-component initial rate studies for reactions with **1** and **2**. For an example data set, see Table S1.



^a The conversion was determined by ¹H NMR of the crude reaction mixture.

Kinetic Isotope Effect Experiments:

Scheme S3: Synthetic route to 12.

ortho-tolyl-d₇ bromide (S-1): To a 250 mL round bottom flask equipped with a magnetic stir bar, were added *ortho*-xylene-d₁₀ (2.4 mL, 20 mmol, 99+% atom D), NBS (5.3 g, 30 mmol), and CCl4 (125 mL). The flask was fitted with a reflux condenser, and the solution was heated to refluxing conditions. Catalytic AIBN (75 mg) was added. The reaction was monitored every 20 minutes by GC-MS. After 1 h, it was observed that the reaction had progressed to completion (70% desired mono-brominated product, 30% undesired di-brominated byproduct). The solution was allowed to cool to room temperature, at which point it was filtered to remove the succinimide precipitate. The filtrate was concentrated *in vacuo*, and the crude product mixture was obtained as a colorless oil. This mixture was taken on to the next step without further purification.

ortho-tolyl-d₉ cyanide (S-2): To a 100 mL round bottom flask equipped with a magnetic stir bar, were added crude S-1 (20 mmol), NaCN (1.5 g, 30 mmol), EtOH (50 mL), and distilled H₂O (10 mL). (CAUTION: NaCN is an extremely toxic chemical and should be handled in a well maintained fume hood. The operator should have appropriate protection at all times.) The solution was stirred under refluxing conditions for 12 h. Following careful removal of EtOH by distillation, the reaction mixture was extracted with EtOAc (3 × 50 mL). The combined organic layers were washed with water (3 × 100 mL) and brine (100 mL). The solution was concentrated in vacuo, and the crude product. Purification by column chromatography (15:1 hexanes:EtOAc) gave pure S-2 as a colorless oil (1.48 g, 47% yield over two steps).

ortho-tolyl- d_7 acetic acid (12): To a 250 mL round bottom flask equipped with a magnetic stir bar, were added S-2, KOH (1.7 g, 30 mmol), EtOH (100 mL), and distilled H₂O (25 mL). The solution was heated to reflux for 12 h, at which point EtOH was removed *via* distillation. The resulting aqueous solution was extracted with EtOAc (3 × 50 mL), and the organic layers were discarded. Concentrated HCl was added to the aqueous layer until the pH had reached 2 (as monitored by pH paper). The solution was extracted with EtOAc (3 × 100 mL). The organic layers were combined, dried over anhydrous Na₂SO₄, filtered, and concentrated *in vacuo*. Purification via column chromatography (3:1 hexanes:EtOAc, 3% HOAc) gave 12 as a white solid (448 mg, 28% yield). ¹H NMR (400 MHz, CDCl₃) δ 3.69 (s, 2 H); ¹³C NMR (100 MHz, CDCl₃) δ 179.37, 136.95, 130.20 (t, J_{C-D} = 23.2 Hz), 130.07 (t, J_{C-D} = 23.9 Hz), 25.8 Hz, 127.38 (t, J_{C-D} = 25.8 Hz), 125.91 (t, J_{C-D} = 24.5 Hz), 39.07 (Note: the CD₃ signal could not be seen in the ¹³C NMR spectrum due to its complex splitting.); IR (neat) v 2918, 1690 (br), 1416, 1327, 1230, 908 cm⁻¹; HRMS (ESI-TOF) m/z Calcd for C₉H₄D₇O₂ [M+H]⁺ 158.1186, found 158.1190.

Procedure for intermolecular kinetic isotope experiments between compounds 2 and 12: To a 20 mL scintillation vial were added, 2 (15.0 mg, 0.1 mmol), 12 (15.7 mg, 0.1 mmol), and CDCl₃ (0.5 mL). The solution was stirred until homogenous, and a small aliquot was taken for ¹H NMR analysis to ensure that the weighed quantities corresponded to a mixture with $50\% \pm 2\%$ of each component. The solvent was removed in vacuo, and the mixture of 2 and 12 was transferred in t-AmylOH (1 mL) to a 50 mL Schlenk-type sealed tube (with a Teflon high pressure valve and side arm) equipped with a magnetic stir bar. Pd(OAc)₂ (2.2 mg, 0.01 mmol), KHCO₃ (40.0 mg, 0.4 mmol), ligand (0.02 mmol), and ethyl acrylate (43 µL, 0.4 mmol) were then added. The reaction tube was capped, then evacuated briefly under high vacuum and charged with O₂ (1 atm, balloon) (×3). The reaction mixture was stirred at room temperature for 5 min, then at 90 °C for the appropriate time. The reaction vessel was then cooled to 0 °C in an ice bath. A 2.0 N HCl solution (5 mL) and diethyl ether (10 mL) were added. A small aliquot of the organic phase was taken, concentrated in vacuo, and analyzed by ¹H NMR. The conversion of 2a, X_{2a} , was determined by integration of the methyl proton signals, which appear as singlets (approximately 2.32 ppm for 2 and 2.36 ppm for 2a). The total conversion X_{total} was determined by integration of the benzylic methylene group signals, which also appear as singlets (3.67 ppm for 2/12 and 3.84 ppm for 2a/12a). The conversion of 12a, X_{12a} , could then be determined from the following formula:

$$X_{12a} = 2 \cdot X_{total} - X_{2a}$$

The experiments were repeated three times without ligand, three times with Boc-Val-OH, and five times with Ac-Ile-OH. The results are shown in Table S13.

Table S13: Results from the kinetic isotope experiments with 2 and 12.^a

Entry ^a	Ligand	Time (min)	% Conv. 2a	% Conv. 12a	k _H ([M]/min)	k _D ([M]/min)	k _H / k _D
1	_	120	42.5	7.9	3.5 x 10 ⁻⁴	6.6 x 10 ⁻⁵	5.38
2	_	120	26.1	5.1	2.2 x 10 ⁻⁴	4.3 x 10 ⁻⁵	5.12
3	_	120	29.1	3.8	2.4 x 10 ⁻⁴	3.2 x 10 ⁻⁵	7.67
4	Boc-Val-OH	10	23.7	3.7	2.4 x 10 ⁻³	3.7 x 10 ⁻⁴	6.41
5	Boc-Val-OH	10	18.3	3.7	1.8 x 10 ⁻³	3.7 x 10 ⁻⁴	4.90
6	Boc-Val-OH	10	18.0	3.4	1.8 x 10 ⁻³	3.4 x 10 ⁻⁴	5.29
7	Ac-IIe-OH	10	26.0	22.0	2.6 x 10 ⁻³	2.2 x 10 ⁻³	1.18
8	Ac-lle-OH	10	26.4	11.0	2.6 x 10 ⁻³	1.1 x 10 ⁻³	2.40
9	Ac-lle-OH	10	26.1	19.5	2.6 x 10 ⁻³	2.0 x 10 ⁻³	1.34
10	Ac-IIe-OH	10	23.5	11.1	2.4 x 10 ⁻³	1.1 x 10 ^{−3}	2.11
11	Ac-lle-OH	10	21.4	18.2	2.1 x 10 ⁻³	1.8 x 10 ⁻³	1.43

^a The conversion was determined by ¹H NMR analysis of the crude reaction mixture, as discussed in the preceding section.

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