

Practical Access to Metallo-Thiophenes: Regioselective Synthesis of 2,4-Disubstituted Thiophenes

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Supporting Information

General Methods

All solvents and reagents were obtained from commercial suppliers and used without purification. Reactions were conducted under an atmosphere of nitrogen with a suitable outlet to accommodate modest pressure changes. Reaction temperatures were monitored by internal thermocouple. Reaction progress and compound purity were determined by HPLC analysis, using an Eclipse XDB C8, 4.6 x 150 mm, 5 μ m column, with a gradient method using 0.1% (v/v) 70% HClO₄/water and acetonitrile as mobile phase. Assay yield and purity were assessed using HPLC comparison to high purity reference standards, and confirmed by quantitative ¹H NMR in DMSO-*d*₆ (vs. internal standards benzyl benzoate, or *N*-benzyl benzamide). ¹H NMR spectra were obtained using a Bruker 400 MHz spectrometer; chemical shifts are reported in ppm using the solvent internal standard (CDCl₃: δ 7.27, DMSO-*d*₆: δ 2.50, CD₃OD: δ 3.31). ¹³C NMR spectra were recorded on a 100 MHz spectrometer with complete proton decoupling; chemical shifts are reported in ppm with the solvent as the internal reference (CDCl₃: δ 77.0, DMSO-*d*₆: δ 39.5). HRMS (ESI-TOF) spectra were obtained using Agilent 1100 systems. Liquid chromatography purification was performed on an ISCO Combiflash Companion apparatus; chromatographic purifications and isolated yields are unoptimized.

General Procedures:

Titration of isopropylmagnesium chloride:

Commercial isopropylmagnesium chloride is available as a 2 M solution in THF (Aldrich 230111). The quality of this material can be confirmed by titration¹ versus stoichiometric (\pm)-menthol (CAS [89-78-1]) as follows:

1. To a flask with stir bar, charge 1.000 g (6.399 mmol) of (\pm)-menthol, 15-20 mg of 1,10-phenanthroline, and 10.0mL (10 Vol) THF.
2. Purge the flask with nitrogen, and cool to 0 °C.
3. Charge isopropylmagnesium chloride solution in THF drop-wise, until a purple color persists for greater than one minute.
4. Calculate molarity of isopropylmagnesium chloride solution as follows:

$$M (i\text{-PrMgCl in THF}) = \frac{\text{weight of } (\pm)\text{-menthol (in grams)}}{156.27 \text{ g/mol menthol}} \times \frac{1000 \text{ mL /L}}{\text{vol } i\text{-PrMgCl (in mL)}}$$

5. Acceptable molarity range for isopropylmagnesium chloride is determined to be 1.85 to 2.2 M solution, and amount used should be calculated accordingly.²

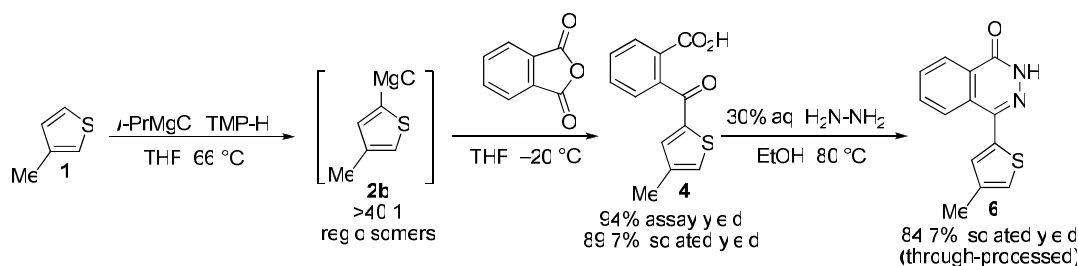
General method for magnesiation of heterocycles:

To a flask charge heterocycle (1.40 equiv) and THF (4 volumes, relative to heterocycle). Initiate stirring, purge the flask with nitrogen, and add 2,2,6,6-tetramethylpiperidine (0.10 equiv) in one portion at 20-25 °C. Add dropwise over ~10 min, *iso*-propylmagnesium chloride (2.00 M solution in THF, 1.00 equiv), at <30 °C. Heat the resulting clear solution to reflux (66 °C) for 18-24 h, until ¹H NMR analysis of a reaction aliquot quenched with CD₃OD indicates >90% deprotonation. The resulting magnesio-heterocycle is cooled to 20 °C prior to use.

¹ Lin, H.-S.; Paquette, L. A. *Synth. Commun.* **1994**, 24, 2503-2506.

² Commercial hexylmagnesium chloride is available as a 2 M solution in THF (Aldrich 64126). This reagent does not provide a well-defined endpoint according to the procedure in reference 1. The quality of this material can be confirmed by potentiometric titration versus stoichiometric 2-butanol. For details, see: Chen, Y.; Wang, T.; Helmy, R.; Zhou, G. X.; LoBrutto, R. *J. Pharm. Biomed. Anal.* **2002**, 29, 393-404.

Experimental:



2-(4-Methylthiophene-2-carbonyl)benzoic acid (4).³ 2,2,6,6-Tetramethylpiperidine (25.2 mL, 150 mmol) was charged in one portion to 3-methylthiophene **1** (147 g, 144 mL, 1500 mmol) in THF (576 mL). *iso*-Propylmagnesium chloride (2.00 M solution in THF, 633 mL, 1270 mmol) was added over 10 min at $<30\text{ }^{\circ}\text{C}$. The resulting solution was heated to reflux at $66\text{ }^{\circ}\text{C}$. After 23 h, ^1H NMR analysis of a reaction aliquot quenched with CD_3OD indicated 98%⁴ conversion to the Mg-thiophene **2b** (96.8% 2-D-4-methylthiophene, 1.2% 2-D-3-methylthiophene of theoretical 0.85 equiv). The Mg-thiophene solution was cooled to $20\text{ }^{\circ}\text{C}$. Phthalic anhydride **3** (170 g, 1150 mmol) in THF (720 mL) was charged to a separate flask and the resulting slurry was cooled to $-20\text{ }^{\circ}\text{C}$. The Mg-thiophene solution (at $20\text{ }^{\circ}\text{C}$) was added to the phthalic anhydride slurry over 45 min, at $-25\text{ }^{\circ}\text{C}$ to $-20\text{ }^{\circ}\text{C}$. After 20 min, the reaction was quenched with H_2O (510 mL) added over 10 min between $-20\text{ }^{\circ}\text{C}$ and $10\text{ }^{\circ}\text{C}$,⁵ followed by 6N HCl (289 mL) to pH 2. The reaction mixture was warmed to $20\text{ }^{\circ}\text{C}$, and MTBE (289 mL) was added. After 10 min, the layers were separated; the upper organic layer assayed to 267 g keto acid **4** (94.1%⁶).

The crude keto acid **4** was concentrated by distillation ($60\text{ }^{\circ}\text{C}$, 350 mbar) to 530-545 mL (2X vs. assay yield), and the resulting pot was maintained at $60\text{ }^{\circ}\text{C}$. Ethanol (1070 mL, 4X) was added, and the solution was distilled again to 530-545 mL total volume. HPLC analysis of the distillate revealed 3-

³ Weinmayr, V. *J. Am. Chem. Soc.* **1952**, 74, 4352-4357.

⁴ 98% conversion is calculated based on limiting reagent *i*-PrMgCl.

⁵ In a power compensation calorimetry experiment conducted at $10\text{ }^{\circ}\text{C}$ isothermal, quenching the reaction with water (4 X over 1 h, 60.6 mmol scale) resulted in a 74 kJ/mol exotherm, uncorrected for heat of mixing.

⁶ Yield is based on limiting reagent phthalic anhydride.

methylthiophene as the only UV-active component. After breaking vacuum, ethanol (800 mL, 3X) was charged and the flask was cooled to 20 °C over 3 h. Water (1330 mL, 5X) was added over 2 h, and then aged for 10 h. The slurry was filtered, and the cake displacement-washed with 25% EtOH:H₂O (535 mL, 2X). The collected mother liquors and wash contained 11.6 g (4.1%) product **4**. The solid was dried on the frit at 20 °C for >24 h, under a nitrogen stream, to provide keto acid **4** as an off-white solid (261 g, 97.3 wt%, 89.7% adjusted yield). ¹H NMR (DMSO-*d*₆, 400 MHz) δ 13.16 (br s, 1H), 7.96 (ddd, *J* = 0.5, 1.5, 7.5 Hz, 1H), 7.64-7.72 (m, 3H), 7.49 (ddq, *J* = 0.6, 1.5, 7.5 Hz, 1H), 7.09 (dq, *J* = 0.4, 1.5 Hz, 1H), 2.18 (dd, *J* = 0.4, 0.6 Hz, 3H); ¹³C NMR (DMSO-*d*₆, 400 MHz) δ 188.5, 166.9, 143.6, 140.7, 138.6, 136.1, 132.1, 130.7, 129.9, 129.9, 129.8, 127.4, 15.0; HRMS calculated for C₁₃H₉O₂S₁ [M + H – H₂O]⁺ 229.0318, found 229.0316; IR (neat): 3050, 2970, 2920, 1690 cm⁻¹; mp: 191 °C.

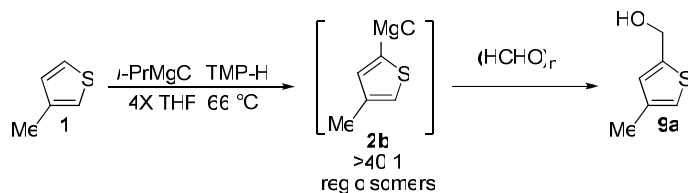
4-(4-Methylthiophene-2-yl)phthalazine-1(2H)-one (6).⁷ Crude keto acid **4** (91.9 g) was synthesized as described above. The crude MTBE/THF layer was concentrated by distillation (60 °C, 300 mbar) to 180-190 mL, and the resulting pot was maintained at 60 °C. Ethanol (367 mL, 4X) was added, and the solution was distilled again to 180-190 mL total volume. After breaking vacuum, ethanol (367 mL, 4X) was charged to the reaction and the flask was cooled to 20-30 °C. To the resulting solution was added hydrazine (35 wt % solution in H₂O, 169 mL, 1870 mmol) over 10 min, at 35 °C. The reaction was heated to 80 °C for 18 h, until HPLC assay of the resulting slurry indicated >95% conversion to product.⁸

The reaction was cooled to 20 °C over 2 h, and then aged at 20 °C for 1 h. The resulting slurry was filtered, and the cake was displacement-washed with 1:1 EtOH:H₂O (180 mL). The cake was dried

⁷ Iwase, N.; Morinaka, Y.; Tamao, Y.; Kanayama, T.; Yamada, K. 3,6-Disubstituted Pyridazine Derivative Blood Platelet Aggregation Inhibitors. Eur. Pat. Appl. EP 534443 19920924, Mar. 31, 1993; *Chem. Abstr.* **1993**, 119, 249963.

⁸ This reaction proceeds below the flash point of 35% hydrazine/water (112.7 °C), and below the boiling point of a hydrazine/water azeotrope (120.3 °C). DSC and ARC scanning of the reaction components at 300 °C and 250 °C, respectively, showed no unsafe thermodynamic events.

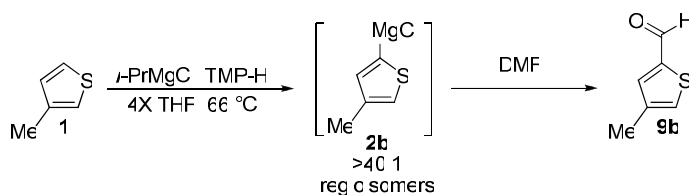
under N₂ stream at 20 °C to give 82.7 g thiophene-phthalazinone **6** as a pale yellow solid (98.6 wt %, 84.7% adjusted yield over two steps). The collected mother liquors and wash contained 1.78 g product **6** (1.9%) and 5.70 g keto acid **7** (6.1%). ¹H NMR (CDCl₃, 400 MHz) δ 2.38 (d, *J* = 0.8, 3H), 7.09 (dq, *J* = 0.8, 1.1, 1H), 7.28 (d, *J* = 1.1, 1H), 7.86 (m, 2H), 8.17 (m, 1H), 8.53 (m, 1H), 10.31 (bs, 1H); ¹³C NMR (CDCl₃, 100 MHz) δ 15.83, 122.91, 126.63, 127.08, 128.22, 129.38, 130.88, 131.71, 133.70, 136.33, 138.14, 142.43, 159.72; mp: 232 °C.



(4-Methylthiophen-2-yl)methanol (**9a**).⁹ Magnesiothiophene **2b** was generated according to the general magnesiation procedure (20.0 mmol scale), cooled to 20 °C, and then added over 30 min to a 0 °C solution of *p*-formaldehyde (0.840 g, 1.40 equiv) in THF (13.8 mL, 5 volumes, relative to heterocycle). The resulting mixture was stirred at 0 °C, until HPLC analysis indicated >95% conversion of the thiophene-Grignard (90 min). The reaction mixture was quenched with water, and then 6N HCl was added until the reaction mixture was slightly acidic (pH 5). MTBE was added to provide a clean phase cut. The layers were separated, and the aqueous layer was washed once with MTBE. HPLC analysis of the combined crude organic layers revealed 2.28 grams (88.9%) assay yield of **9a**. Purification by flash chromatography (ISCO Companion, MTBE/hexanes gradient) and careful concentration *in vacuo* provided the desired compound **9a** as a pale yellow amorphous solid (1.78 g, 82.5 wt%, 57% adjusted isolated yield). ¹H NMR (CDCl₃, 400 MHz) δ 2.24 (s, 3H), 2.39 (s, exchangeable, 1H), 4.73 (app. s, 2H), 6.81 (s, 1H), 6.84 (s, 1H); ¹³C NMR (CDCl₃, 100 MHz) δ 15.6,

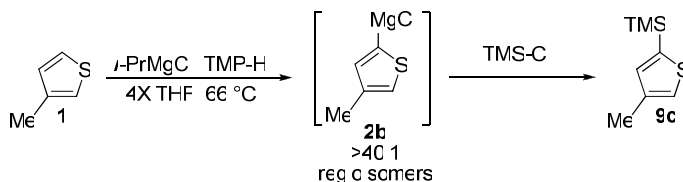
⁹ (a) Lozanova, A. V.; Moiseenkov, A. M.; Semenovskii, A. V. *Izv. Akad. Nauk, Ser. Khim.* **1980**, 958-959. (b) Wang, C.-C.; Chen, H.-C.; Wang, S.-H.; Lin, M.-C.; Shieh, T.-L.; Huang, Y.-H.; Chuang, S.-C.; King, C.-H. R. Preparation of 4-oxoquinazoline derivatives as kinesin inhibitors. U.S. Pat. Appl. US 2008-125094, Nov. 27, 2008; *Chem. Abstr.* **2008**, 150, 5760.

59.9, 120.5, 127.7, 137.4, 143.8; HRMS calculated for $C_6H_7O_1S_1Na_1$ $[M+Na]$ 150.01153, found 150.01150; IR (neat): 3645, 2925, 1020 cm^{-1} .

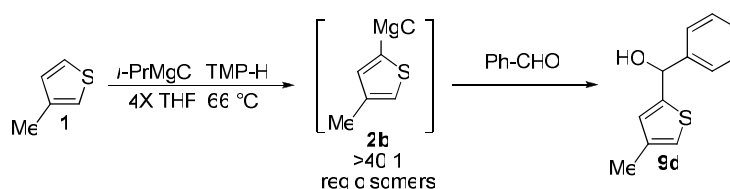


4-Methylthiophene-2-carbaldehyde (9b).¹⁰ Magnesiothiophene **2b** was generated according to the general magnesiation procedure (20.0 mmol scale), cooled to 20 °C, and then added over 30 min to a 0 °C solution of *N,N*-dimethylformamide (1.26 mL, 1.40 equiv) in THF (13.8 mL, 5 volumes, relative to heterocycle). The resulting mixture was stirred at 0 °C, until HPLC analysis indicated >95% conversion of the thiophene-Grignard (15 min). The reaction mixture was quenched with water, and then 6N HCl was added until the reaction mixture was slightly acidic (pH 4-5). MTBE was added to provide a clean phase cut. The layers were separated, and the aqueous layer was washed once with MTBE. HPLC analysis of the combined crude organic layers revealed 2.20 grams (87.2%) assay yield of **9b**. Purification by flash chromatography (ISCO Companion, MTBE/hexanes gradient) and careful concentration *in vacuo* at 0 °C provided the desired compound **9b** as a volatile pale yellow oil (2.05 g, 74.0 wt%, 60.1% adjusted isolated yield). ¹H NMR ($CDCl_3$, 400 MHz) δ 2.33 (s, 3H), 7.37 (s, 1H), 7.58 (s, 1H), 9.88 (s, 1H); ¹³C NMR ($CDCl_3$, 100 MHz) δ 182.9, 143.6, 139.1, 137.9, 131.0, 15.4; HRMS calculated for $C_6H_7O_1S_1$ $[M+H]$ 127.02121, found 127.02101; IR (neat): 2925, 2865, 1765, 1386, 1020 cm^{-1} .

¹⁰ (a) Jean, S. *J. Org. Chem.* **1954**, *19*, 70-3. (b) Smith, K.; Barrat, M. L. *J. Org. Chem.* **2007**, *72*, 1031-1034.



Trimethyl(4-methylthiophen-2-yl)silane (9c).¹¹ Magnesiothiophene **2b** was generated according to the general magnesiation procedure (20.0 mmol scale), cooled to 20 °C, and then added over 30 min to a 0 °C solution of chlorotrimethylsilane (3.55 mL, 1.40 equiv) in THF (13.8 mL, 5 volumes, relative to heterocycle). The resulting mixture was stirred at 0 °C, until HPLC analysis indicated >95% conversion of the thiophene-Grignard (1 h). The reaction mixture was quenched with water, and then 6N HCl was added until the reaction mixture was neutral (pH 7). MTBE was added to provide a clean phase cut. The layers were separated, and the aqueous layer was washed once with MTBE. HPLC analysis of the combined crude organic layers revealed 2.92 grams (85.7%) assay yield of **9c**. Purification by flash chromatography (ISCO Companion, MTBE/hexanes gradient) and careful concentration *in vacuo* at 0 °C provided the desired compound **9c** as a volatile colorless oil (2.14 g, 96.1 wt%, 60.4% adjusted isolated yield). ¹H NMR (CDCl₃, 400 MHz) δ 0.35 (s, 9H), 2.34 (2, 3H), 7.09 (s, 1H), 7.18 (s, 1H); ¹³C NMR (CDCl₃, 100 MHz) δ 140.3, 138.9, 136.5, 126.0, 15.0, -0.1; HRMS calculated for C₈H₁₆S₁Si₁ [M+H] 171.06637, found 171.06602; IR (neat): 2925, 1675, 1438, 1274 cm⁻¹.

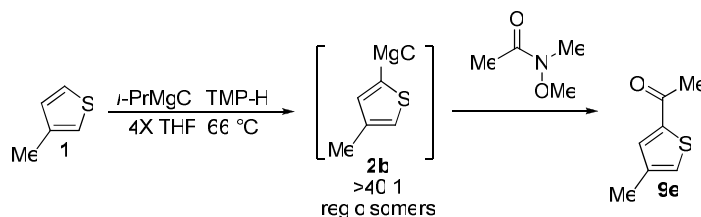


(4-Methylthiophen-2-yl)(phenyl)methanol (9d).¹² Magnesiothiophene **2b** was generated according to the general magnesiation procedure (20.0 mmol scale), cooled to 20 °C, and then added over 30 min to a 0 °C solution of benzaldehyde (2.85 mL, 1.40 equiv) in THF (13.8 mL, 5 volumes, relative to

¹¹ Albertin, L.; Bertarelli, C.; Gallazzi, M. C.; Meille, S. V.; Capelli, S. C. *J. Chem. Soc., Perkin Trans. 2*, **2002**, 1752-1759.

¹² Agarwal, N.; Ravikanth, M. *Tetrahedron*, **2004**, 60, 4739-4747. See also ref. 10b.

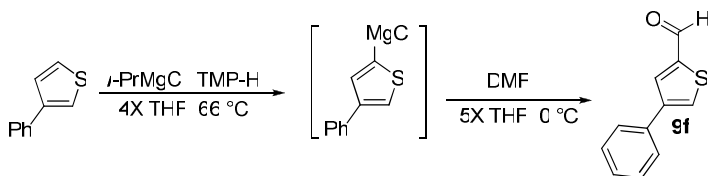
heterocycle). The resulting mixture was stirred at 0 °C, until HPLC analysis indicated >95% conversion of the thiophene-Grignard (18 h). The reaction mixture was quenched with water, and then 6N HCl was added until the reaction mixture was slightly acidic (pH 4). MTBE was added to provide a clean phase cut. The layers were separated, and the aqueous layer was washed once with MTBE. HPLC analysis of the combined crude organic layers revealed 3.73 grams (91.2%) assay yield of **9d**. Purification by flash chromatography (ISCO Companion, MTBE/hexanes gradient) and concentration *in vacuo* provided the desired compound **9d** as an off-white amorphous solid (2.88 g, 86.7 wt%, 61.1% adjusted isolated yield). ¹H NMR (CDCl₃, 400 MHz) δ 2.21 (s, 3H), 2.46 (broad s, 1H), 5.99 (s, 1H), 6.70 (s, 1H), 6.85 (s, 1H), 7.30-7.41 (m, 3H), 7.44-7.48 (m, 2H); ¹³C NMR (CDCl₃, 100 MHz) δ 15.8, 72.5, 120.6, 126.3, 127.0, 128.0, 128.6, 137.4, 143.2, 127.9; HRMS calculated for C₁₂H₁₂S₁O₁Na₁ [M+Na] 227.05011, found 227.05050; IR (neat): 3030, 2984, 1690, 1295, 1180 cm⁻¹.



1-(4-Methylthiophen-2-yl)ethanone (9e).¹³ Magnesiothiophene **2b** was generated according to the general magnesiation procedure (20.0 mmol scale), cooled to 20 °C, and then added over 30 min to a 0 °C solution of *N*-methoxy-*N*-methylacetamide (2.98 mL, 1.40 equiv) in THF (13.8 mL, 5 volumes, relative to heterocycle). The resulting mixture was stirred at 0 °C, until HPLC analysis indicated >95% conversion of the thiophene-Grignard (18 h). The reaction mixture was quenched with water, and then 6N HCl was added until the reaction mixture was slightly acidic (pH 4-5). MTBE was added to provide a clean phase cut. The layers were separated, and the aqueous layer was washed once with MTBE. HPLC analysis of the combined crude organic layers revealed 2.45 grams (87.4%) assay yield of **9e**. Purification by flash chromatography (ISCO Companion, MTBE/hexanes gradient) and careful

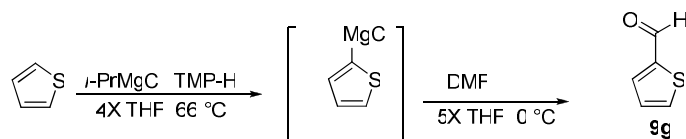
¹³ (a) Hartough, H. D.; Kosak, A. I. *J. Am. Chem. Soc.* **1947**, 69, 3093-3096. (b) Armstrong, A.; Pullin, R.D. C.; Jenner, C. R.; Scutt, J. N. *J. Org. Chem.* **2010**, 75, 3499-3502.

concentration *in vacuo* at 0 °C provided the desired compound **9e** as a volatile yellow oil (2.10 g, 91.8 wt%, 68.8% adjusted isolated yield). ¹H NMR (CDCl₃, 400 MHz) δ 2.26 (s, 3H), 2.49 (s, 3H), 4.73 (app. s, 2H), 7.19 (s, 1H), 7.47 (s, 1H); ¹³C NMR (CDCl₃, 100 MHz) δ 15.4, 26.6, 129.4, 134.3, 138.7, 143.9, 190.5; HRMS calculated for C₇H₈O₁S₁ [M+H] 141.03686; found 141.03690; IR (neat): 3030, 2984, 1820 cm⁻¹.

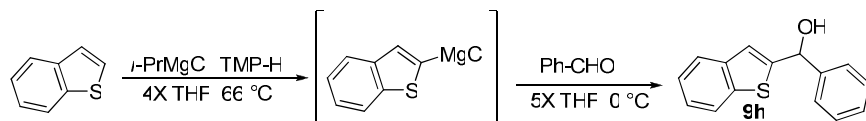


4-Phenylthiophene-2-carbaldehyde (9f).¹⁴ Magnesiothiophene **8a** was generated according to the general magnesiation procedure (20.0 mmol scale, 18 h deprotonation), cooled to 20 °C, and then added over 30 min to a 0 °C solution of *N,N*-dimethylformamide (1.26 mL, 1.40 equiv) in THF (13.8 mL, 5 volumes). The resulting mixture was stirred at 0 °C, until HPLC analysis indicated >95% conversion of the thiophene-Grignard (30 min). The reaction mixture was quenched with water, and then 6N HCl was added until the reaction mixture was slightly acidic (pH 6). MTBE was added to provide a clean phase cut. The layers were separated, and the aqueous layer was washed once with MTBE. HPLC analysis of the combined crude organic layers revealed 3.32 grams (88.3%) assay yield of **9f**. Purification by flash chromatography (ISCO Companion, MTBE/hexanes gradient) and careful concentration *in vacuo* at 0 °C provided the desired compound **9f** as a white amorphous solid (2.07 g, 99.0 wt%, 54.4% adjusted isolated yield). ¹H NMR (CDCl₃, 400 MHz) δ 7.37 (m, 1H), 7.46 (m, 2H), 7.61 (m, 2H), 7.87 (m, 1H), 8.05 (m, 1H), 9.99 (s, 1H); ¹³C NMR (CDCl₃, 100 MHz) δ 126.3, 128.0, 129.1, 129.6, 134.4, 134.7, 143.7, 144.4, 182.9; HRMS calculated for C₁₁H₈O₁S₁ [M+H] 189.03686, found 189.03723; HRMS calculated for C₁₁H₇O₁S₁Na [M+Na] 211.01881, found 211.01881; IR (neat): 3030, 2984, 2848, 1780, 1390 cm⁻¹.

¹⁴ Johnson, A. L. *J. Org. Chem.* **1976**, *41*, 1320-1324.



Thiophene-2-carbaldehyde (9g).¹⁵ Magnesiothiophene **8b** was generated according to the general magnesiation procedure (20.0 mmol scale, 20 h deprotonation), cooled to 20 °C, and then added over 30 min to a 0 °C solution of *N,N*-dimethylformamide (1.26 mL, 1.40 equiv) in THF (13.8 mL, 5 volumes). The resulting mixture was stirred at 0 °C, until HPLC analysis indicated >95% conversion of the thiophene-Grignard (10 min). The reaction mixture was quenched with water, and then 6N HCl was added until the reaction mixture was slightly acidic (pH 6). MTBE was added to provide a clean phase cut. The layers were separated, and the aqueous layer was washed once with MTBE. HPLC analysis of the combined crude organic layers revealed 2.07 grams (92.3%) assay yield of **9g**. Purification by flash chromatography (ISCO Companion, MTBE/hexanes gradient) and careful concentration *in vacuo* at 0 °C provided the desired compound **9g** as a colorless volatile oil (1.20 g, 99.0 wt%, 53.0% adjusted isolated yield). ¹H NMR (CDCl₃, 400 MHz) δ 7.24 (m, 1H), 7.79 (m, 2H), 9.96 (s, 1H); ¹³C NMR (CDCl₃, 100 MHz) δ 182.6, 143.5, 136.2, 134.8, 128.0; HRMS calculated for C₅H₅O₁S₁ [M+H] 113.00556, found 113.00587; IR (neat): 2970, 1750, 1420 cm⁻¹.

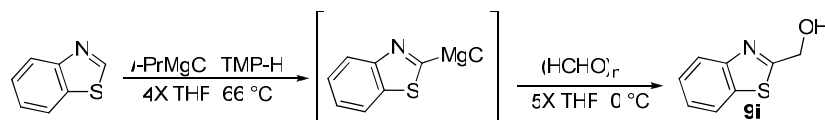


Benzo[b]thiophen-2-yl(phenyl)methanol (9h).¹⁶ Magnesiothiophene **8c** was generated according to the general magnesiation procedure (20.0 mmol scale, 24 h deprotonation), cooled to 20 °C, and then added over 30 min to a 0 °C solution of benzaldehyde (2.85 mL, 1.40 equiv) in THF (13.8 mL, 5 volumes). The resulting mixture was stirred at 0 °C, until HPLC analysis indicated >95% conversion of

¹⁵ CAS No [98-03-3]

¹⁶ (a) Shirley, D. A.; Cameron, M. D. *J. Am. Chem. Soc.* **1952**, 74, 664-665. (b) Rohbogner, C. J.; Wunderlich, S. H.; Clososki, G. C.; Knochel, P. *Eur. J. Org. Chem.* **2009**, 1781-1795.

the thiophene-Grignard (16 h). The reaction mixture was quenched with water, and then 6N HCl was added until the reaction mixture was slightly acidic (pH 6). MTBE was added to provide a clean phase cut. The layers were separated, and the aqueous layer was washed once with MTBE. HPLC analysis of the combined crude organic layers revealed 3.90 grams (81.1%) assay yield of **9h**. Purification by flash chromatography (ISCO Companion, MTBE/hexanes gradient) and concentration *in vacuo* provided the desired compound **9g** as a white solid (3.30 g, 93.8 wt%, 64.4% adjusted isolated yield). ¹H NMR (CDCl₃, 400 MHz) δ 2.73 (s, exchangeable, 1H), 6.11 (m, 1H), 7.13 (m, 1H), 7.29-7.44 (m, 5H), 7.51 (m, 2H), 7.71 (m, 1H), 7.81 (m, 1H); ¹³C NMR (CDCl₃, 100 MHz) δ 148.7, 142.6, 139.8, 139.5, 128.7, 128.6, 126.5, 124.3, 124.2, 123.7, 122.7, 121.3, 73.1; HRMS calculated for C₁₅H₁₂O₁S₁Na [M+Na] 263.05011, found 263.05045; IR (neat): 3020, 2984, 2640, 1690, 1270, 1100 cm⁻¹.

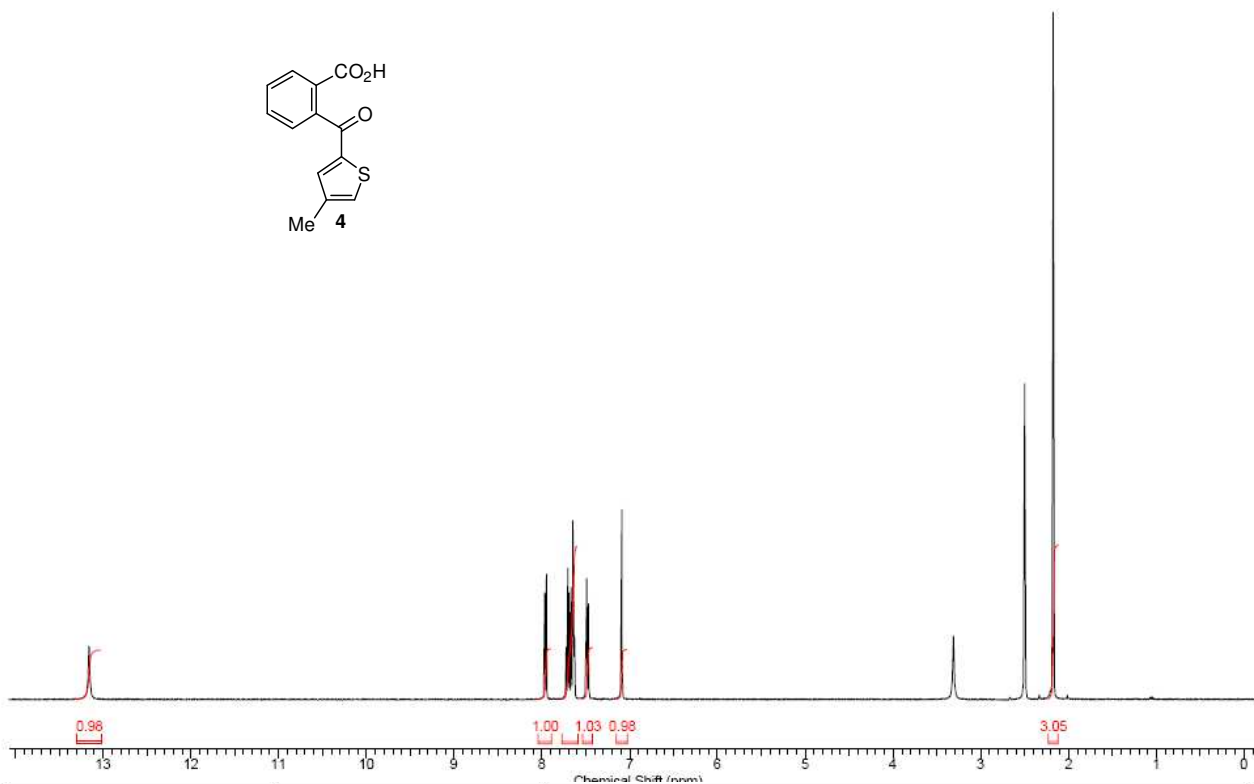
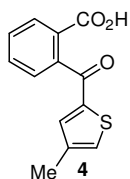


Benzo[d]thiazol-2-ylmethanol (9i).¹⁷ Magnesiothiophene **8c** was generated according to the general magnesiation procedure (20.0 mmol scale, 30 h deprotonation), cooled to 20 °C, and then added over 30 min to a 0 °C solution of benzaldehyde (2.85 mL, 1.40 equiv) in THF (13.8 mL, 5 volumes). The resulting mixture was stirred at 0 °C, until HPLC analysis indicated >95% conversion of the thiazole-Grignard (24 h). The reaction mixture was quenched with water, and then 6N HCl was added until the reaction mixture was acidic (pH 4). MTBE was added to provide a clean phase cut. The layers were separated, and the aqueous layer was washed once with MTBE. HPLC analysis of the combined crude organic layers revealed 2.36 grams (71.4%) assay yield of **9i**. Purification by flash chromatography (ISCO Companion, MTBE/hexanes gradient) and concentration *in vacuo* provided the desired compound **9i** as a yellow oil (2.27 g, 82.5 wt%, 56.6% adjusted isolated yield). ¹H NMR (CDCl₃, 400 MHz) δ 3.10 (t, *J* = 6.1 Hz, 1H), 5.04 (d, *J* = 6.1 Hz, 2H), 7.35 (m, 2H), 7.84 (m, 2H); ¹³C NMR (CDCl₃,

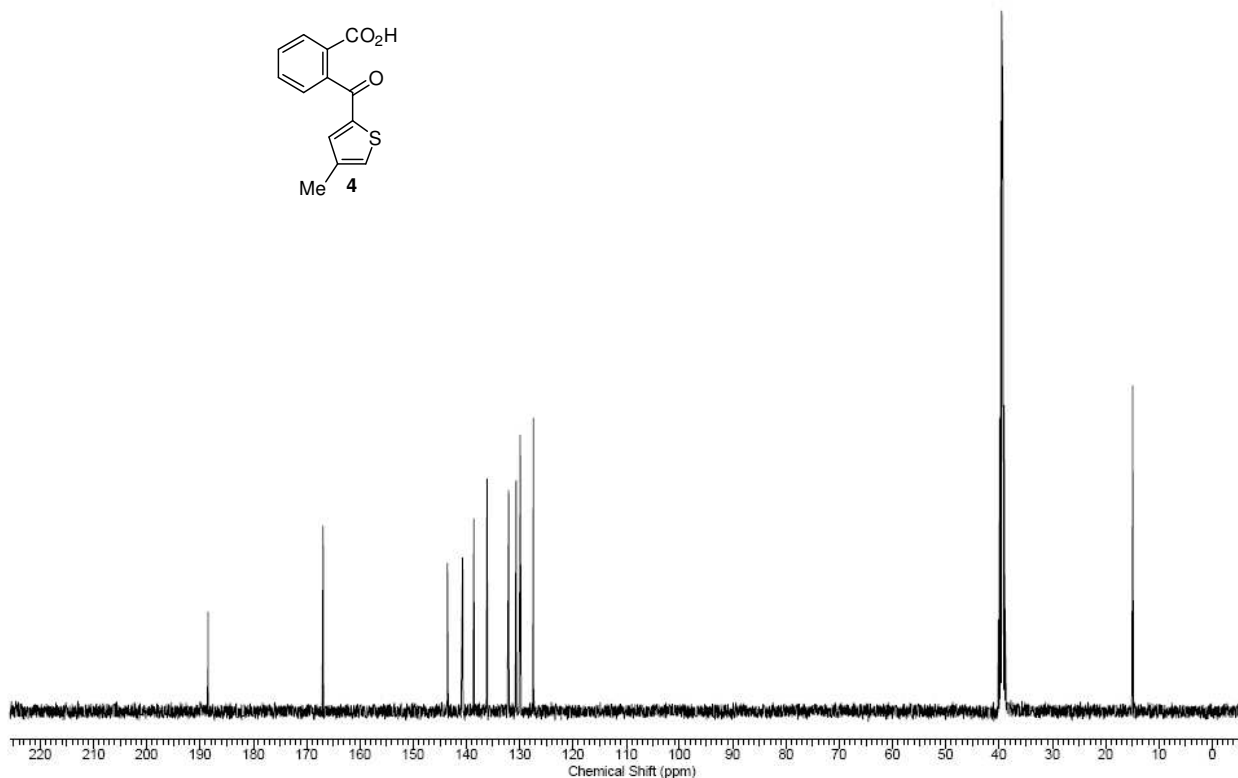
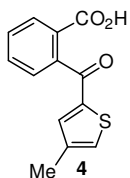
100 MHz) δ 170.2, 153.0, 136.4, 124.6, 124.5, 123.4, 122.4, 59.9. HRMS calculated for $C_8H_7N_1O_1S_1Na$ [M+Na] 188.01460. Found 188.01400; mp 102 °C; IR (neat): 3190, 3065, 2988, 2940, 2340, 1050 cm^{-1} .

¹⁷ (a) Courtot, C.; Tchelitcheff, S. *Compt. Rend.* **1943**, 217, 201-3. (b) Jeffreys, R. A. *J. Chem. Soc.* **1954**, 503-505.

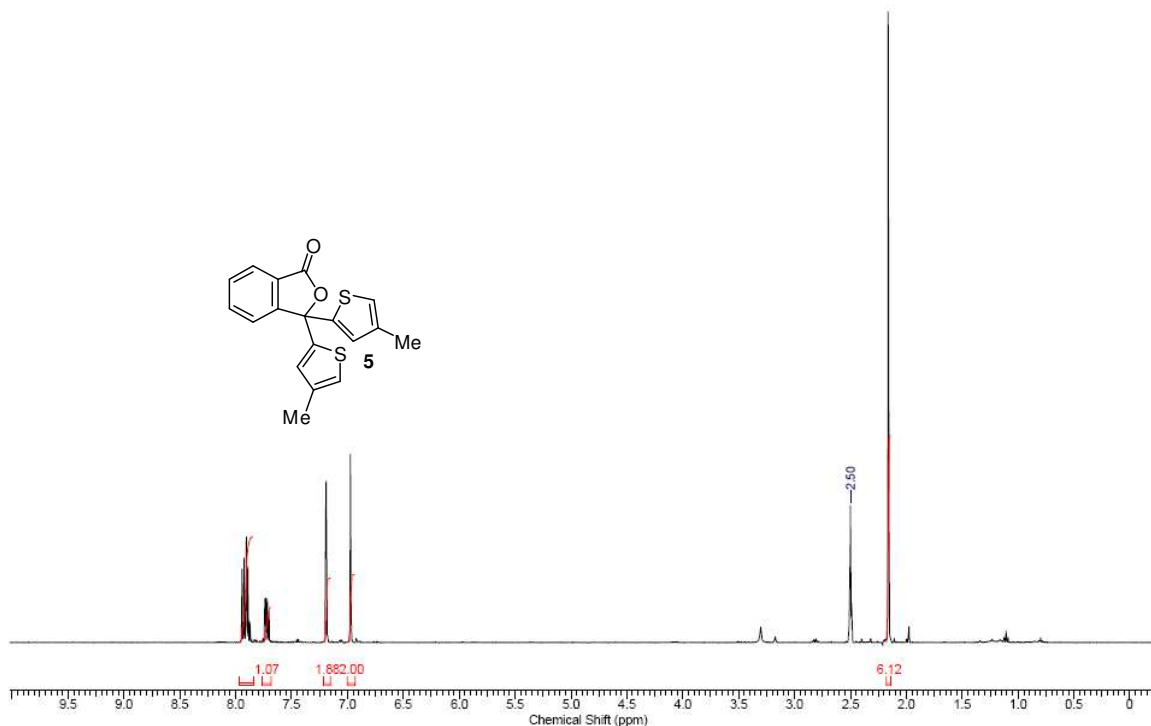
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Nucleus	1H	Number of Transients	16	Origin	Bruker BioSpin GmbH		
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SW(cyclical) (Hz)	6410.16	Solvent	DMSO-d6	Spectrum Offset (Hz)	2798.0542	Sweep Width (Hz)	6410.06
Temperature (degree C)	28.560						



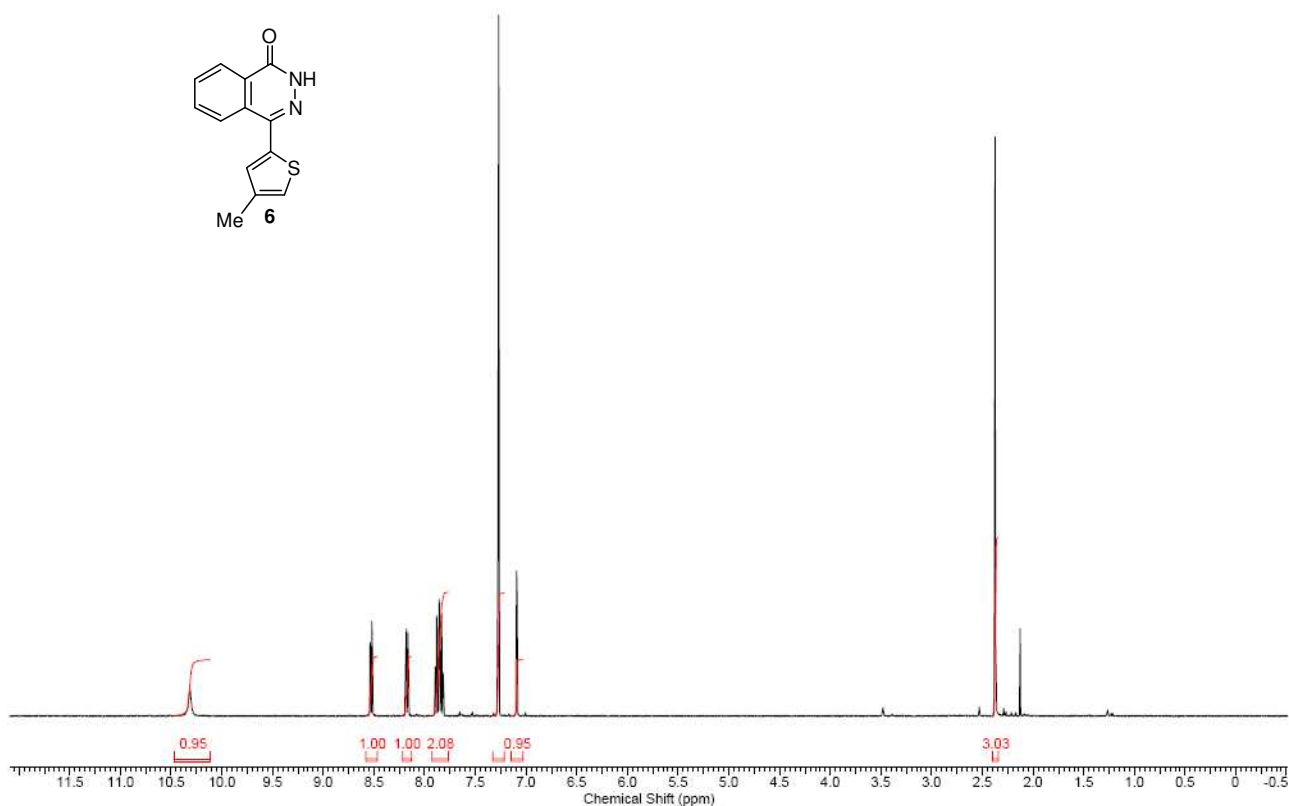
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Nucleus		13C		Number of Transients		256		Origin		Bruker BioSpin GmbH			
Original Points Count		65536		Owner		shr-usam-cc10046				Points Count		65536	
SW(cyclical) (Hz)		28408.66		Solvent		DMSO-d6		Spectrum Offset (Hz)		12015.6895		Sweep Width (Hz)	28408.22
Temperature (degree C)		28.760											



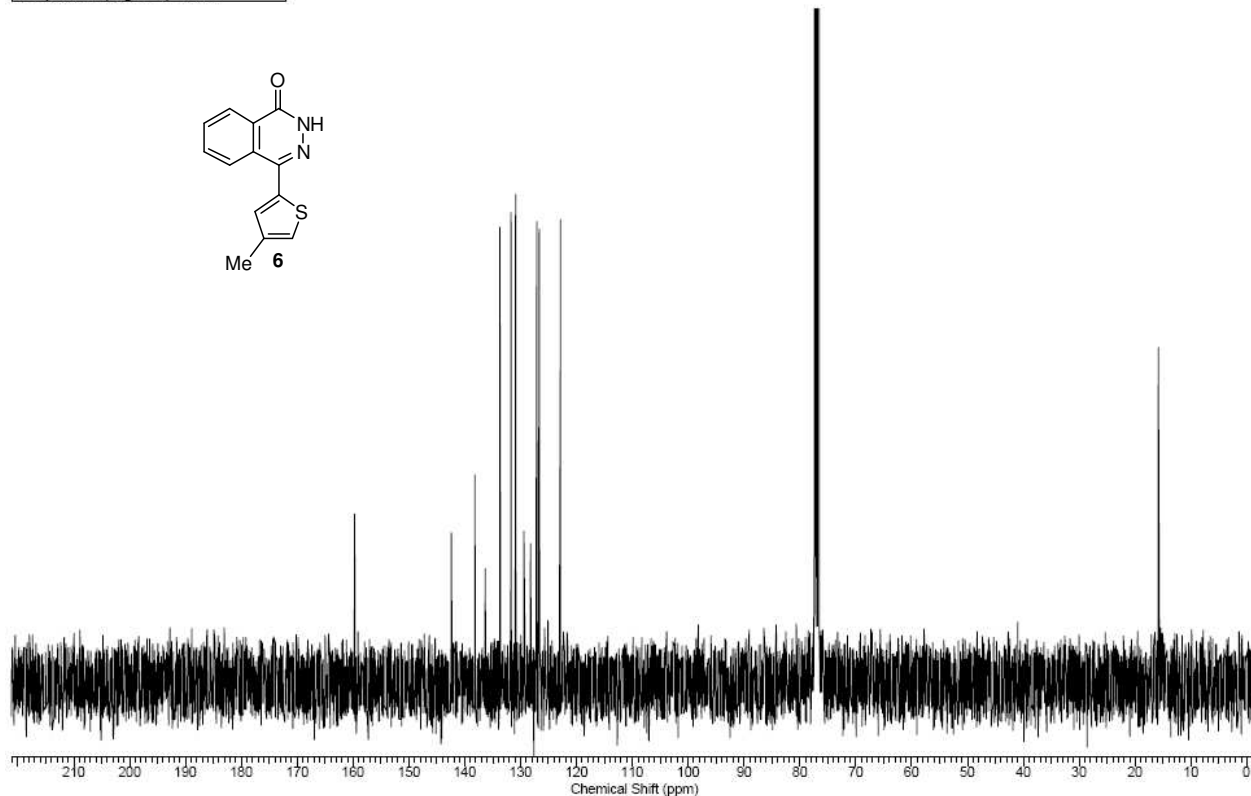
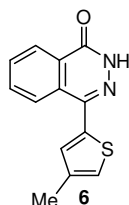
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Nucleus	¹ H	Number of Transients	4	Origin	Bruker BioSpin GmbH
Original Points Count	65536	Owner	shr-usam-cc10046	Points Count	65536
SW(cyclical) (Hz)	6410.16	Solvent	DMSO-d6	Spectrum Offset (Hz)	2495.3164
Temperature (degree C)	27.000			Sweep Width (Hz)	6410.06



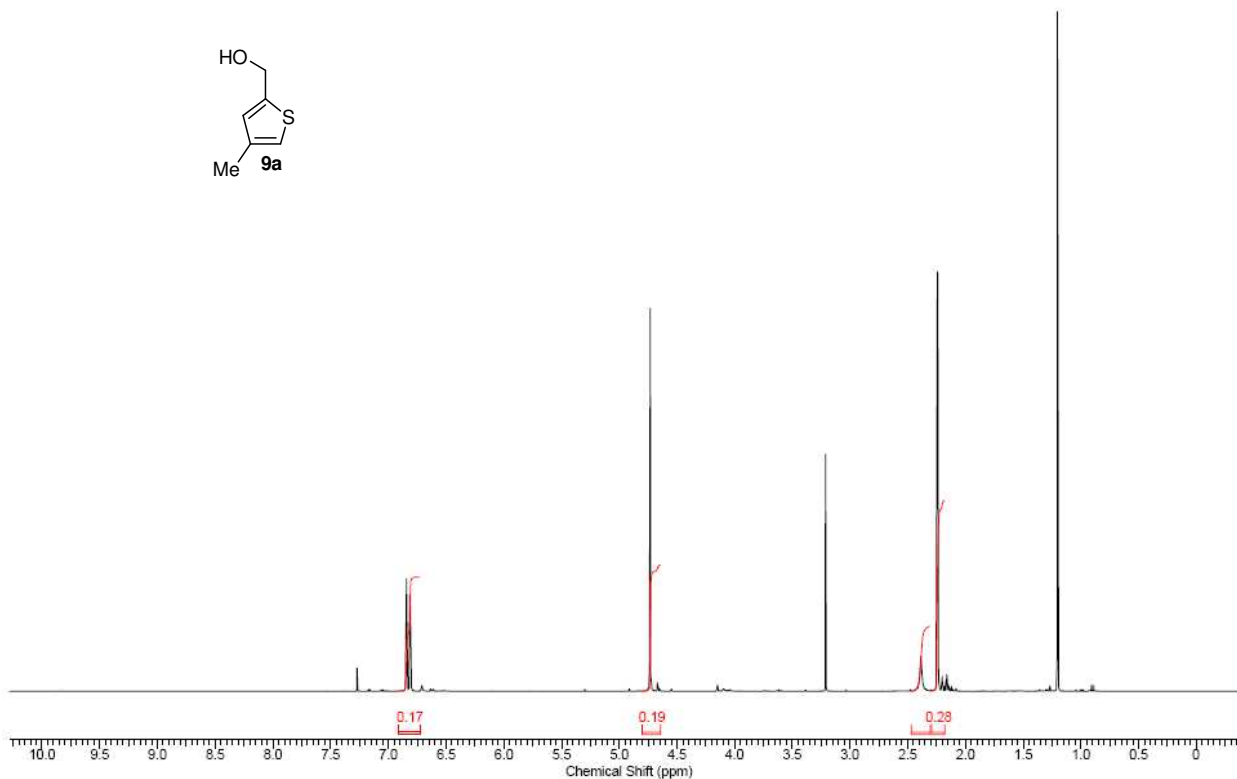
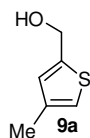
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Nucleus	¹ H	Number of Transients	16	Origin	Bruker BioSpin GmbH
Original Points Count	65536	Owner	smallmolecules	Points Count	65536
Solvent	CHLOROFORM-d			SW(cyclical) (Hz)	6410.16
Temperature (degree C)	27.000			Spectrum Offset (Hz)	2493.0339
				Sweep Width (Hz)	6410.06



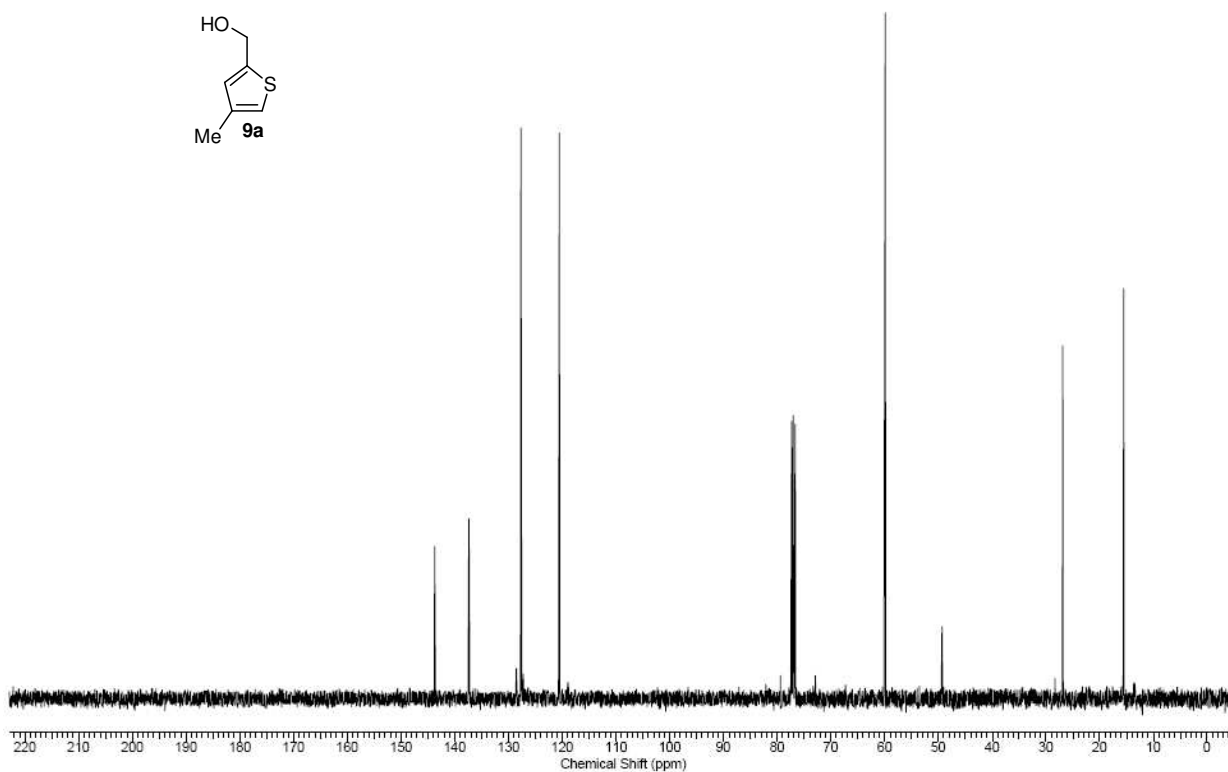
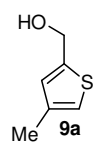
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Nucleus	13C	Number of Transients	1024	Origin	Bruker BioSpin GmbH
Original Points Count	65536	Owner	smallmolecules	Points Count	65536
Solvent	CHLOROFORM-d			SW(cyclical) (Hz)	28408.66
Temperature (degree C)	27.000			Spectrum Offset (Hz)	12066.2695
				Sweep Width (Hz)	28408.22



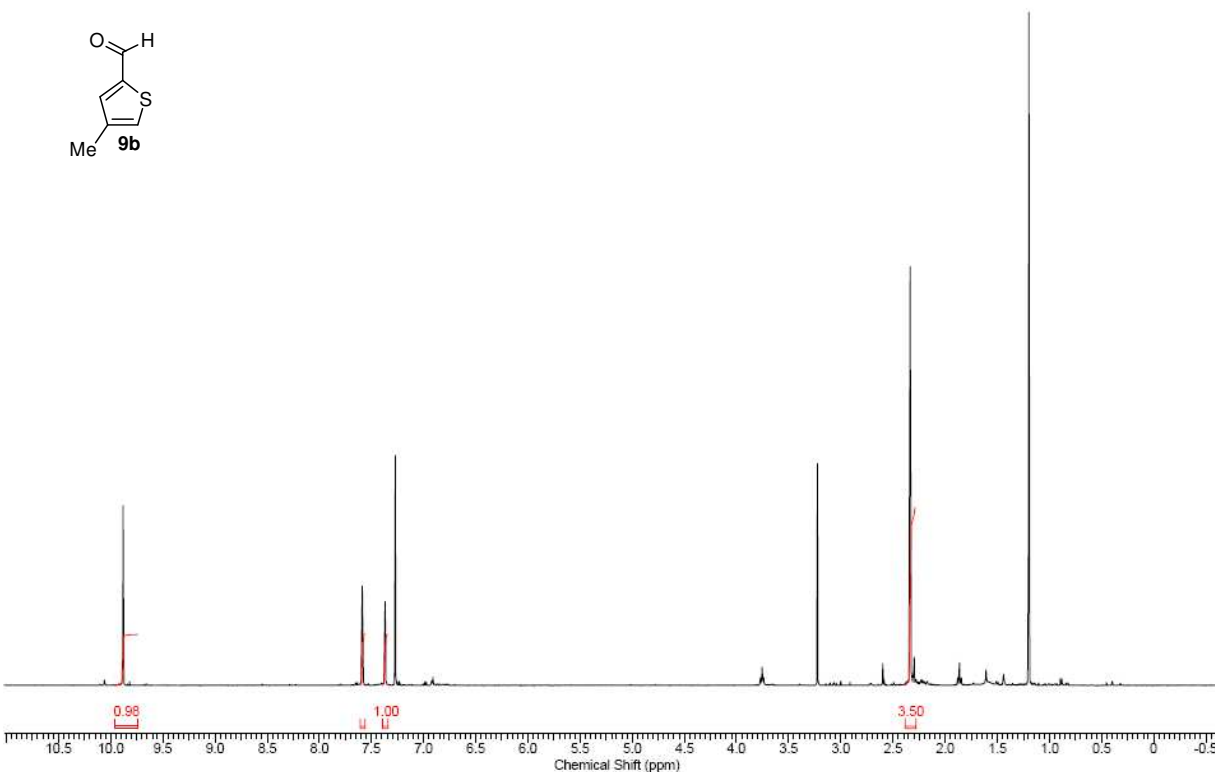
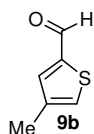
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Nucleus	1H	Number of Transients	16	Origin	Bruker BioSpin GmbH
Original Points Count	65536	Owner	shr-usam-cc10046	Points Count	65536
SW(cyclical) (Hz)	6410.16	Solvent	CHLOROFORM-d	Spectrum Offset (Hz)	2493.1318
Sweep Width (Hz)	6410.06	Temperature (degree C)	27.000		



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Nucleus	¹³ C	Number of Transients	128	Origin	Bruker BioSpin GmbH
Original Points Count	65536	Owner	shr-usam-cc10046	Points Count	65536
SW(cyclical) (Hz)	28408.66	Solvent	CHLOROFORM-d	Spectrum Offset (Hz)	12057.1670
Sweep Width (Hz)	28408.22	Temperature (degree C)	27.000		

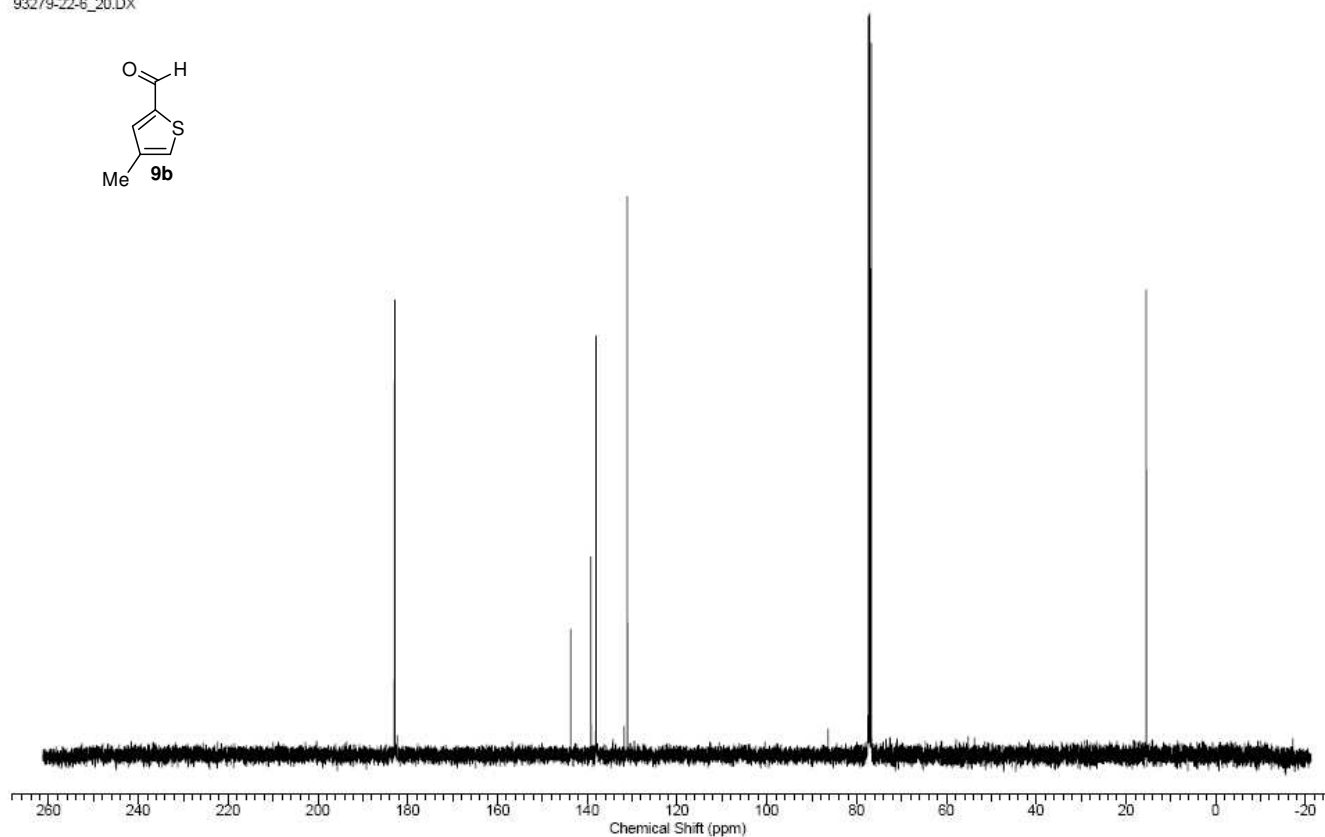
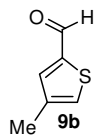


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Original Points Count	65536	Owner	shr-usam-cc10046	Points Count	65536
SW(cyclical) (Hz)	6410.16	Solvent	CHLOROFORM-d	Spectrum Offset (Hz)	2492.8384
Sweep Width (Hz)	6410.06	Temperature (degree C)	27.000		

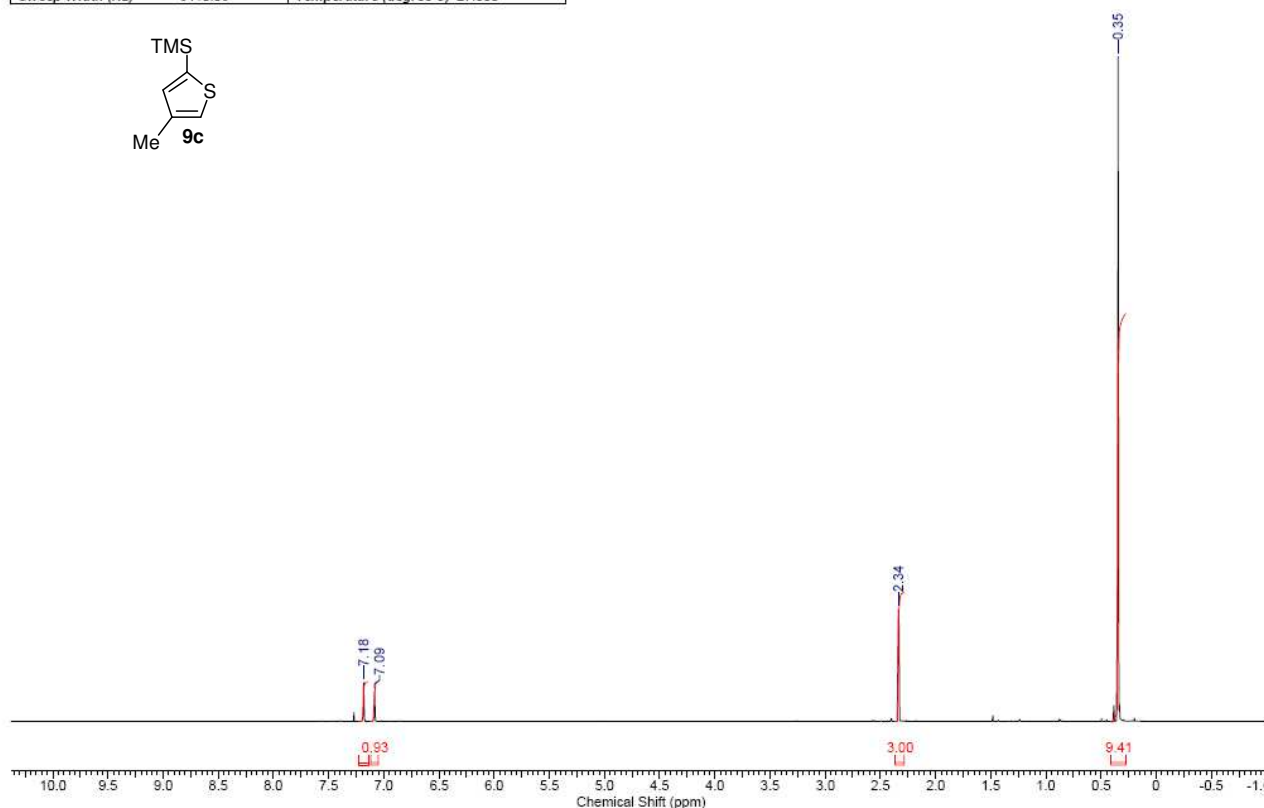
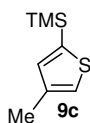


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Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	512
Original Points Count	65536	Owner	shr-usam-cc10046	Origin	Bruker BioSpin GmbH
SW(cyclical) (Hz)	28408.66	Solvent	CHLOROFORM-d	Points Count	65536
Sweep Width (Hz)	28408.22	Temperature (degree C)	27.000	Spectrum Offset (Hz)	12070.8818

93279-22-6_20.DX

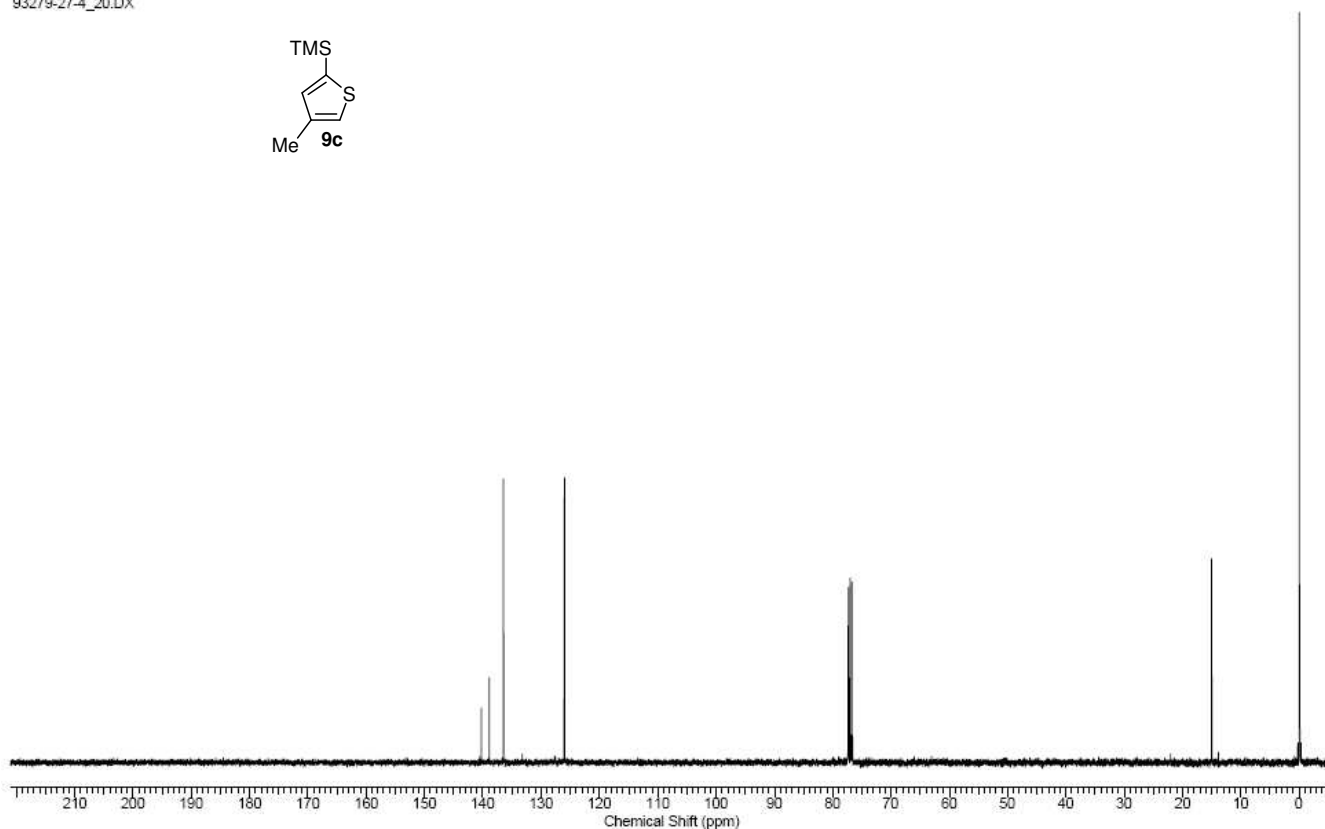
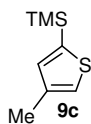


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Nucleus	¹ H	Number of Transients	16	Origin	Bruker BioSpin GmbH
Original Points Count	65536	Owner	shr-usam-cc10046	Points Count	65536
SW(cyclical) (Hz)	6410.16	Solvent	CHLOROFORM-d	Spectrum Offset (Hz)	2493.0339
Sweep Width (Hz)	6410.06	Temperature (degree C)	27.000		

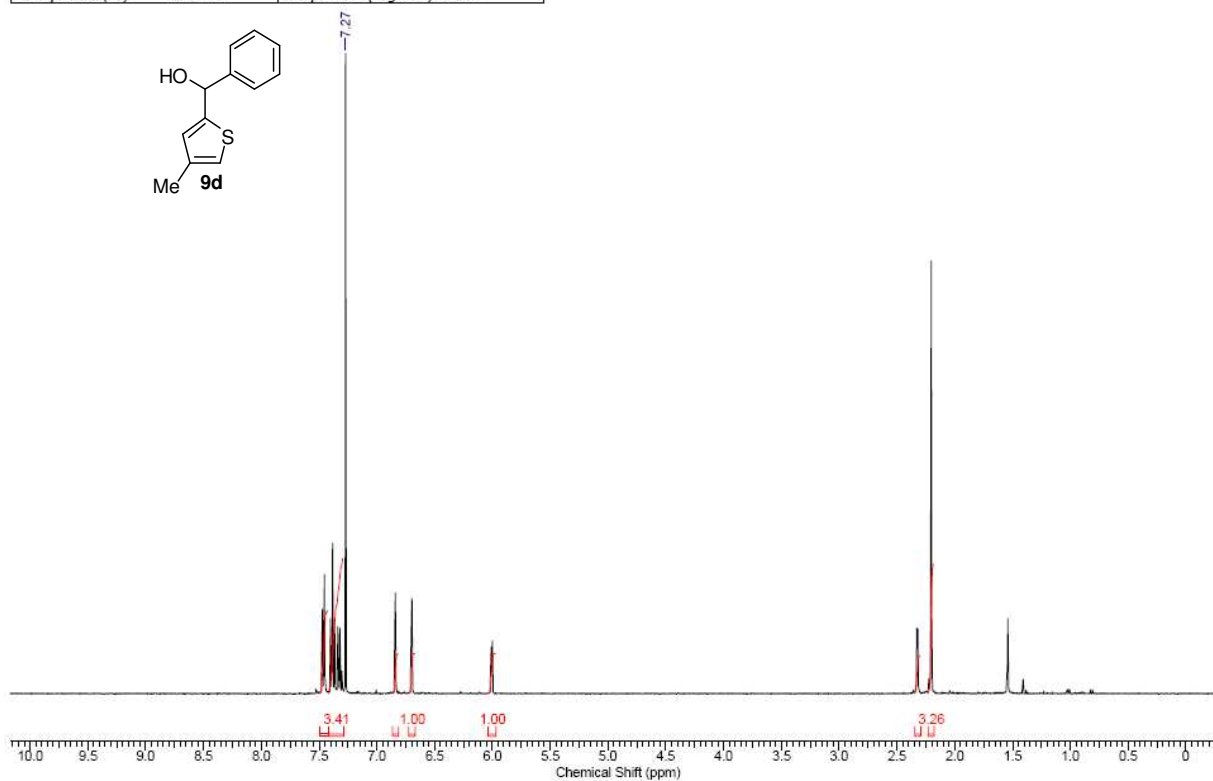
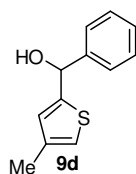


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Date Stamp	18 Aug 2008 14:27:04	File Name	\\CHOWDER\TEAMS\NMR\JCAMP\KNGAI\2008\93279-27-4_20.DX		
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Original Points Count	65536	Owner	shr-usam-cc10046	Origin	Bruker BioSpin GmbH
SW(cyclical) (Hz)	26408.66	Solvent	CHLOROFORM-d	Points Count	65536
Sweep Width (Hz)	26408.22	Temperature (degree C)	27.000	Spectrum Offset (Hz)	12064.1025

93279-27-4_20.DX

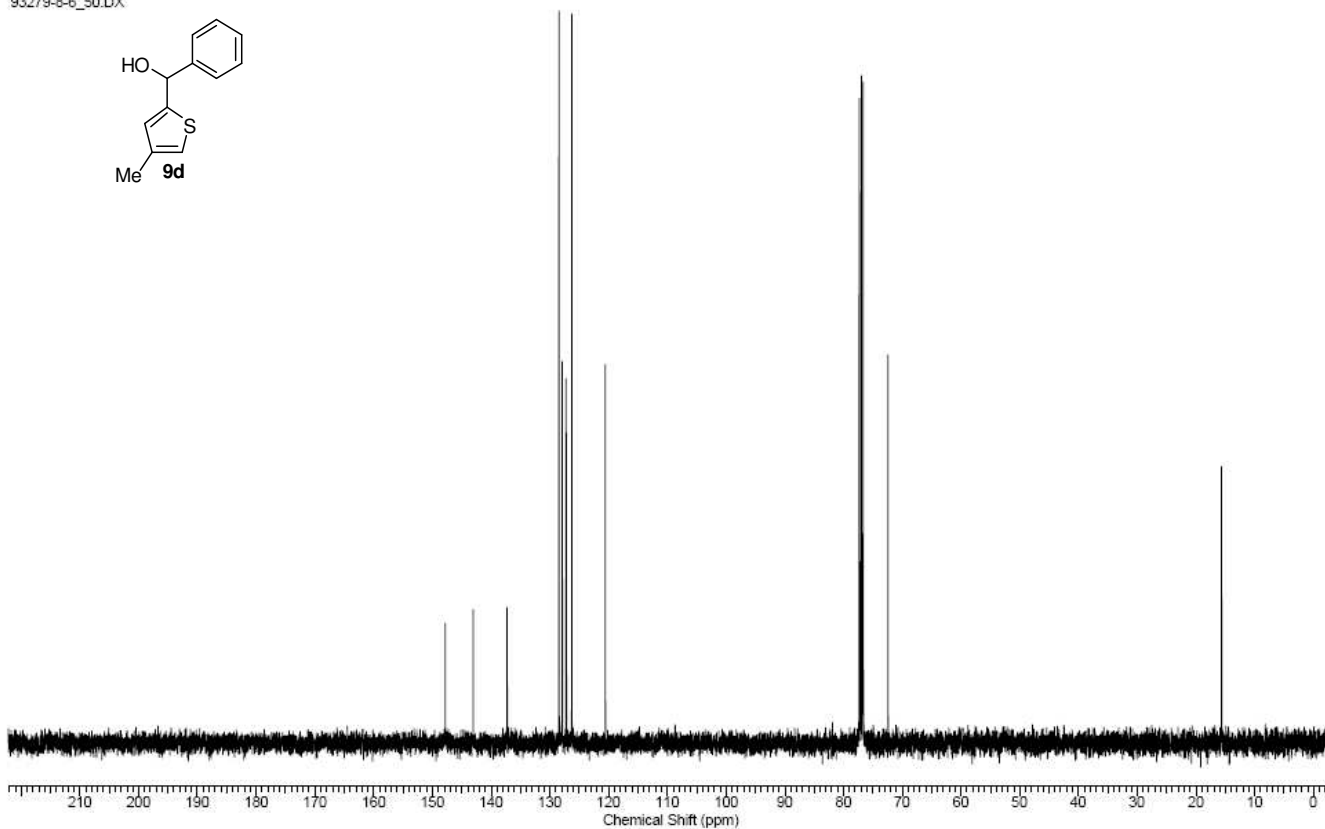
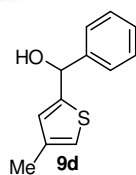


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Nucleus	¹ H	Number of Transients	16	Origin	Bruker BioSpin GmbH
Original Points Count	65536	Owner	shr-usam-cc10046	Points Count	65536
SW(cyclical) (Hz)	6410.16	Solvent	CHLOROFORM-d	Spectrum Offset (Hz)	2795.1621
Sweep Width (Hz)	6410.06	Temperature (degree C)	27.000		

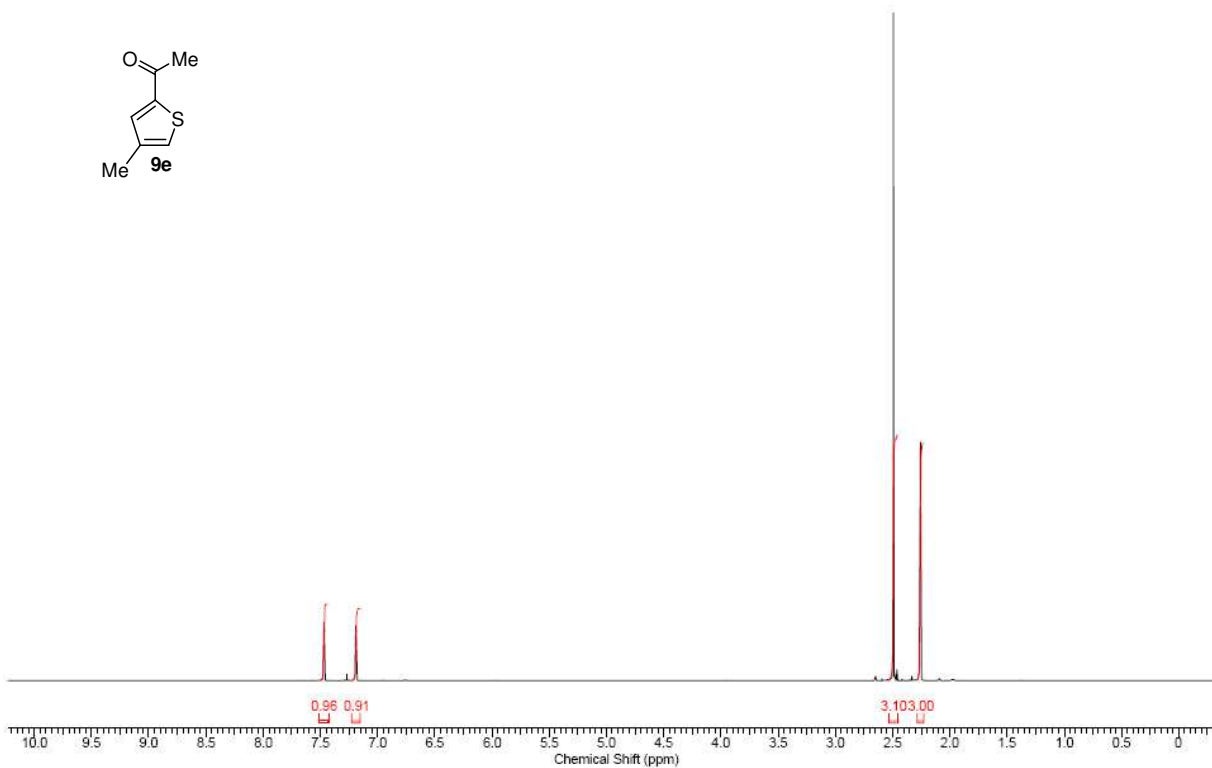
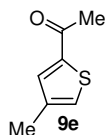


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Date Stamp	20 Aug 2008 10:31:38			File Name	WCHOWDER\TEAMS\NMR\JCAMP\KNGAI\2008\93279-8-6_50.DX
Frequency (MHz)	100.57	Nucleus	¹³ C	Number of Transients	128
Original Points Count	65536	Owner	shr-usam-cc10046	Origin	Bruker BioSpin GmbH
SW(cyclical) (Hz)	28406.66	Solvent	CHLOROFORM-d	Points Count	65536
Sweep Width (Hz)	28406.22	Temperature (degree C)	27.000	Spectrum Offset (Hz)	12061.5010

93279-8-6_50.DX

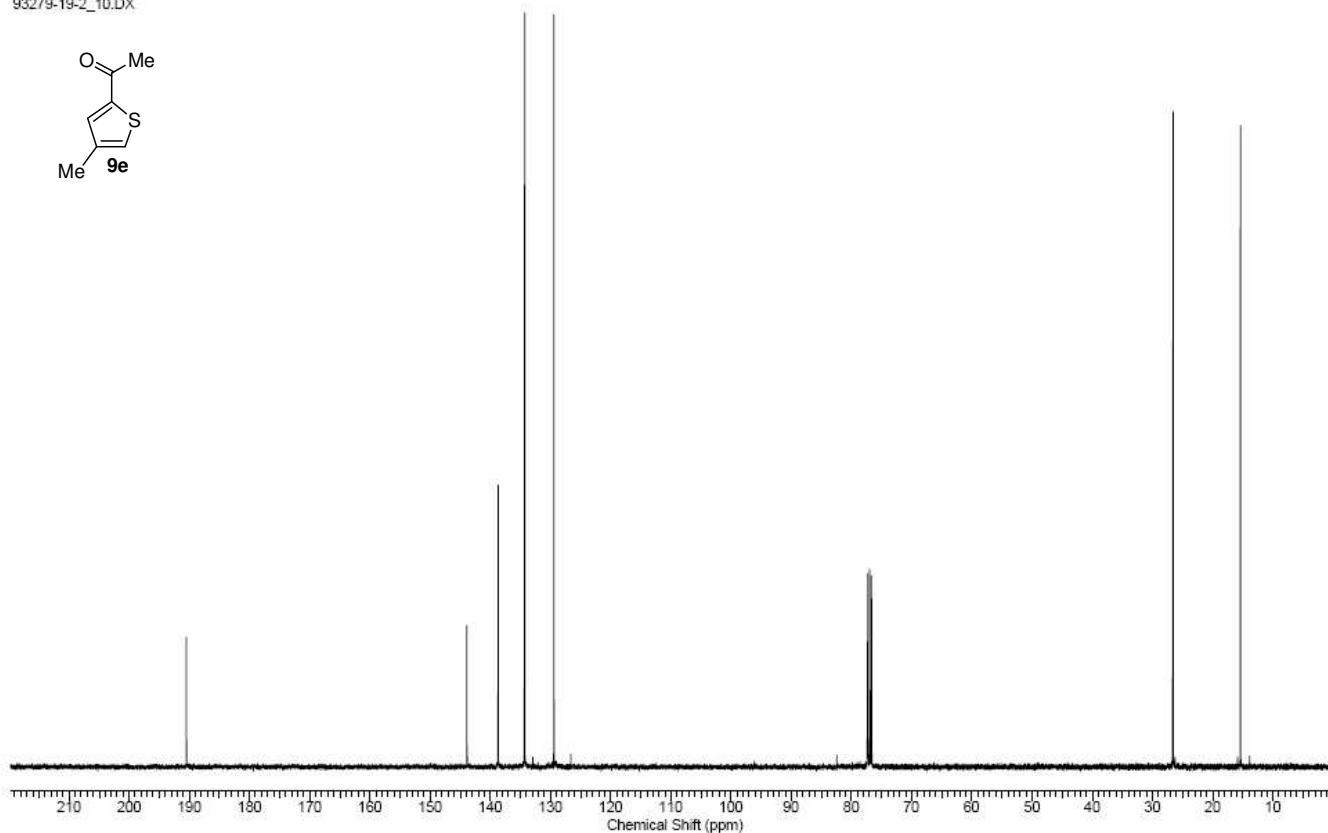
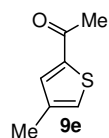


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Nucleus	¹ H	Number of Transients	16	Origin	Bruker BioSpin GmbH
Original Points Count	65536	Owner	shr-usam-cc10046	Points Count	65536
SW(cyclical) (Hz)	6410.16	Solvent	CHLOROFORM-d	Spectrum Offset (Hz)	2795.6511
Sweep Width (Hz)	6410.06	Temperature (degree C)	27.000		

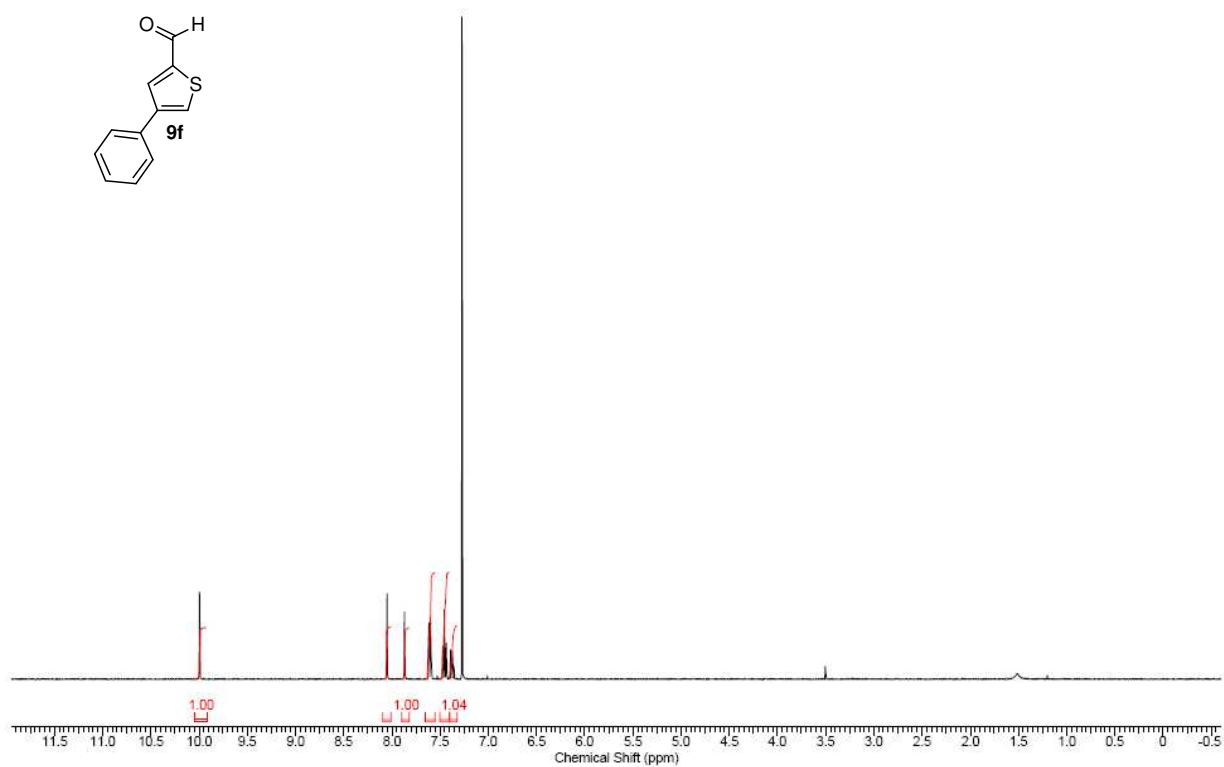
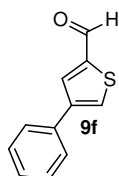


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Original Points Count	65536	Owner	shr-usam-cc10046	Origin	Bruker BioSpin GmbH
SW(cyclical) (Hz)	28408.66	Solvent	CHLOROFORM-d	Points Count	65536
Sweep Width (Hz)	28408.22	Temperature (degree C)	27.000	Spectrum Offset (Hz)	12053.2646

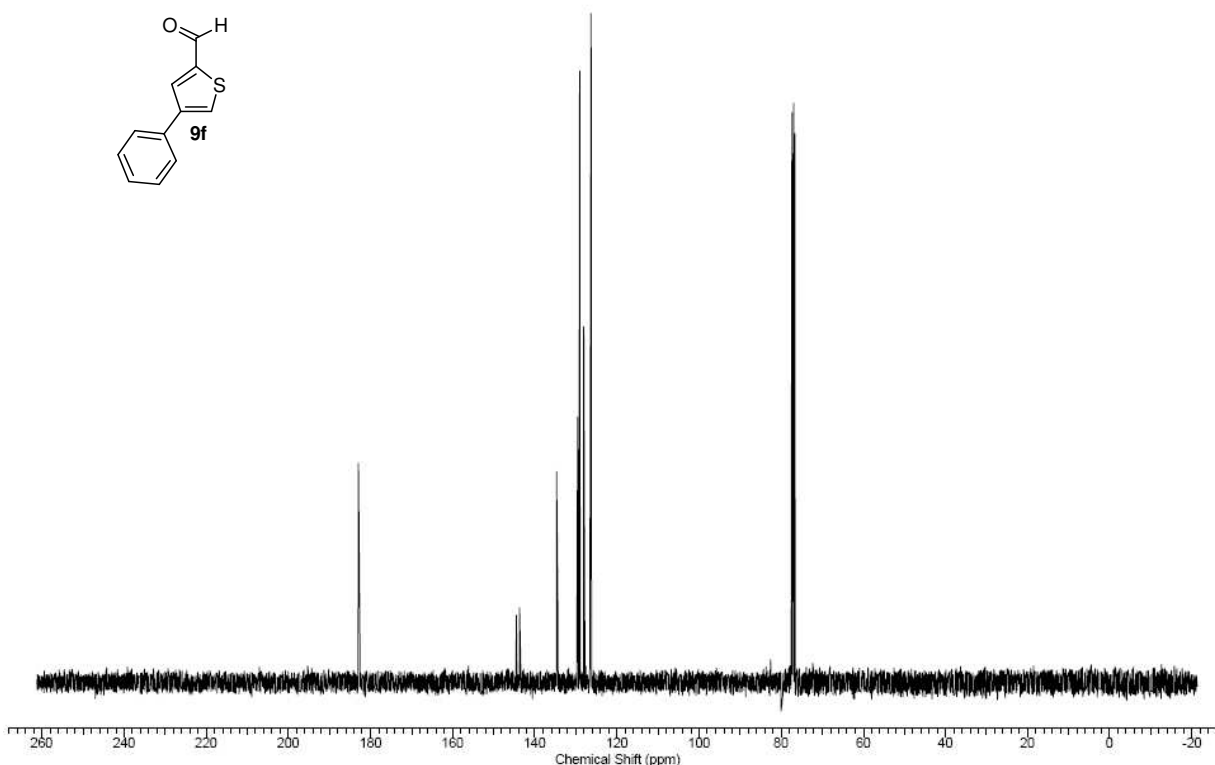
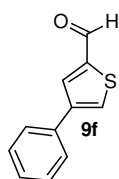
93279-19-2_10.DX



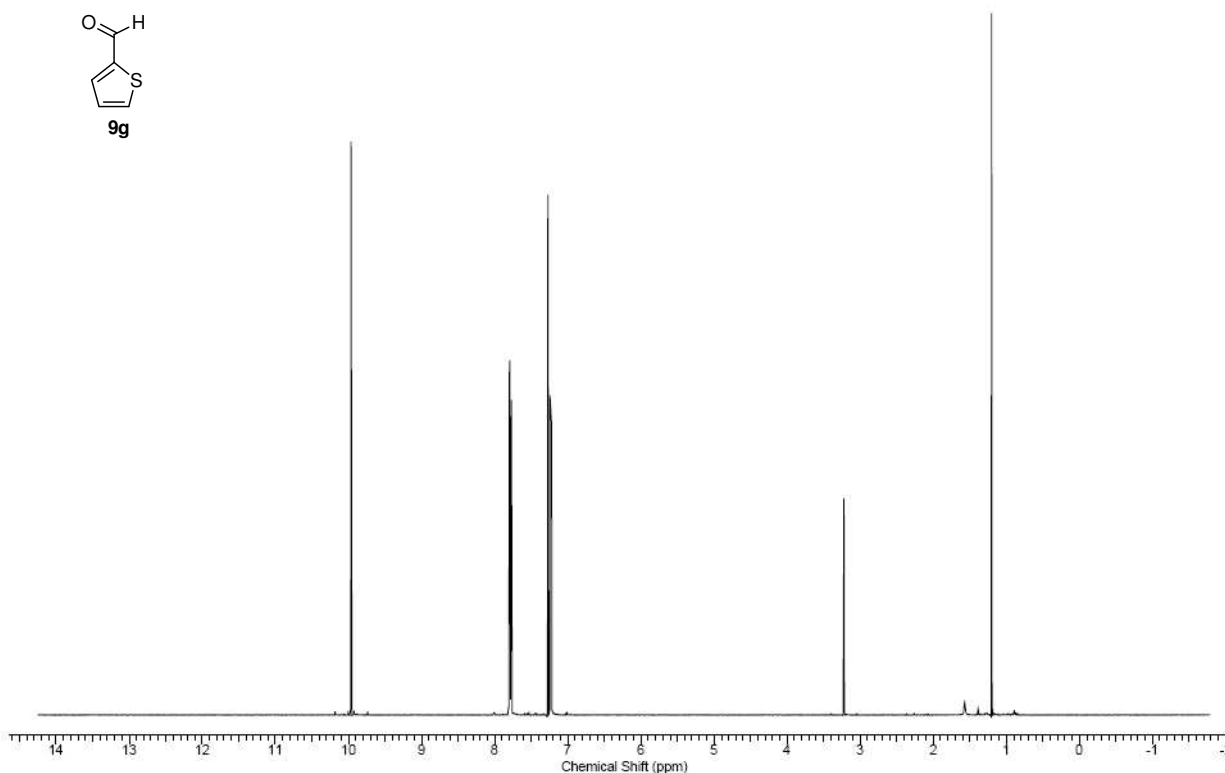
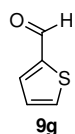
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Original Points Count	65536	Owner	shr-usam-cc10046	Points Count	65536
SW(cyclical) (Hz)	6410.16	Solvent	CHLOROFORM-d	Spectrum Offset (Hz)	2492.8384
Sweep Width (Hz)	6410.06	Temperature (degree C)	27.000		



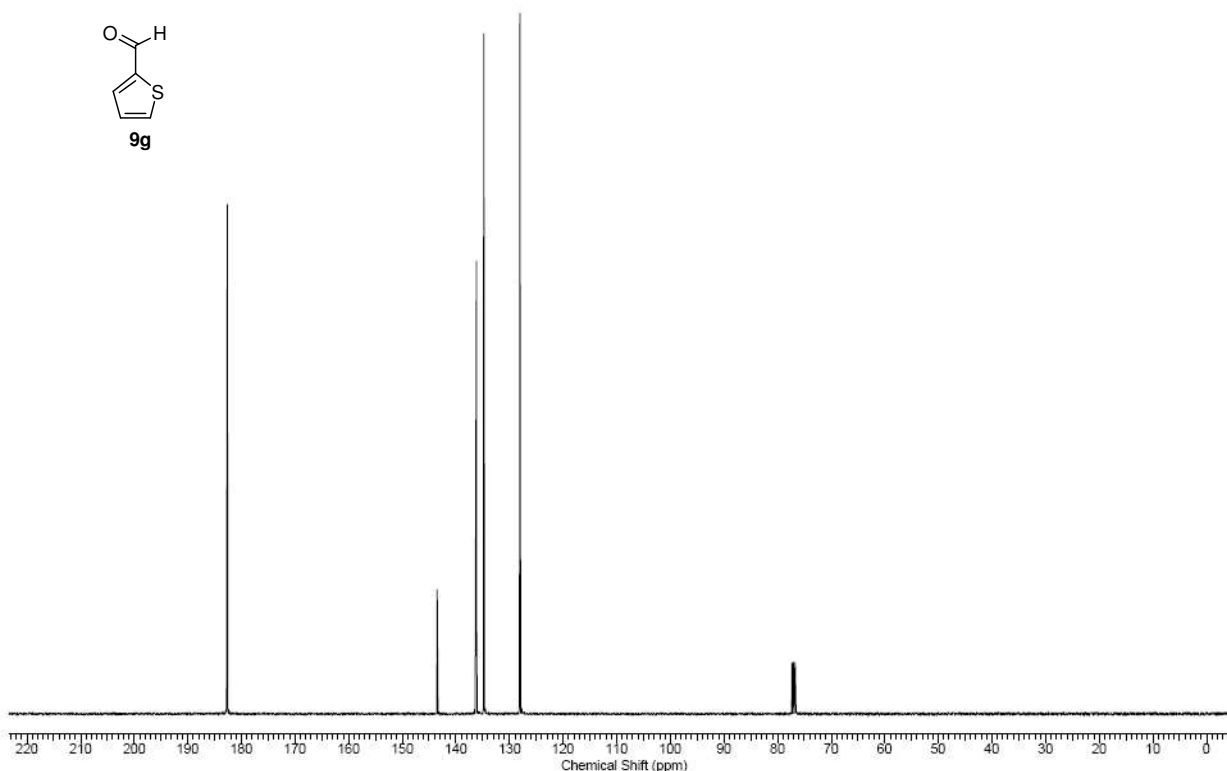
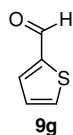
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Nucleus	¹³ C	Number of Transients	512	Origin	Bruker BioSpin GmbH
Original Points Count	65536	Owner	shr-usam-cc10046	Points Count	65536
SW(cyclical) (Hz)	28408.66	Solvent	CHLOROFORM-d	Spectrum Offset (Hz)	12064.9688
Sweep Width (Hz)	28408.22	Temperature (degree C)	27.000		



Acquisition Time (sec)	10.2239	Comment	kgai	Date	05 Aug 2008 19:30:12
File Name	WCHOWDER\TEAMS\NMR\JCAMP\KNGAI\2008\94406-5-4_10.DX	Frequency (MHz)	399.93		
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Original Points Count	65536	Owner	shr-usam-cc10046	Points Count	65536
SW(cyclical) (Hz)	6410.16	Solvent	CHLOROFORM-d	Spectrum Offset (Hz)	2492.8381
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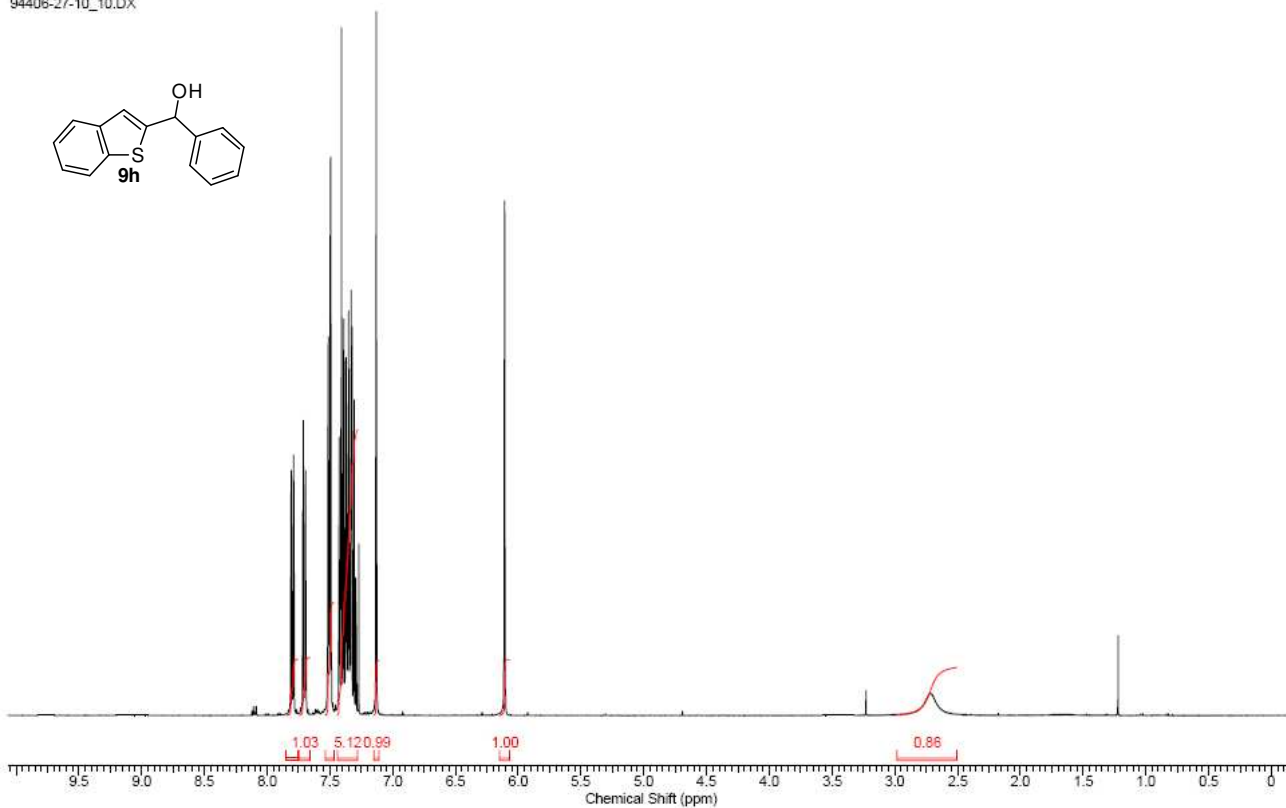
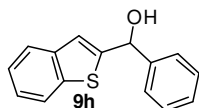


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Original Points Count	65536	Owner	shr-usam-cc10046	Points Count	65536	
SW(cyclical) (Hz)	28408.66	Solvent	CHLOROFORM-d	Spectrum Offset (Hz)	12028.1230	
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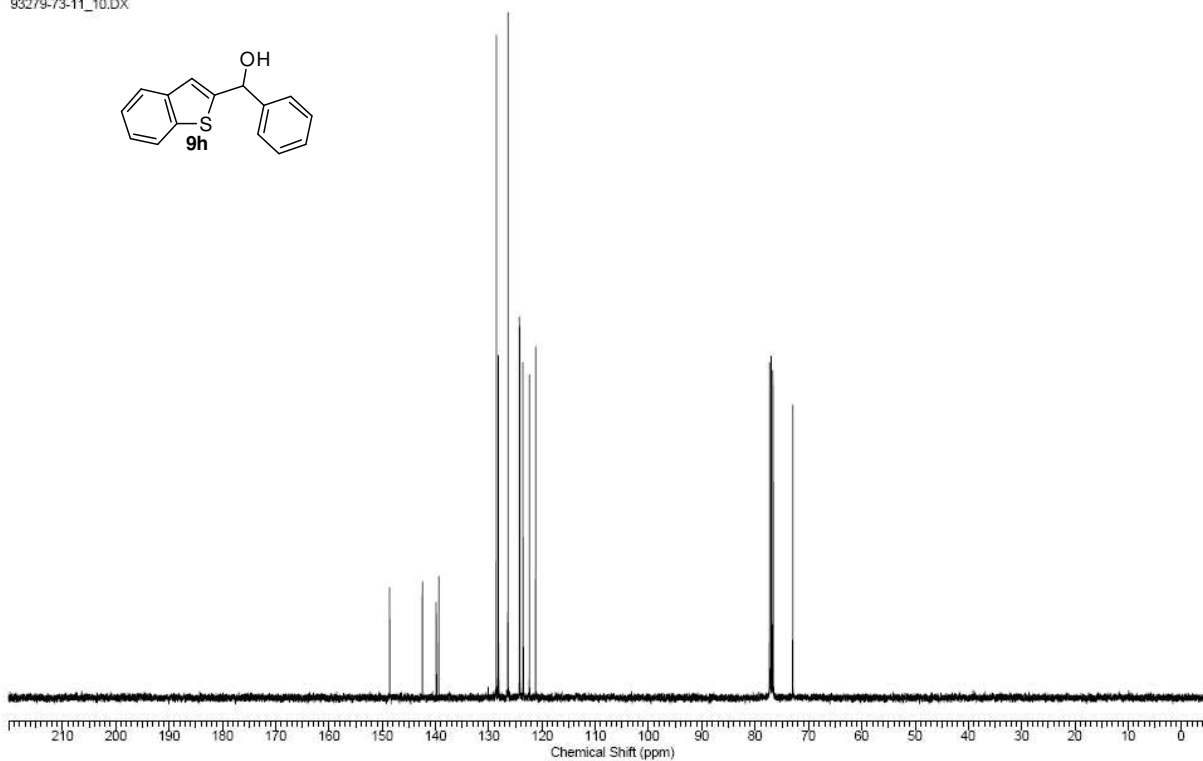
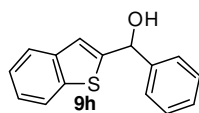
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Original Points Count	65536	Owner	shr-usam-cc10046	Points Count	65536
SW(cyclical) (Hz)	6410.16	Solvent	CHLOROFORM-d	Spectrum Offset (Hz)	2795.3579
Sweep Width (Hz)	6410.06	Temperature (degree C)	27.000		

94406-27-10_10.DX

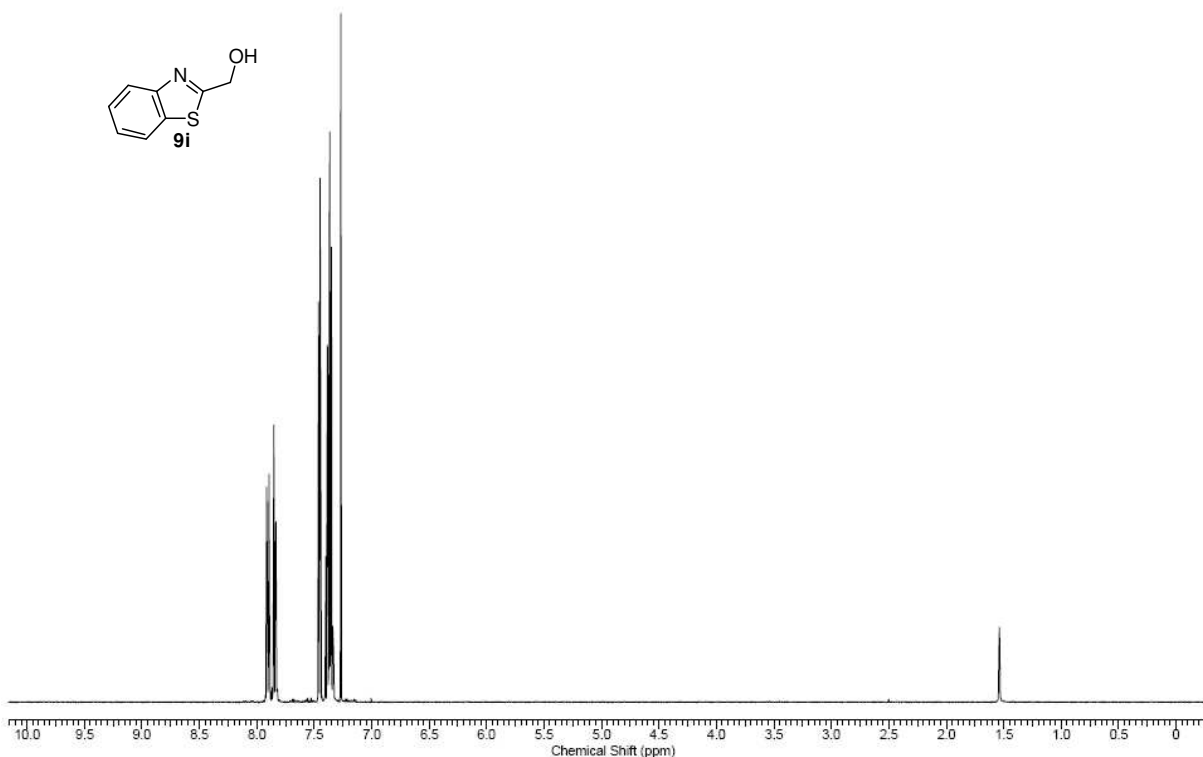
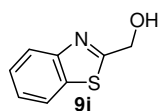


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Original Points Count	65536	Owner	shr-usam-cc10046	Origin	Bruker BioSpin GmbH
SW(cyclical) (Hz)	28408.66	Solvent	CHLOROFORM-d	Points Count	65536
Sweep Width (Hz)	28408.22	Temperature (degree C)	27.000	Spectrum Offset (Hz)	12061.0391

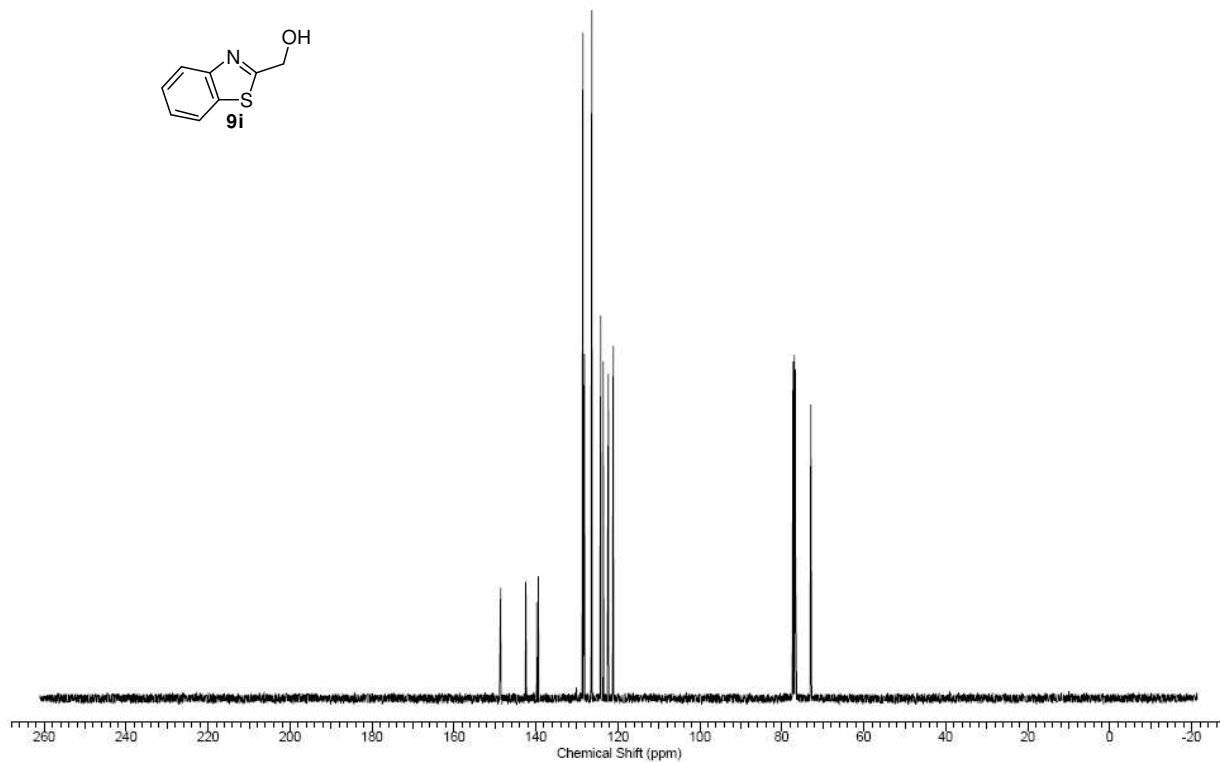
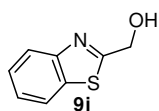
93279-73-11_10.DX



Acquisition Time (sec)	10.2239	Comment	knai	Date	21 Jul 2006 16:14:25
File Name	\CHOWDER\TEAMS\NMR\JCAMP\KNGAI\2008\93279-73-01_10.DX	Frequency (MHz)	399.93		
Nucleus	¹ H	Number of Transients	16	Origin	Bruker BioSpin GmbH
Original Points Count	65536	Owner	shr-usam-cc10046	Points Count	65536
SW(cyclical) (Hz)	6410.16	Solvent	CHLOROFORM-d	Spectrum Offset (Hz)	2492.8381
Sweep Width (Hz)	6410.06	Temperature (degree C)	27.000		



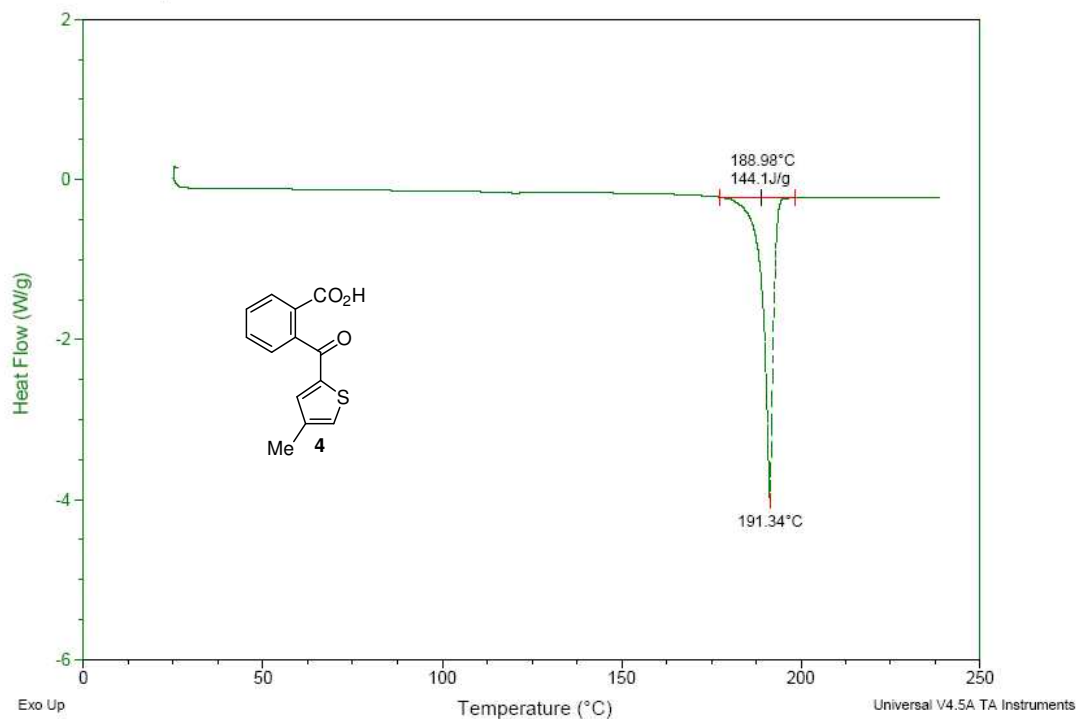
Acquisition Time (sec)	2.3069	Comment	krngai	Date	27 Aug 2008 14:47:05
File Name	\\CHOWDER\TEAMS\NMR\JCAMP\KNGAI\2008\93279-73-11	10.DX		Frequency (MHz)	100.62
Nucleus	¹³ C	Number of Transients	512	Origin	Bruker BioSpin GmbH
Original Points Count	65536	Owner	shr-usam-cc10046	Points Count	65536
SW(cyclical) (Hz)	28408.66	Solvent	CHLOROFORM-d	Spectrum Offset (Hz)	12061.0391
Sweep Width (Hz)	28408.22	Temperature (degree C)	27.000		



Sample: 111129-03-8
Size: 5.5300 mg
Method: 25C to 240C at 5C/min
Comment: keto-acid product

DSC

File: E:\DSC\2010\March\NFL\111129-03-8.007
Operator: NFL
Run Date: 29-Mar-2010 23:03
Instrument: DSC Q200 V24.3 Build 115



Sample: 110999-49-10
Size: 3.0900 mg
Method: 25C to 240C at 10C/min
Comment: Final solid

DSC

File: E:\DSC\2010\March\Steve\110999-49-10.001
Operator: SMM
Run Date: 09-Mar-2010 16:49
Instrument: DSC Q200 V24.3 Build 115

