

Organocatalytic Approach to Polysubstituted Piperidines and Tetrahydropyrans

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Supporting Information

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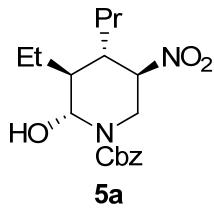
Experimental

General information. For thin-layer chromatography (TLC), Silica gel plates GF254 were used and compounds were visualized by irradiation with UV light, I_2 , or by treatment with a solution of phosphomolybdic acid in ethanol followed by heating. Optical rotations were measured by used a Perkin–Elmer 241MC polarimeter in the solvent indicated. FT-IR spectra were recorded on an AVATAR-360 spectrophotometer. 1H and ^{13}C NMR spectra were recorded on MERCURY300, Bruker DRX-400, and Bruker AV-500 spectrometers with TMS as the internal standard. HRMS were recorded by using either FTMS-7 or IonSpec 4.7 spectrometers. HPLC was carried out using a Water 515 pump, waters 2487 UV detector, Millennium workstation, Phenomenex (Lux Amylose-2, Lux Cellulose-1, Lux Cellulose-2), Daicel Chiralpak HPLC column. Yields refer to pure compounds, unless otherwise indicated.

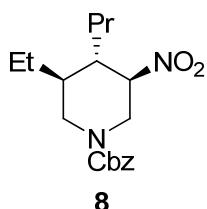
General procedure for cascade Michael-aminalization process of aldehydes with substituted nitroalkenes: Aldehyde **2** (0.40 mmol) was added to a suspension of catalyst **1** (0.02-0.03 mmol), nitroalkenes **3** (0.20 mmol) and benzoic acid (0.06-0.09 mmol) in water (0.40 mL) at room temperature. The stirring was continued until complete conversion of **3** (monitored by TLC). The reaction mixture was diluted with ethyl acetate, dried over Na_2SO_4 and concentrated. Purification by flash column chromatography (silica gel, petroleum ether/AcOEt) afforded the the corresponding substituted piperidine. The enantiomeric excess (ee) was determined by HPLC on a chiral phase. *Noteworthy is that these substituted piperidines could isomerize at the chiral center of hemiaminal part during standing in $CDCl_3$, and therefore some additional peaks (for isomer) were observed in ^{13}C NMR spectra if ^{13}C NMR determination took longer time.*

Reduction of **3 with triethylsilane and $BF_3\cdot OEt_2$.** To a mixture of the above crude product (0.15 mmol) and triethylsilane (72 μ L, 0.45 mmol) in CH_2Cl_2 (1.5 mL) was added $BF_3\cdot OEt_2$ (23 μ L, 0.3 mmol) at 0 °C. The reaction mixture was stirred for 1-3 h.

Purification by flash column chromatography (silica gel, petroleum ether/AcOEt) afforded the reduced product. The enantiomeric excess (ee) was determined by HPLC on a chiral phase.

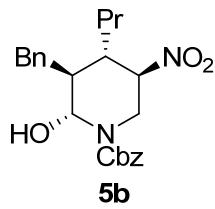


(2R,3S,4S,5R)-3-Ethyl-2-hydroxy-5-nitro-4-propyl-piperidine-1-carboxylic acid benzyl ester 5a. $[\alpha]_D^{28} = -8.4$ (*c* 1.00, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.39-7.32 (m, 5H), 5.74 (s, 1H), 5.15 (s, 2H), 4.47 (dt, *J* = 4.4, 10.4 Hz, 1H), 4.18 (d, *J* = 10.0 Hz, 1H), 3.78 (t, *J* = 10.8 Hz, 1H), 3.00-2.80 (br, 1H), 2.46-2.39 (m, 1H), 1.63-1.56 (m, 1H), 1.48-1.37 (m, 3H), 1.33-1.21 (m, 3H), 1.00 (t, *J* = 6.8 Hz, 3H), 0.89 (t, *J* = 6.8 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 155.12, 135.89, 128.81, 128.60, 128.27, 85.39, 74.97, 68.07, 42.79, 41.22, 37.59, 30.28, 21.27, 17.17, 14.49, 11.38; IR (neat): ν = 3432 br, 2962 m, 2934 m, 1690 s, 1550 s, 1431 m, 1145 m, 963 m, 748 m, 698 m cm^{-1} ; ESI-MS *m/z* 373.4 ($\text{M} + \text{Na}$) $^+$, HRMS (MALDI) calcd for $\text{C}_{18}\text{H}_{26}\text{N}_2\text{O}_5\text{Na}$ ($\text{M} + \text{Na}$) $^+$ *m/z* 373.1734, found 373.1733; HPLC: PHENOMENEX LUX AMYLOSE-2 (PA-2) column, hexane/*i*-PrOH = 70:30, flow rate 0.7 mL/min, 214 nm, major isomer: $t_R = 8.5$ min (major), $t_R = 7.7$ min (minor), *ee*=98%. (Noteworthy is that some additional peaks (for minor isomer) were observed in ^{13}C NMR spectra due to isomerization)

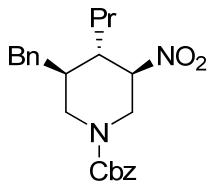


(3S,4S,5R)-3-Ethyl-5-nitro-4-propyl-piperidine-1-carboxylic acid benzyl ester 8. $[\alpha]_D^{28} = -40.6$ (*c* 1.00, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.39-7.31 (m, 5H), 5.20-5.07 (m, 2H), 4.65-4.38 (m, 2H), 4.30-4.10 (m, 1H), 3.30-3.10 (m, 1H),

2.63-2.45 (m, 1H), 1.97 (tt, J = 3.2, 10.8 Hz, 1H), 1.70-1.60 (m, 1H), 1.60-1.46 (m, 1H), 1.45-1.35 (m, 1H), 1.35-1.15 (m, 4H), 0.93 (t, J = 7.2 Hz, 3H), 0.88 (t, J = 6.8 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 154.88, 136.35, 128.72, 128.39, 128.14, 85.57, 67.78, 47.80, 47.16, 43.73, 38.43, 30.37, 22.71, 17.65, 14.56, 10.75; IR (neat): ν = 2962 m, 2875 m, 1694 s, 1546 s, 1440 s, 1222 m, 1146 m, 1098 w, 754 w, 699 m cm⁻¹; ESI-MS m/z 335.2 (M + H)⁺, 357.3 (M + Na)⁺, HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{26}\text{N}_2\text{O}_4\text{Na}$ (M + Na)⁺ m/z 357.1785, found 357.1793; HPLC: PHENOMENEX LUX AMYLOSE-2 (PA-2) column, hexane/*i*-PrOH = 70:30, flow rate 0.7 mL/min, 214 nm, t_{R} = 12.5 min (major), t_{R} = 11.0 min (minor), *ee* = 98%.

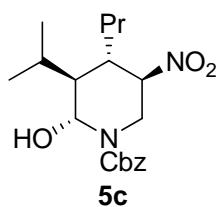


(2R,3S,4S,5R)-3-Benzyl-2-hydroxy-5-nitro-4-propyl-piperidine-1-carboxylic acid benzyl ester 5b. $[\alpha]_D^{27} = -8.7$ (*c* 1.02, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ major isomer: 7.31-7.19 (m, 10H), 5.26 (d, J = 2.0 Hz, 1H), 5.08-4.98 (m, 2H), 4.50 (dt, J = 4.4, 10.4 Hz, 1H), 4.05-4.22 (m, 1H), 3.80 (t, J = 10.8 Hz, 1H), 2.91 (dd, J = 4.0, 13.6 Hz, 1H), 2.65-2.57 (m, 2H), 1.78 (tt, J = 3.6, 14.8 Hz, 1H), 1.61-1.55 (m, 1H), 1.45-1.36 (m, 3H), 0.93 (t, J = 6.8 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ major isomer: 154.67, 138.93, 135.76, 129.09, 128.78, 128.72, 128.43, 127.92, 126.66, 85.17, 74.73, 67.85, 43.51, 41.17, 37.49, 34.66, 30.39, 17.24, 14.50; IR (neat): ν = 3419 br, 2959 m, 2932 m, 1705 s, 1550 s, 1432 m, 1305 m, 1143 m, 954 w, 737 w, 699 m cm⁻¹; ESI-MS m/z 435.5 (M + Na)⁺, HRMS (MALDI) calcd for $\text{C}_{23}\text{H}_{28}\text{N}_2\text{O}_5\text{Na}$ (M + Na)⁺ m/z 435.1890, found 435.1889. (Noteworthy is that some additional peaks (for minor isomer) were observed in ^{13}C NMR spectra due to isomerization)



reduction product of **5b**

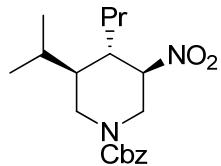
(3S,4S,5R)-3-Benzyl-5-nitro-4-propyl-piperidine-1-carboxylic acid benzyl ester (reduction preoduct of 5b). $[\alpha]_D^{28} = -16.8$ (*c* 1.20, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.35-7.21 (m, 7H), 7.12-7.06 (m, 3H), 5.10 (d, *J* = 11.2 Hz, 1H), 4.99 (d, *J* = 11.2 Hz, 1H), 4.55-4.45 (m, 2H), 3.95 (d, *J* = 7.2 Hz, 1H), 3.20-3.30 (m, 1H), 3.02 (d, *J* = 11.6 Hz, 1H), 2.54 (t, *J* = 11.6 Hz, 1H), 2.20-2.35 (m, 1H), 2.10-2.04 (m, 1H), 1.76-1.61 (m, 2H), 1.42-1.32 (m, 3H), 0.92 (t, *J* = 7.0 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 154.65, 138.58, 136.18, 128.91, 128.81, 128.64, 128.18, 127.58, 126.72, 85.23, 67.61, 48.06, 47.06, 44.29, 39.28, 36.76, 30.58, 17.71, 14.56; IR (neat): ν = 2959 m, 2932 m, 2873 m, 1708 s, 1538 s, 1470 m, 1455 m, 1374 m, 1254 m, 1222 s, 1139 s, 736 m, 699 m cm^{-1} ; ESI-MS *m/z* 419.4 ($\text{M} + \text{Na}$) $^+$, HRMS (ESI) calcd for $\text{C}_{23}\text{H}_{28}\text{N}_2\text{O}_4\text{Na}$ ($\text{M} + \text{Na}$) $^+$ *m/z* 419.1941, found 419.1950; HPLC: PHENOMENEX LUX CELLULOSE-2 (PC-2) column, hexane/*i*-PrOH = 70:30, flow rate 0.7 mL/min, 214 nm, t_R = 14.5 min (major), t_R = 10.6 min (minor), *ee*=99%.



5c

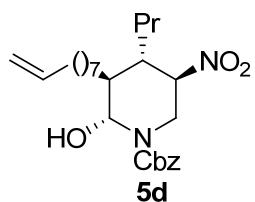
(2R,3S,4S,5R)-2-Hydroxy-3-isopropyl-5-nitro-4-propyl-piperidine-1-carboxylic acid benzyl ester 5c. $[\alpha]_D^{26} = -8.6$ (*c* 0.97, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ major isomer: 7.32-7.30 (m, 5H), 5.82 (s, 1H), 5.12 (s, 2H), 4.51 (dt, *J* = 4.4, 10.0 Hz, 1H), 4.16 (d, *J* = 8.8, Hz, 1H), 3.82 (t, *J* = 11.4, Hz, 1H), 2.73-2.67 (m, 1H), 1.98-1.90 (m, 1H), 1.46-1.43 (m, 1H), 1.42-1.40 (m, 1H), 1.37-1.28 (m, 2H), 1.24-1.16 (m, 1H), 1.09 (d, *J* = 7.2, Hz, 3H), 1.02 (d, *J* = 7.2 Hz, 3H), 0.90 (t, *J* = 6.8 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ major isomer: 154.90, 135.89, 128.78, 128.55, 128.19, 85.34, 75.46, 67.98, 44.90, 40.97, 35.31, 29.80, 26.62, 20.88, 17.70, 16.39, 14.57; IR (neat): ν = 3437 br, 2962 s, 2934 m, 1702 s, 1552 s, 1429 m, 1266 m, 985 m, 739 s, 702 m

cm^{-1} ; ESI-MS m/z 387.3 ($\text{M} + \text{Na}$) $^+$, HRMS (ESI) calcd for $\text{C}_{19}\text{H}_{28}\text{N}_2\text{O}_5\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 387.1890, found 387.1883. (Noteworthy is that some additional peaks (for isomer) were observed in ^{13}C NMR spectra due to isomerization)



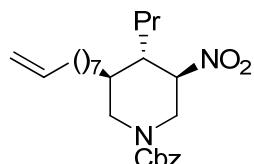
reduction product of **5c**

(3S,4S,5R)-3-Isopropyl-5-nitro-4-propyl-piperidine-1-carboxylic acid benzyl ester (reduction product of 5c). $[\alpha]_D^{25} = -34.4$ (c 0.97, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.39-7.31 (m, 5H), 5.19-5.09 (m, 2H), 4.54-4.42 (m, 2H), 4.20-4.08 (m, 1H), 3.26-3.14 (m, 1H), 2.68-2.56 (m, 1H), 2.15 (tt, $J = 3.2, 10.8$ Hz, 1H), 2.00-1.92 (m, 1H), 1.54-1.48 (m, 1H), 1.38-1.16 (m, 4H), 1.06-0.98 (m, 3H), 0.88 (t, $J = 6.8$ Hz, 3H), 0.86 (t, $J = 6.8$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 155.00, 136.37, 128.72, 128.39, 128.11, 85.67, 67.78, 47.28, 43.45, 41.87, 29.85, 25.76, 21.42, 17.12, 15.71, 14.60; IR (neat): $\nu = 2961$ s, 2933 m, 1709 s, 1549 s, 1470 m, 1435 m, 1374 m, 1292 m, 1223 s, 1147 m, 1098 m, 970 w, 745 m, 698 w cm^{-1} ; ESI-MS m/z 349.3 ($\text{M} + \text{H}$) $^+$, 371.3 ($\text{M} + \text{Na}$) $^+$, HRMS (ESI) calcd for $\text{C}_{19}\text{H}_{28}\text{N}_2\text{O}_4\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 397.1734, found 397.1741; HPLC: PHENOMENEX LUX AMYLOSE-2 (PA-2) column, hexane/*i*-PrOH = 50:50, flow rate 0.7 mL/min, 214 nm, $t_R = 9.3$ min (major), $t_R = 8.7$ min (minor), *ee*>99%.



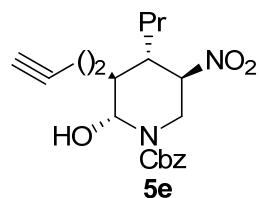
(2R,3S,4S,5R)-2-Hydroxy-5-nitro-3-non-8-enyl-4-propyl-piperidine-1-carboxylic acid benzyl ester 5d. $[\alpha]_D^{25} = -13.5$ (c 1.00, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.37-7.35 (m, 5H), 5.84-5.76 (m, 1H), 5.70 (s, 1H), 5.15 (s, 2H), 4.99 (d, $J = 16.8$ Hz, 1H), 4.93 (d, $J = 10.0$ Hz, 1H), 4.46 (dt, $J = 4.4, 10.4$ Hz, 1H), 4.17 (d, $J = 8.4$ Hz, 1H), 3.78 (t, $J = 11.2$ Hz, 1H), 2.42 (m, 1H), 2.33 (t, $J = 7.6$ Hz, 1H), 2.06-2.01 (m, 2H), 1.64-1.61 (m, 1H), 1.45-1.26 (m, 16H), 0.90 (t, $J = 6.4$ Hz, 3H); ^{13}C NMR (100 MHz,

CDCl₃) δ 155.15, 139.27, 135.88, 128.81, 128.61, 128.28, 114.34, 85.37, 75.24, 68.09, 41.24, 37.61, 33.90, 30.33, 29.82, 29.53, 29.18, 29.03, 28.30, 26.70, 24.84, 17.21, 14.50; IR (neat): ν = 3448 br, 2928 s, 1686 s, 1551 s, 1336 w, 1143 m, 968 m, 910 m, 697 m cm⁻¹; ESI-MS *m/z* 469.5 (M + Na)⁺, HRMS (ESI) calcd for C₂₅H₃₈N₂O₅Na (M + Na)⁺ *m/z* 469.2673, found 469.2682. (Noteworthy is that some additional peaks (for isomer) were observed in ¹³C NMR spectra due to isomerization)



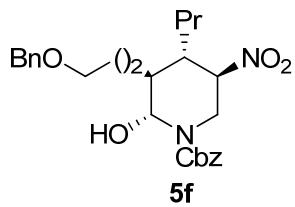
reduction product of **5d**

(3R,4S,5S)-3-Nitro-5-non-8-enyl-4-propyl-piperidine-1-carboxylic acid benzyl ester (reduction product of 5d). [α]_D²⁷ = -33.7 (c 1.00, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.39-7.32 (m, 5H), 5.86-5.76 (m, 1H), 5.18-5.10 (m, 2H), 5.00 (dd, *J* = 1.6, 17.2 Hz, 1H), 4.94 (d, *J* = 10.4 Hz, 1H), 4.54-4.38 (m, 2H), 4.27-4.10 (m, 1H), 3.27-3.15 (m, 1H), 2.60-2.45 (m, 1H), 2.04 (q, *J* = 6.8 Hz, 2H), 1.97-1.92 (m, 1H), 1.56-1.50 (m, 2H), 1.39-1.12 (m, 15H), 0.88 (t, *J* = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 154.83, 139.23, 136.35, 128.72, 128.40, 128.13, 114.35, 85.48, 67.79, 48.23, 47.16, 44.11, 37.16, 33.87, 30.42, 29.82, 29.44, 29.12, 29.00, 26.32, 17.68, 14.57; IR (neat): ν = 2928 s, 2855 s, 1710 s, 1550 s, 1467 s, 1433 s, 1376 m, 1290 m, 1221 s, 1142 m, 970 m, 911 m, 754 m, 698 m cm⁻¹; ESI-MS *m/z* 431.5 (M + H)⁺, 453.5 (M + Na)⁺, HRMS (MALDI) calcd for C₂₅H₃₈N₂O₄Na (M + Na)⁺ *m/z* 453.2724, found 453.2719; HPLC: PHENOMENEX LUX AMYLOSE-2 (PA-2) column, hexane/*i*-PrOH = 50:50, flow rate 0.7 mL/min, 214 nm, t_R = 9.2 min (major), t_R = 7.8 min (minor), *ee*>99%.



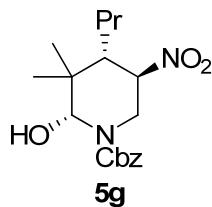
(2R,3S,4S,5R)-3-But-3-ynyl-2-hydroxy-5-nitro-4-propyl-piperidine-1-carboxylic acid benzyl ester 5e. [α]_D²⁵ = -26.2 (c 1.05, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ

7.38-7.31 (m, 5H), 5.75 (s, 1H), 5.18-5.10 (m, 2H), 4.50 (dt, J = 4.0, 10.0 Hz, 1H), 4.11-4.24 (m, 1H), 3.77 (t, J = 11.2 Hz, 1H), 2.45 (t, J = 10.4 Hz, 1H), 2.35-2.31 (m, 2H), 1.97 (brs, 1H), 1.76-1.66 (m, 3H), 1.48-1.42 (m, 1H), 1.34-1.26 (m, 4H), 0.90 (t, J = 6.8 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 155.13, 135.78, 128.80, 128.61, 128.26, 85.15, 83.27, 74.64, 69.58, 68.13, 41.11, 39.72, 37.30, 30.20, 26.79, 17.05, 15.68, 14.46; IR (neat): ν = 3435 br, 3297 s, 2959 s, 2932 s, 1694 s, 1549 m, 1430 m, 1337 m, 1144 m, 1095 m, 981 s, 698 m, 646 s cm^{-1} ; ESI-MS m/z 397.3 ($\text{M} + \text{Na}$) $^+$, HRMS (ESI) calcd for $\text{C}_{20}\text{H}_{26}\text{N}_2\text{O}_5\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 397.1734, found 397.1741; HPLC: sino-AD column, hexane/*i*-PrOH = 90:10, flow rate 0.7 mL/min, 214 nm, major isomer: t_{R} = 10.2 min (major), t_{R} = 12.6 min (minor), *ee*=99%. (Noteworthy is that some additional peaks (for isomer) were observed in ^{13}C NMR spectra due to isomerization)

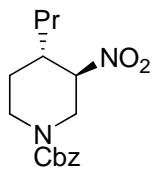


(2R,3S,4S,5R)-3-(3-Benzyl-1-oxo-propyl)-2-hydroxy-5-nitro-4-propyl-piperidine-1-carboxylic acid benzyl ester 5f. $[\alpha]_{\text{D}}^{25} = -11.7$ (c 1.22, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ major isomer: 7.35-7.29 (m, 10H), 5.71 (s, 1H), 5.12 (s, 2H), 4.47 (s, 2H), 4.43 (dd, J = 4.0, 10.0 Hz, 1H), 4.21-4.11 (m, 1H), 3.73 (t, J = 11.6 Hz, 1H), 3.47 (t, J = 6.0 Hz, 2H), 2.46-2.37 (m, 1H), 1.86-1.75 (m, 1H), 1.61-1.59 (m, 2H), 1.47-1.42 (m, 2H), 1.28-1.24 (m, 4H), 0.88 (t, J = 6.4 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ major isomer: 154.98, 138.38, 135.87, 128.76, 128.54, 128.49, 128.24, 127.78, 127.74, 85.33, 75.04, 73.05, 70.18, 68.00, 41.13, 37.58, 30.21, 29.78, 26.59, 24.94, 17.15, 14.44; IR (neat): ν = 3418 br, 2958 s, 2931 s, 2872 s, 1705 s, 1549 s, 1455 m, 1337 w, 1102 m, 968 m, 738 s, 698 m cm^{-1} ; ESI-MS m/z 493.5 ($\text{M} + \text{Na}$) $^+$, HRMS (MALDI) calcd for $\text{C}_{26}\text{H}_{34}\text{N}_2\text{O}_6\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 493.2309, found 493.2311; HPLC: PHENOMENEX LUX AMYLOSE-2 (PA-2) column, hexane/*i*-PrOH = 70:30, flow rate 0.7 mL/min, 214 nm, major isomer: t_{R} = 18.3 min (major), t_{R} = 10.5 min (minor),

ee>99%. (Noteworthy is that some additional peaks (for isomer) were observed in ^{13}C NMR spectra due to isomerization).



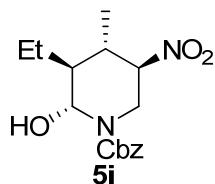
(2R,4S,5R)-2-Hydroxy-3,3-dimethyl-5-nitro-4-propyl-piperidine-1-carboxylic acid benzyl ester 5g. $[\alpha]_D^{28} = -8.7$ (*c* 0.67, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ major isomer: 7.40-7.32 (m, 5H), 5.26 (s, 1H), 5.14 (d, *J* = 7.2 Hz, 2H), 4.48 (dt, *J* = 4.8, 11.2 Hz, 1H), 4.39-4.17 (m, 1H), 3.70 (t, *J* = 12.0 Hz, 1H), 2.36-2.32 (m, 1H), 1.82-1.56 (m, 1H), 1.49-1.42 (m, 1H), 1.24-1.18 (m, 2H), 1.10 (s, 3H), 0.85 (t, *J* = 6.8 Hz, 3H), 0.82 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ major isomer: 155.30, 135.89, 128.82, 128.62, 128.26, 86.06, 82.44, 68.09, 41.62, 39.25, 30.20, 24.75, 23.24, 19.67, 14.47; IR (neat): ν = 3428 br, 2962 m, 2918 m, 1690 s, 1552 s, 1431 s, 1331 m, 1213 m, 1055 m, 968 m, 739 m, 698 m cm^{-1} ; ESI-MS *m/z* 373.4 ($\text{M} + \text{Na}$) $^+$, HRMS (MALDI) calcd for $\text{C}_{18}\text{H}_{26}\text{N}_2\text{O}_5\text{Na}$ ($\text{M} + \text{Na}$) $^+$ *m/z* 373.1734, found 373.1741. HPLC: PHENOMENEX LUX CELLULOSE-2 (PC-2) column, hexane/*i*-PrOH = 90:10, flow rate 0.7 mL/min, 214 nm, major isomer: $t_R = 16.1$ min (major), $t_R = 9.7$ min (minor), *ee* = 90%, minor isomer: $t_R = 8.4$ min (major), $t_R = 11.6$ min (minor), *ee* = 92%. (Noteworthy is that some additional peaks (for isomer) were observed in ^{13}C NMR spectra due to isomerization)



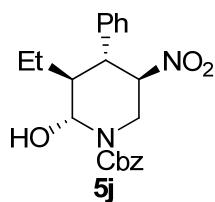
reduction product of **5g**

(3R,4S)-3-Nitro-4-propyl-piperidine-1-carboxylic acid benzyl ester (reduction product of 5h). $[\alpha]_D^{29} = -17.17$ (*c* 0.83, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.39-7.32 (m, 5H), 5.14-5.13 (m, 2H), 4.60-4.47 (m, 1H), 4.27-4.23 (m, 1H), 4.20-4.15 (m, 1H), 3.30-3.15 (m, 1H), 2.86 (t, *J* = 11.2 Hz, 1H), 2.14-2.12 (m, 1H), 1.91 (d, *J* = 13.2 Hz, 1H), 1.43-1.34 (m, 2H), 1.25-1.18 (m, 3H), 0.89 (t, *J* = 6.8 Hz,

3H); ^{13}C NMR (100 MHz, CDCl_3) δ 154.95, 136.31, 128.75, 128.45, 128.24, 87.06, 67.86, 47.01, 43.65, 39.73, 34.22, 28.61, 19.03, 14.06; IR (neat): ν = 2959 s, 2929 s, 1705 s, 1548 s, 1431 s, 1212 s, 1155 m, 1091 m, 973 m, 751 m, 698 m cm^{-1} ; ESI-MS m/z 307.3 ($\text{M} + \text{H}$) $^+$, 329.3 ($\text{M} + \text{Na}$) $^+$, HRMS (ESI) calcd for $\text{C}_{16}\text{H}_{22}\text{N}_2\text{O}_4\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 329.1472, found 329.1472; HPLC: PHENOMENEX LUX AMYLOSE-2 (PA-2) column, hexane/*i*-PrOH = 80:20, flow rate 0.7 mL/min, 214 nm, major isomer: $t_{\text{R}} = 20.1$ min (major), $t_{\text{R}} = 21.7$ min (minor), *ee* = 86%.

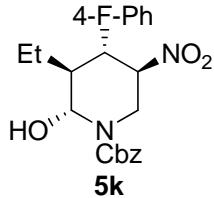


(2R,3S,4S,5R)-3-Ethyl-2-hydroxy-4-methyl-5-nitro-piperidine-1-carboxylic acid benzyl ester 5i. $[\alpha]_{\text{D}}^{29} = -26.8$ (*c* 0.75, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.36-7.32 (m, 5H), 5.74 (d, *J* = 2.0 Hz, 1H), 5.14 (s, 2H), 4.27-4.21 (m, 2H), 3.67 (t, *J* = 12.0 Hz, 1H), 2.45-2.38 (m, 1H), 1.70-1.63 (m, 1H), 1.46-1.38 (m, 1H), 1.26-1.20 (m, 1H), 1.00 (t, *J* = 7.2 Hz, 3H), 0.98 (d, *J* = 6.8 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 155.06, 135.83, 128.81, 128.62, 128.24, 88.71, 74.91, 68.10, 46.04, 41.27, 33.82, 21.50, 15.25, 11.34; IR (neat): ν = 3466 br, 2962 m, 2929 s, 1698 s, 1669 s, 1549 s, 1536 s, 1472 m, 1439 m, 1337 m, 1246 m, 1149 m, 1019 m, 729 m cm^{-1} ; ESI-MS m/z 345.3 ($\text{M} + \text{Na}$) $^+$, HRMS (ESI) calcd for $\text{C}_{16}\text{H}_{22}\text{N}_2\text{O}_5\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 345.1421, found 345.1424; HPLC: PHENOMENEX LUX AMYLOSE-2 (PA-2) column, hexane/*i*-PrOH = 70:30, flow rate 0.7 mL/min, 214 nm, major isomer: $t_{\text{R}} = 10.3$ min (major), $t_{\text{R}} = 8.2$ min (minor), *ee* = 98%. (Noteworthy is that some additional peaks (for isomer) were observed in ^{13}C NMR spectra due to isomerization)

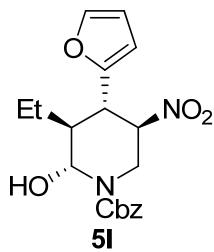


(2R,3S,4R,5R)-3-Ethyl-2-hydroxy-4-phenylpiperidine-1-carboxylic acid benzyl ester 5j. $[\alpha]_{\text{D}}^{25} = +33.0$ (*c* 1.32, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.37-7.32 (m, 5H), 7.30-7.23 (m, 3H), 7.15-7.13 (m, 2H), 5.90 (d, *J* = 1.6 Hz, 1H),

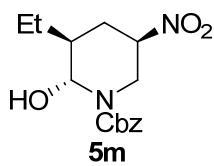
5.17 (s, 2H), 4.76 (dt, J = 4.8, 11.2 Hz, 1H), 4.42 (d, J = 8.0 Hz, 1H), 3.79 (t, J = 12.0 Hz, 1H), 3.48 (t, J = 11.6 Hz, 1H), 1.80-1.74 (m, 1H), 1.38-1.29 (m, 1H), 1.09-1.03 (m, 1H), 0.83 (t, J = 7.2 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 155.10, 137.49, 135.72, 129.08, 128.83, 128.67, 128.28, 128.01(2C), 87.86, 74.95, 68.23, 46.19, 45.65, 41.56, 22.04, 11.22; IR (neat): ν = 3396 br, 2962 w, 1675 s, 1434 s, 1357 m, 1336 m, 1288 m, 1193 m, 1151 m, 1109 m, 965 m, 765 w, 701w cm^{-1} ; ESI-MS m/z 407.4 ($\text{M} + \text{Na}$) $^+$, HRMS (ESI) calcd for $\text{C}_{21}\text{H}_{24}\text{N}_2\text{O}_5\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 407.1577, found 407.1578; HPLC: PHENOMENEX LUX CELLULOSE-2 (PC-2) column, hexane/*i*-PrOH = 70:30, flow rate 0.7 mL/min, 214 nm, major isomer: t_{R} = 11.1 min (major), t_{R} = 6.7 min (minor), *ee* >99%. (Noteworthy is that some additional peaks (for isomer) were observed in ^{13}C NMR spectra due to isomerization)



(3S,4R,5R)-3-Ethyl-4-(4-fluorophenyl)-5-nitro-piperidine-1-carboxylic acid benzyl ester 5k. $[\alpha]_{\text{D}}^{23} = +27.6$ (c 1.43, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.39-7.34 (m, 5H), 7.13-7.10 (m, 2H), 7.02-6.97 (m, 2H), 5.90 (s, 1H), 5.17 (s, 2H), 4.70 (dt, J = 4.4, 11.2 Hz, 1H), 4.43 (d, J = 7.6 Hz, 1H), 3.79 (t, J = 11.6 Hz, 1H), 3.48 (t, J = 11.6 Hz, 1H), 1.72 (tt, J = 2.8, 11.2 Hz, 1H), 1.37-1.29 (m, 1H), 1.07-1.01 (m, 1H), 0.83 (t, J = 7.2 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 162.35 (J = 245.7 Hz), 155.10, 135.63, 133.21 (J = 3.6 Hz), 129.55, 128.83, 128.69, 128.28, 116.07 (J = 21.1 Hz), 87.84, 74.86, 68.28, 45.74, 45.49, 41.49, 21.98, 11.19; ^{19}F NMR (300 MHz, CDCl_3) δ -114.50; IR (neat): ν = 3418 br, 1683 s, 1549 s, 1513 m, 1435 m, 1335 m, 1226 m, 833 m, 733 m cm^{-1} ; ESI-MS m/z 425.2 ($\text{M} + \text{Na}$) $^+$, HRMS (ESI) calcd for $\text{C}_{21}\text{H}_{23}\text{N}_2\text{O}_5\text{FNa}$ ($\text{M} + \text{Na}$) $^+$ m/z 425.1483, found 425.1501; HPLC: PHENOMENEX LUX CELLULOSE-2 (PC-2) column, hexane/*i*-PrOH = 80:20, flow rate 0.7 mL/min, 214 nm, major isomer: t_{R} = 16.7 min (major), t_{R} = 8.6 min (minor), *ee* >99%. (Noteworthy is that some additional peaks (for isomer) were observed in ^{13}C NMR spectra due to isomerization)

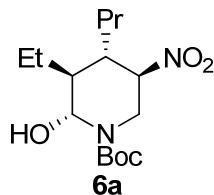


(2R,3S,4S,5R)-3-Ethyl-4-furan-2-yl-2-hydroxy-5-nitro-piperidine-1-carboxylic acid benzyl ester 5l. $[\alpha]_D^{26} = +40.0$ (*c* 1.16, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.40-7.34 (m, 6H), 6.26 (dd, *J* = 2.0, 2.8 Hz, 1H), 6.15 (d, *J* = 2.8 Hz, 1H), 5.86 (d, *J* = 2.0 Hz, 1H), 5.17 (s, 2H), 4.81 (dt, *J* = 4.8, 11.2 Hz, 1H), 4.40 (d, *J* = 11.6 Hz, 1H), 3.76 (t, *J* = 12.0 Hz, 1H), 3.62 (t, *J* = 11.6 Hz, 1H), 1.92-1.85 (m, 1H), 1.42-1.36 (m, 1H), 1.17-1.10 (m, 1H), 0.86 (t, *J* = 7.2 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 155.03, 150.48, 142.62, 135.74, 128.85, 128.68, 128.30, 110.41, 109.54, 85.36, 74.93, 68.25, 43.98, 41.14, 40.14, 22.37, 11.18; IR (neat): ν = 3419 s, 2965 m, 1678 s, 1556 s, 1435 w, 1335 w, 1192 m, 1151 m, 965 m, 883 m, 758 s cm^{-1} ; ESI-MS *m/z* 397.3 ($\text{M} + \text{Na}$) $^+$, HRMS (ESI) calcd for $\text{C}_{19}\text{H}_{22}\text{N}_2\text{O}_6\text{Na}$ ($\text{M} + \text{Na}$) $^+$ *m/z* 397.1370, found 397.1370; HPLC: PHENOMENEX LUX CELLULOSE-2 (PC-2) column, hexane/*i*-PrOH = 70:30, flow rate 0.7 mL/min, 214 nm, major isomer: $t_R = 8.3$ min (major), $t_R = 6.6$ min (minor), *ee*>99%. (Noteworthy is that some additional peaks (for isomer) were observed in ^{13}C NMR spectra due to isomerization)

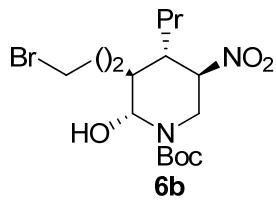


(2R,3S,5R)-3-Ethyl-2-hydroxy-5-nitro-piperidine-1-carboxylic acid benzyl ester 5m. $[\alpha]_D^{29} = +0.71$ (*c* 0.38, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.40-7.33 (m, 5H), 5.87 (s, 1H), 5.16 (s, 2H), 4.43 (tt, *J* = 4.0, 12.0 Hz, 2H), 3.56 (t, *J* = 12.8 Hz, 1H), 2.80-2.60 (br s, 1H), 2.29-2.25 (m, 1H), 2.06 (q, *J* = 12.4 Hz, 1H), 1.58-1.55 (m, 2H), 1.42-1.38 (m, 1H), 0.98 (t, *J* = 7.2 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 155.26, 135.92, 128.84, 128.64, 128.35, 80.25, 75.69, 68.11, 40.92, 40.59, 29.31, 24.74, 11.38; IR (neat): ν = 3418 br, 2963 m, 2924 m, 1701s, 1545 s, 1328 m, 1268 m, 1164 m, 1004 m, 958 m, 901 m, 751 m, 698 m cm^{-1} ; ESI-MS *m/z* 331.3 ($\text{M} + \text{Na}$) $^+$, HRMS

(ESI) calcd for $C_{15}H_{20}N_2O_5Na$ ($M + Na$)⁺ m/z 331.1264, found 331.1270; HPLC: PHENOMENEX LUX AMYLOSE-2 (PA-2) column, hexane/*i*-PrOH = 70:30, flow rate 0.5 mL/min, 214 nm, major isomer: t_R = 10.0 min (major), t_R = 12.0 min (minor), *ee* = 79%. (Noteworthy is that some additional peaks (for isomer) were observed in ¹³C NMR spectra due to isomerization)

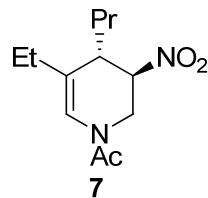


(2R,3S,4S,5R)-3-Ethyl-2-hydroxy-5-nitro-4-propyl-piperidine-1-carboxylic acid *tert*-butyl ester 6a. $[\alpha]_D^{27} = -18.4$ (*c* 1.88, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 5.69 (s, 1H), 4.46 (dt, *J* = 4.8, 10.8 Hz, 1H), 4.10 (d, *J* = 10.4 Hz, 1H), 3.70 (t, *J* = 10.8 Hz, 1H), 3.27 (brs, 1H), 2.44-2.39 (m, 1H), 1.62-1.56 (m, 1H), 1.47 (s, 9H), 1.43-1.22 (m, 6H), 1.01 (t, *J* = 6.8 Hz, 3H), 0.89 (t, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 154.43, 85.69, 81.72, 74.35, 42.75, 41.37, 37.66, 30.26, 28.39, 21.28, 17.19, 14.49, 11.32; IR (neat): ν = 3419 s, 2962 m, 2932 m, 1683 s, 1557 m, 1540 m, 1430 m, 1387 w, 1162 s, 1007 w, 884 w cm⁻¹; ESI-MS m/z 339.3 (M + Na)⁺, HRMS (ESI) calcd for $C_{15}H_{28}N_2O_3Na$ ($M + Na$)⁺ m/z 339.1890, found 339.1891; HPLC: sino-AD column, hexane/*i*-PrOH = 90:10, flow rate 0.7 mL/min, 214 nm, major isomer: t_R = 7.9 min (major), t_R = 6.5 min (minor), *ee*>99%. (Noteworthy is that some additional peaks (for isomer) were observed in ¹³C NMR spectra due to isomerization)

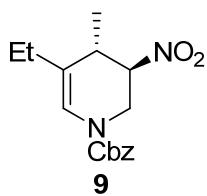


(2R,3S,4S,5R)-3-(3-Bromo-propyl)-2-hydroxy-5-nitro-4-propyl-piperidine-1-carboxylic acid *tert*-butyl ester 6b. $[\alpha]_D^{29} = -27.7$ (*c* 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 5.61 (s, 1H), 4.47 (dt, *J* = 4.4, 10.4 Hz, 1H), 4.09 (d, *J* = 10.0 Hz, 1H), 3.69 (t, *J* = 12.4 Hz, 1H), 3.46-3.42 (br, 1H), 3.44 (t, *J* = 6.4 Hz, 2H), 2.47-2.42 (m, 1H), 2.11-2.03 (m, 1H), 1.91-1.83 (m, 1H), 1.72-1.66 (m, 1H), 1.62-1.54 (m, 1H), 1.51-1.46 (m, 1H), 1.46 (s, 9H), 1.37-1.23 (m, 4H), 0.91 (t, *J* = 6.8 Hz, 3H); ¹³C NMR

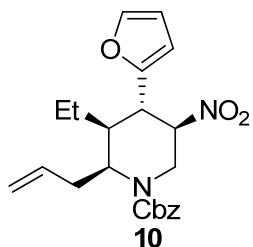
(100 MHz, CDCl_3) δ 154.37, 85.40, 81.94, 74.48, 41.23, 40.50, 37.45, 33.64, 30.21, 29.70, 28.37, 26.93, 17.14, 14.47; IR (neat): ν = 3445 br, 2959 m, 2932 m, 1678 s, 1557 m, 1538 m, 1472 m, 1389 m, 1165 m, 1006 w, 917 w cm^{-1} ; ESI-MS m/z 431.4 ($\text{M} + \text{Na}$) $^+$, HRMS (ESI) calcd for $\text{C}_{16}\text{H}_{29}\text{BrN}_2\text{O}_5\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 431.1152, found 431.1156; HPLC: Chiralpak IC column, hexane/*i*-PrOH = 98:2, flow rate 0.7 mL/min, 214 nm, major isomer: t_R = 24.1 min (major), t_R = 19.3 min (minor), *ee*>99%. (Noteworthy is that some additional peaks (for isomer) were observed in ^{13}C NMR spectra due to isomerization)



1-((3R,4S)-5-Ethyl-3-nitro-4-propyl-3,4-dihydro-2H-pyridin-1-yl)-ethanone 7. $[\alpha]_D^{26} = +11.8$ (c 1.0, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 6.35 & 7.02 (both s, due to rotamers, 1H), 4.76 & 4.47 (ddd, J = 1.6, 3.2, 15.2 Hz, & dd, J = 3.2, 13.2 Hz, due to rotamers, 1H), 4.67 & 4.59 (q, J = 3.2 Hz & q, J = 2.8 Hz, due to rotamers, 1H), 3.51 & 3.67 (dd, J = 3.2, 14.8 Hz & dd, J = 3.2, 13.6 Hz, due to rotamers, 1H), 2.96 & 3.02 (t, J = 4.4 Hz & t, J = 4.4 Hz, due to rotamers, 1H), 2.19 & 2.23 (both s, due to rotamers, 3H), 2.09 (q, J = 7.6 Hz, 2H), 1.69-1.60 (m, 2H), 1.46-1.35 (m, 2H), 1.04 (t, J = 7.6 Hz, 3H), 0.98 (t, J = 6.8 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 168.32 & 167.59 (due to rotamers), 121.27 & 121.70 (due to rotamers), 119.86 & 118.29 (due to rotamers), 80.83 & 80.89 (due to rotamers), 38.85 & 42.26 (due to rotamers), 38.44 & 37.58 (due to rotamers), 34.74 & 34.93 (due to rotamers), 26.03 & 25.81 (due to rotamers), 21.60 & 22.00 (due to rotamers), 19.98 & 20.04 (due to rotamers), 14.05, 12.57 & 12.55 (due to rotamers); IR (neat): ν = 3418 m, 2964 s, 2934 s, 2875 m, 1652 s, 1549 s, 1410 m, 1316 m, 1210 m, 1179 m, 1017 m, 740 m, 611 m cm^{-1} ; ESI-MS m/z 241.2 ($\text{M} + \text{H}$) $^+$, HRMS (ESI) calcd for $\text{C}_{12}\text{H}_{20}\text{N}_2\text{O}_3\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 263.1366, found 263.1372; HPLC: PHENOMENEX LUX AMYLOSE-2 (PA-2) column, hexane/*i*-PrOH = 70:30, flow rate 0.7 mL/min, 214 nm, t_R = 15.2 min (major), t_R = 21.8 min (minor), *ee*=93%.



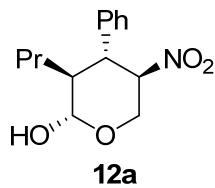
(3R,4S)-5-Ethyl-4-methyl-3-nitro-3,4-dihydro-2H-pyridine-1-carboxylic acid benzyl ester 9. To a solution of **4i** (32 mg, 0.10 mol) in 2 mL CH_2Cl_2 was added several drops of TFA at rt. The reation mixture was stirred at this temperature until **4i** was consumed as monitored by TLC. Direct concentration of the reaction mixture gave **9** (30 mg, 100%), which is pure enough for analysis. $[\alpha]_D^{23} = +26.5$ (*c* 0.83, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.38-7.33 (m, 5H), 6.69 & 6.58 (both s, due to rotamers, 1H), 5.26-5.16 (m, 2H), 4.50-4.39 (m, 2H), 3.81-3.76 (m, 1H), 3.04 (s, 1H), 2.10-2.02 (m, 2H), 1.17 (d, *J* = 7.2 Hz, 3H), 1.04 & 1.02 (both t, *J* = 7.6 Hz, due to rotamers, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 153.00 & 152.95 (due to rotamers), 136.05, 128.74, 128.47, 128.26, 120.93 & 120.51, 119.00 & 118.65 (due to rotamers), 83.82 & 83.86 (due to rotamers), 68.07 & 68.12 (due to rotamers), 40.98 & 40.88 (due to rotamers), 33.09 & 33.27 (due to rotamers), 25.37, 18.31 & 18.23 (due to rotamers), 12.54 & 12.67 (due to rotamers); IR (neat): ν = 2918 m, 1708 s, 1546 m, 1215 m, 1139 m, 1029 m, 977 m, 936 m, 692 m, 600 m cm^{-1} ; ESI-MS *m/z* 327.2 ($\text{M} + \text{Na}$) $^+$, HRMS (ESI) calcd for $\text{C}_{16}\text{H}_{20}\text{N}_2\text{O}_4\text{Na}$ ($\text{M} + \text{Na}$) $^+$ *m/z* 327.1315, found 327.1322.



(2S,3S,4S,5R)-2-Allyl-3-ethyl-4-furan-2-yl-5-nitro-piperidine-1-carboxylic acid benzyl ester 10. To a solution of **4l** (37 mg, 0.10 mmol) and allyl trimethylsilane (60 μL , 0.5 mmol) in CH_2Cl_2 (2 ml) was added $\text{BF}_3\cdot\text{OEt}_2$ (25 μL , 0.2 mmol) at -78 °C. The reation mixture was stirred at this temperature until **4l** was consumed as monitored by TLC. Purification by flash column chromatography (silica gel,

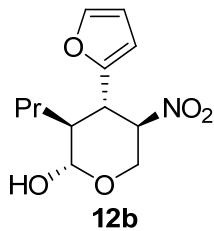
petroleum ether/AcOEt 20:1 to 15:1) afforded 37 mg (93%) of **10**. $[\alpha]_D^{23} = +31.6$ (*c* 0.93, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.39-7.32 (m, 6H), 6.29-6.28 (m, 1H), 6.17 (d, *J* = 2.8 Hz, 1H), 5.82-5.72 (m, 1H), 5.17-5.09 (m, 2H), 5.05 (d, *J* = 3.6 Hz, 1H), 5.02 (s, 1H), 4.91-4.88 (m, 1H), 4.60 (d, *J* = 15.6 Hz, 1H), 4.15 (m, 1H), 3.55 (dd, *J* = 6.4, 15.6 Hz, 1H), 3.47 (dd, *J* = 9.2, 11.6 Hz, 1H), 2.46-2.42 (m, 1H), 2.29-2.21 (m, 1H), 1.86-1.79 (m, 1H), 1.44-1.35 (m, 1H), 1.27-1.21 (m, 1H), 0.72 (t, *J* = 7.6 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 155.74, 151.28, 142.57, 136.34, 133.86, 128.64, 128.27, 128.19, 118.40, 110.64, 109.03, 86.06, 67.82, 54.96, 42.19, 39.77, 39.69, 36.95, 23.53, 9.60; IR (neat): ν = 2964 m, 2933 m, 1703 s, 1552 s, 1424 s, 1243 s, 1114 m, 1011 m, 925 m, 737 s, 598 m cm^{-1} ; ESI-MS *m/z* 399.2 ($\text{M} + \text{H}$) $^+$, 421.3 ($\text{M} + \text{Na}$) $^+$, HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{26}\text{N}_2\text{O}_5\text{Na}$ ($\text{M} + \text{Na}$) $^+ m/z$ 421.1734, found 421.1738.

General procedure for cascade Michael-acetalization process of aldehydes with hydroxymethyl substituted nitroolefins: Valeraldehyde (0.40 mmol) was added to a suspension of catalyst **1** (20 mol %), hydroxymethyl substituted nitroolefins **11** (0.20 mmol) and benzoic acid (60 mol %) in water (0.40 mL) at room temperature, and stirred until complete conversion of the starting materials (monitored by TLC). The reaction mixture was diluted with ethyl acetate, dried over Na_2SO_4 and concentrated. Purification by flash column chromatography afforded **12**. The enantiomeric excess (ee) was determined by HPLC on a chiral phase.



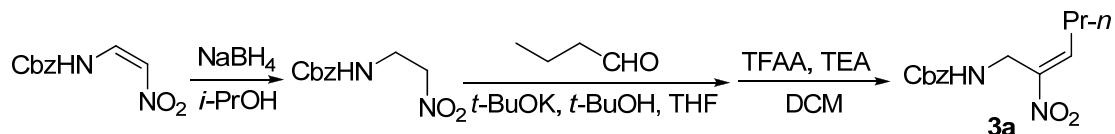
(2S,3S,4R,5R)-5-Nitro-4-phenyl-3-propyl-tetrahydro-pyran-2-ol 12a. $[\alpha]_D^{28} = -8.43$ (*c* 0.42, MeOH); ^1H NMR (400 MHz, CDCl_3) δ 7.34-7.24 (m, 3H), 7.20-7.18 (m, 2H), 5.31 (s, 1H), 4.87 (dq, *J* = 4.8, 11.2 Hz, 1H), 4.43 (t, *J* = 10.8 Hz, 1H), 4.04 (dd, *J* = 4.8, 10.4 Hz, 1H), 3.52 (t, *J* = 11.6 Hz, 1H), 2.68 (brs, 1H), 2.03-1.98 (m, 1H), 1.36-1.27 (m, 2H), 1.11-1.04 (m, 1H), 0.97-0.90 (m, 1H), 0.75 (t, *J* = 7.2 Hz, 3H); ^1H

NMR (400 MHz, MeOD) δ 7.35-7.32 (m, 2H), 7.28-7.25 (m, 3H), 5.23 (d, J = 2.8 Hz, 1H), 5.03 (dt, J = 5.2, 11.2 Hz, 1H), 4.40 (t, J = 10.4 Hz, 1H), 4.00 (dd, J = 4.8, 10.4 Hz, 1H), 3.48 (t, J = 12.0 Hz, 1H), 2.06-2.01 (m, 1H), 1.38-1.25 (m, 2H), 1.11-1.03 (m, 1H), 0.93-0.84 (m, 1H), 0.74 (t, J = 7.6 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 137.55, 129.39, 129.07, 128.10, 127.97, 92.20, 87.76, 60.36, 45.35, 43.35, 30.90, 19.71, 14.13; ^{13}C NMR (100 MHz, MeOD) δ 139.59, 129.76, 128.63, 92.84, 89.11, 61.14, 47.00, 44.81, 32.29, 20.60, 14.43; IR (neat): ν = 3419 br, 2950 m, 1542 s, 1455m, 1352 m, 1059 m, 1033 m, 930 m, 692 m cm^{-1} ; EI-MS m/z 265 M^+ , HRMS (EI) calcd for $\text{C}_{14}\text{H}_{19}\text{NO}_4$ M^+ m/z 265.1314, found 265.1319; HPLC: IC column, hexane/*i*-PrOH = 90:10, flow rate 0.7 mL/min, 214 nm, t_{R} = 8.3 min (major), t_{R} = 7.5 min (minor), *ee*>99%. (Noteworthy is that some additional peaks (for isomer) were observed in ^{13}C NMR spectra due to isomerization)



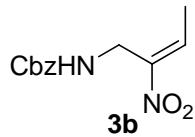
(2S,3S,4S,5R)-4-Furan-2-yl-5-nitro-3-propyl-tetrahydro-pyran-2-ol 12b. $[\alpha]_D^{28}$ = -0.01 (*c* 1.96, MeOH); ^1H NMR (400 MHz, CDCl_3) δ 7.37-7.36 (m, 1H), 6.30-6.26 (m, 1H), 6.18-6.16 (m, 1H), 5.27 (d, J = 2.8 Hz, 1H), 4.94 (dt, J = 4.8, 11.2 Hz, 1H), 4.37 (t, J = 10.8 Hz, 1H), 4.02 (dd, J = 4.8, 10.4 Hz, 2H), 3.65 (t, J = 11.6 Hz, 1H), 2.13-2.07 (m, 1H), 1.40-1.25 (m, 2H), 1.14-0.97 (m, 2H), 0.81 (t, J = 6.8 Hz, 3H); ^1H NMR (400 MHz, MeOD) δ 7.44-7.42 (m, 1H), 6.32-6.30 (m, 1H), 6.19-6.18 (m, 1H), 5.18 (d, J = 2.8 Hz, 1H), 4.96 (dt, J = 5.2, 11.2 Hz, 1H), 4.34 (t, J = 10.8 Hz, 1H), 3.97 (dd, J = 4.8, 10.4 Hz, 1H), 3.60 (t, J = 11.2 Hz, 1H), 2.07-2.01 (m, 1H), 1.39-1.28 (m, 2H), 1.10-1.03 (m, 1H), 1.01-0.93 (m, 1H), 0.71 (t, J = 6.8 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) 150.55, 142.51, 110.41, 109.35, 92.04, 85.20, 59.97, 41.74, 39.36, 31.30, 19.68, 14.20; ^{13}C NMR (100 MHz, MeOD) δ 152.66, 143.45, 111.20, 109.63, 92.65, 86.71, 60.62, 43.52, 40.82, 32.58, 20.54, 14.48; IR (neat): ν = 3396 br, 2954 m, 2873 w, 1541 s, 1466 w, 1357 w, 1277 w, 1152 m, 1032 w, 1009 m, 928 m, 908 w, 726 m cm^{-1} ; EI-MS m/z 255 M^+ , HRMS (EI) calcd for $\text{C}_{12}\text{H}_{17}\text{NO}_5$ M^+ m/z

255.1107, found 255.1113; HPLC: AD-H column, hexane/*i*-PrOH = 95:5, flow rate 0.7 mL/min, 214 nm, major isomer: t_R = 28.0 min (major), t_R = 48.5 min (minor), $ee > 99\%$. (Noteworthy is that some additional peaks (for isomer) were observed in ^{13}C NMR spectra due to isomerization)

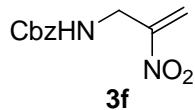


Typical procedure for assembly of 1-protected aminomethyl 2-alkyl substituted nitroalkenes 3a. To a solution of ((*Z*)-2-nitrovinyl)-carbamic acid benzyl ester (2.24 g, 10 mmol) in 50 mL of *i*-PrOH were added NaBH₄ (760 mg, 2.0 eq, 20 mmol) at 0 °C. The resultant solution was warmed to rt. After stirring at rt for 2 h, 1 mL acetone and 20 mL saturate NH₄Cl aqueous solution was added slowly, the aqueous phase was extracted with ethyl acetate. The combined organic layers were washed with brine and dried over Na₂SO₄ and concentrated to give reduced product as light yellow solid, which was dissolved in 10 mL *t*-BuOH and 10 mL THF. To this solution was added *n*-butylaldehyde (15 mmol, 1.5 equiv.) and *t*-BuOK (112 mg, 1 mmol, 0.1 equiv.) at 0 °C. The resultant solution was warmed to rt. After stirring at rt for 5 h, the reaction mixture was diluted with ethyl ether and washed with water, the aqueous phase was extracted with ethyl ether three times. The combined organic phase was washed with brine and dried over Na₂SO₄ and concentrated to give condensation product, which was dissolved in 20 mL CH₂Cl₂. To this solution was added TEA (20 mmol, 2 equiv.) and TFAA (15 mmol) slowly at -20 °C. After stirring at -20 °C for 1.5 h, the reaction mixture was diluted with CH₂Cl₂ and washed with water. The aqueous phase was extracted with CH₂Cl₂ three times. The combined organic phase was washed with brine and dried over Na₂SO₄ and concentrated. Purification of the residue by flash column chromatography (silica gel, 10:1 petroleum ether/ethyl acetate as eluent) afforded the corresponding (*E*)-2-nitro-hex-2-enylcarbamic acid benzyl ester **3a** (2.06 g, 74% for 3 steps). ¹H NMR (400 MHz, CDCl₃) δ 7.33-7.24 (m, 5H), 5.47 (brs, 1H), 5.08 (s, 2H), 4.28 (d, *J* = 6.4 Hz, 2H), 2.50-2.45 (m, 2H), 1.58-1.53 (m, 2H), 0.98 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 156.17, 148.20, 140.87,

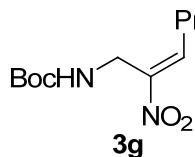
136.25, 128.64, 128.32, 128.15, 67.11, 36.36, 29.96, 21.89, 13.79; IR (neat): ν = 3334 br, 2962 m, 1714 s, 1520 s, 1332 m, 1238 s, 1136 s, 738 m, 698 m cm^{-1} ; ESI-MS m/z 279.2 ($\text{M} + \text{H}$) $^+$, 301.2 ($\text{M} + \text{Na}$) $^+$, HRMS (ESI) calcd for $\text{C}_{14}\text{H}_{18}\text{N}_2\text{O}_4\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 301.1159, found 301.1164.



((E)-2-Nitrobut-2-enyl)carbamic acid benzyl ester 3b. 49% yield for three steps. ^1H NMR (400 MHz, CDCl_3) δ 7.28-7.24 (m, 6H), 5.45 (brs, 1H), 4.99 (s, 2H), 4.18 (d, J = 6.4 Hz, 2H), 2.01 (d, J = 7.6 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 156.27, 148.85, 136.34, 136.15, 128.58, 128.26, 128.08, 67.06, 36.01, 13.74; IR (neat): ν = 3333 br, 2956 m, 1714 s, 1515 s, 1341 m, 1241 s, 1137 m, 1080 m, 975 m, 846 m, 777 m, 698 m cm^{-1} ; ESI-MS m/z 273.3 ($\text{M} + \text{Na}$) $^+$, HRMS (ESI) calcd for $\text{C}_{12}\text{H}_{14}\text{N}_2\text{O}_4\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 273.0846, found 273.0846.

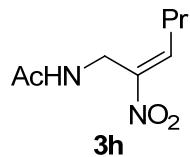


(2-Nitroallyl)carbamic acid benzyl ester 3f. 40% for three steps. ^1H NMR (400 MHz, CDCl_3) δ 7.38-7.32 (m, 5H), 6.58 (s, 1H), 5.92 (s, 1H), 5.38 (brs, 1H), 5.11 (s, 2H), 4.27 (d, J = 6.4 Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 156.22, 153.90, 136.10, 128.76, 128.52, 128.34, 120.80, 67.41, 40.50; ESI-MS m/z 236.1 ($\text{M} + \text{H}$) $^+$, 259.1 ($\text{M} + \text{Na}$) $^+$.

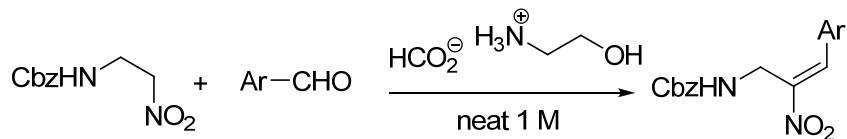


((E)-2-Nitrohex-2-enyl)carbamic acid *tert*-butyl ester 3g. Compound 3g was prepared from ((Z)-2-nitrovinyl)carbamic acid *tert*-butyl ester and *n*-butylaldehyde in 59% overall yield according to typical procedure. ^1H NMR (400 MHz, CDCl_3) δ 7.24 (t, J = 8.0 Hz, 1H), 5.19 (brs, 1H), 4.23 (d, J = 6.0 Hz, 2H), 2.48 (q, J = 7.6 Hz, 2H), 1.61-1.52 (m, 2H), 1.42 (s, 9H), 0.98 (t, J = 7.6 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 155.50, 148.78, 140.47, 79.99, 35.88, 29.93, 28.40, 21.92, 13.72; IR (neat):

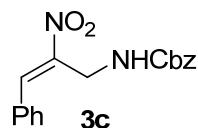
$\nu = 3419$ br, 2966 s, 2935 m, 1716 s, 1526 s, 1367 m, 1332 m, 1249 m, 1167 s, 932 w, 857 w cm^{-1} ; ESI-MS m/z 267.3 ($\text{M} + \text{Na}$) $^+$, HRMS (ESI) calcd for $\text{C}_{11}\text{H}_{20}\text{N}_2\text{O}_4\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 267.1315, found 267.1319.



***N*-(*(E*)-2-Nitrohex-2-enyl)acetamide **3h**.** Compound **3h** was prepared from *N*-(*(Z*)-2-nitrovinyl)acetamide and *n*-butylaldehyde in 50% overall yield according to typical procedure. ^1H NMR (400 MHz, CDCl_3) δ 7.26 (t, $J = 8.0$ Hz, 1H), 6.30 (br s, 1H), 4.35 (d, $J = 6.4$ Hz, 2H), 2.50 (q, $J = 7.6$ Hz, 2H), 1.98 (s, 3H), 1.61-1.52 (m, 2H), 0.99 (t, $J = 7.6$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 170.18, 148.03, 140.95, 34.65, 29.93, 22.93, 21.80, 13.73; IR (neat): $\nu = 3255$ s, 2962 m, 1668 m, 1644 m, 1516 s, 1338 m, 1288 m, 1095 w, 1075 w, 748 w cm^{-1} ; ESI-MS m/z 187.1 ($\text{M} + \text{H}$) $^+$, 219.2 ($\text{M} + \text{Na}$) $^+$, HRMS (MALDI) calcd for $\text{C}_8\text{H}_{14}\text{N}_2\text{O}_3\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 209.0897, found 209.0897.

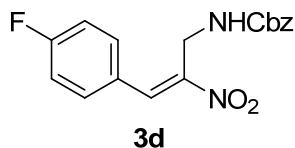


Typical procedure for assembly of 1-protected aminomethyl 2-aryl substituted nitroalkenes. A mixture of (2-nitroethyl)carbamic acid benzyl ester (1 mmol) and arylaldehyde (1 mmol) in 1 mL 2-HEAF was stirred overnight at rt until the starting material disappeared. The mixture was partitioned between water and ethyl acetate. The aqueous phase was extracted with ethyl acetate. The combined organic phase was washed with brine, dried over Na_2SO_4 and concentrated. Purification of the residue by flash column chromatography (silica gel, 15:1 to 10:1 petroleum ether/ethyl acetate as eluent) afforded the corresponding nitroalkene.

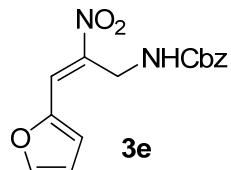


(*E*)-2-Nitro-3-phenylallyl carbamic acid benzyl ester **3c.** 85% yield. ^1H NMR

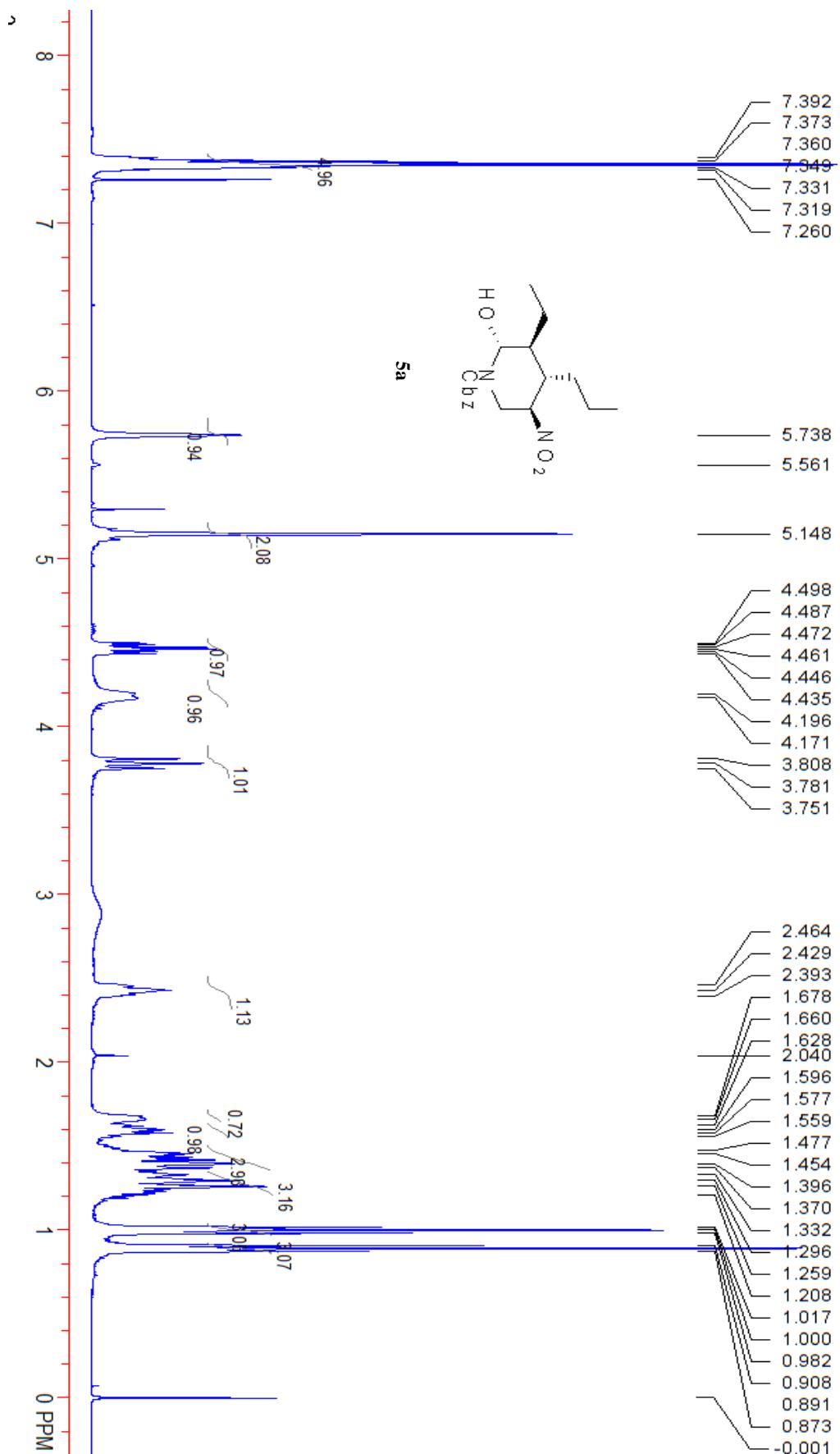
(400 MHz, CDCl_3) δ 8.17 (s, 1H), 7.72-7.66 (m, 2H), 7.52-7.45 (m, 3H), 7.40-7.30 (m, 5H), 5.50 (brs, 1H), 5.12 (s, 2H), 4.58 (d, J = 6.0 Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 156.20, 147.27, 137.21, 136.20, 131.10, 131.00, 130.48, 129.34, 128.70, 128.40, 128.24, 67.29, 37.79; ESI-MS m/z 335.3 ($\text{M} + \text{Na}$) $^+$, HRMS (MALDI) calcd for $\text{C}_{17}\text{H}_{16}\text{N}_2\text{O}_4\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 335.1002, found 335.1004.

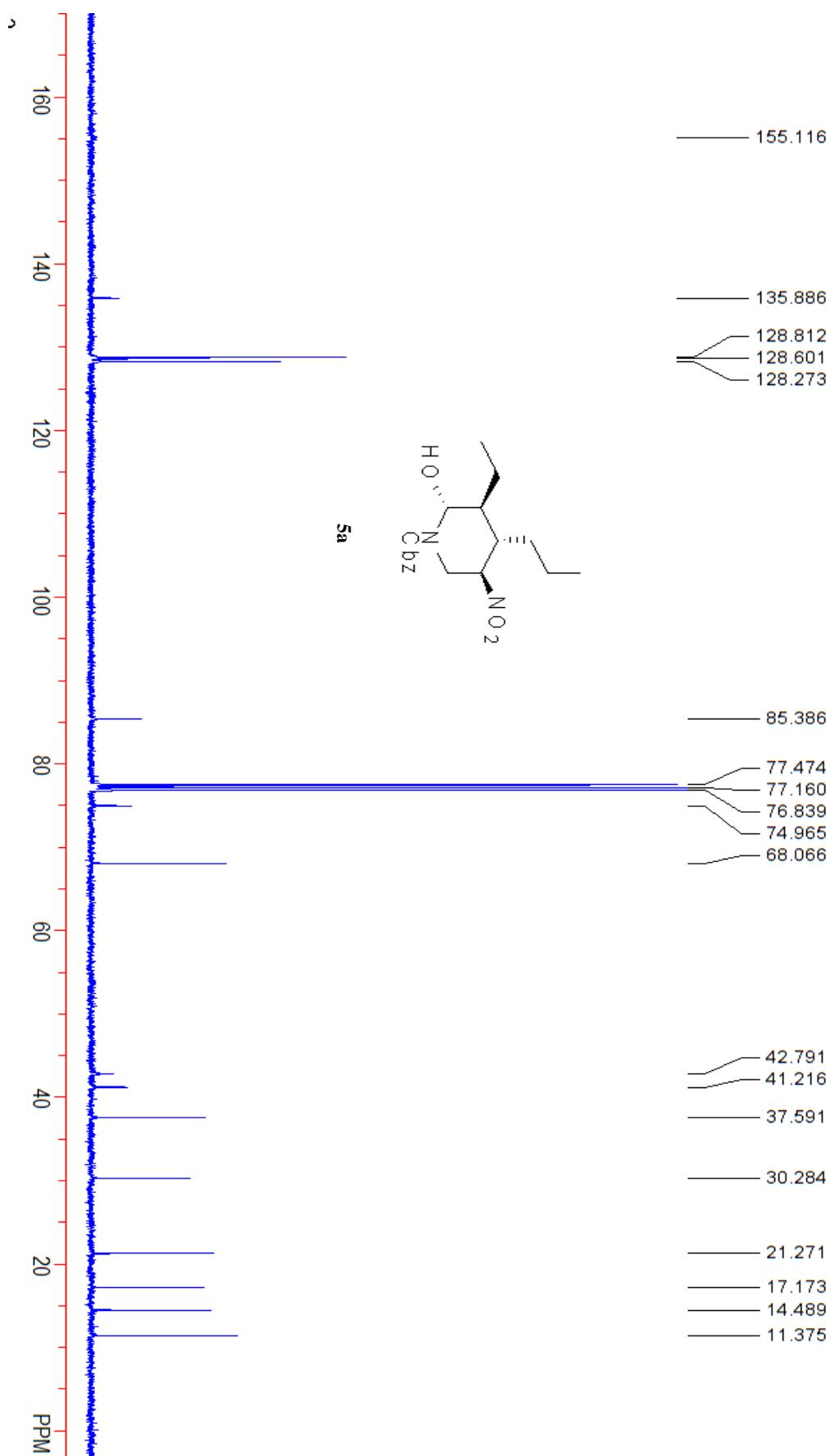


((E)-3-(4-Fluorophenyl)-2-nitroallyl)carbamic acid phenethyl ester 3d. 83% yield, ^1H NMR (400 MHz, CDCl_3) δ 8.12 (s, 1H), 7.78-7.70 (m, 2H), 7.38-7.30 (m, 5H), 7.19-7.15 (m, 2H), 5.55 (brs, 1H), 5.12 (s, 2H), 5.54 (d, J = 6.0 Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 164.21 (d, J = 252.4 Hz), 156.20, 146.96, 136.06, 135.92, 132.78 (d, J = 8.0 Hz), 128.62, 128.24 (d, J = 20.0 Hz), 127.03 (d, J = 3.7 Hz), 116.55 (d, J = 21.8 Hz), 67.25, 37.64; IR (neat): ν = 3249 w, 3144 w, 1728 s, 1600 m, 1516 m, 1407 m, 1347 m, 1242 m, 1033 m, 858 w, 755 w, 696 w cm^{-1} ; ESI-MS m/z 353.3 ($\text{M} + \text{Na}$) $^+$, HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{15}\text{FN}_2\text{O}_4\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 353.0908, found 353.0907.



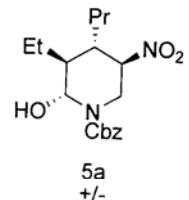
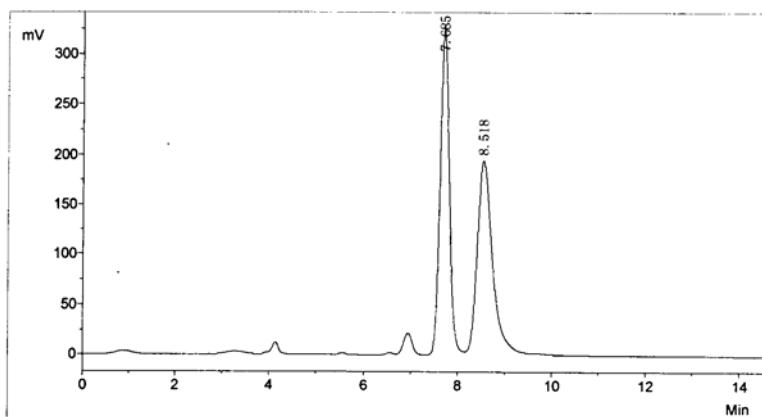
((E)-3-Furan-2-yl-2-nitroallyl)carbamic acid benzyl ester 3e. 86% yield. ^1H NMR (400 MHz, CDCl_3) δ 7.91 (s, 1H), 7.73 (s, 1H), 7.40-7.27 (m, 5H), 7.13 (s, 1H), 6.62 (s, 1H), 5.44 (brs, 1H), 5.09 (s, 2H), 4.83 (d, J = 6.4 Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 156.22, 147.75, 146.82, 143.49, 136.24, 128.59, 128.26, 128.16, 123.09, 121.88, 113.43, 67.08, 37.63; IR (neat): ν = 3318 s, 2922 s, 2851 m, 1686 s, 1498 m, 1311 m, 1269 m, 981 w, 770 m, 699 w cm^{-1} ; ESI-MS m/z 325.3 ($\text{M} + \text{Na}$) $^+$, HRMS (ESI) calcd for $\text{C}_{15}\text{H}_{14}\text{N}_2\text{O}_5\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 325.0795, found 325.0799.





HPLC REPORT

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Date:2010-06-01 Time :09:21
Operator:



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HPLC REPORT

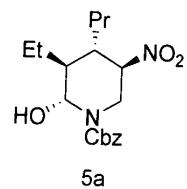
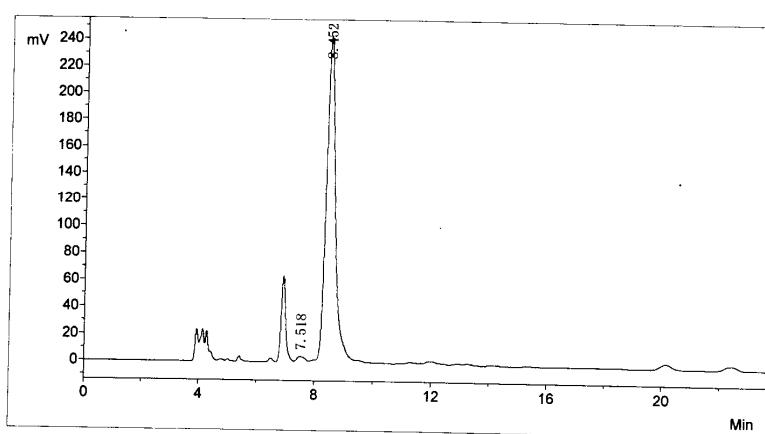
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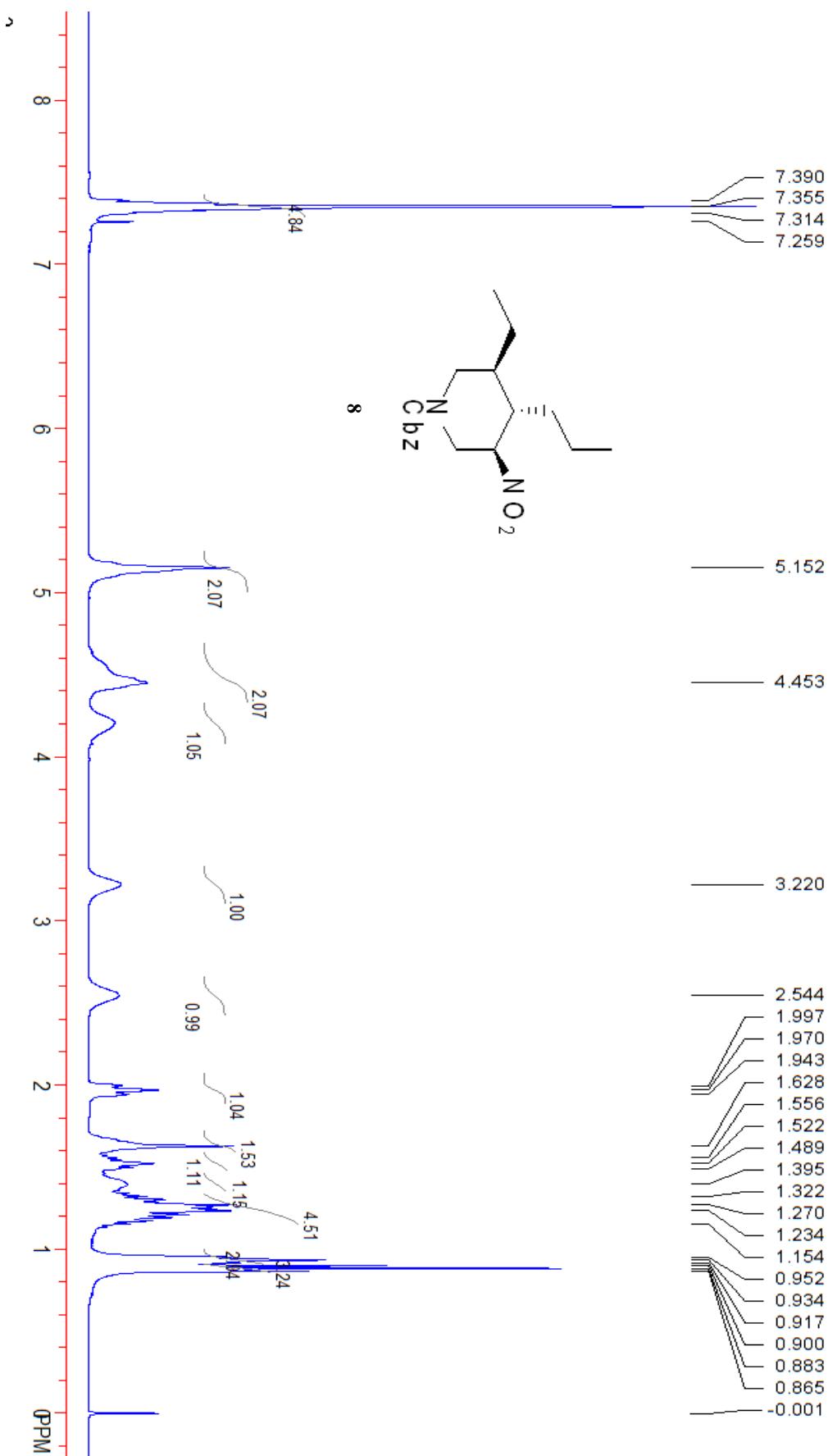
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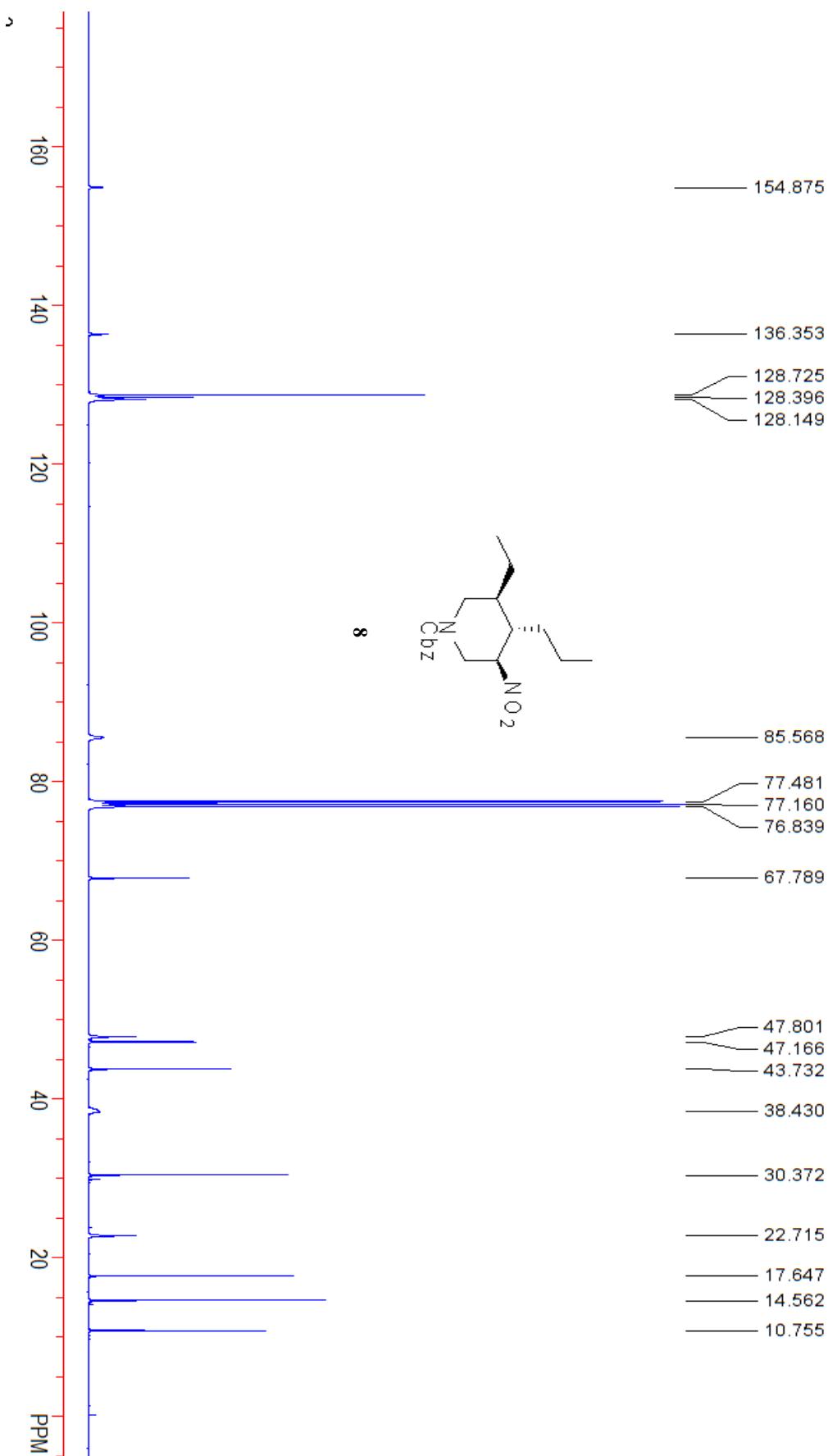
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5a

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HPLC Report

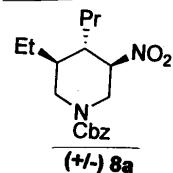
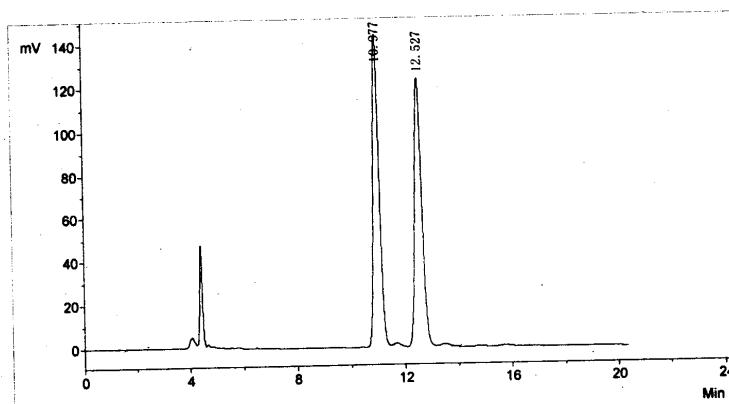
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Date:2010-07-07

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HPLC Report

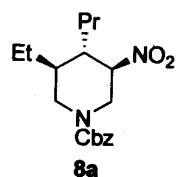
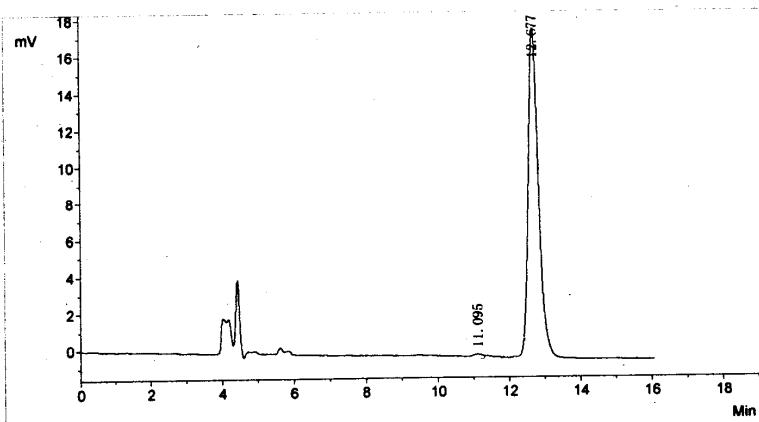
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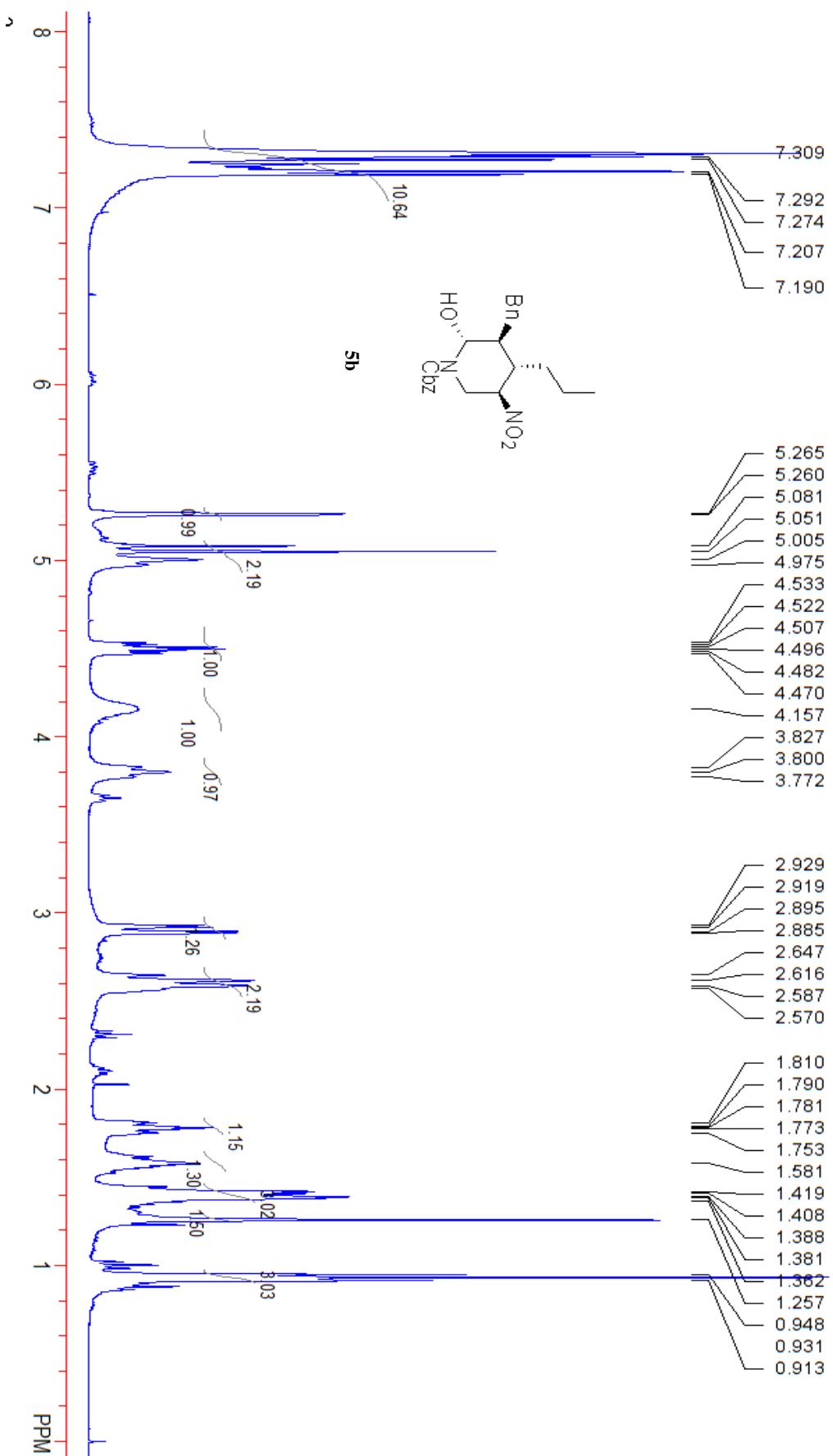
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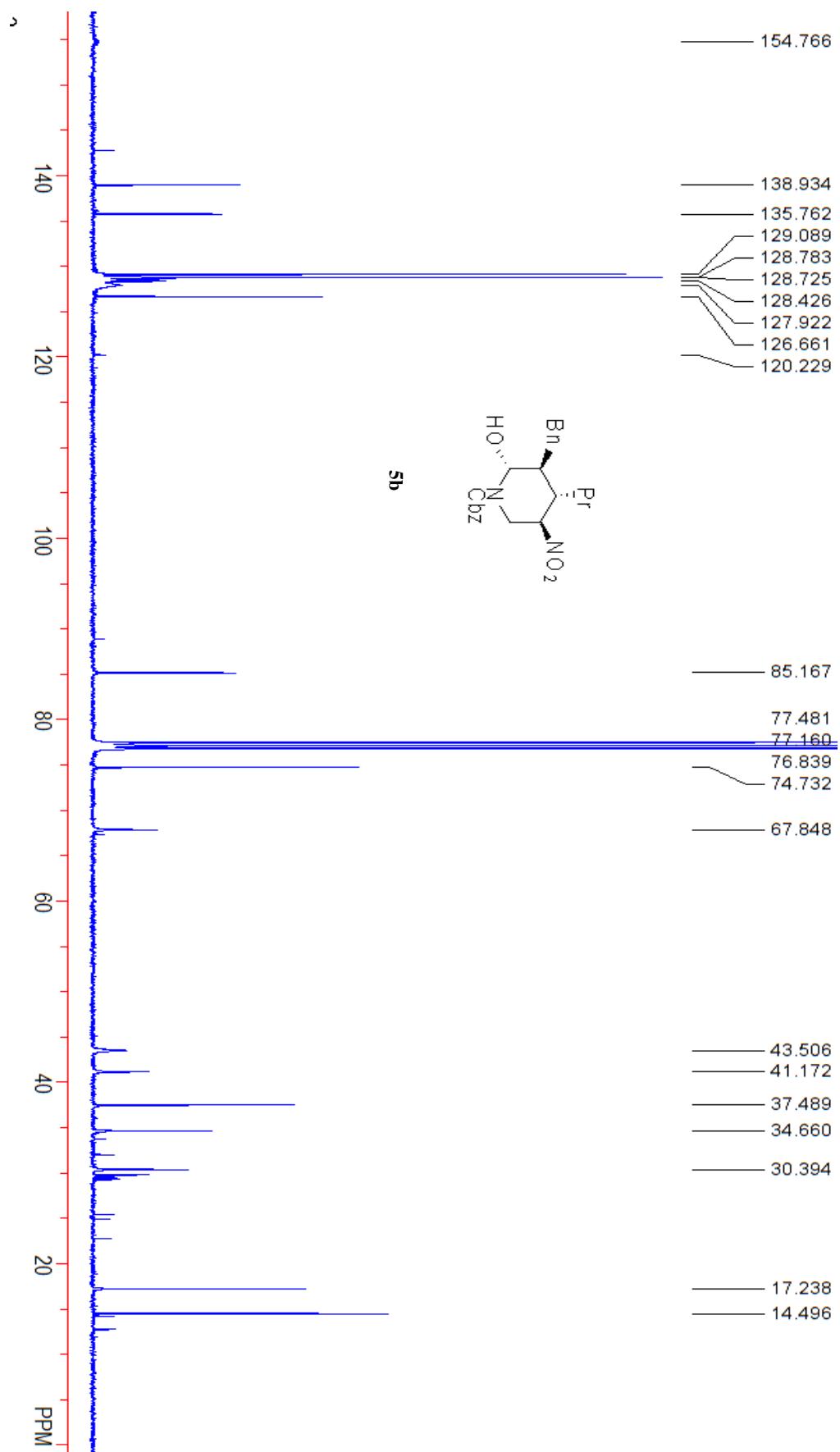
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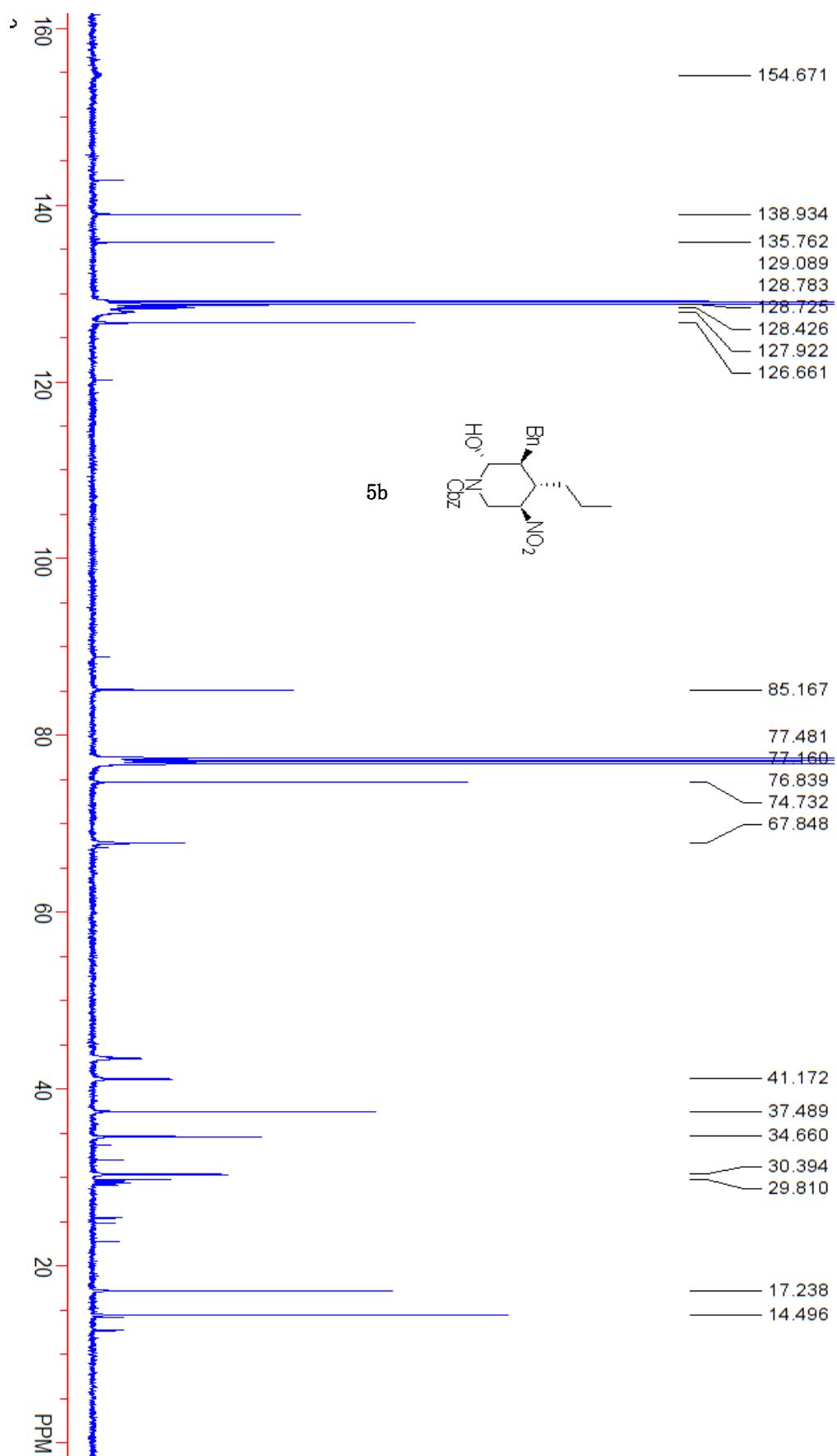
Time:14:15

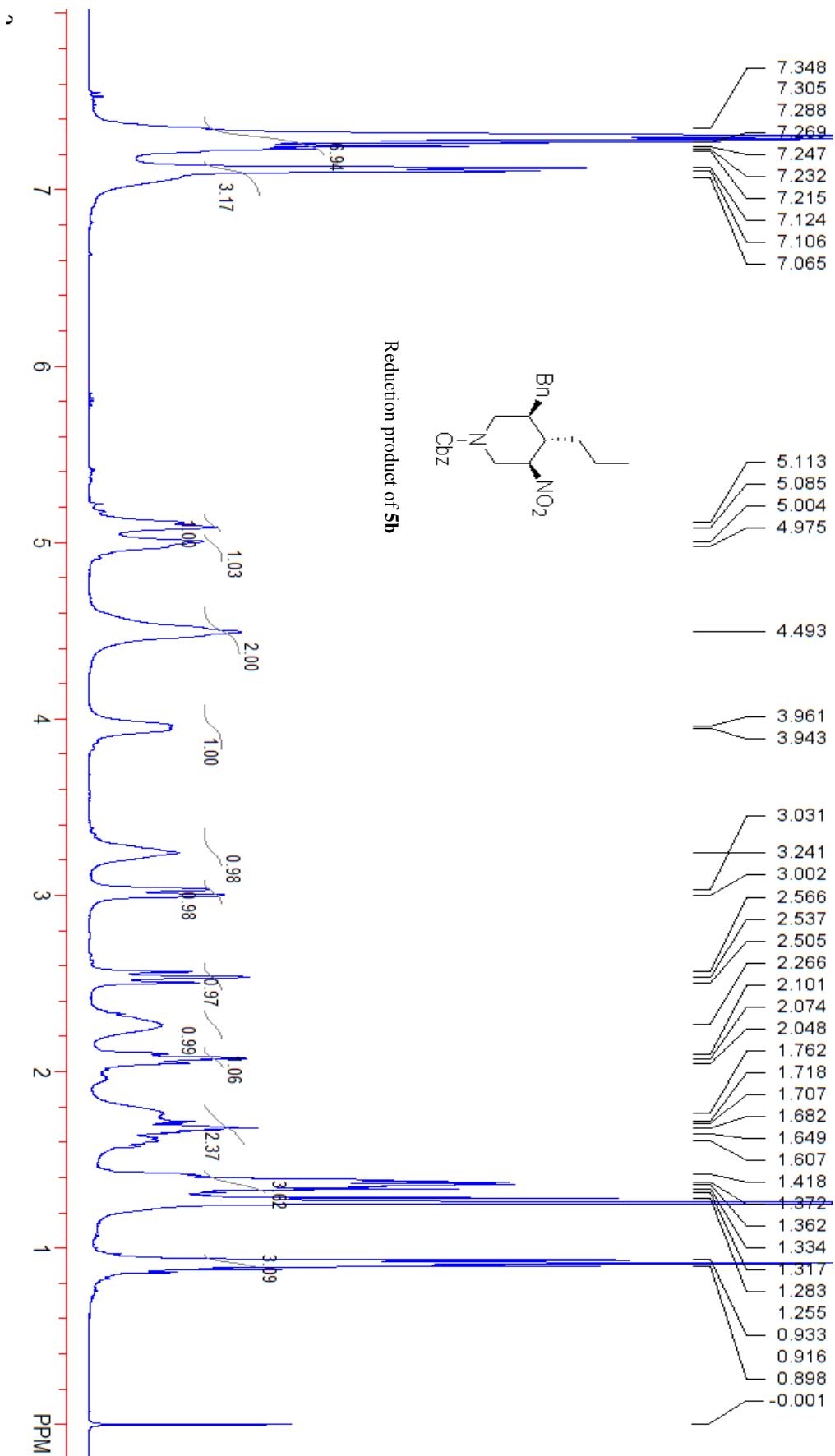


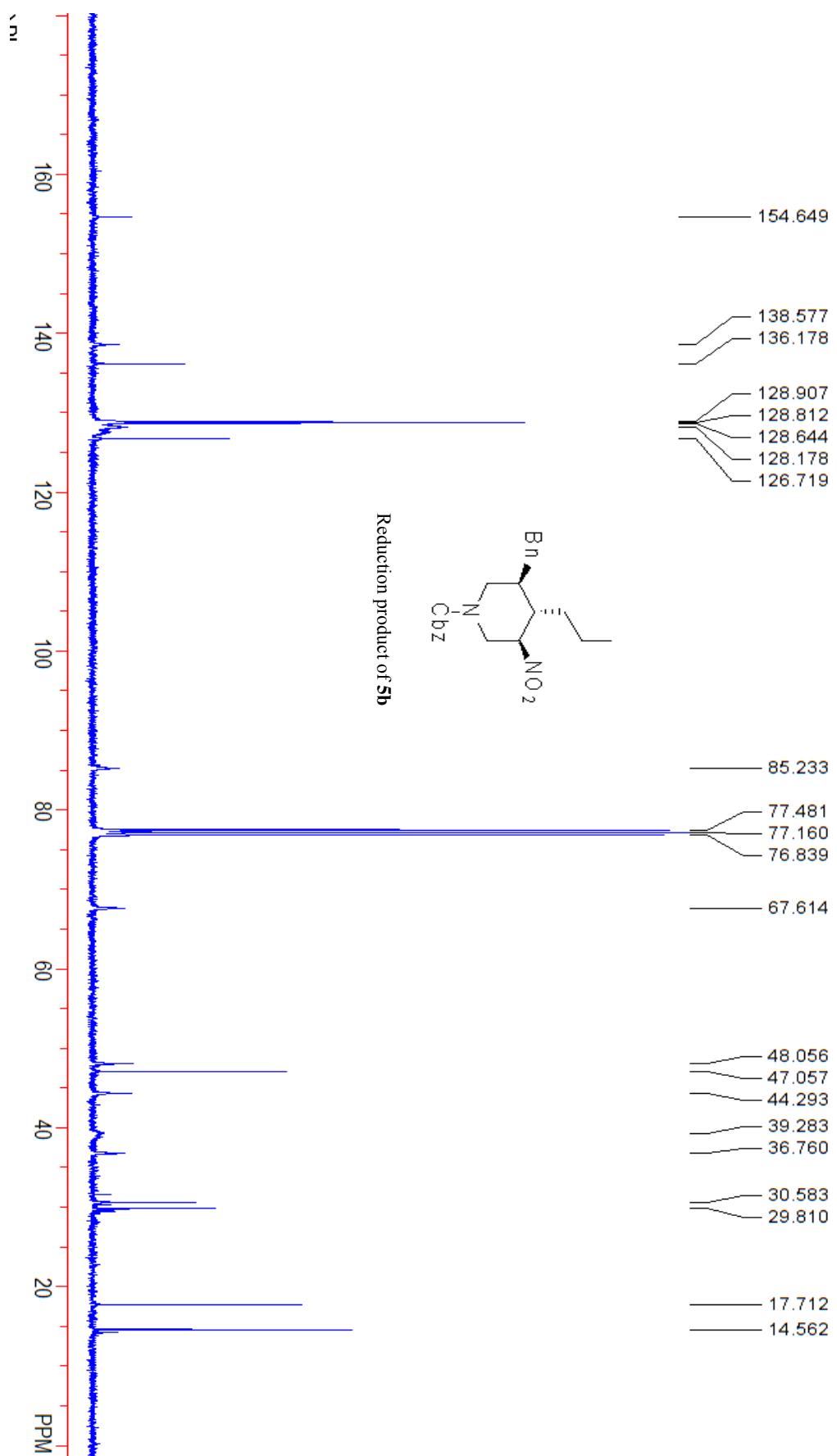
No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	11.095	136.6	2559.5	0.7148
2	2	12.677	17767.1	355502.8	99.2852
	Total		17903.7	358062.3	100.0000











HPLC Report

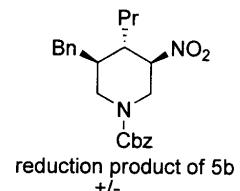
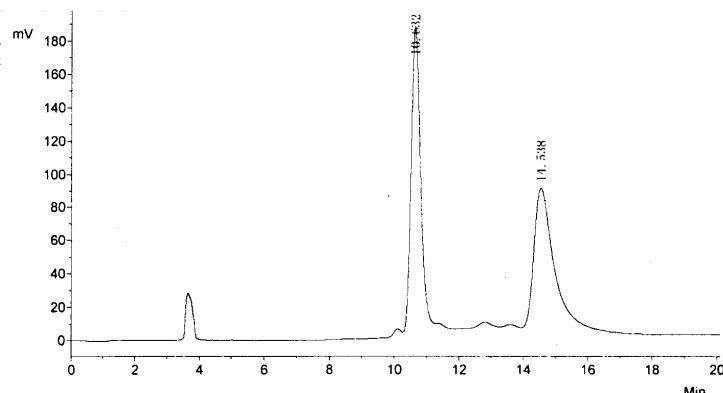
Sample Name:

Data File:ZSL17-29-2+-PC-2-730.7214.che

Operator:

Date:2010-09-07

Time:13:51



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	10.632	181398.8	3576778.5	49.7194
2	2	14.538	74437.0	3617148.2	50.2806
	Total		255835.8	7193926.7	100.0000

HPLC Report

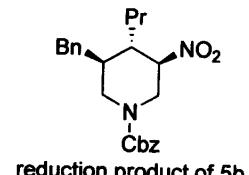
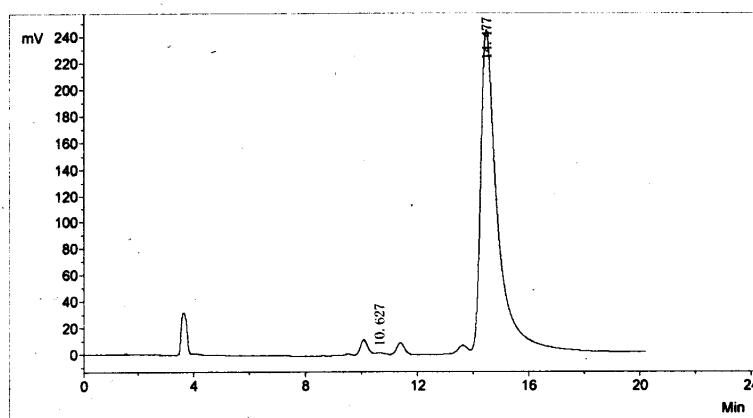
Sample Name:

Data File:ZSL6-33-2.che

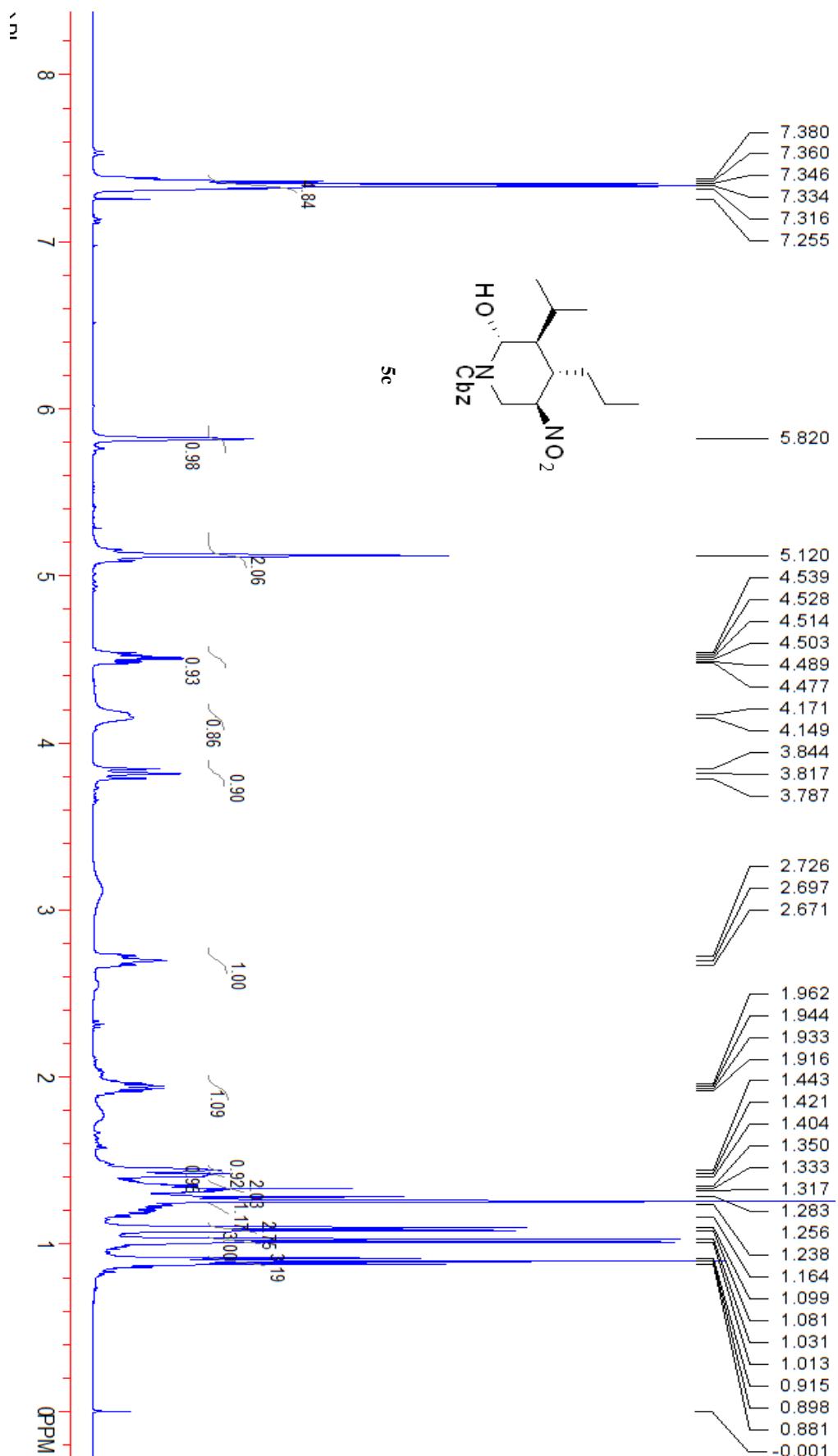
Operator:

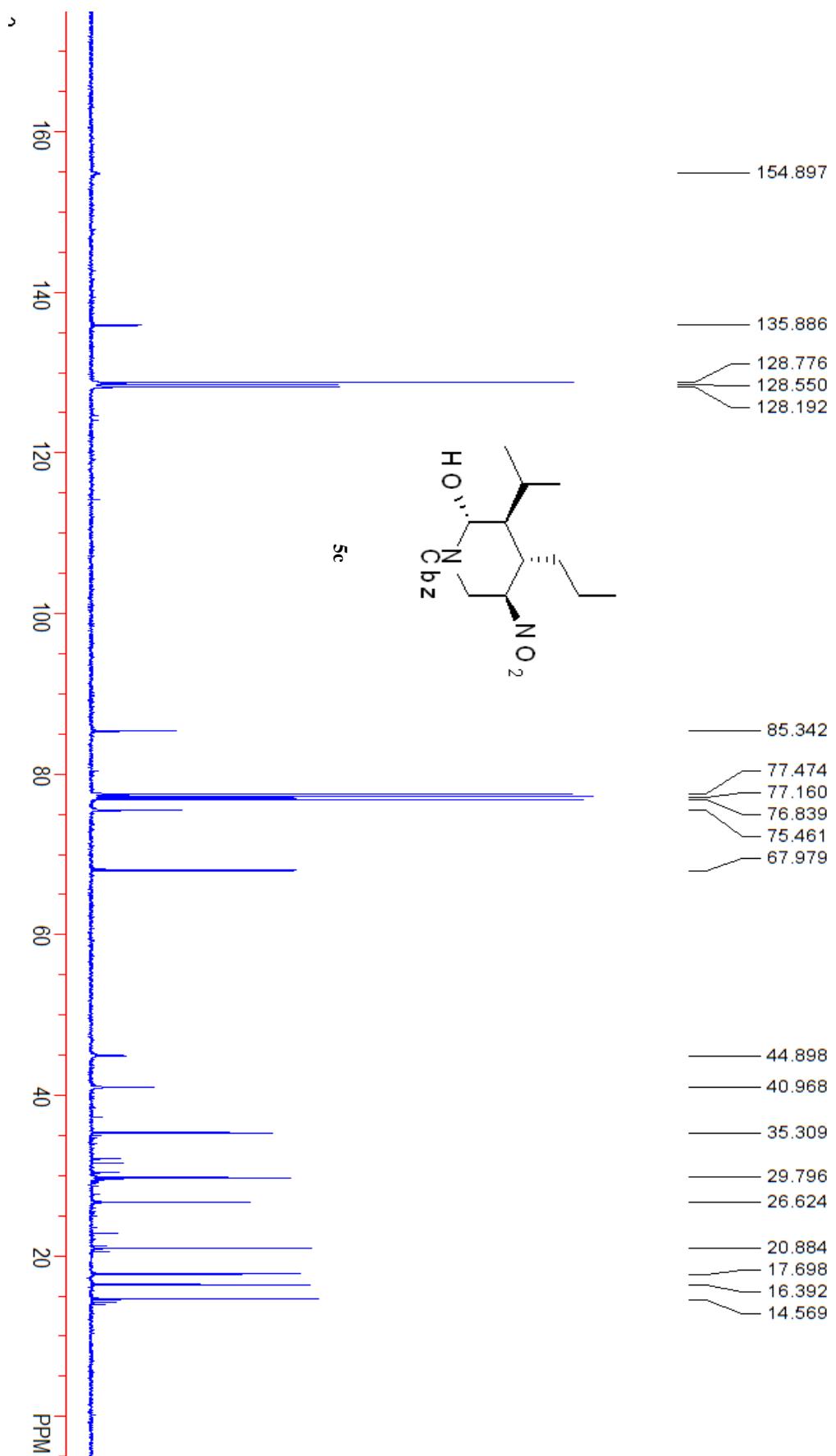
Date:2010-09-07

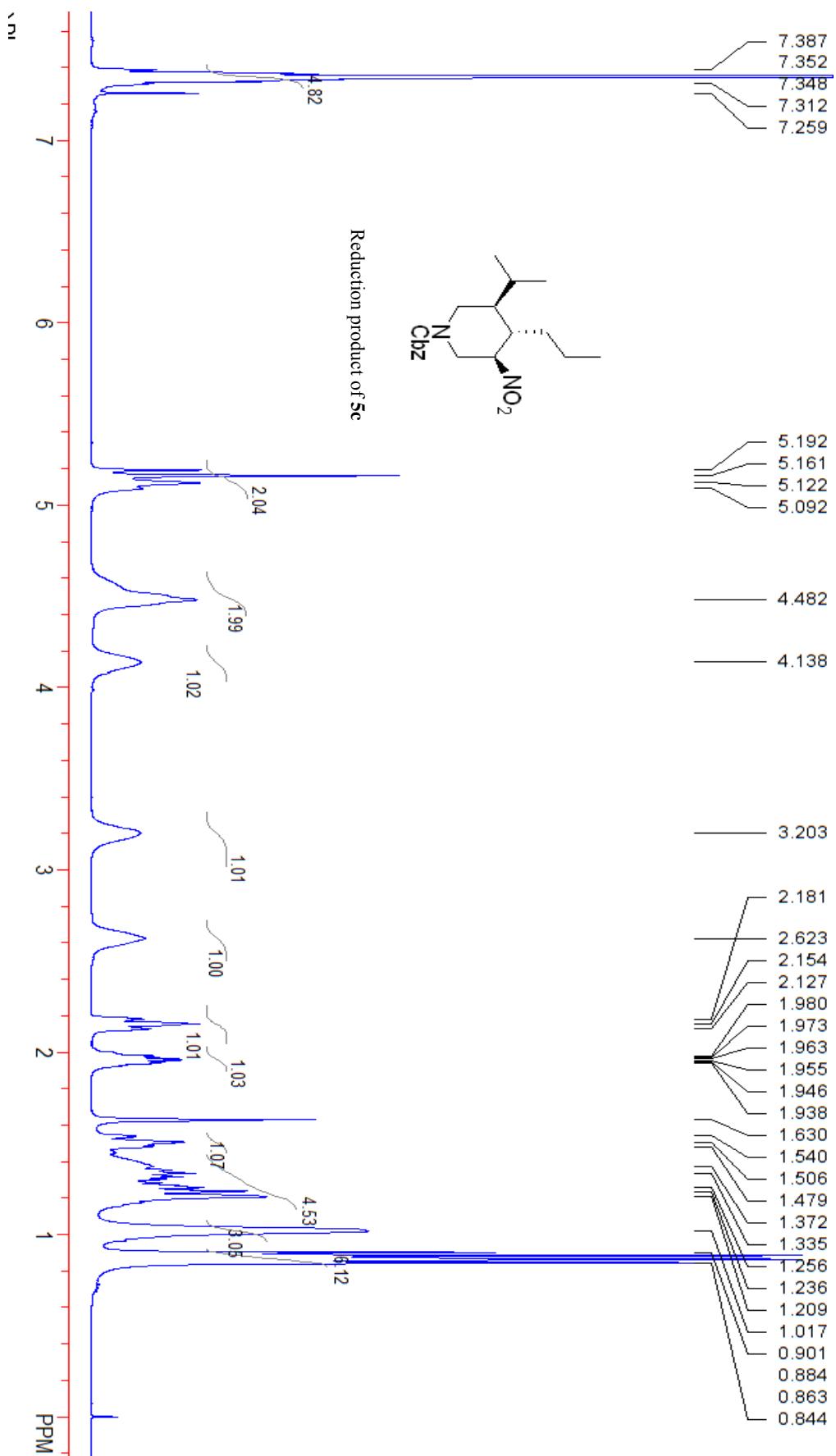
Time:14:16

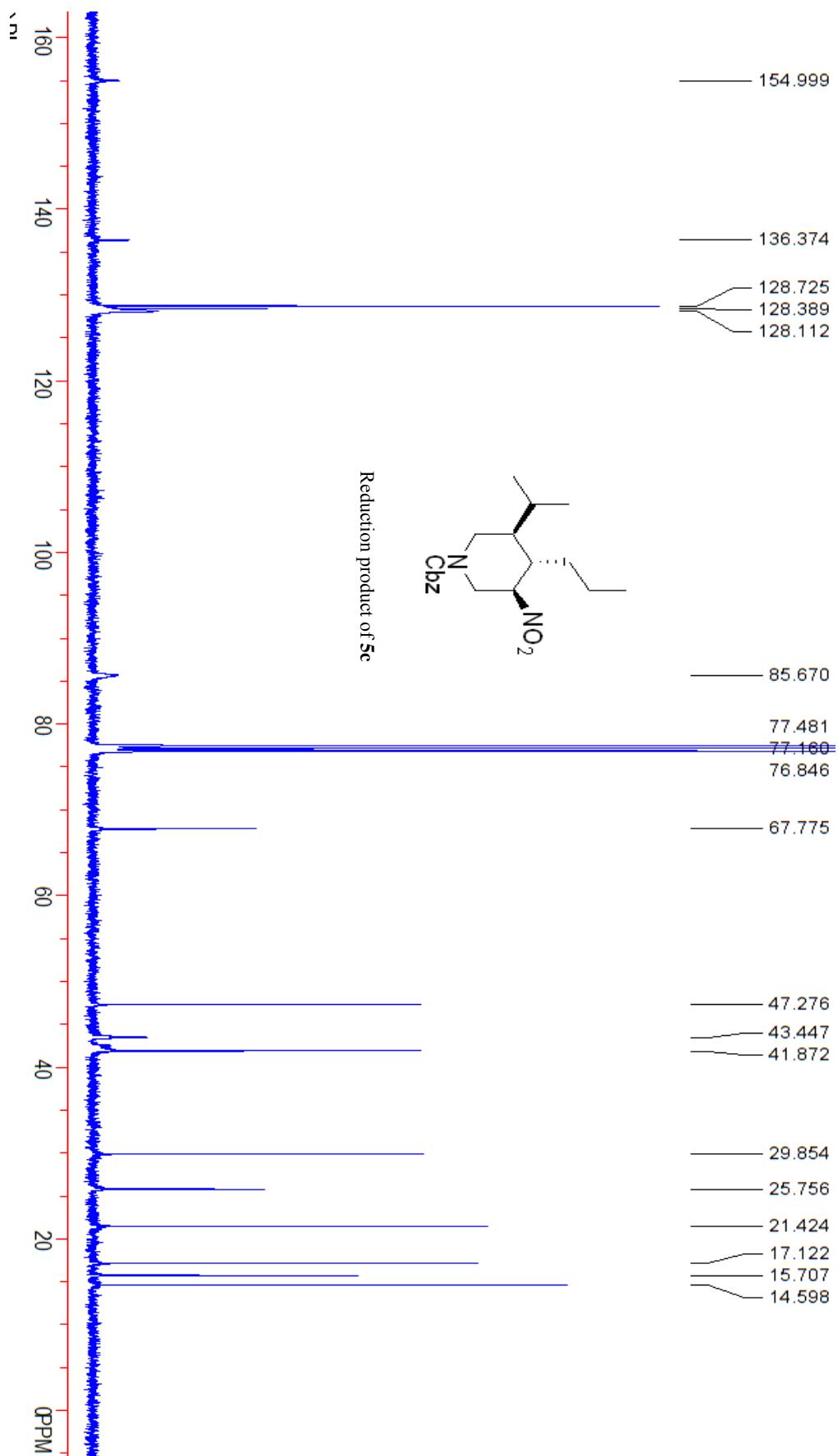


No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	10.627	621.7	8241.4	0.0831
2	2	14.477	241752.0	9907979.4	99.9169
	Total		242373.7	9916220.9	100.0000









HPLC Report

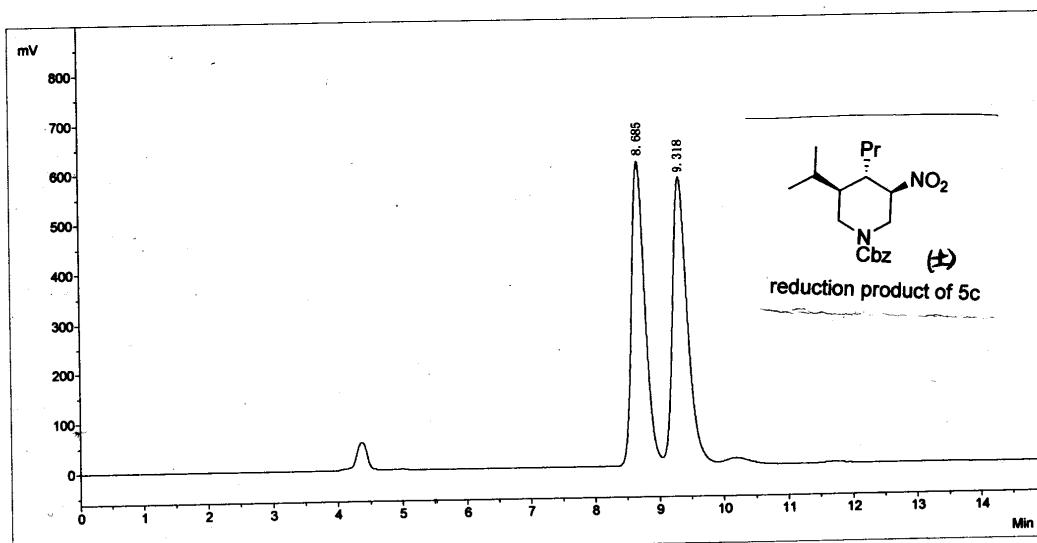
Sample Name:

Data File:ZSL-6-16-2+- PA-2 55 214 0.7.che

Operator:

Date:2010-07-20

Time:14:01



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		8.685	612982.1	8794990.6	49.1811
2	2		9.318	574153.1	9087883.3	50.8189
Total				1187135.2	17882873.9	100.0000

HPLC Report

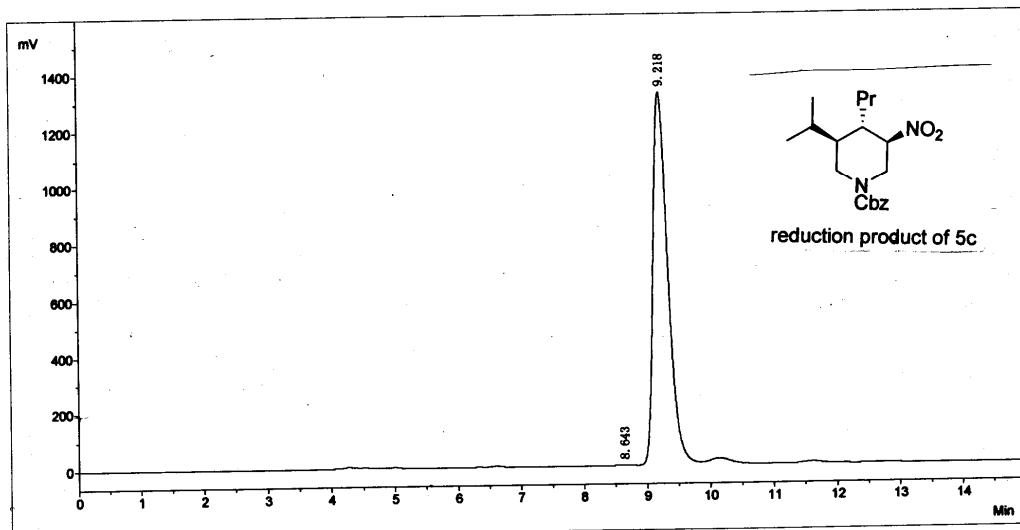
Sample Name:

Data File:ZSL-6-30-2.che

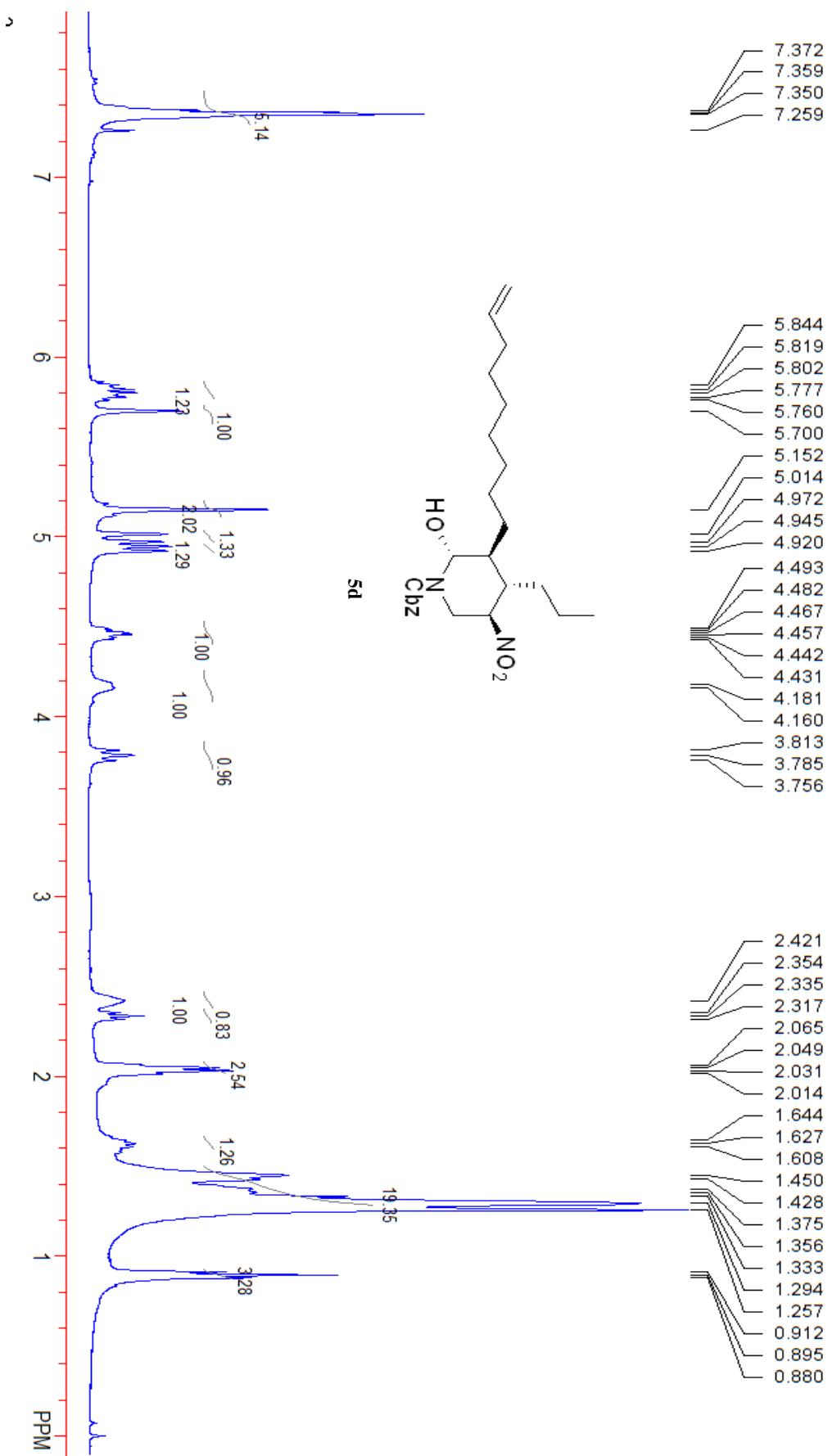
Operator:

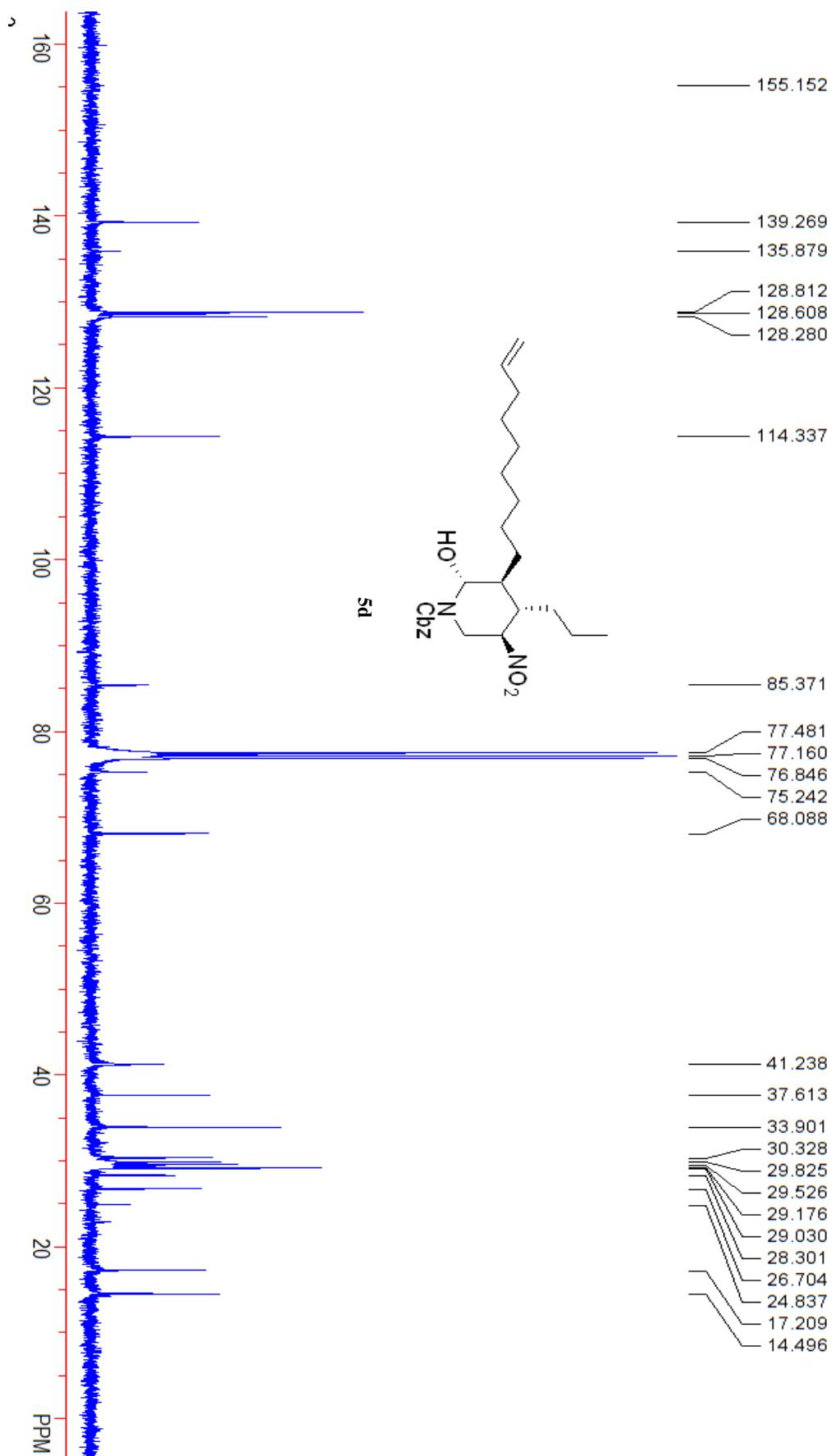
Date:2010-07-20

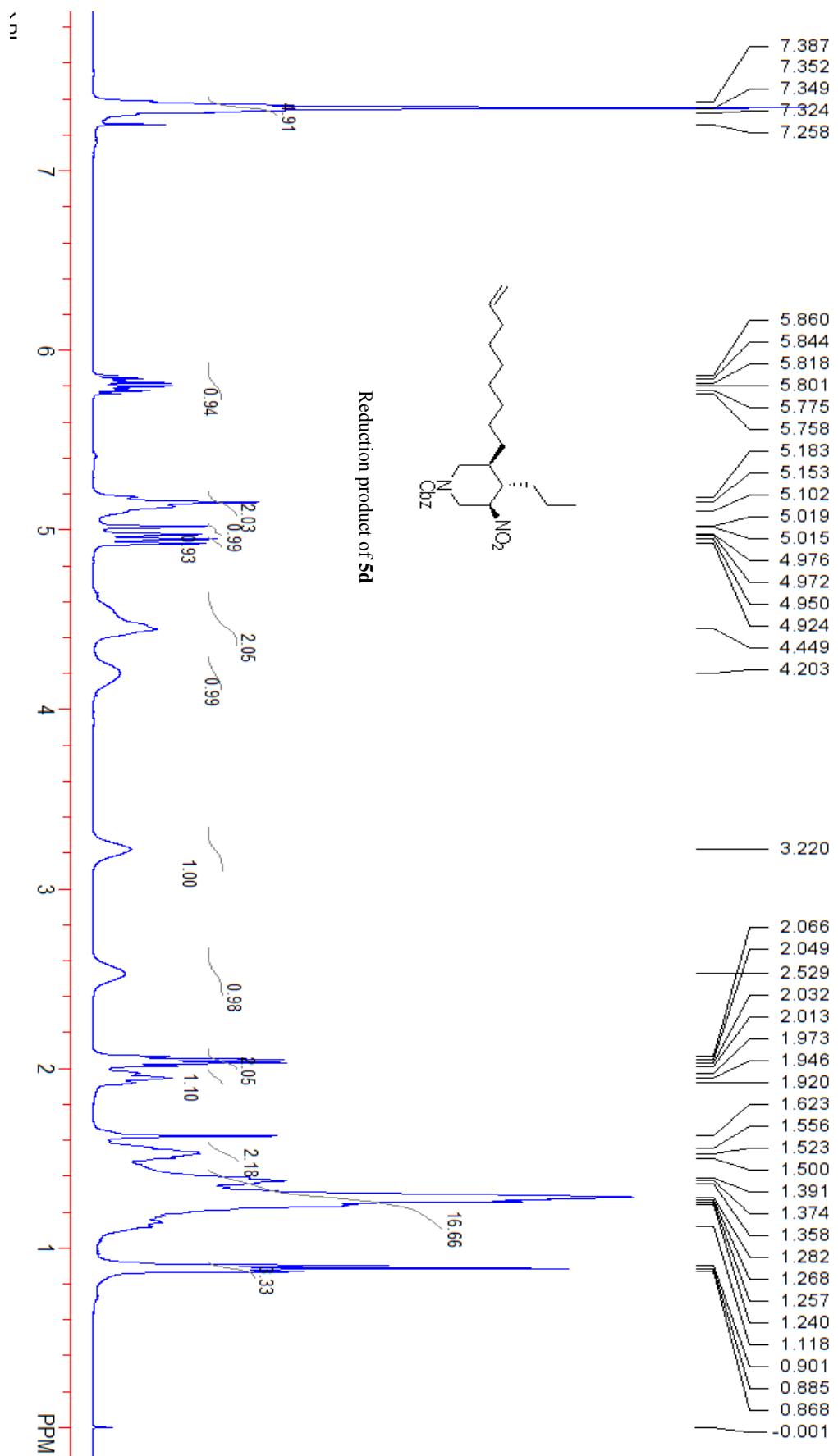
Time: 14:27

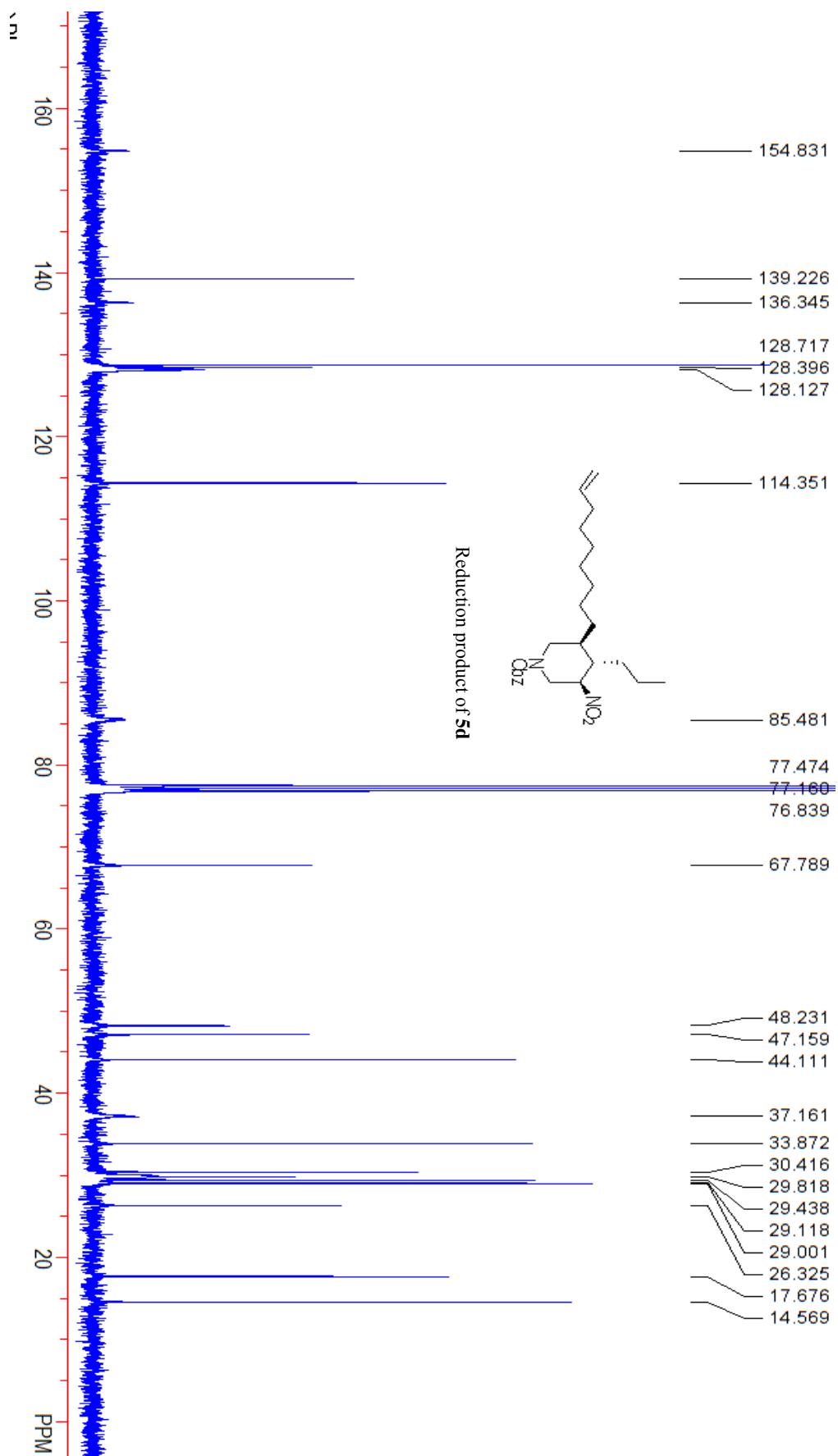


No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		8.643	2207.4	26436.7	0.1249
2	2		9.218	1318725.3	21136472.8	99.8751
		Total		1320932.7	21162909.5	100.0000









HPLC Report

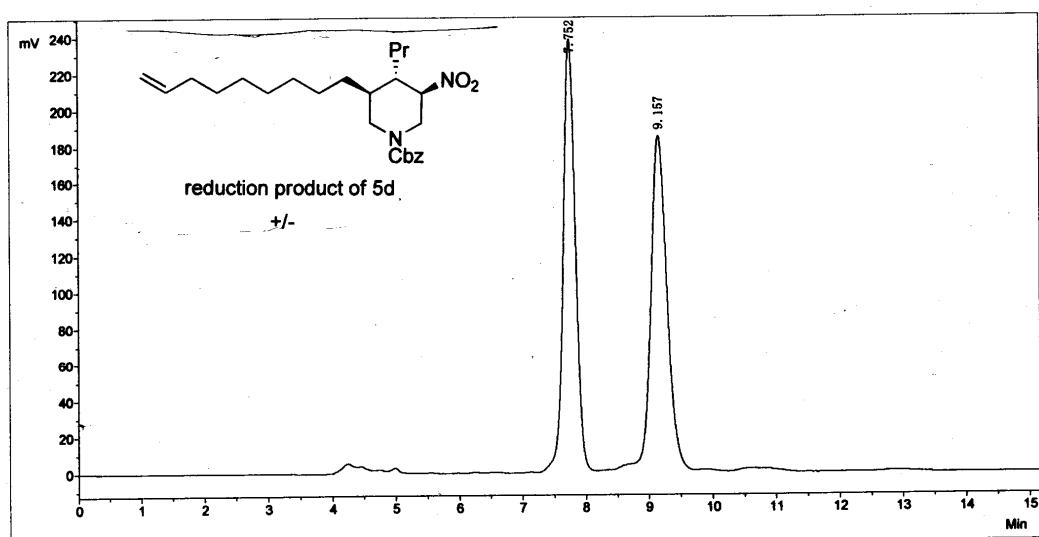
Sample Name:

Data File:ZSL-6-19-2+- PA-2 55 214 0.7.che

Operator:

Date:2010-07-20

Time:13:25



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		7.752	238116.7	3174916.5	50.4255
2	2		9.157	182479.9	3121331.3	49.5745
Total				420596.5	6296247.8	100.0000

HPLC Report

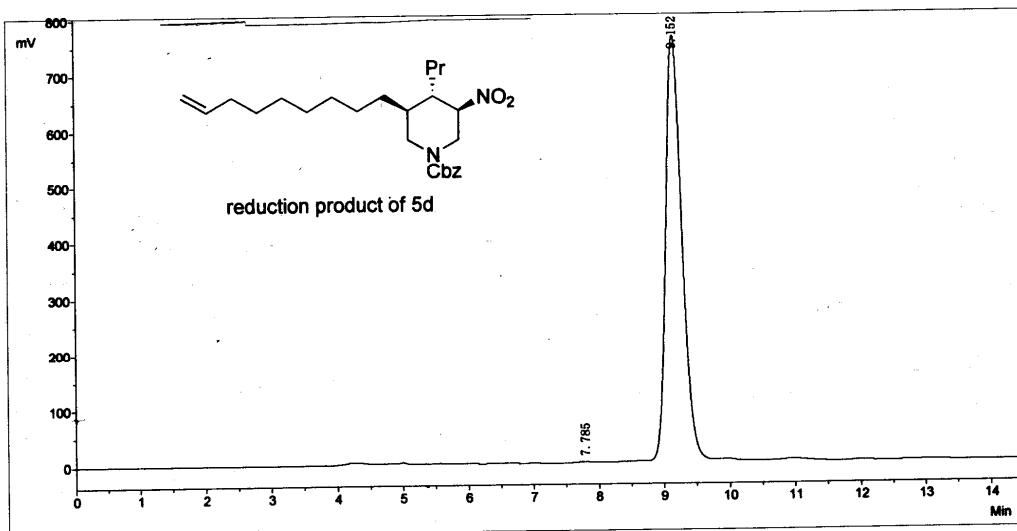
Sample Name:

Data File:ZSL-6-29.che

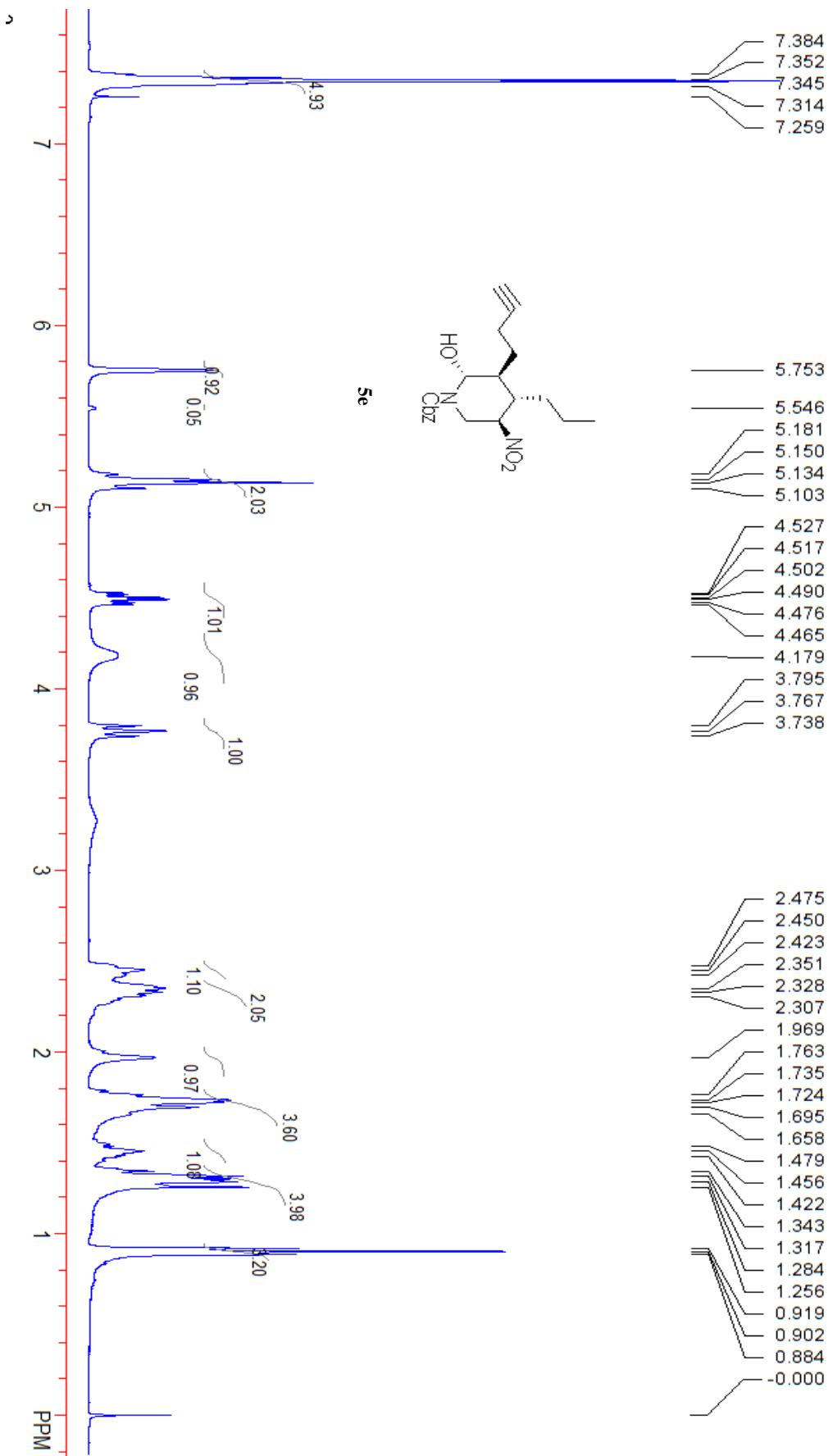
Operator:

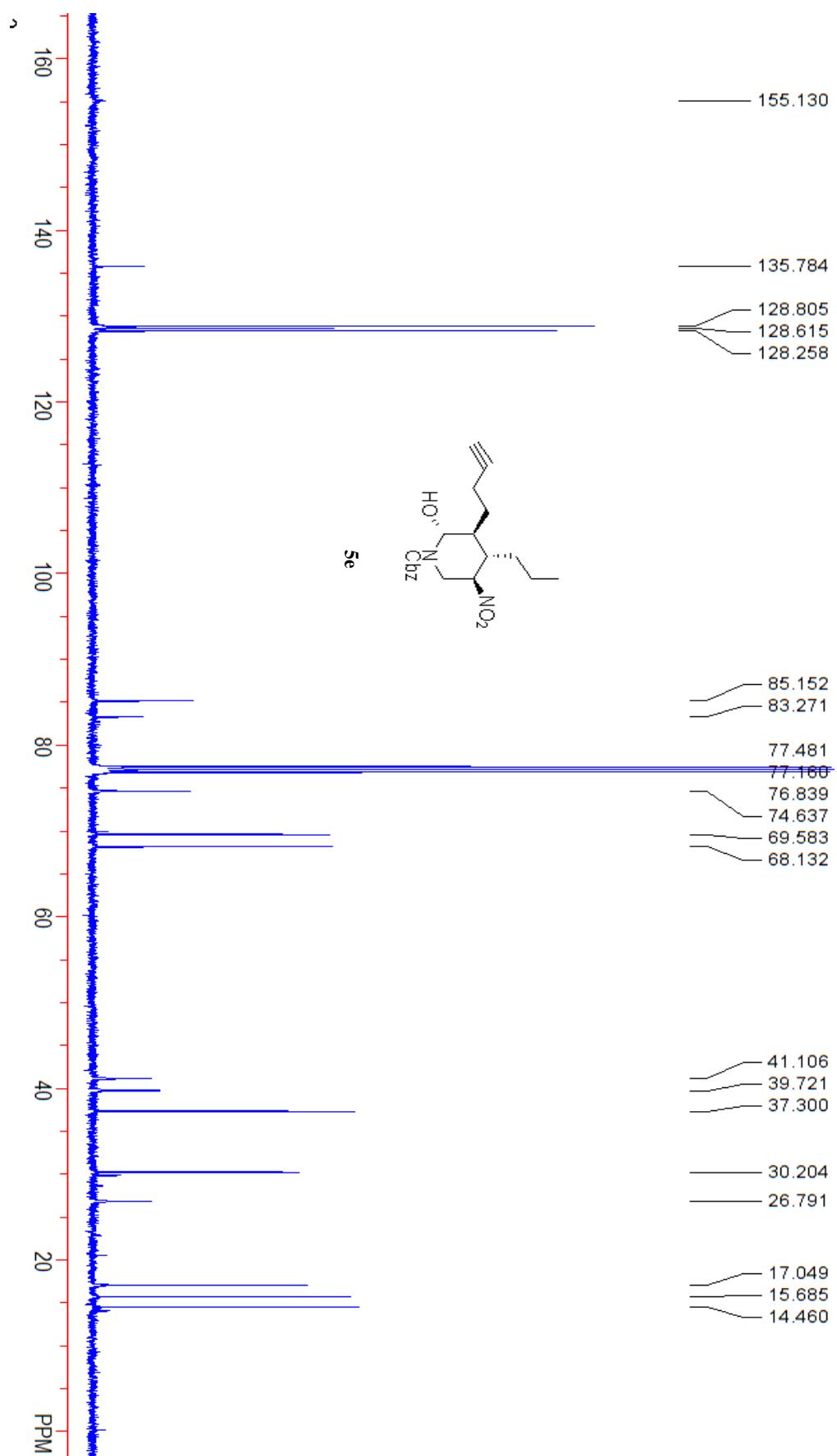
Date:2010-07-20

Time:13:43



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		7.785	1439.2	16491.4	0.1275
2	2		9.152	760472.5	12914286.9	99.8725
Total				761911.7	12930778.3	100.0000





HPLC Report

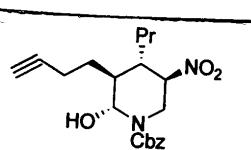
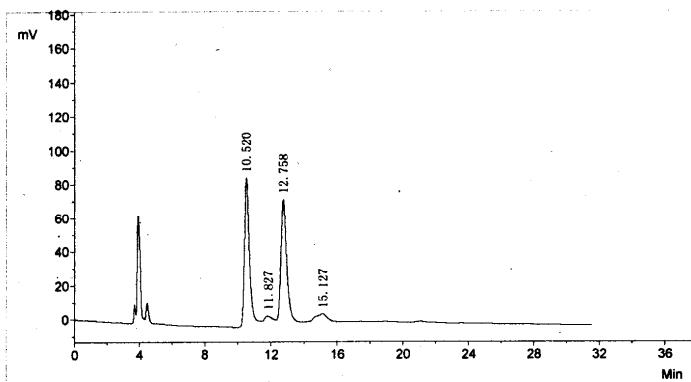
Sample Name:

Data File:ZSL6-20+-SINO-AD-910.7214.che

Operator:

Date:2010-08-19

Time:09:04



5e
+/-

No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	10.520	86975.5	1719927.1	46.1434
2	2	11.827	2685.0	69032.4	1.8521
3	3	12.758	71819.8	1698966.1	45.5811
4	4	15.127	4827.4	239425.0	6.4235
Total		166307.7	3727350.6	100.0000	

HPLC Report

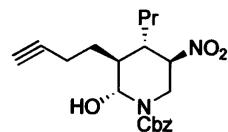
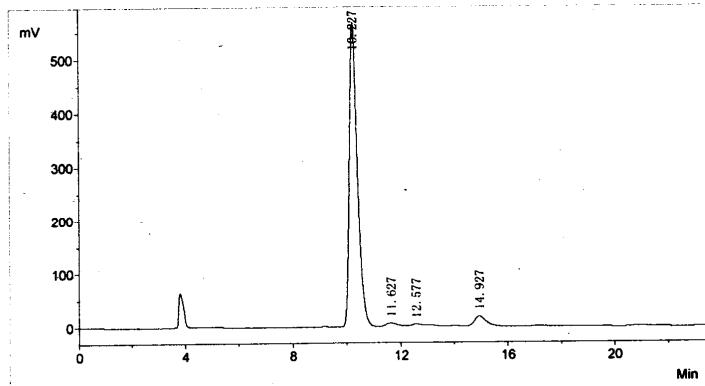
Sample Name:

Data File:ZSL6-28.che

Operator:

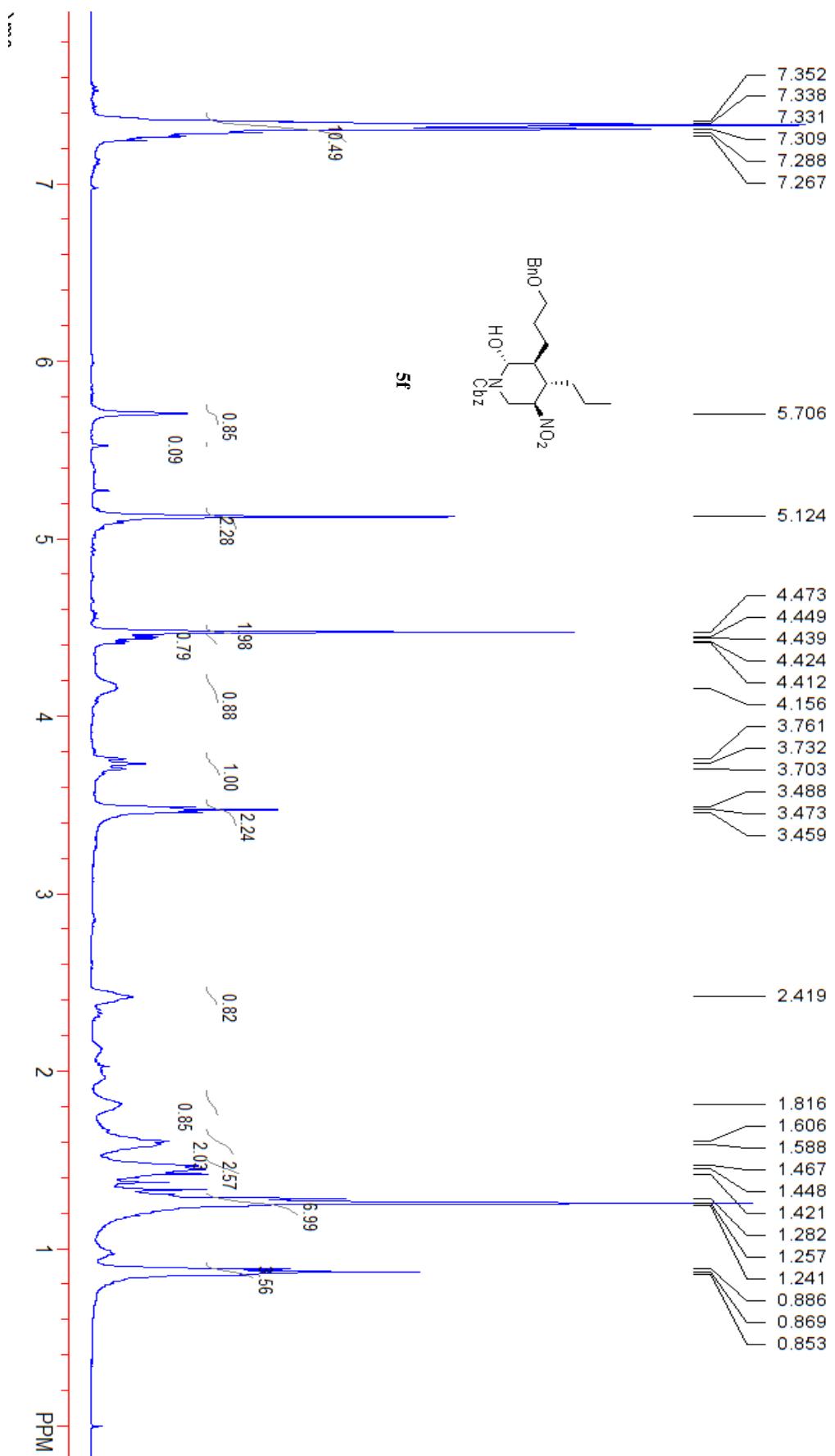
Date:2010-08-19

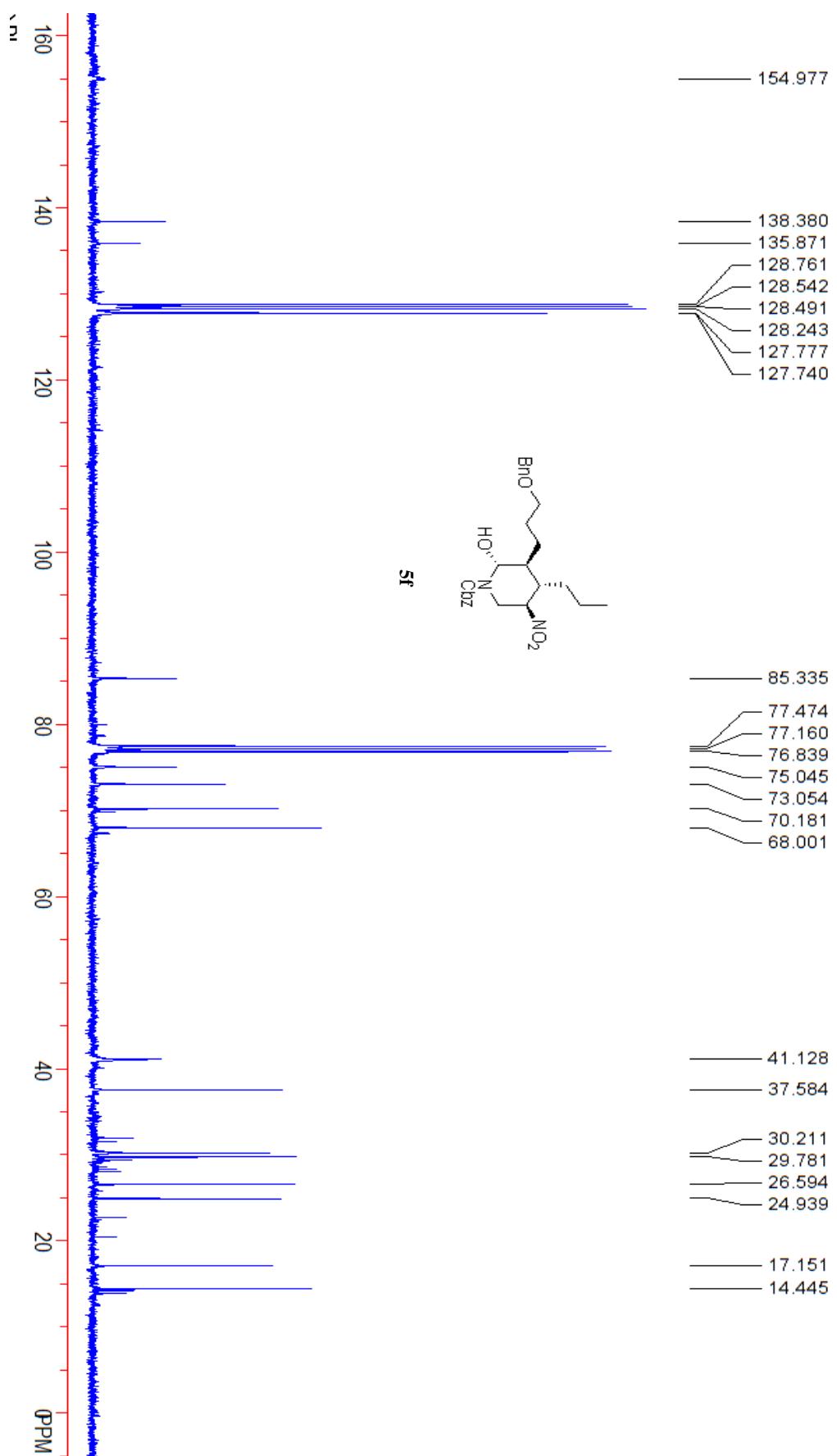
Time:09:41



5e

No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	10.227	564408.8	11776232.0	93.7569 ✓
2	2	11.627	5774.7	161741.2	1.2877
3	3	12.577	3127.3	59477.1	0.4735 ✓
4	4	14.927	19044.9	562943.1	4.4819
Total		592355.7	12560393.3	100.0000	





HPLC Report

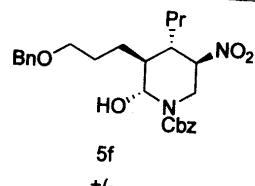
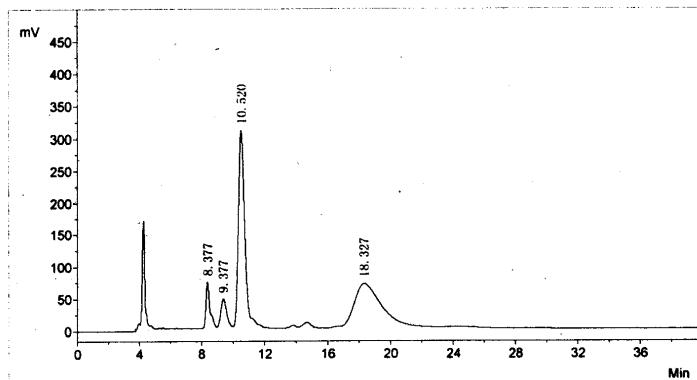
Sample Name:

Data File:ZSL-17-49+- PA-2 73 214 0.7.che

Operator:

Date:2010-08-16

Time:16:35



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	8.377	72153.0	1264820.3	7.0741
2	2	9.377	44074.8	1111179.7	6.2165
3	3	10.520	300193.5	7770978.7	43.4628
4	4	18.327	66426.7	7732317.3	43.2466
Total			482848.0	17879596.0	100.0000

HPLC Report

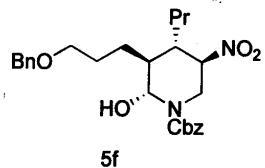
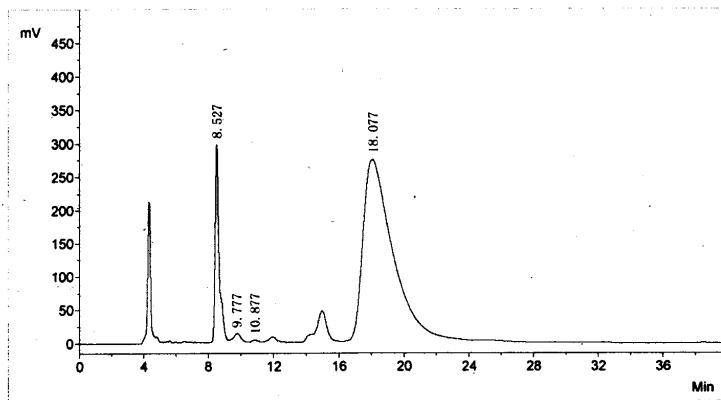
Sample Name:

Data File:ZSL-6-32.che

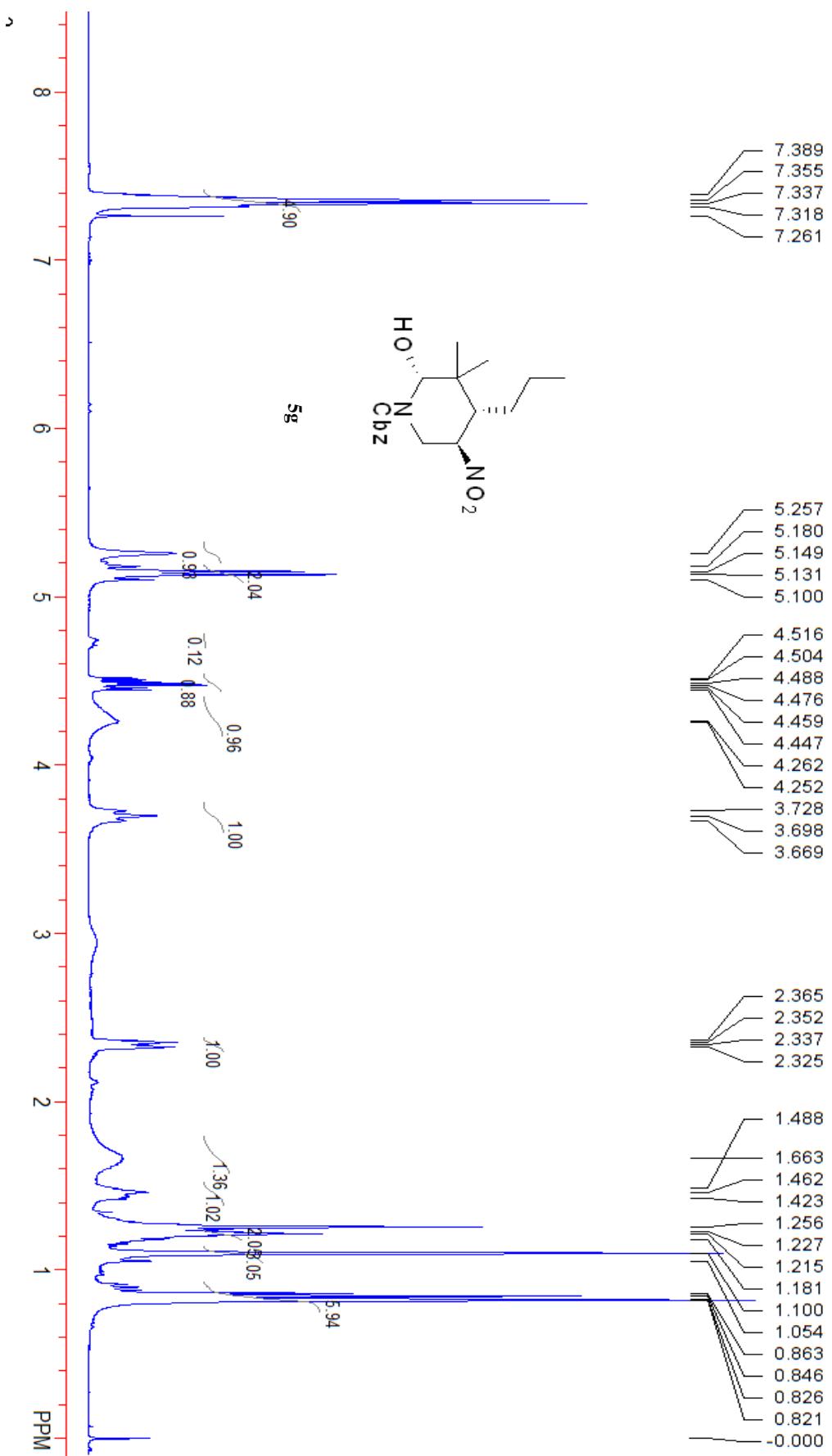
Operator:

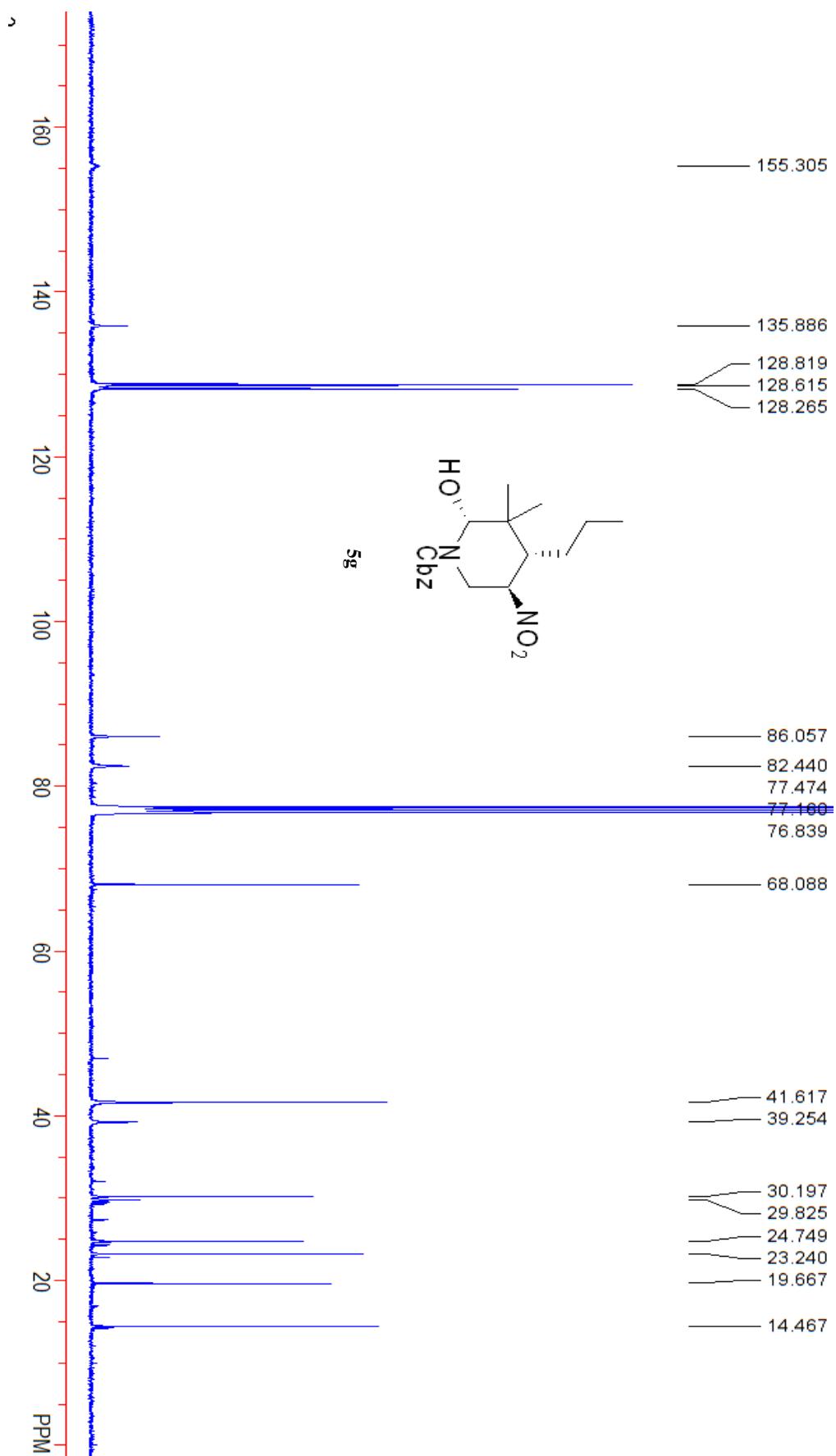
Date:2010-08-16

Time:17:33



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	8.527	297646.4	5204237.6	12.9060
2	2	9.777	13661.7	434562.6	1.0777
3	3	10.877	3312.1	82949.2	0.2057
4	4	18.077	273054.8	34602347.9	85.8106
Total			587675.0	40324097.2	100.0000





HPLC Report

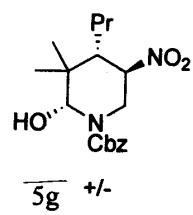
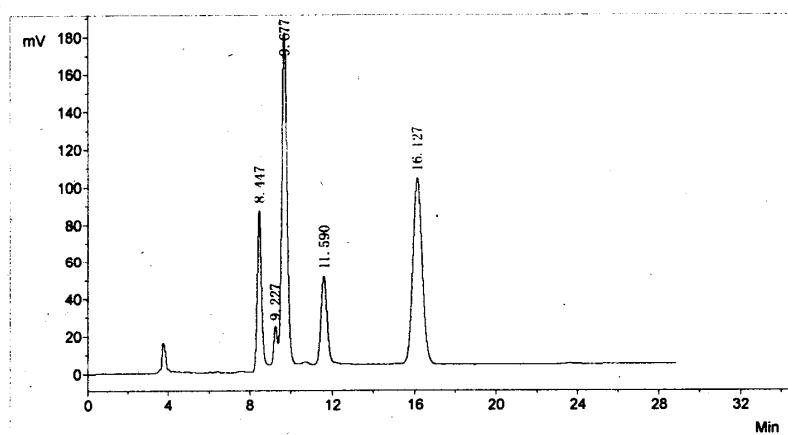
Sample Name:

Data File:ZSL-6-12+- PC-2 91 214 0.7.che

Operator:

Date:2010-09-01

Time:10:18



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	8.447	84949.7	1170842.7	13.4630
2	2	9.227	20003.7	240992.5	2.7711
3	3	9.677	177394.2	3136117.0	36.0608
4	4	11.590	46931.6	957040.2	11.0046
5	5	16.127	99869.3	3191744.0	36.7005
Total			429148.5	8696736.4	100.0000

HPLC Report

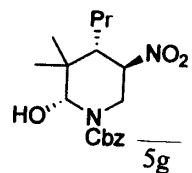
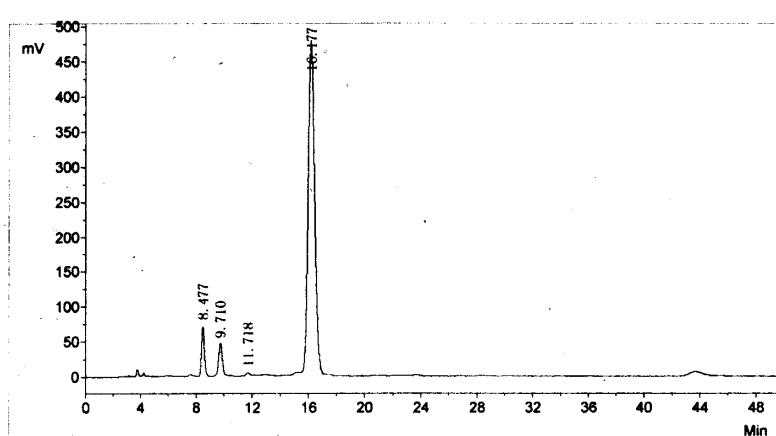
Sample Name:

Data File:ZSL-6-58-1.che

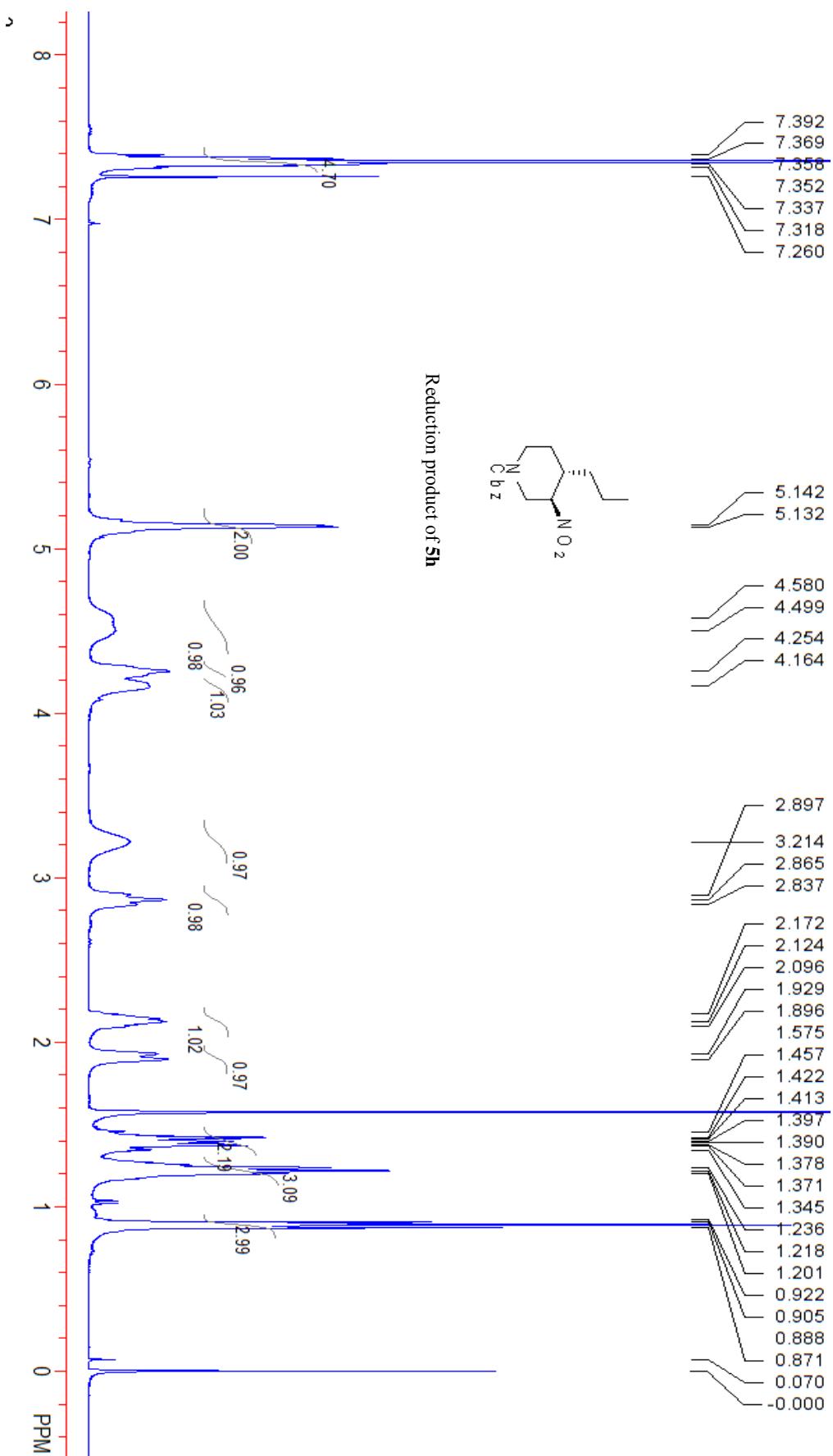
Operator:

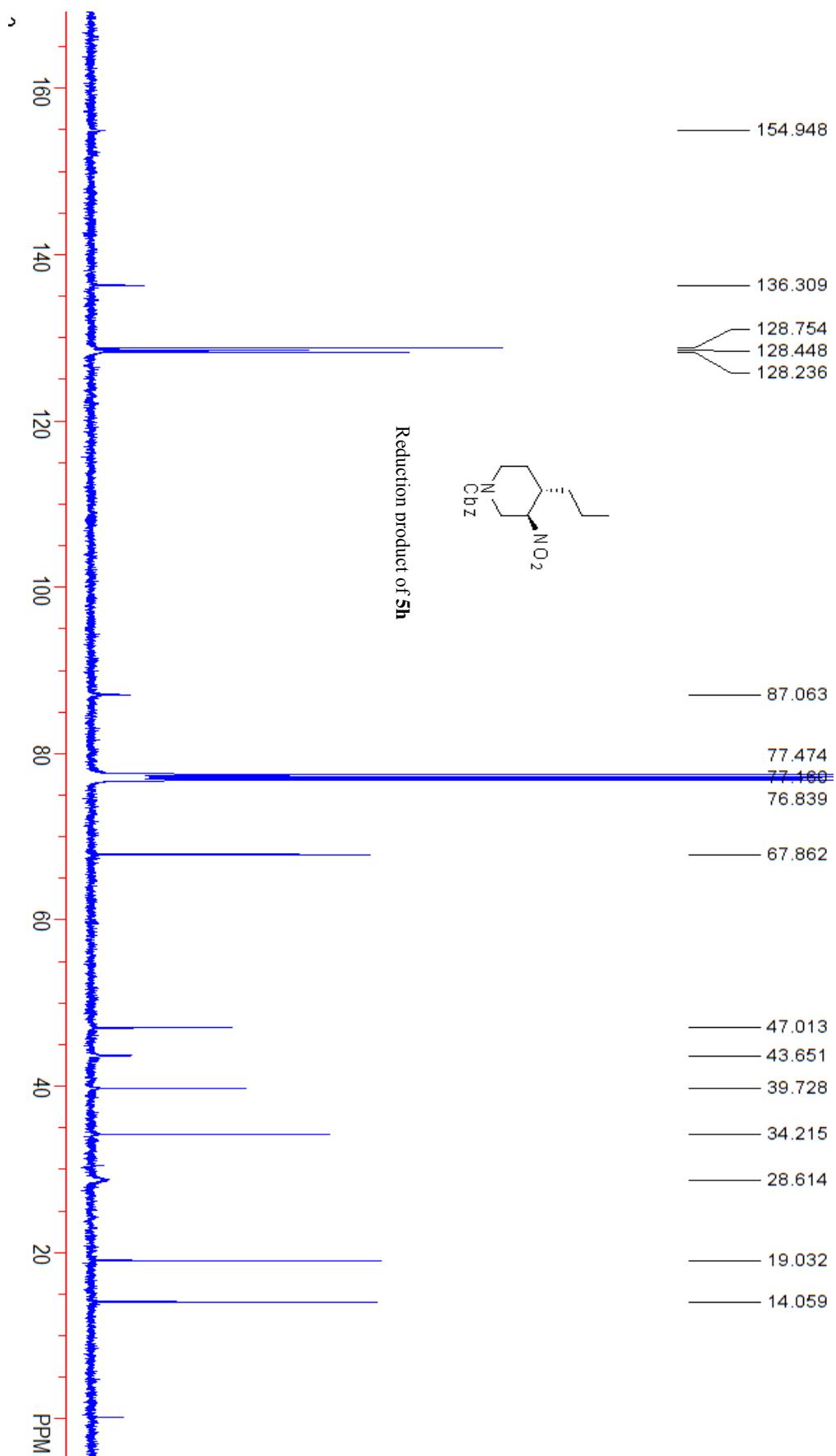
Date:2010-09-01

Time:10:56



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	8.477	71228.2	1025711.5	6.0447
2	2	9.710	43662.8	785706.3	4.6303
3	3	11.718	2788.2	41095.3	0.2422
4	4	16.177	475484.4	15116135.5	89.0827
Total			593163.6	16968648.5	100.0000





HPLC Report

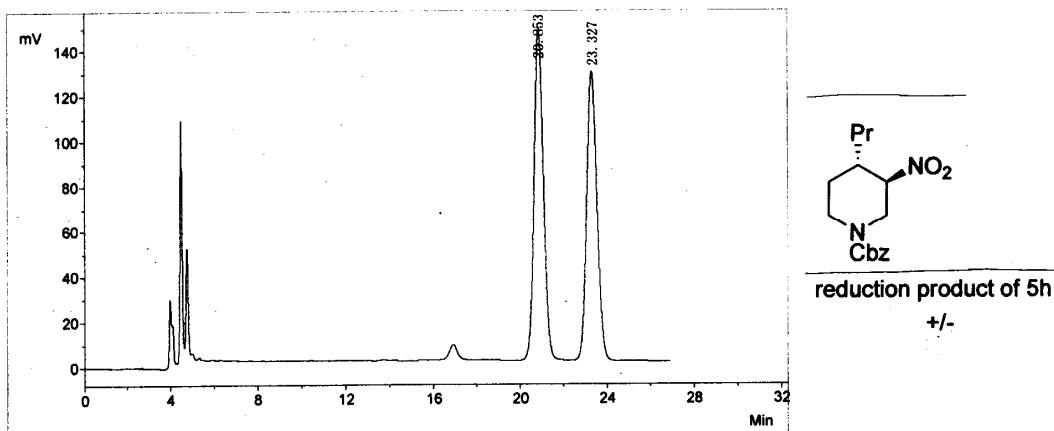
Sample Name:

Data File:ZSL6-82+-.che

Operator:

Date:2010-10-18

Time:08:49



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	20.853	145686.3	4283260.4	50.4646
2	2	23.327	127371.2	4204392.6	49.5354
	Total		273057.6	8487652.9	100.0000

HPLC Report

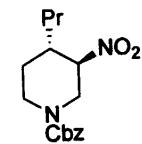
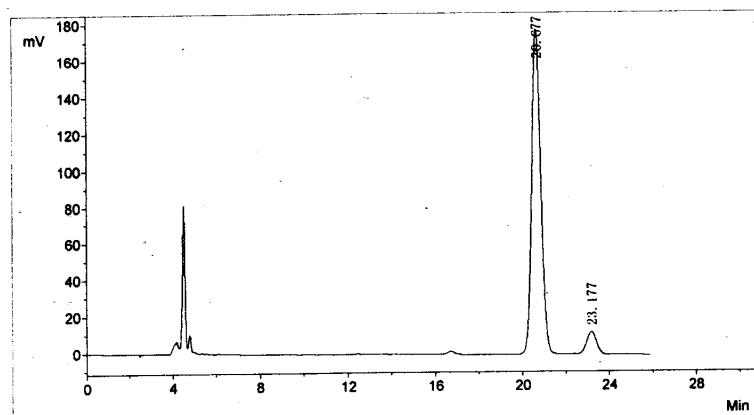
Sample Name:

Data File:ZSL6-89.che

Operator:

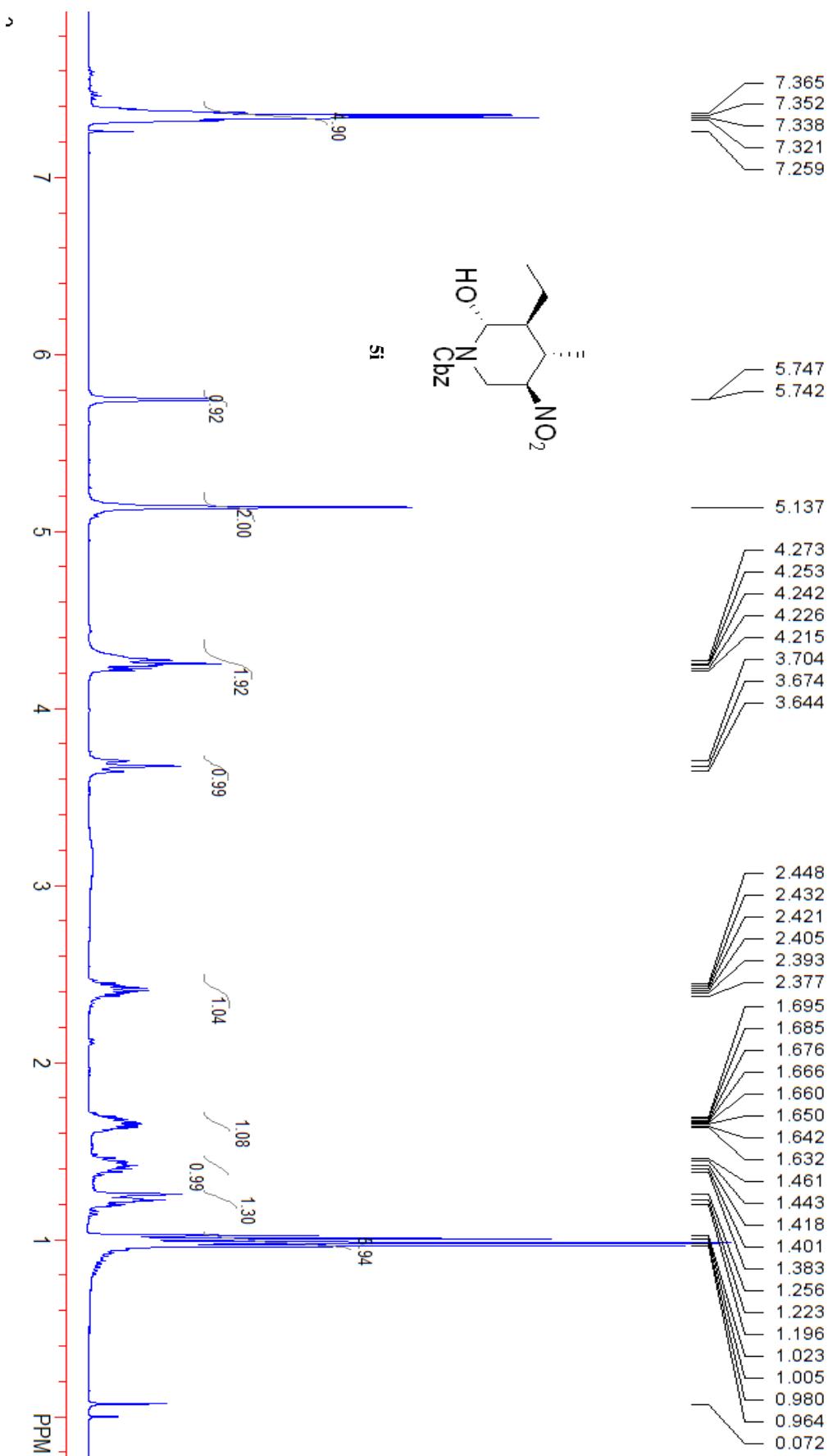
Date:2010-10-18

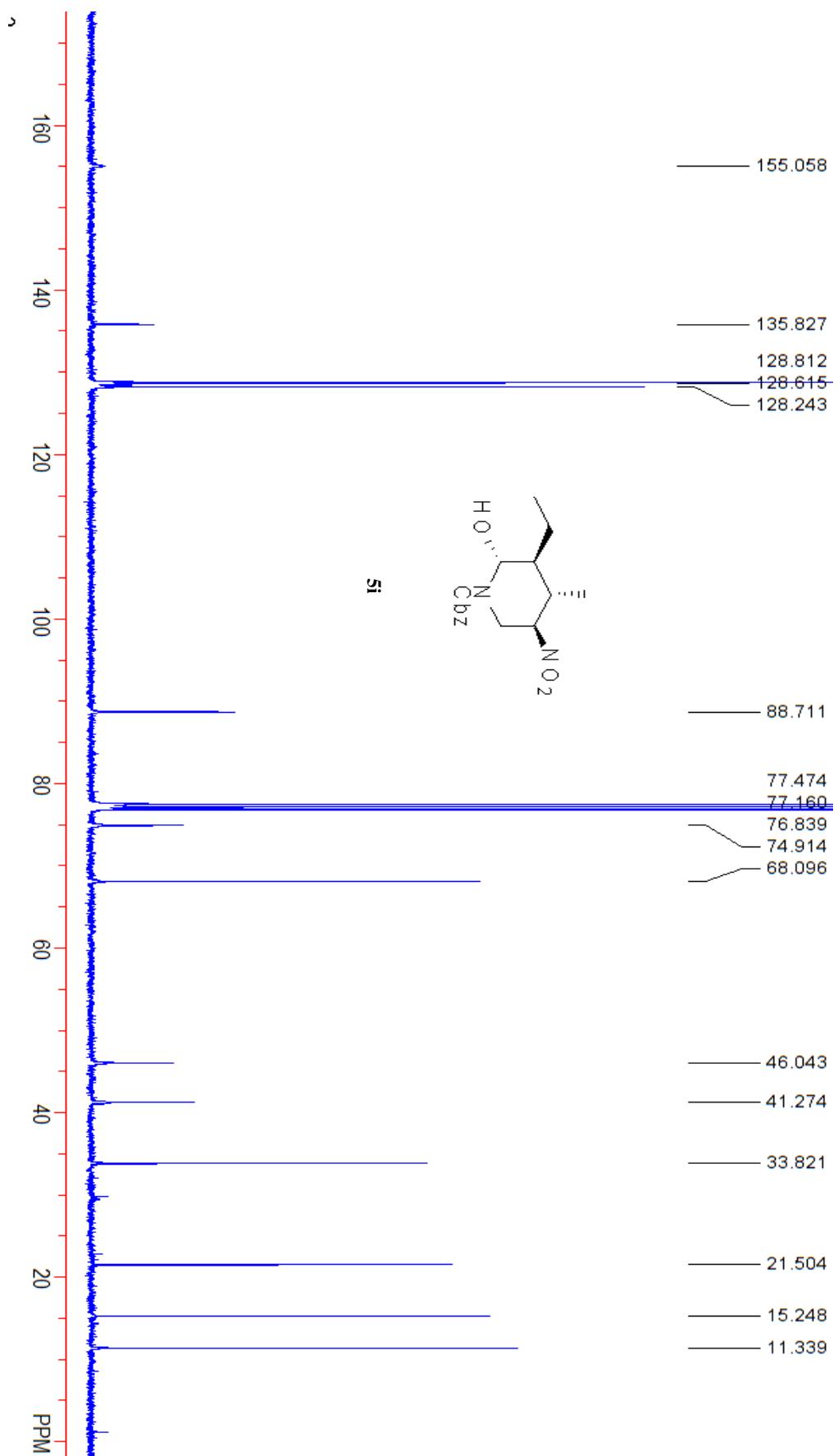
Time:09:52



reduction product of 5h

No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	20.677	177490.1	5171404.2	92.9367
2	2	23.177	12366.4	393031.5	7.0633
Total			189856.4	5564435.8	100.0000





HPLC Report

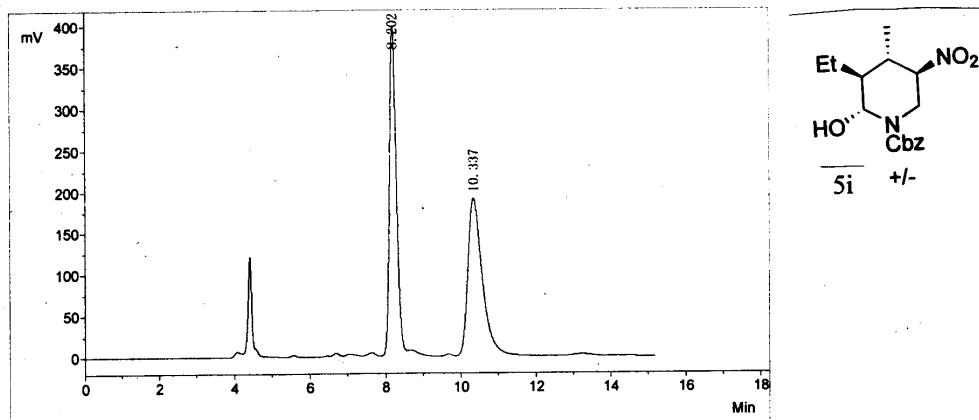
Sample Name:

Data File:ZSL6-88+-PA-2730.7214.che

Operator:

Date:2010-10-20

Time:13:45



No.	PeakNo	R. Time	PeakHeight	PeakArea	Percent
1	1	8.202	390934.8	4789758.2	50.5280
2	2	10.337	187029.8	4689654.3	49.4720
Total			577964.5	9479412.5	100.0000

HPLC Report

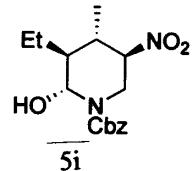
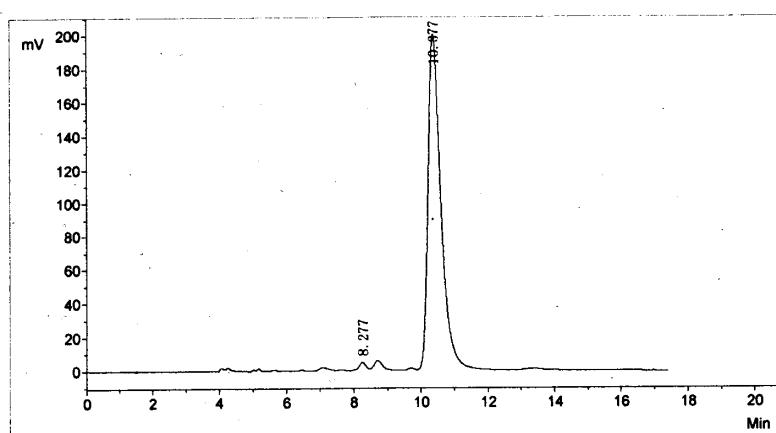
Sample Name:

Data File:ZSL6-90.che

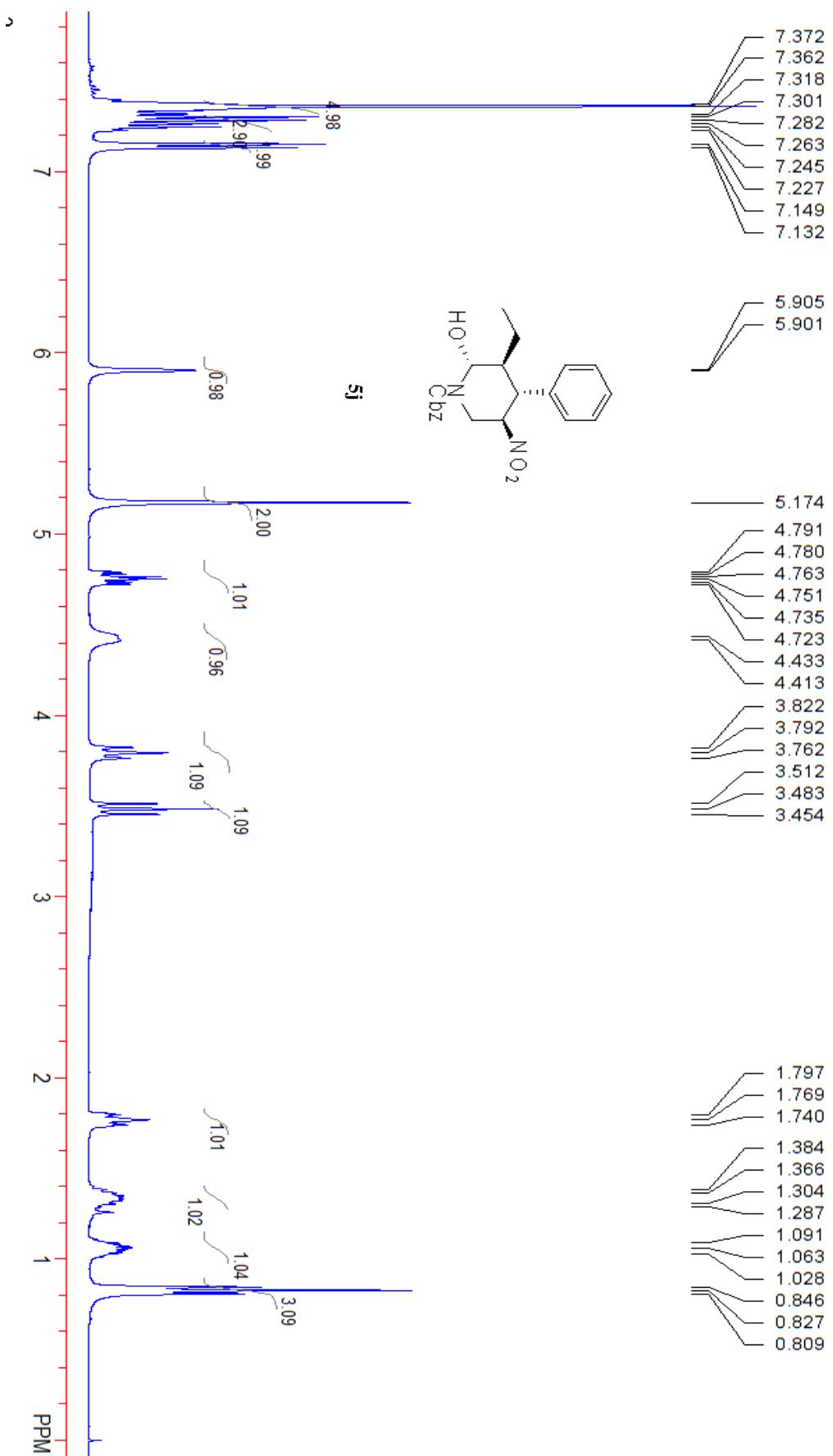
Operator:

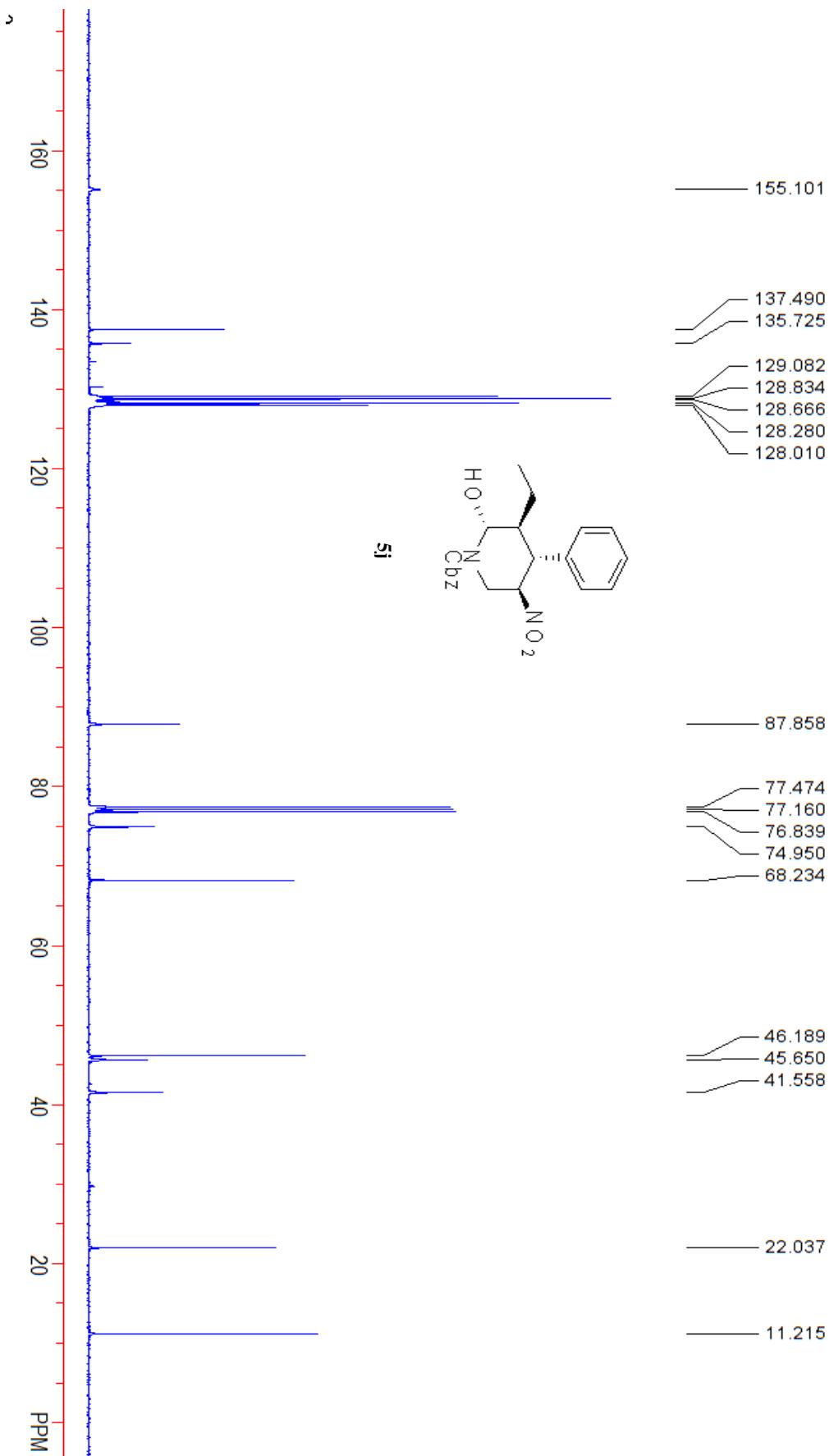
Date:2010-10-20

Time:14:07



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	8.277	4068.2	50828.0	0.9310
2	2	10.377	200075.3	5408880.0	99.0690
Total			204143.6	5459707.9	100.0000





HPLC Report

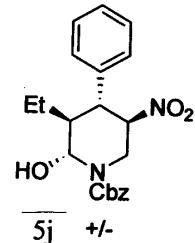
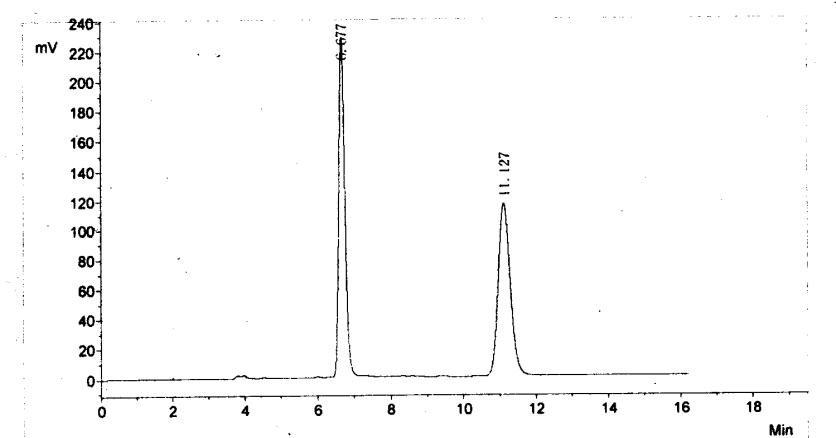
Sample Name:

Data File:ZSL6-17+-PC-2730.7214.che

Operator:

Date:2010-08-25

Time:09:09



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	6.677	227041.6	2713102.9	50.0982
2	2	11.127	116279.6	2702463.8	49.9018
Total			343321.3	5415566.6	100.0000

HPLC Report

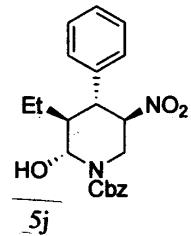
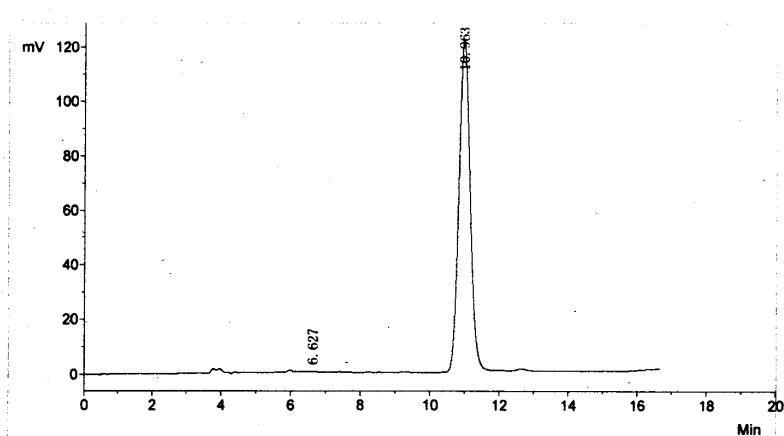
Sample Name:

Data File:ZSL6-60.che

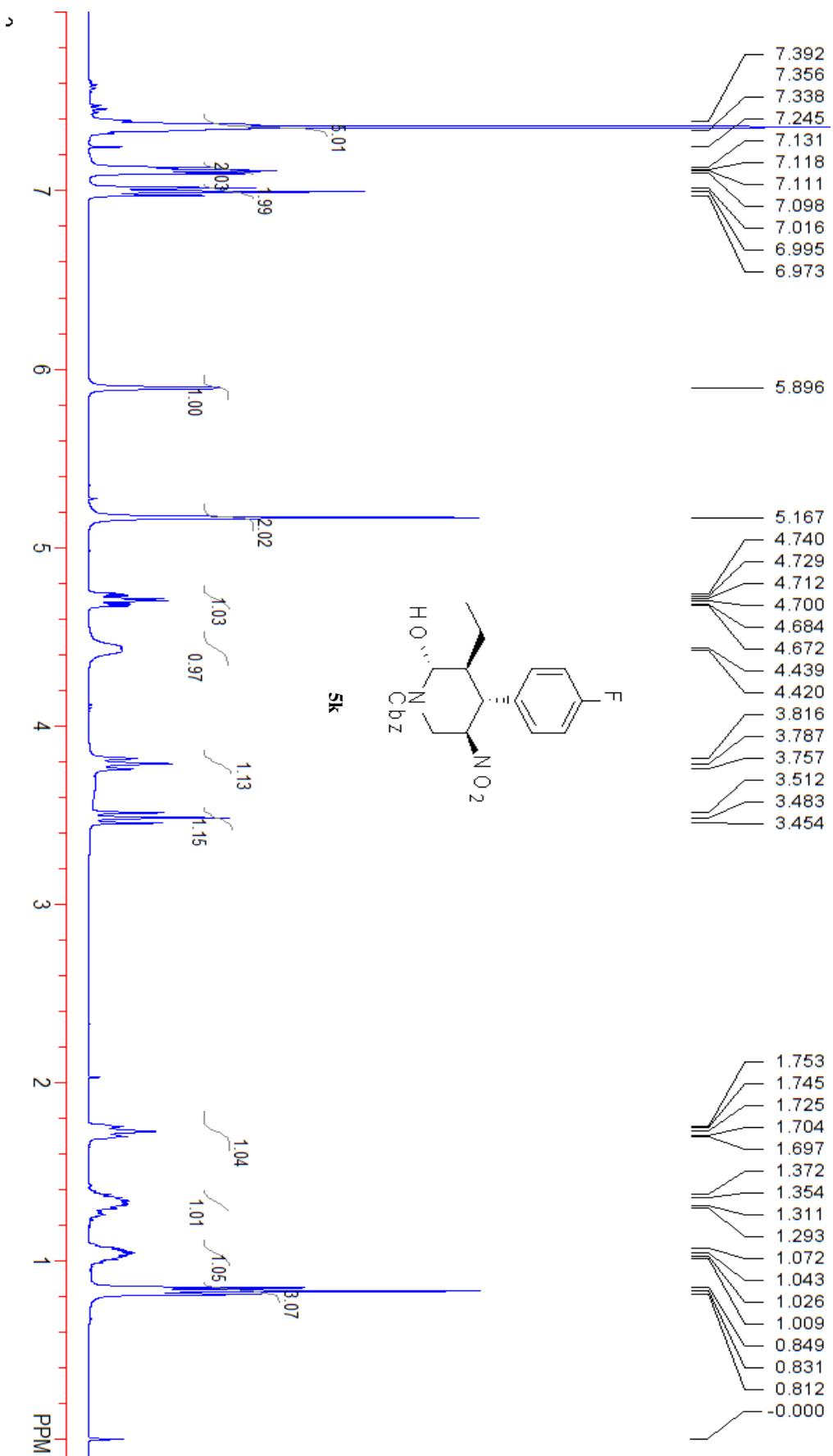
Operator:

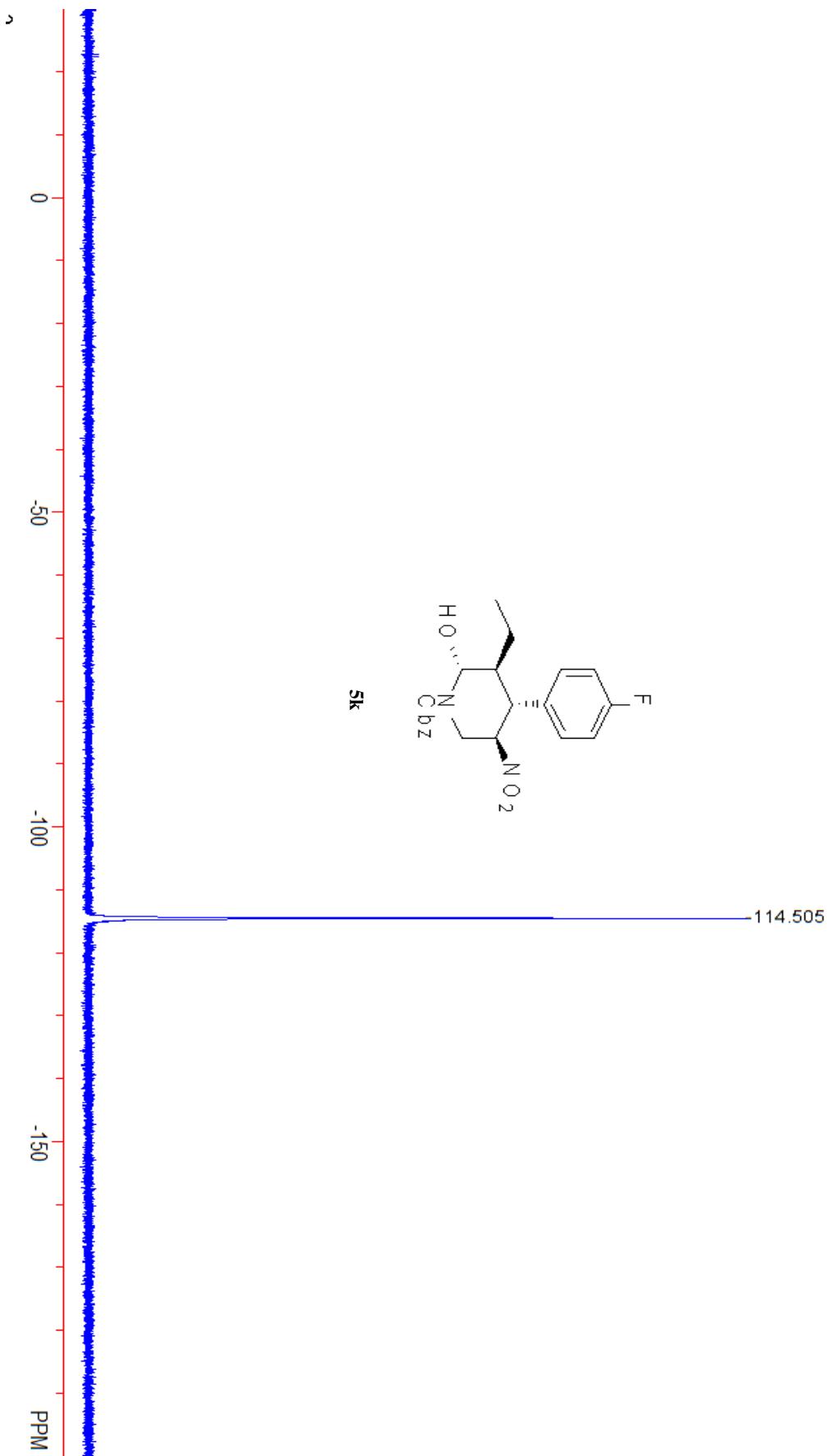
Date:2010-08-25

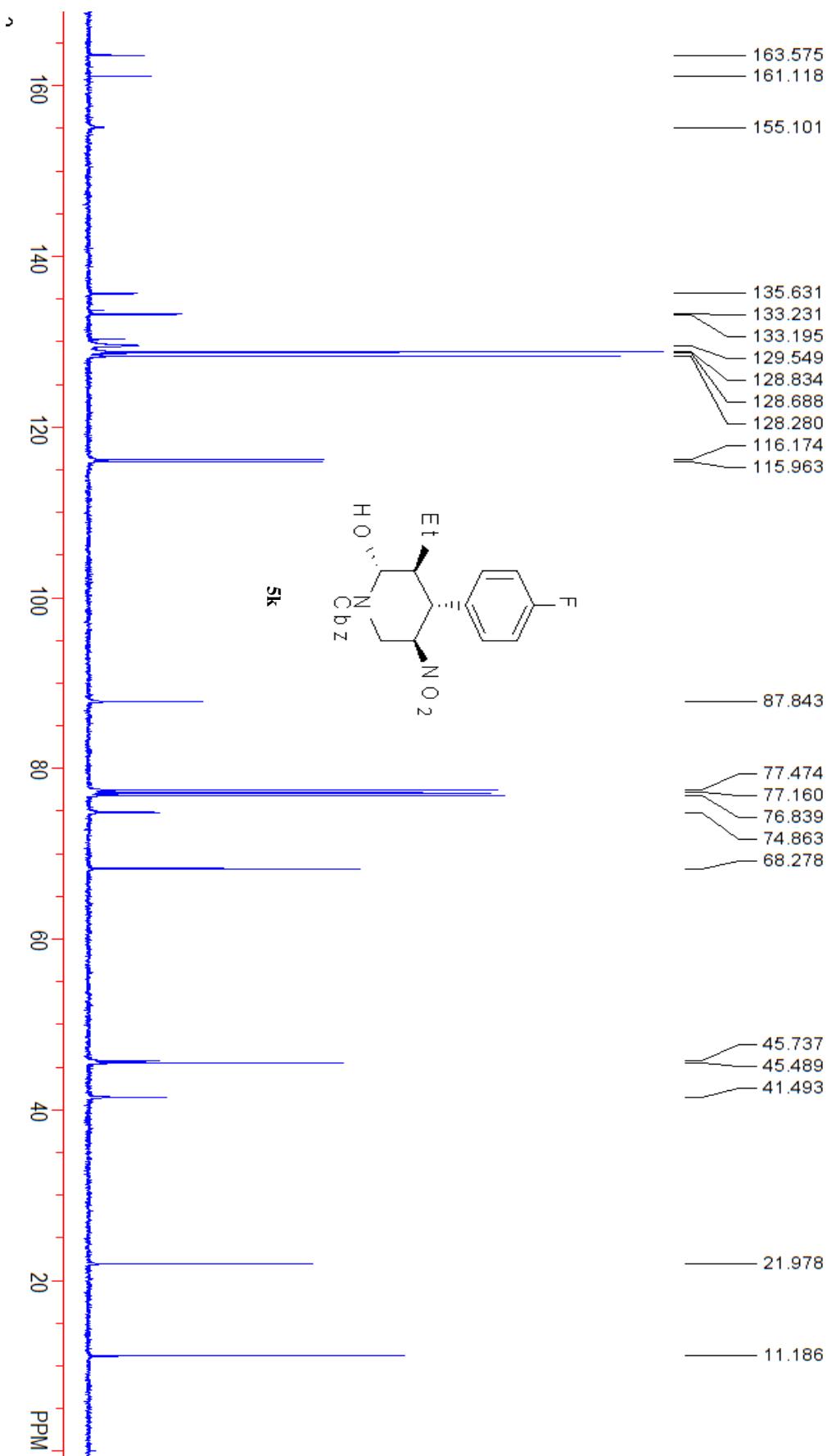
Time:09:31



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	6.627	272.0	5650.9	0.2090
2	2	10.963	121082.4	2698644.4	99.7910
Total			121354.4	2704295.3	100.0000







HPLC Report

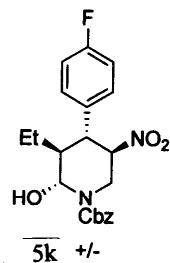
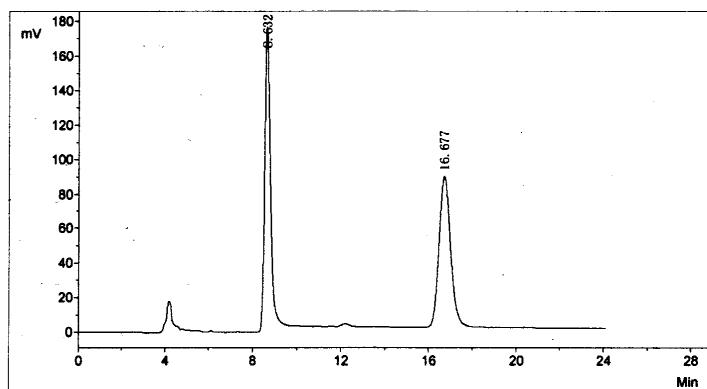
Sample Name:

Data File:ZSL6-97+-PC-2820.7214.che

Operator:

Date:2010-11-23

Time:13:29



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	8.632	174752.3	3111513.3	49.1262
2	2	16.677	87305.2	3222207.0	50.8738
Total			262057.5	6333720.3	100.0000

HPLC Report

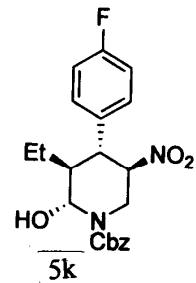
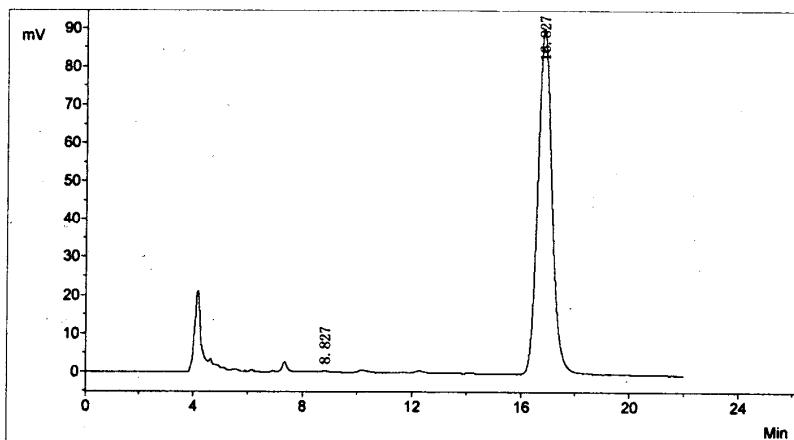
Sample Name:

Data File: ZSL6-98.che

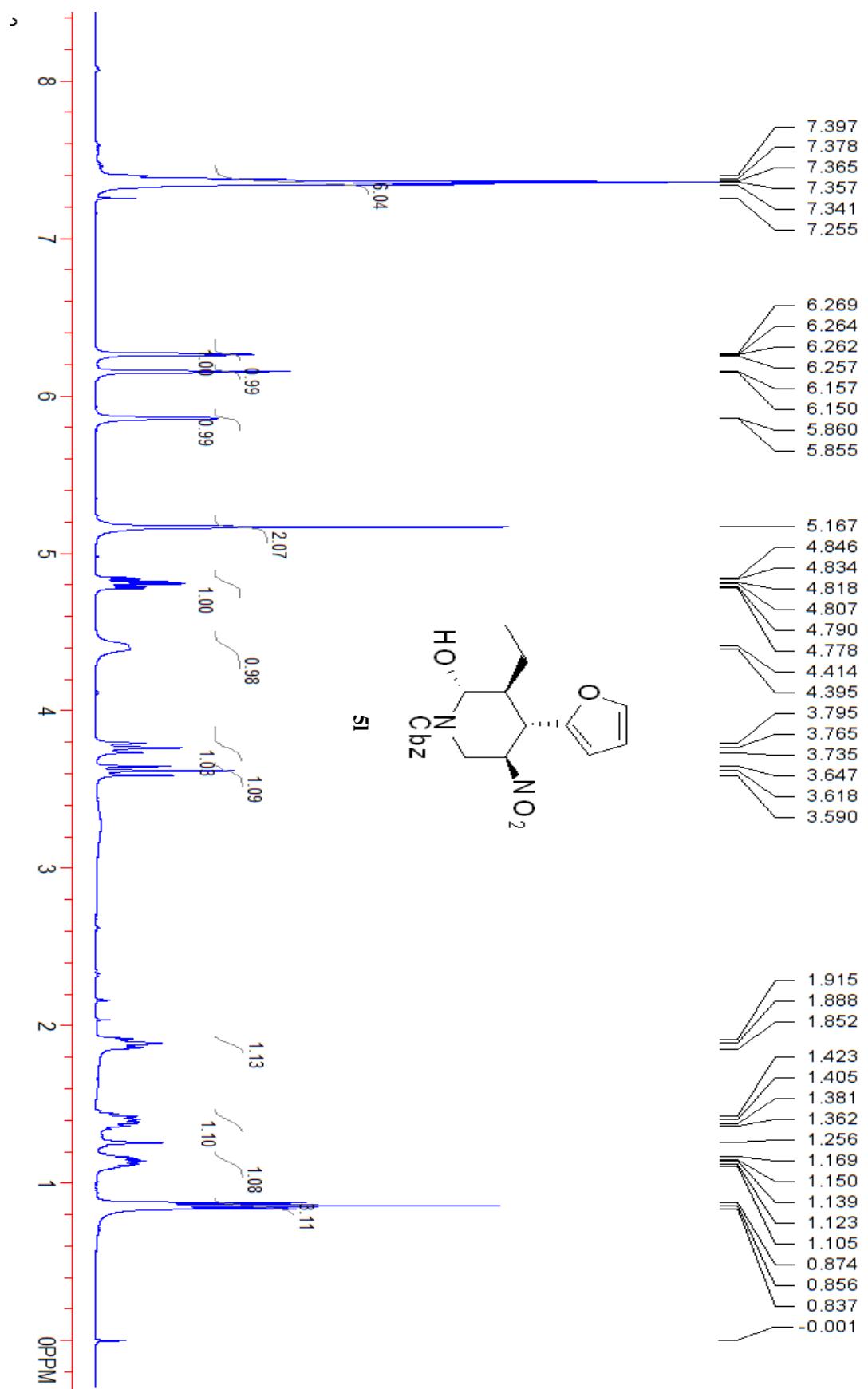
Operator:

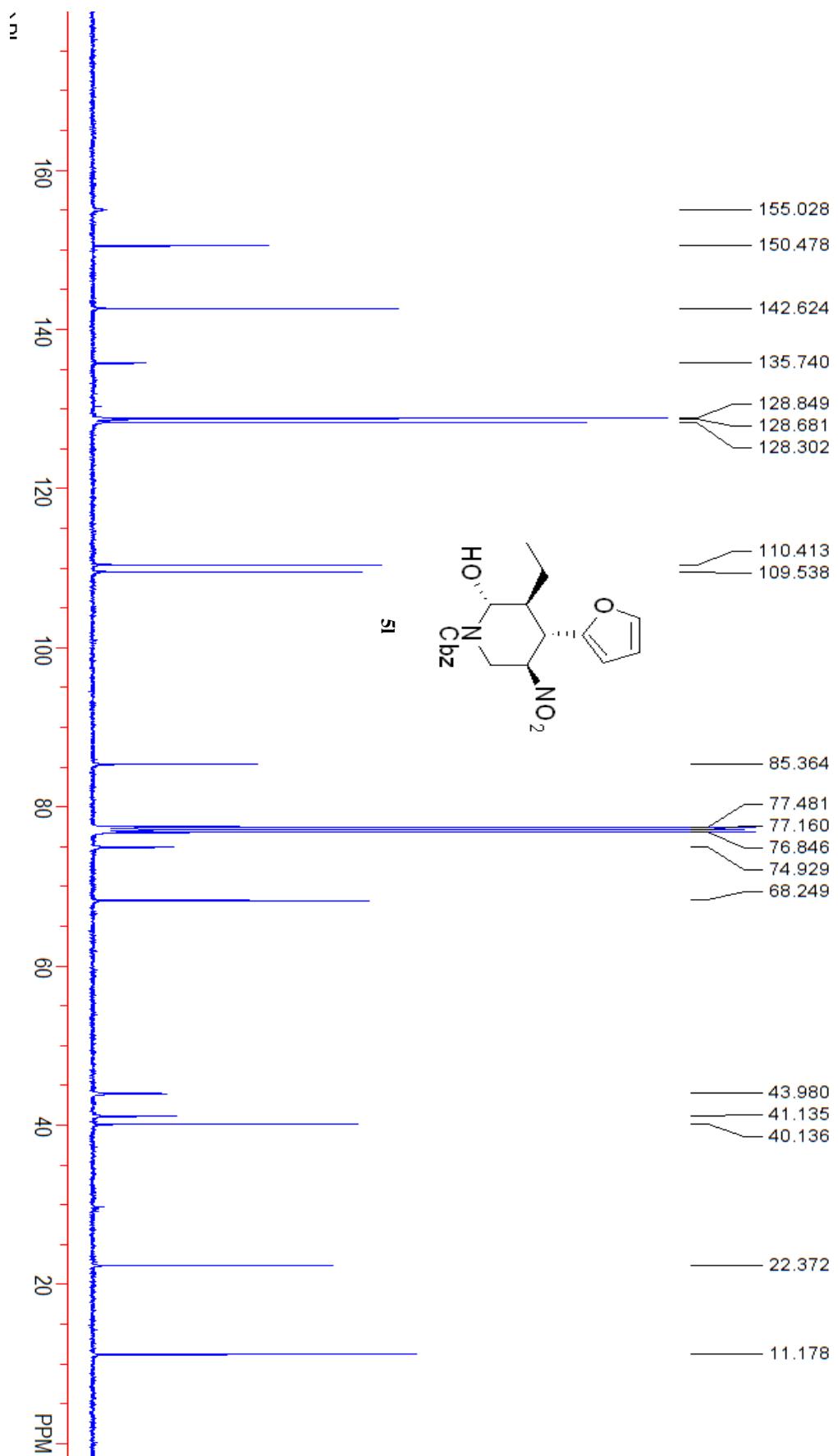
Date: 2010-11-23

Time: 13:57



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	8.827	246.4	1609.5	0.0489
2	2	16.827	90385.6	3292203.7	99.9511
Total			90632.0	3293813.2	100.0000





HPLC Report

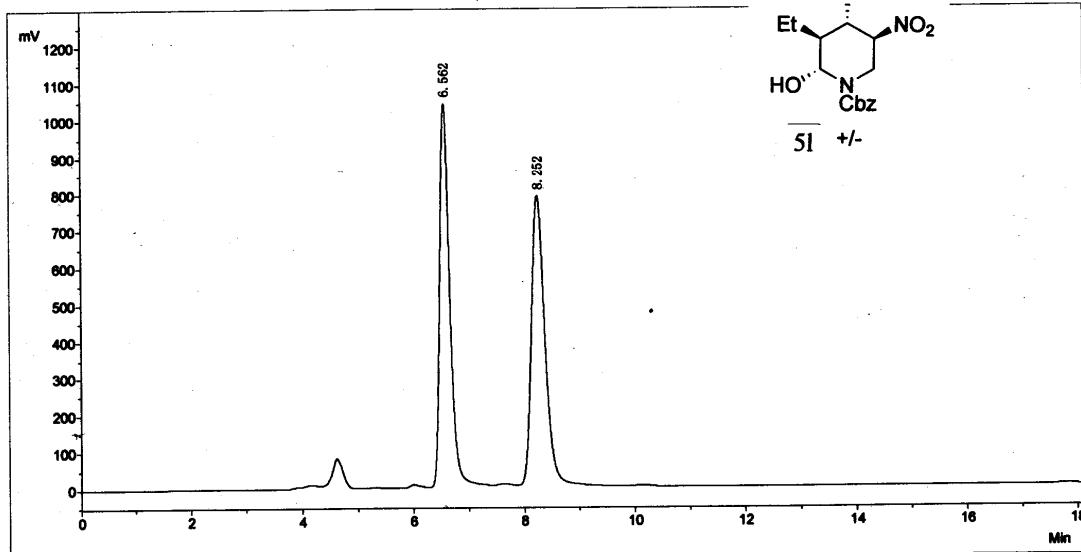
Sample Name:

Data File:ZSL-6-52+- PC-2 73 214 0.7.che

Operator:

Date:2010-08-20

Time:09:32



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		6.562	1039130.9	12968537.7	49.4847
2	2		8.252	783219.4	13238615.3	50.5153
Total				1822350.3	26207153.0	100.0000

HPLC Report

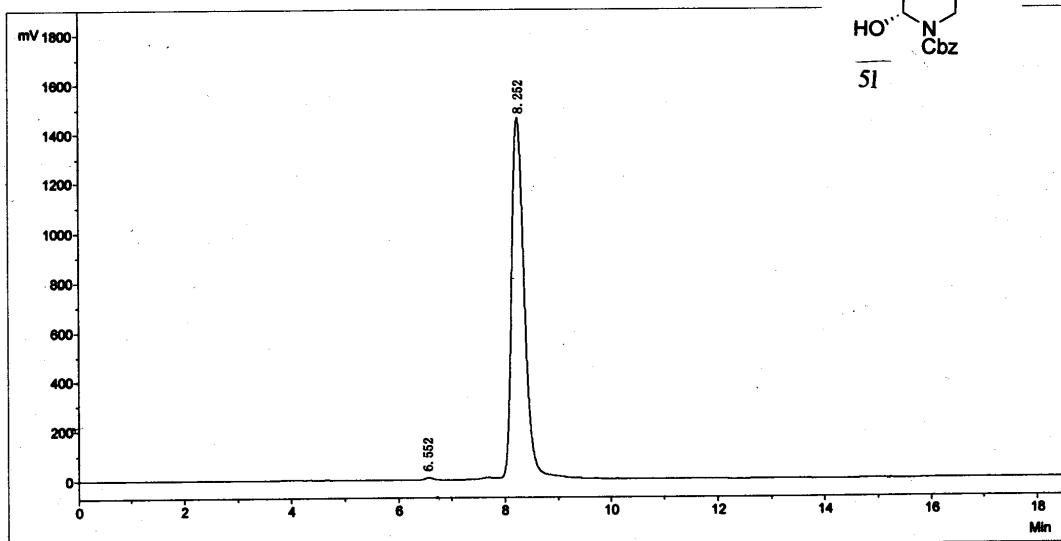
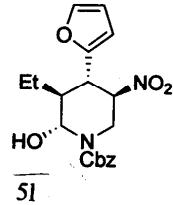
Sample Name:

Data File:ZSL-6-57.che

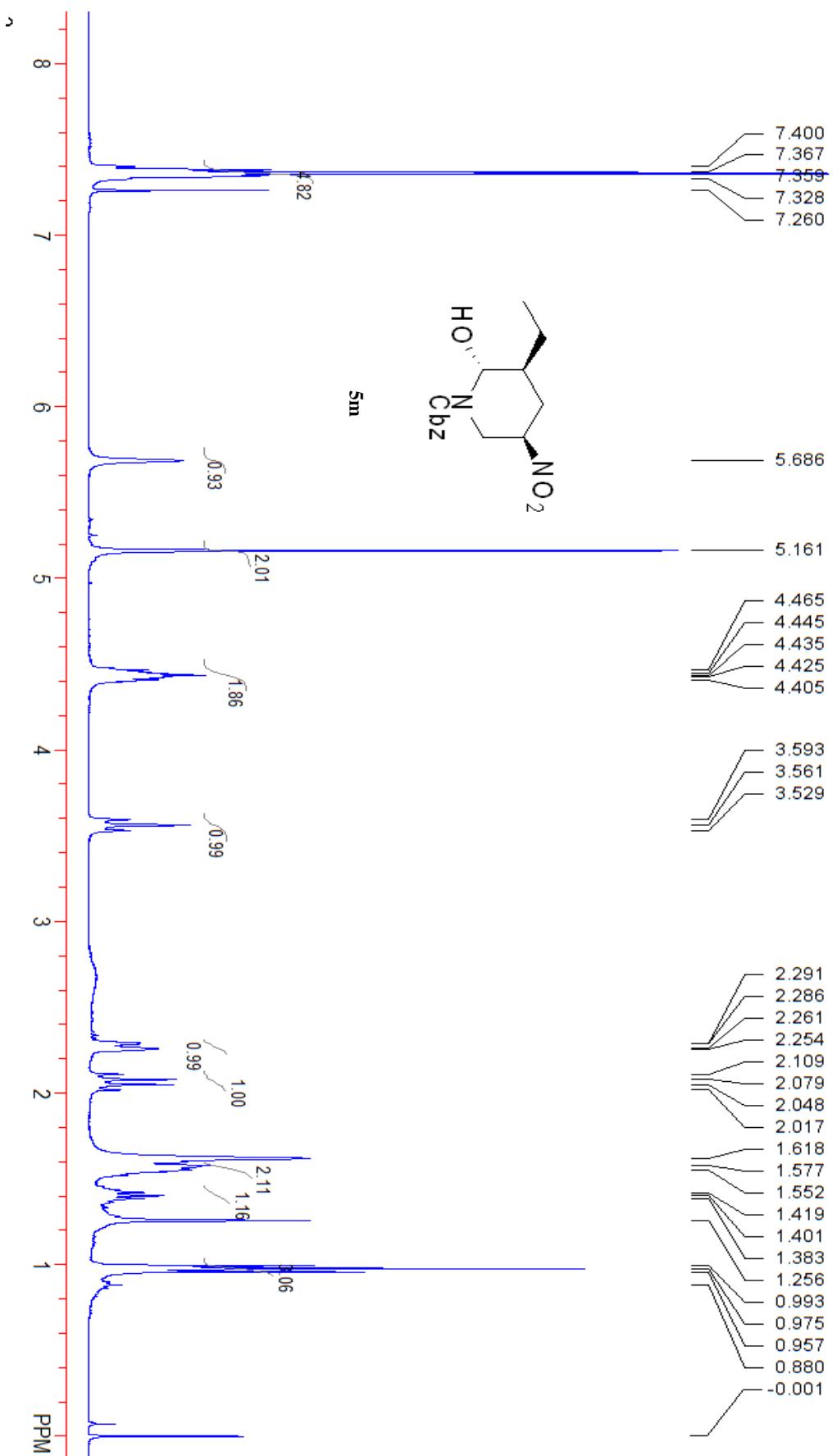
Operator:

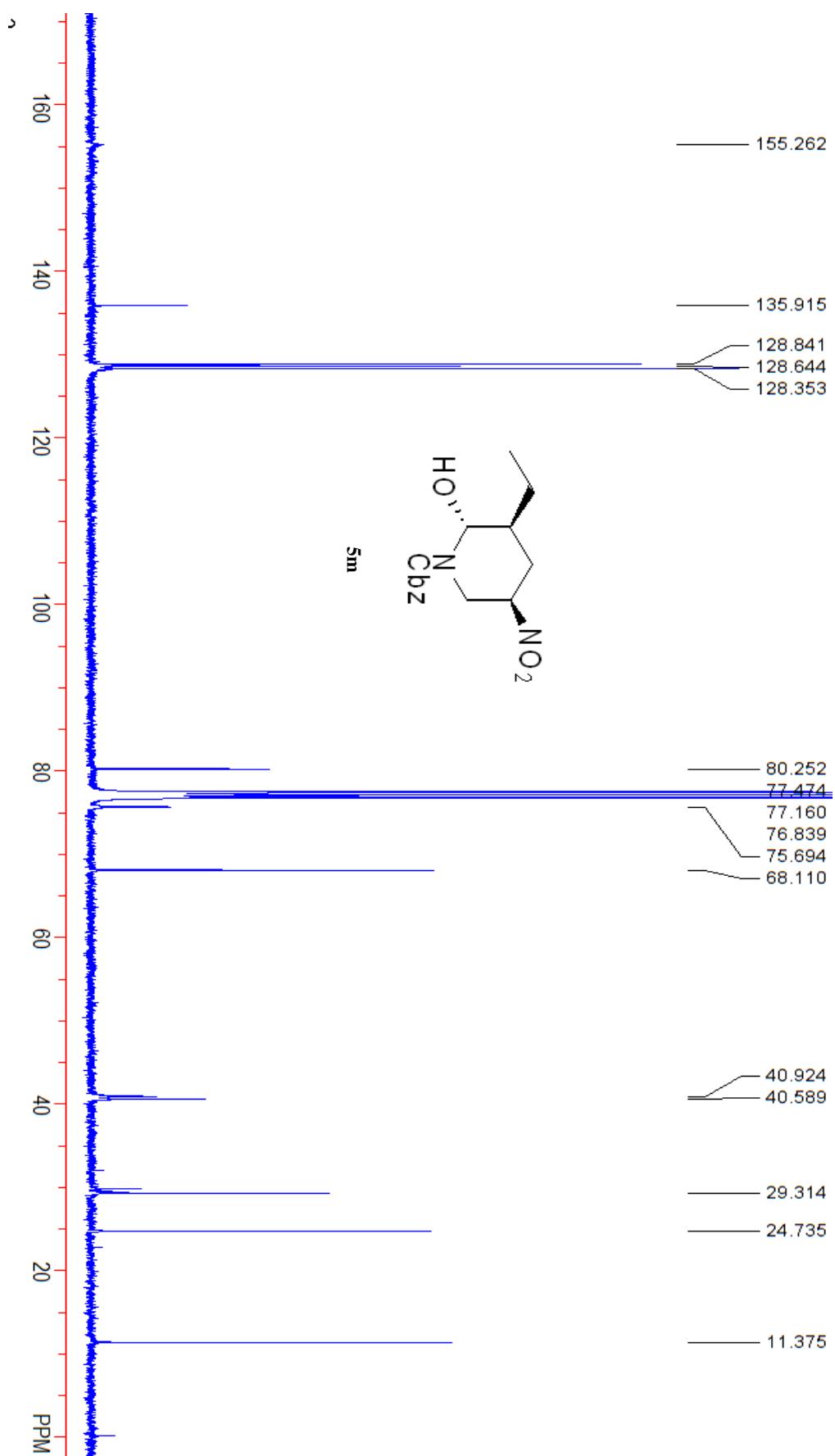
Date:2010-08-20

Time:09:53



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		6.552	9232.6	106097.7	0.4436
2	2		8.252	1448408.9	23811995.8	99.5564
Total				1457641.5	23918093.5	100.0000





HPLC Report

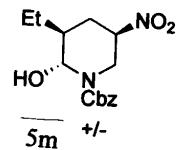
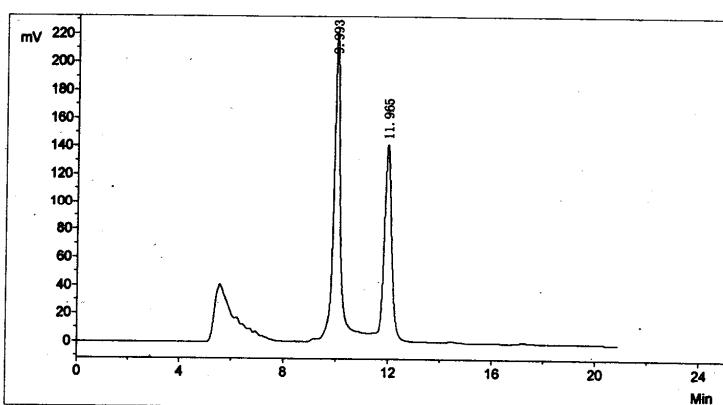
Sample Name:

Data File:ZSL6-75+-PA-2730.5214.che

Operator:

Date:2010-10-13

Time:10:01



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		9.993	215869.9	3477952.2	57.8622
2	2		11.965	136573.4	2532800.5	42.1378
Total				352443.2	6010752.6	100.0000

HPLC Report

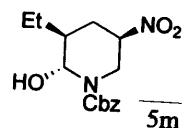
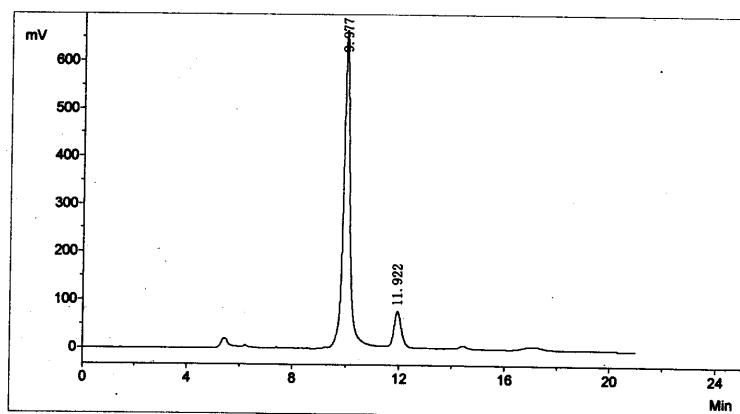
Sample Name:

Data File:ZSL6-78.che

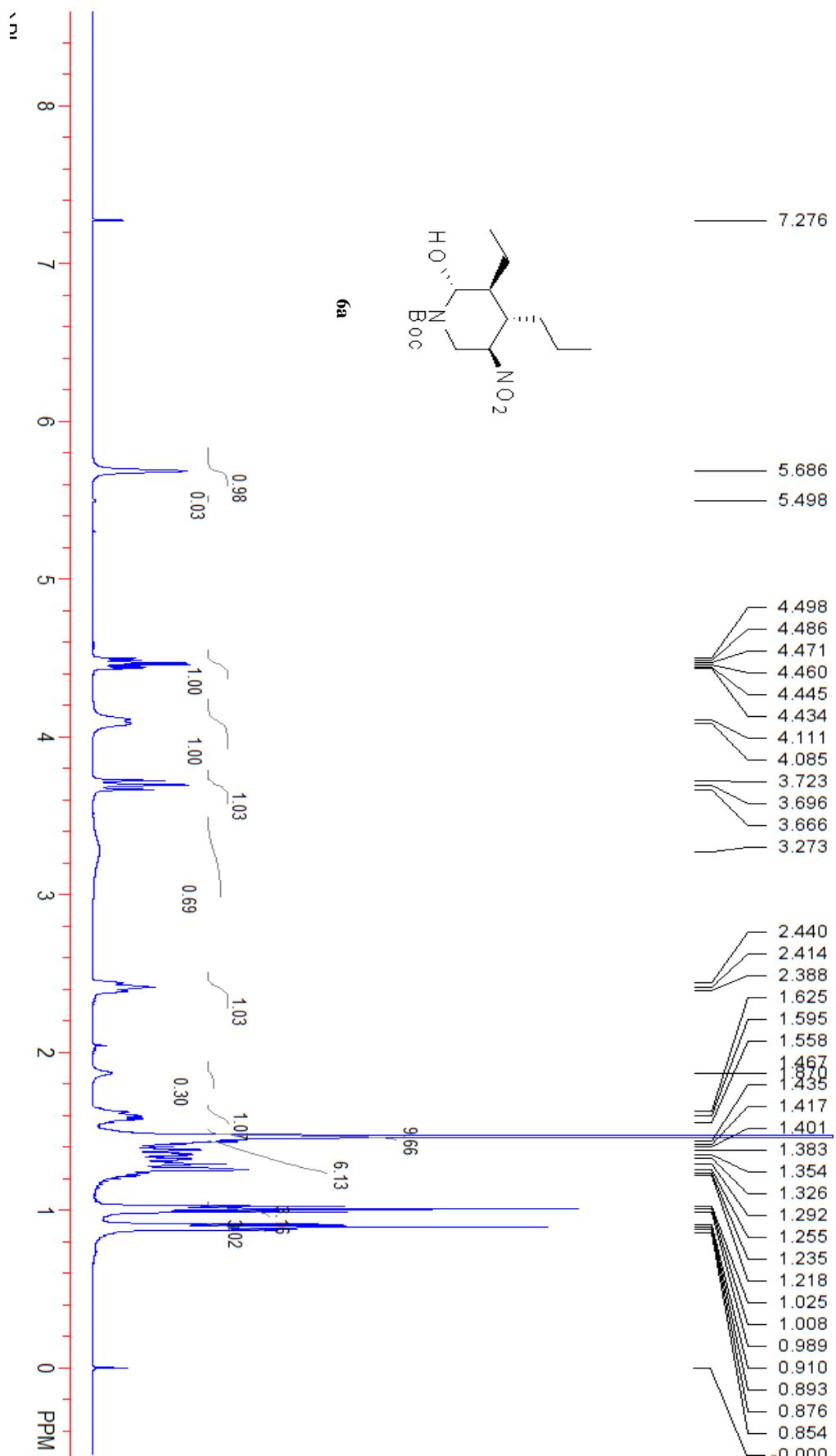
Operator:

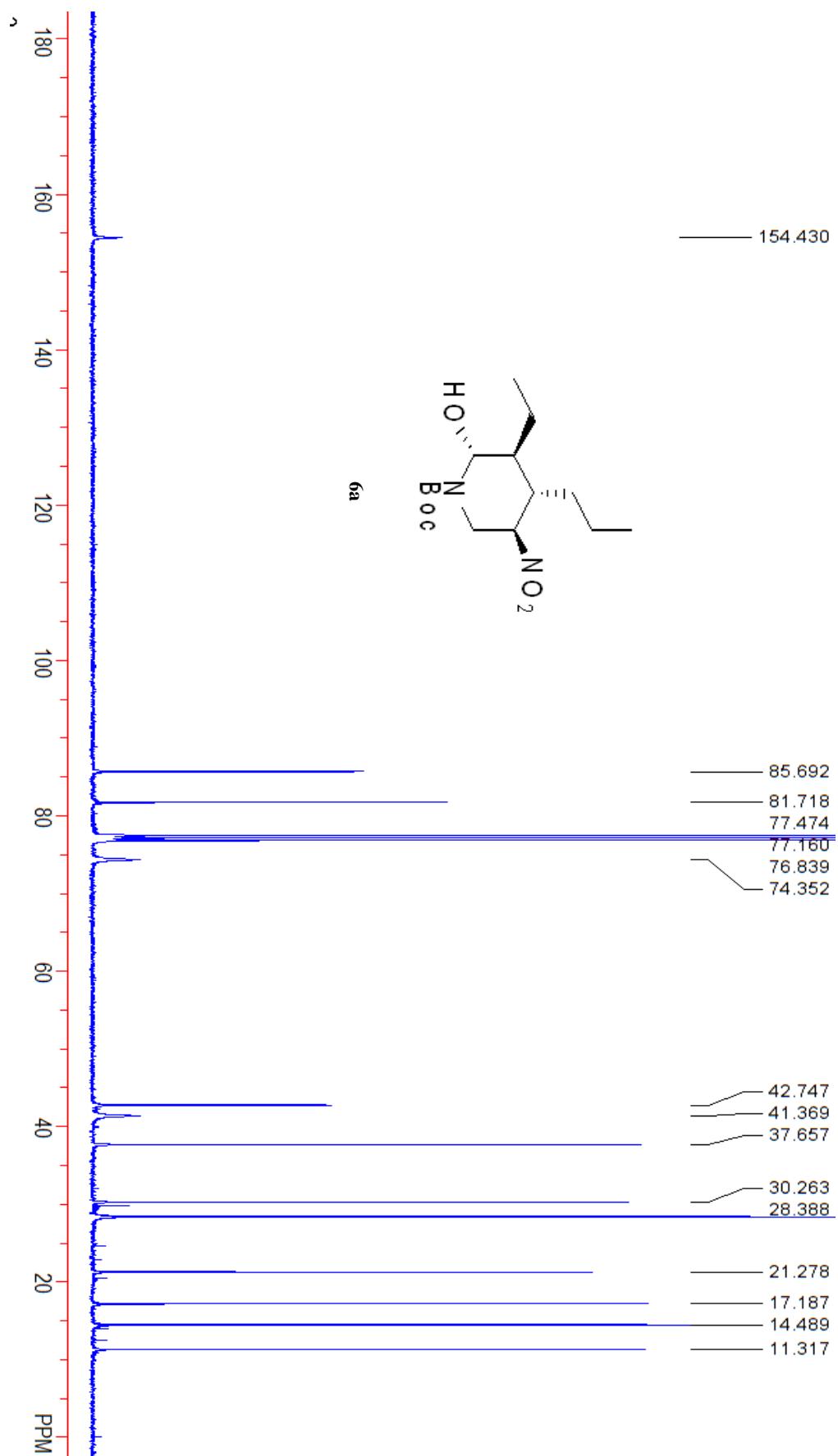
Date:2010-10-13

Time:10:26



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		9.977	659599.3	11319465.2	89.2951
2	2		11.922	71528.3	1357006.8	10.7049
Total				731127.6	12676472.0	100.0000





HPLC REPORT

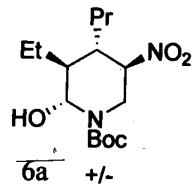
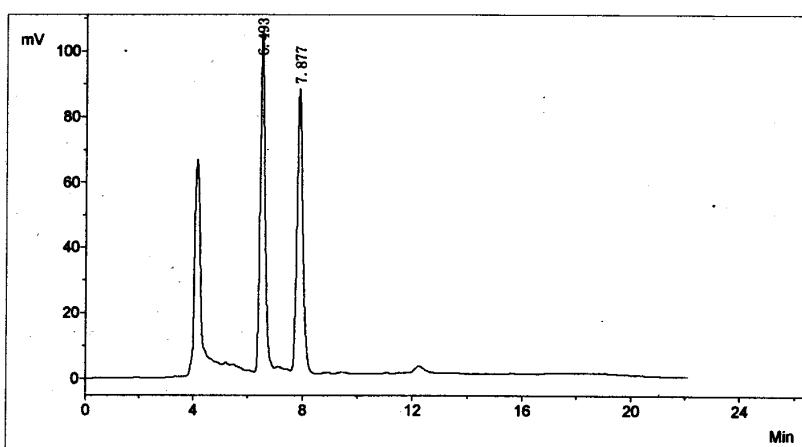
Data File:ZSL6-39-2+-SINO-AD910.7214.che

Sample name:

Date:2010-08-26

Time :08:58

Operator:



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		6.493	102897.1	1258809.9	50.0411
2	2		7.877	85796.0	1256741.0	49.9589
Total				188693.0	2515550.9	100.0000

HPLC REPORT

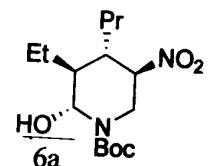
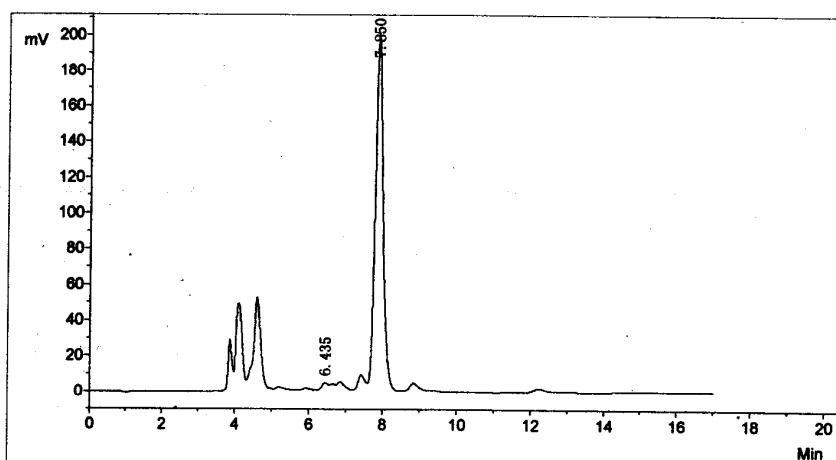
Data File:6-41-2.che

Sample name:

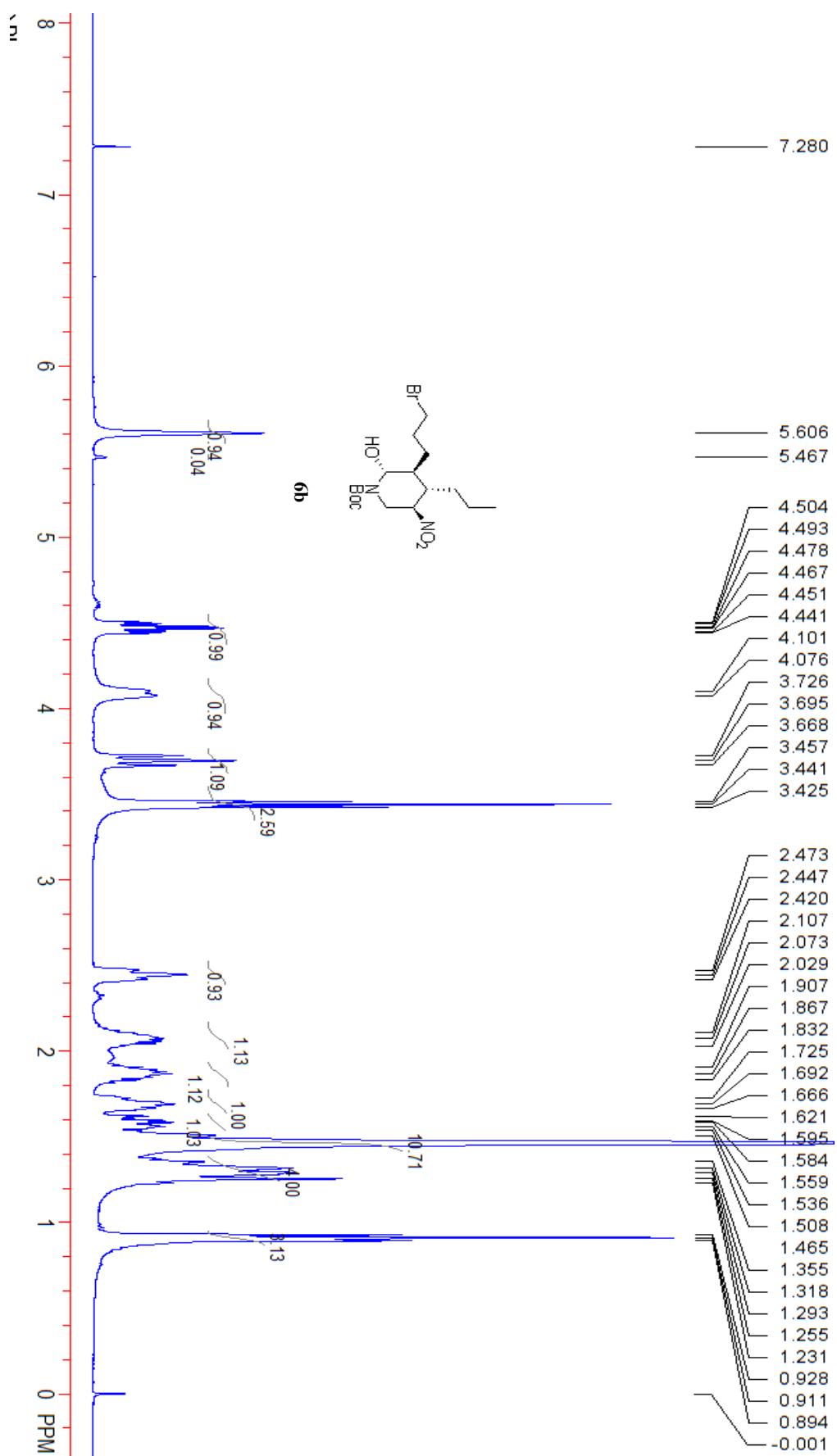
Date:2010-08-26

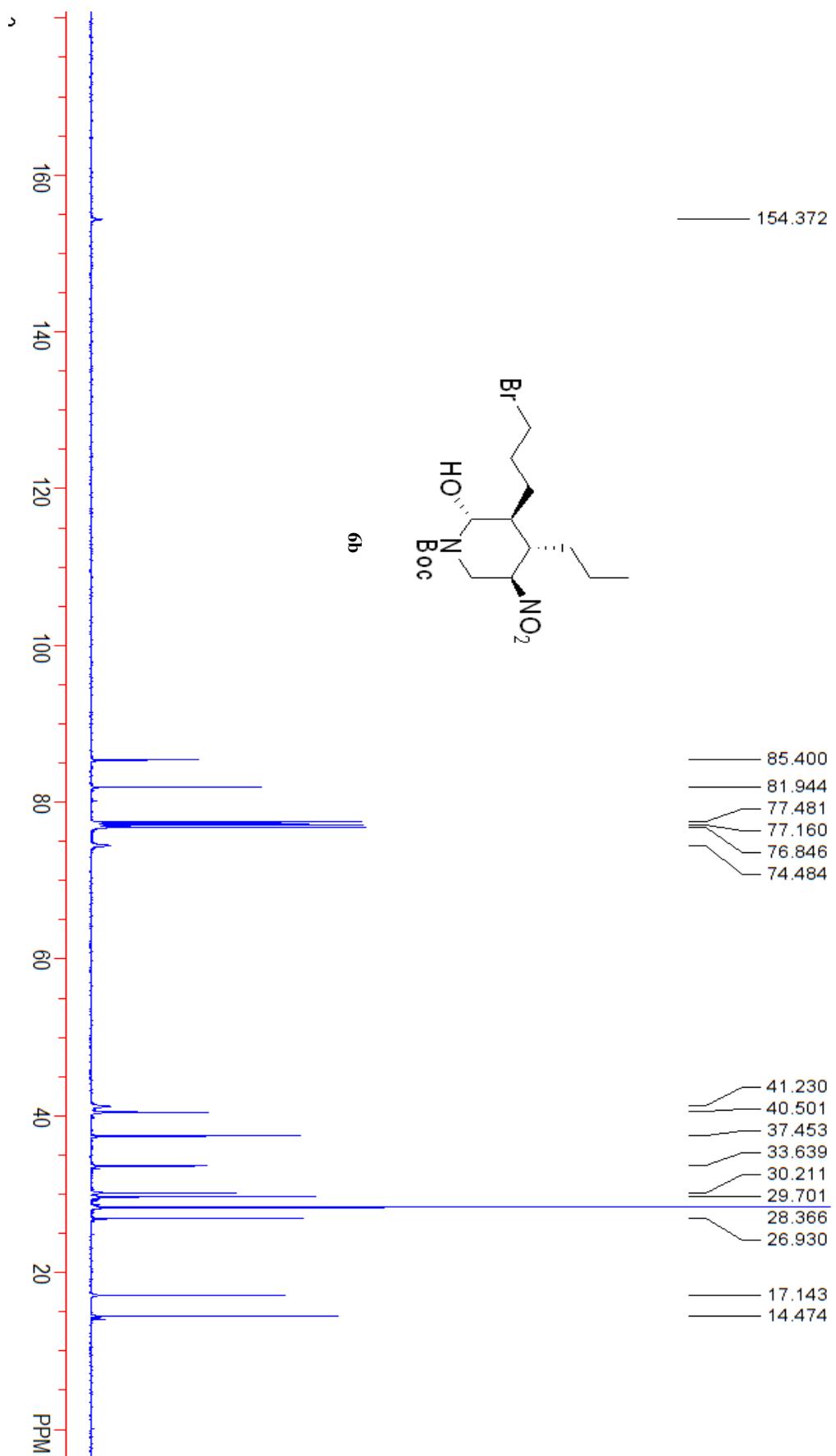
Time :09:27

Operator:



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		6.435	807.1	3981.3	0.1446
2	2		7.850	196129.3	2748652.6	99.8554
Total				196936.4	2752633.9	100.0000





HPLC REPORT

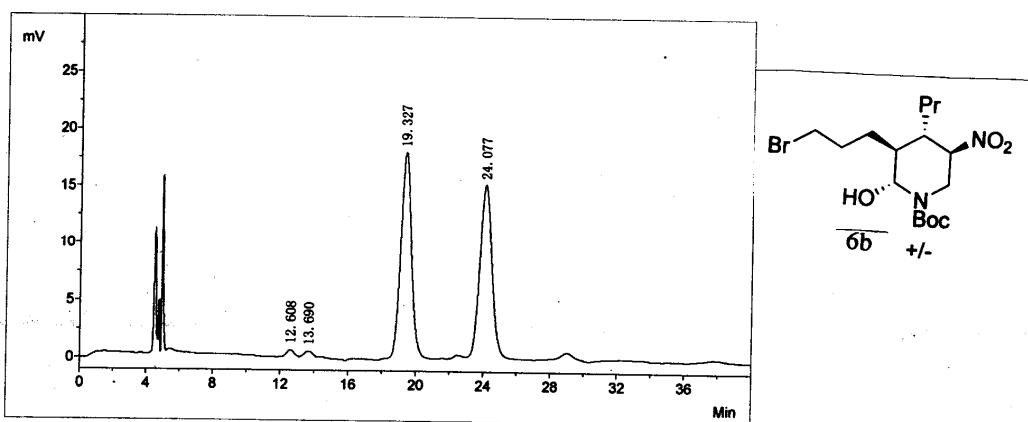
Data File:ZSL-6-44+- IC 982 214 0.7.che

Sample name:

Date:2010-08-16

Time :15:56

Operator:



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		12.608	579.8	17664.9	1.1128
2	2		13.690	504.5	16716.8	1.0531
3	3		19.327	18018.8	799430.5	50.3596
4	4		24.077	15054.6	753631.6	47.4745
Total				34157.7	1587443.7	100.0000

HPLC REPORT

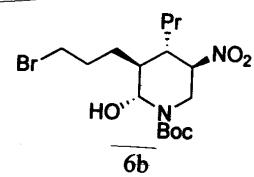
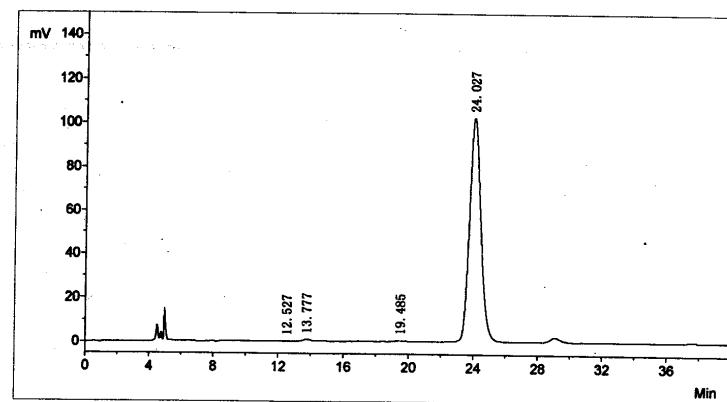
Data File:ZSL-6-42.che

Date:2010-08-16

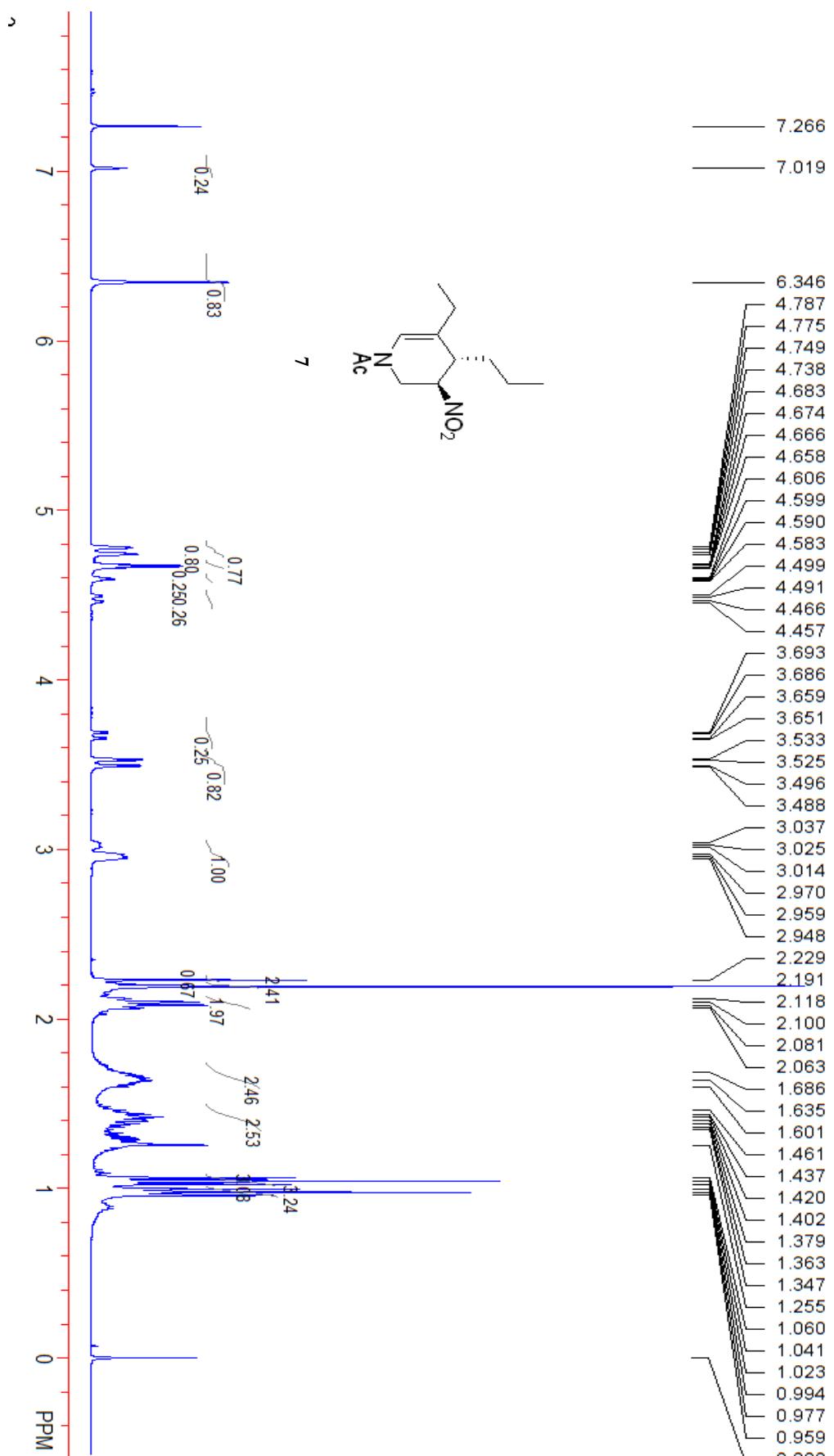
Operator:

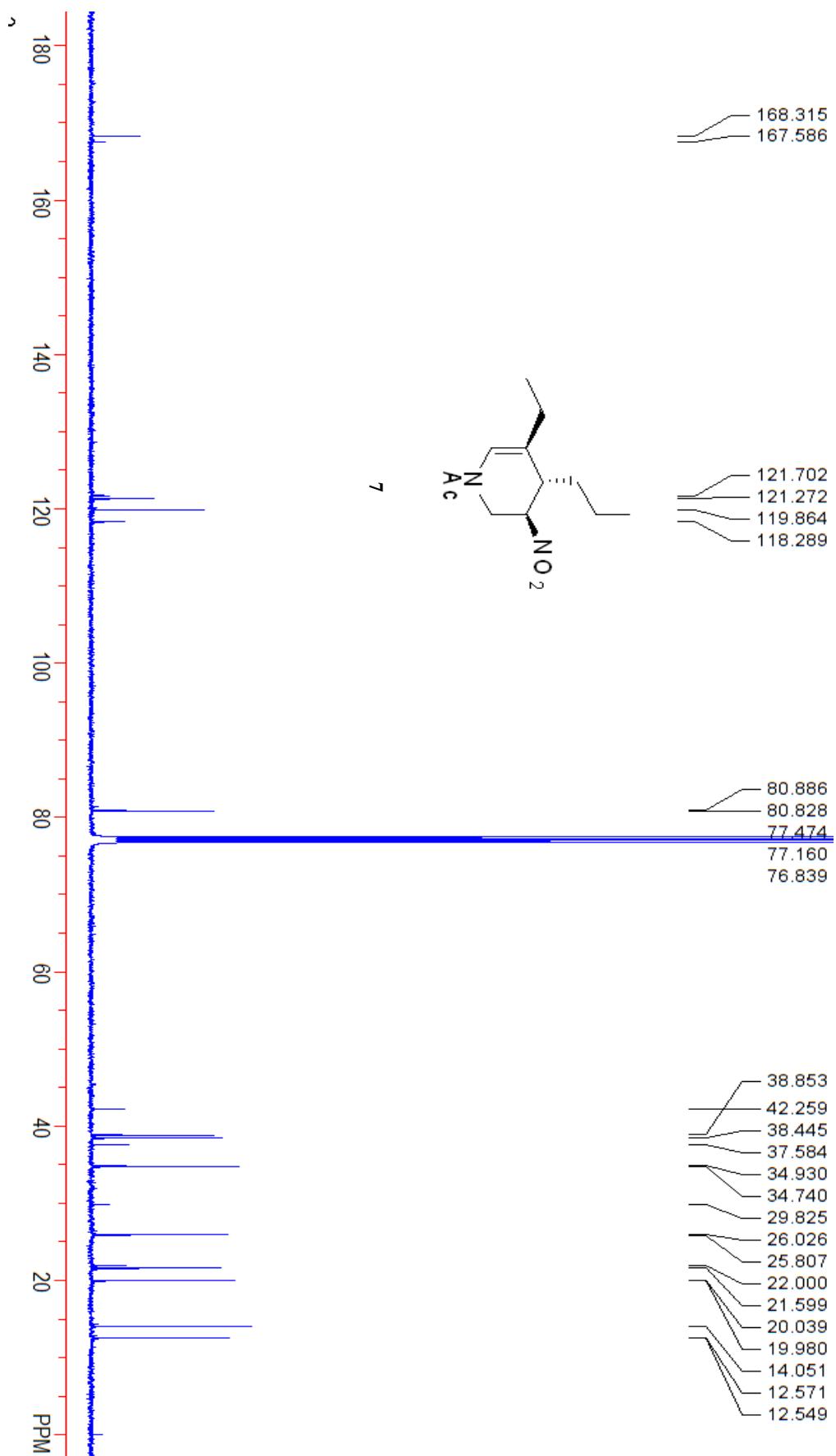
Sample name:

Time :16:40



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		12.527	55.6	1831.4	0.0367
2	2		13.777	817.9	24950.0	0.4996
3	3		19.485	340.9	19602.6	0.3925
4	4		24.027	101902.7	4947892.8	99.0713
Total				103117.1	4994276.8	100.0000





HPLC Report

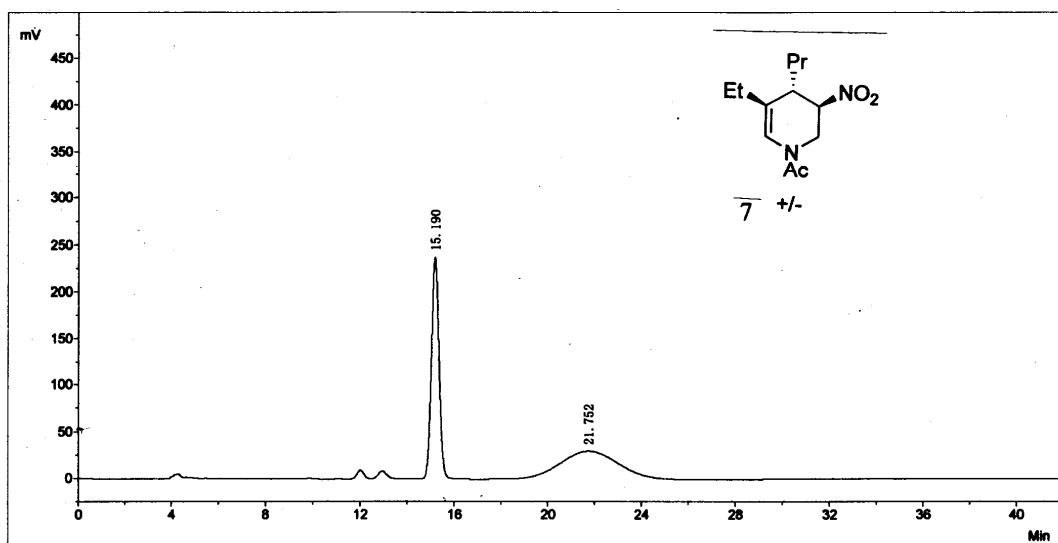
Sample Name:

Data File:ZSL-6-45-0+- PA-2 73 214 0.7.che

Operator:

Date:2010-08-20

Time:13:53



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		15.190	237616.4	5339516.5	50.7161
2	2		21.752	30110.3	5188736.6	49.2839
Total				267726.7	10528253.1	100.0000

HPLC Report

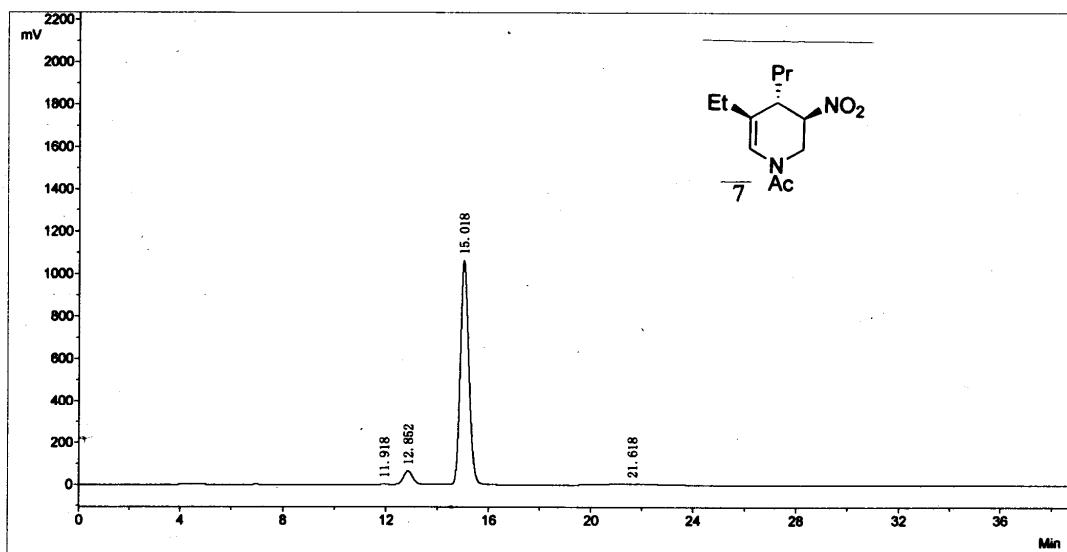
Sample Name:

Data File:ZSL-6-49-0..che

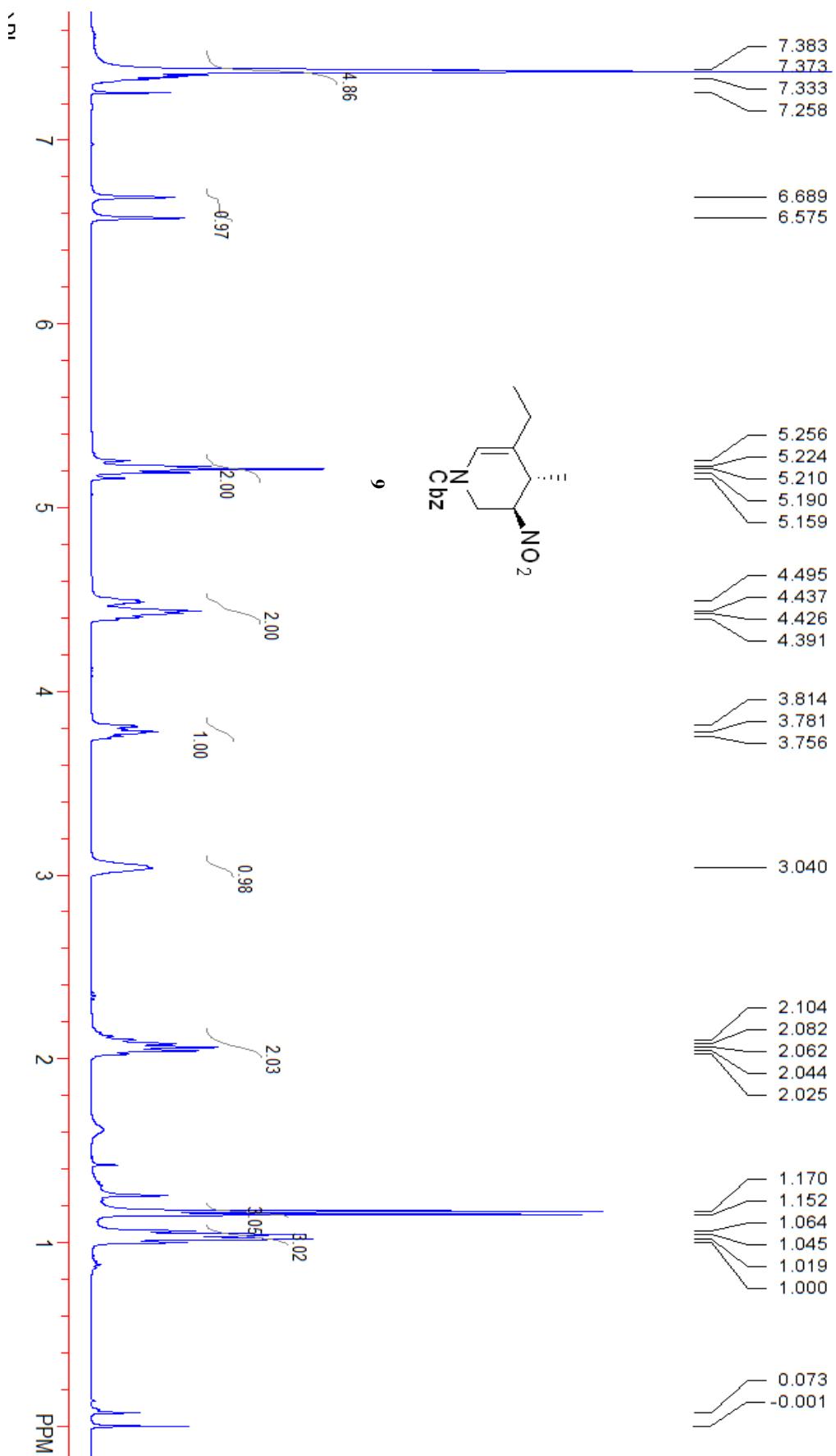
Operator:

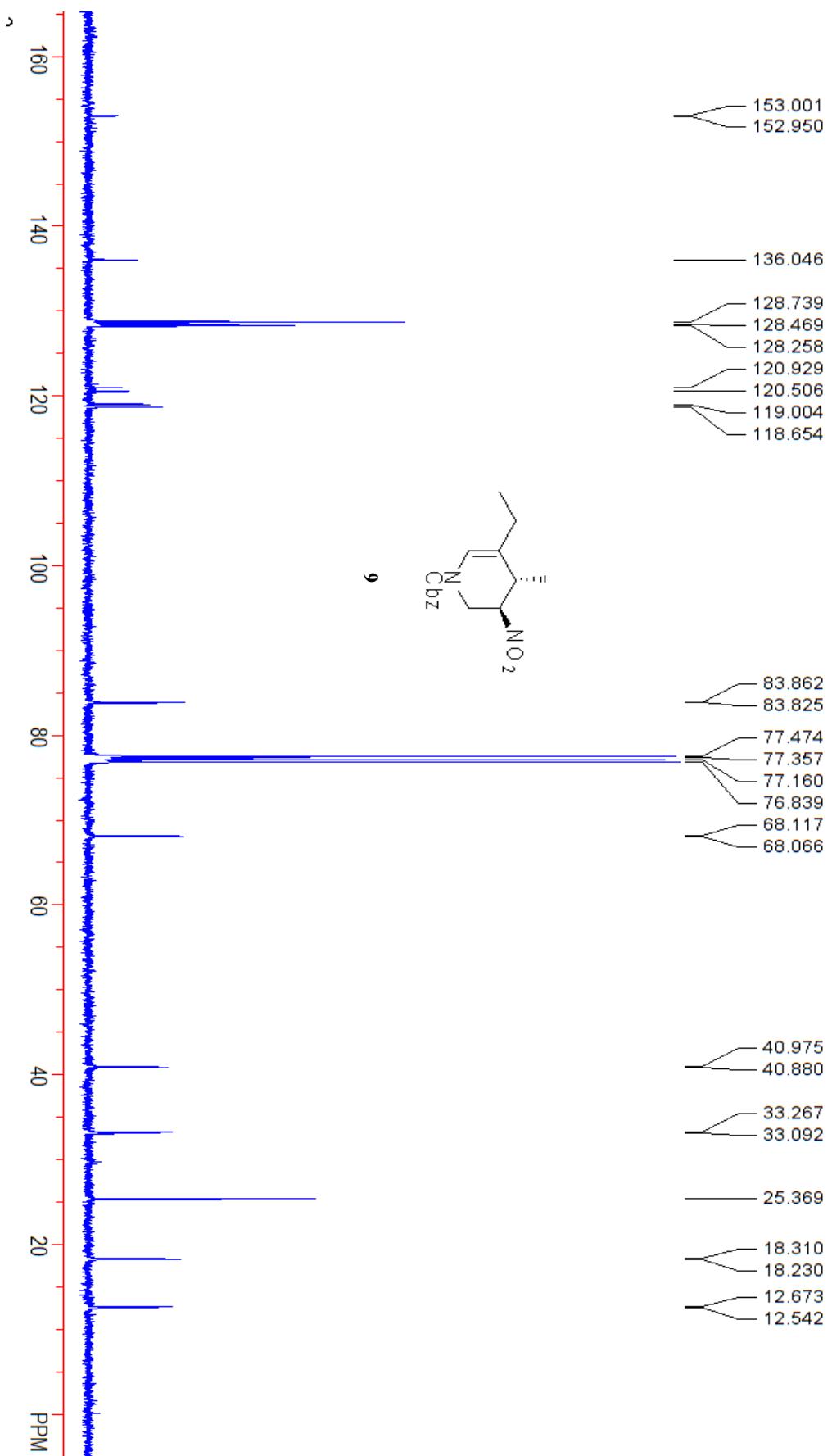
Date:2010-08-20

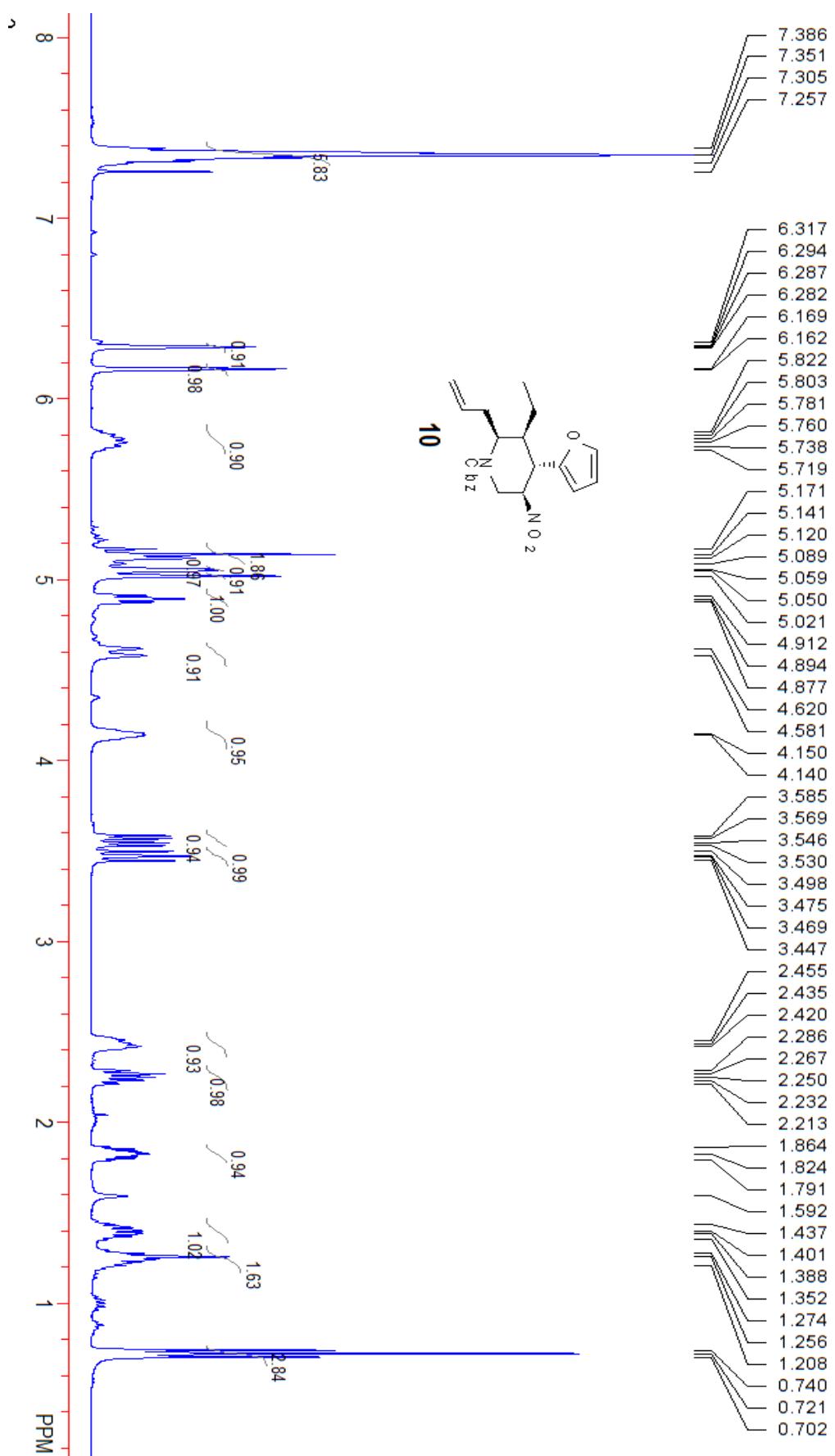
Time:14:37

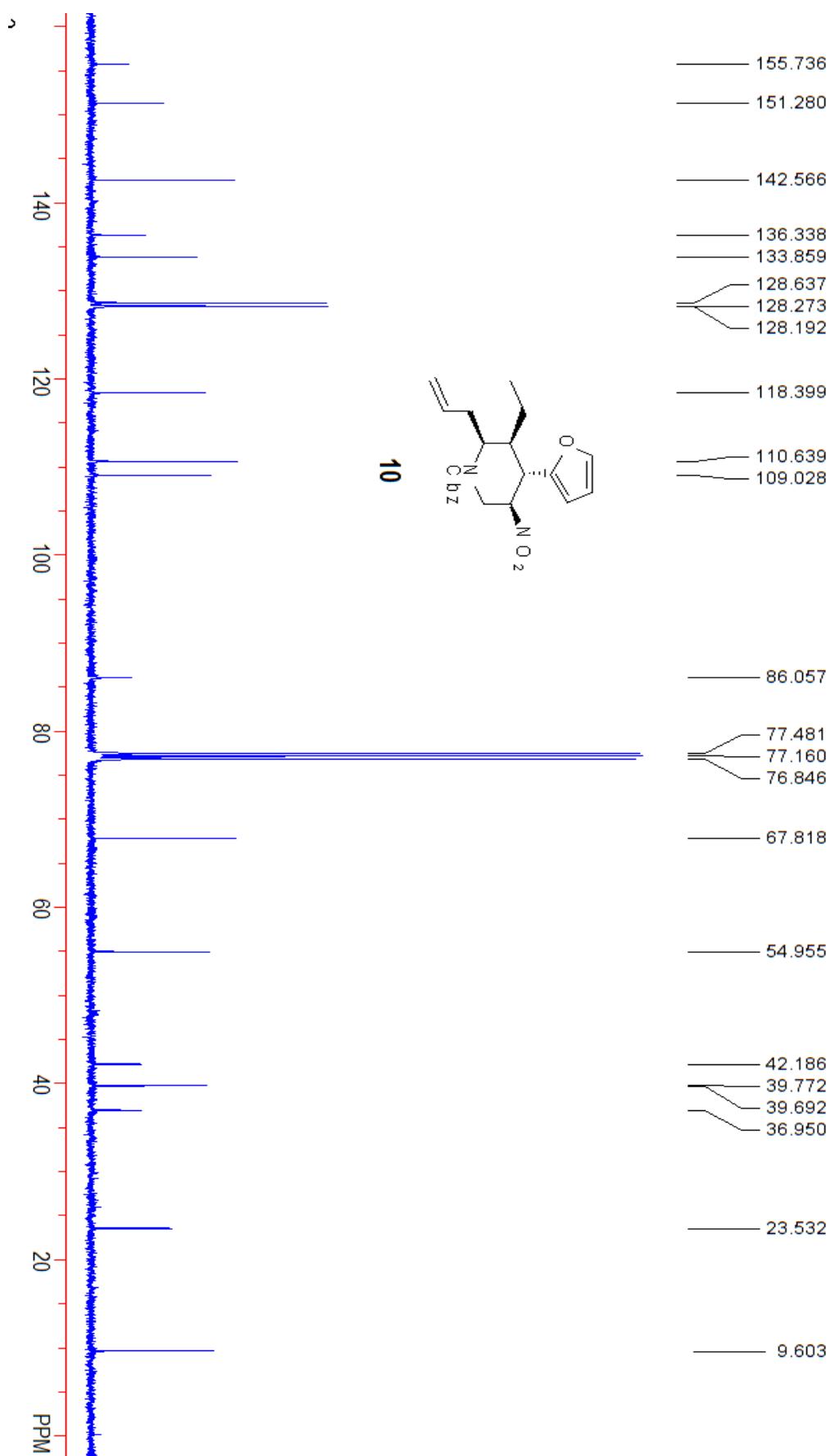


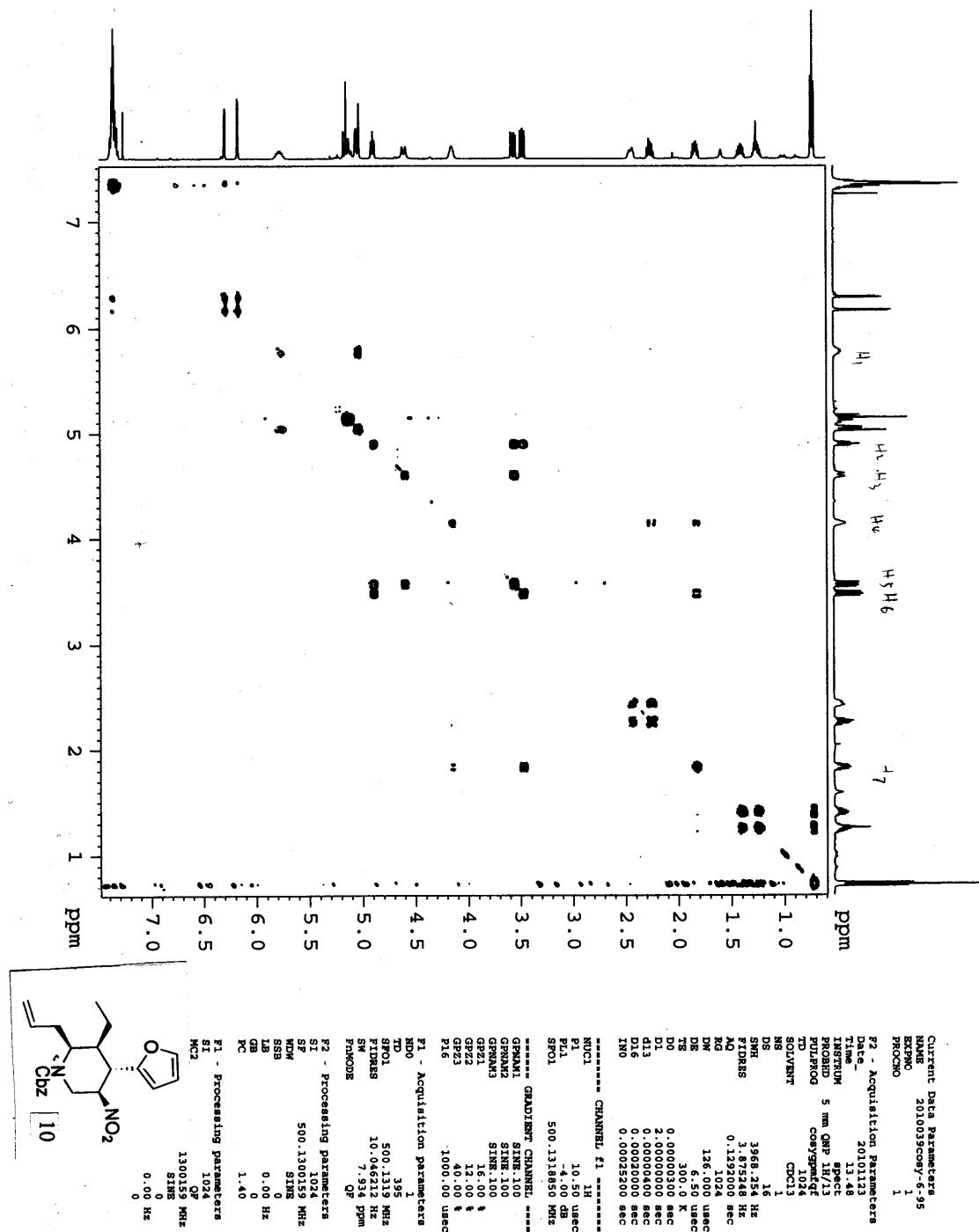
No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		11.918	1116.1	22220.3	0.0819
2	2		12.852	64588.6	1637058.0	6.0356
3	3		15.018	1061127.9	24532317.4	90.4475
4	4		21.618	5505.1	931660.9	3.4349
Total				1132337.8	27123256.6	100.0000

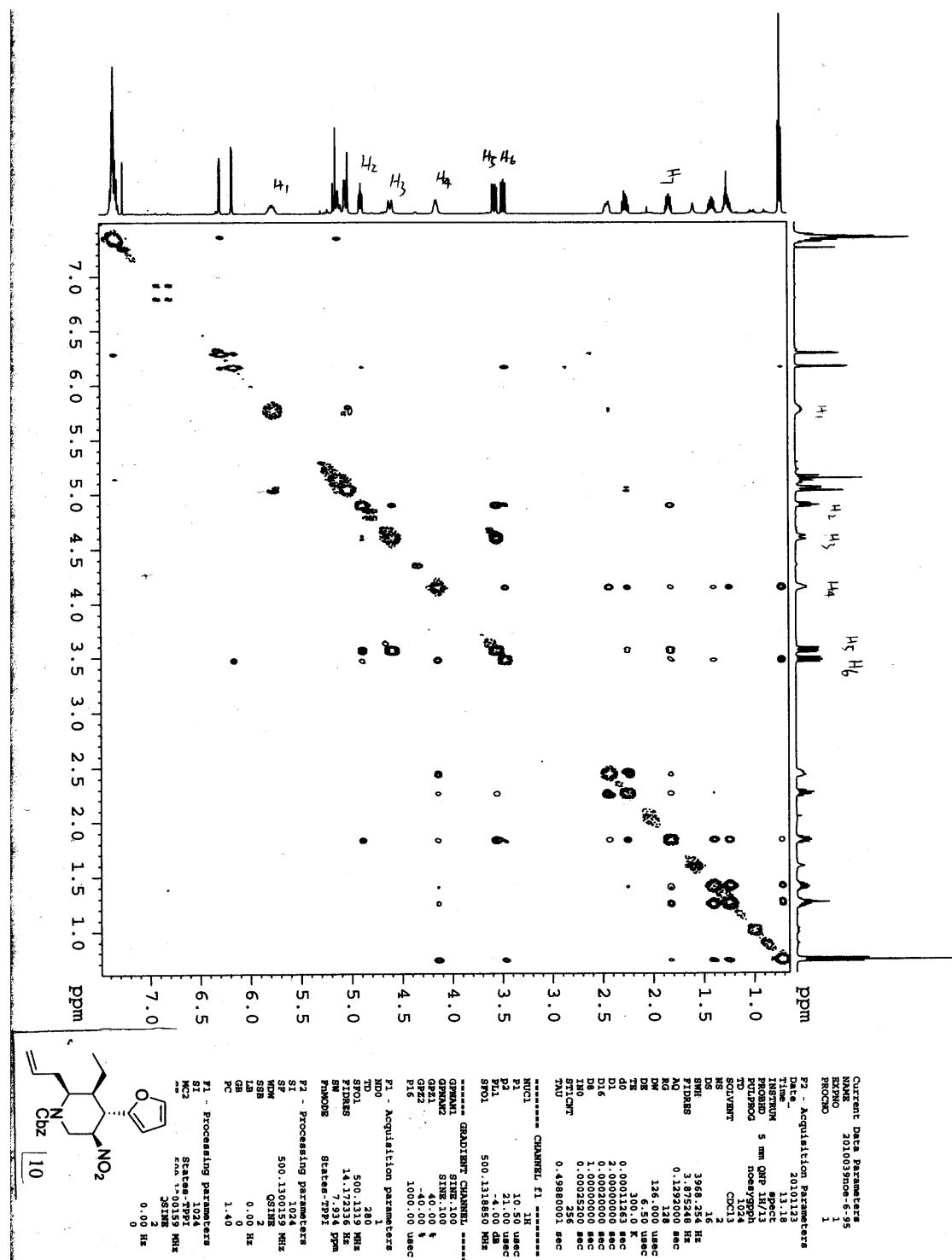


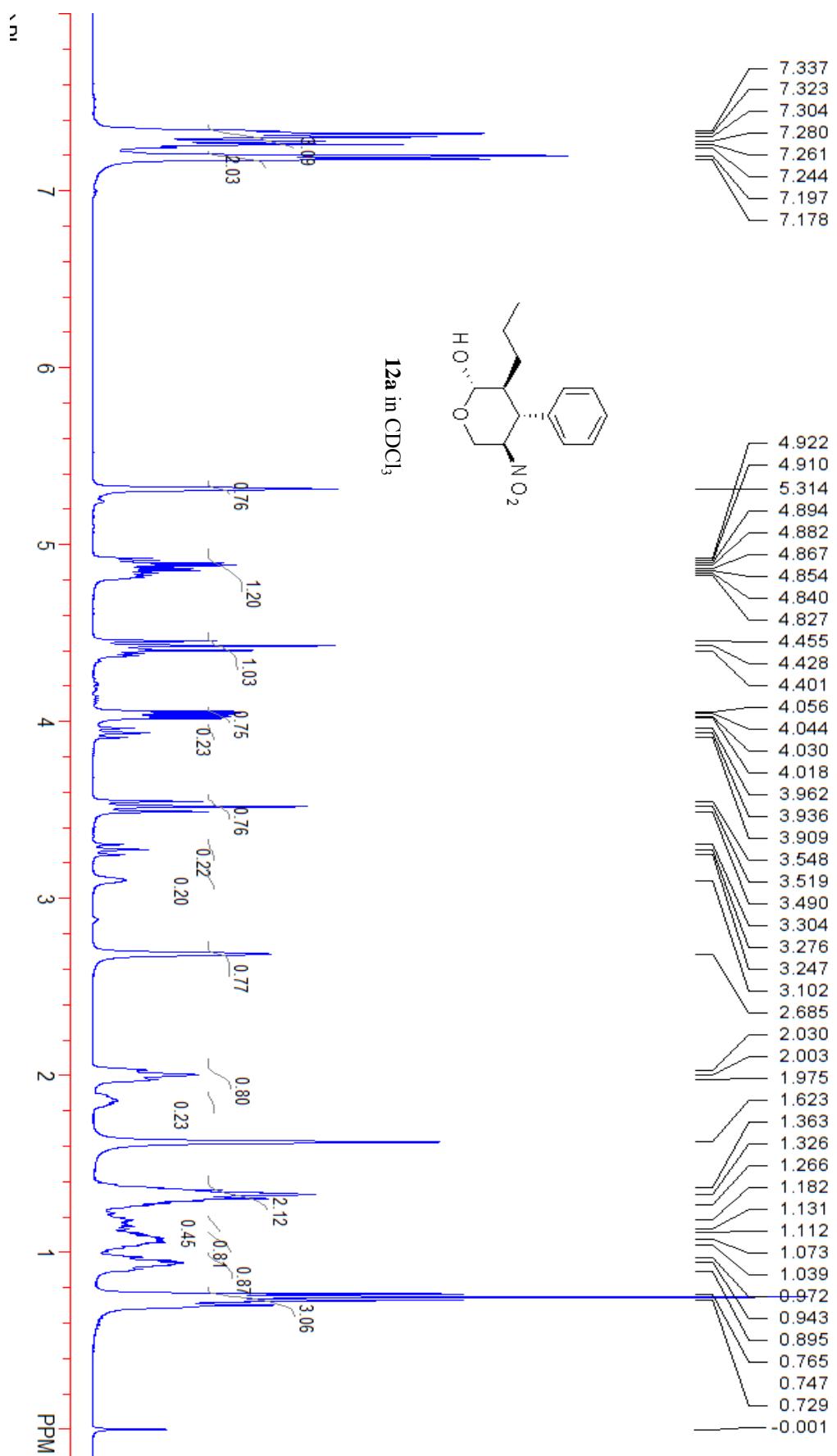


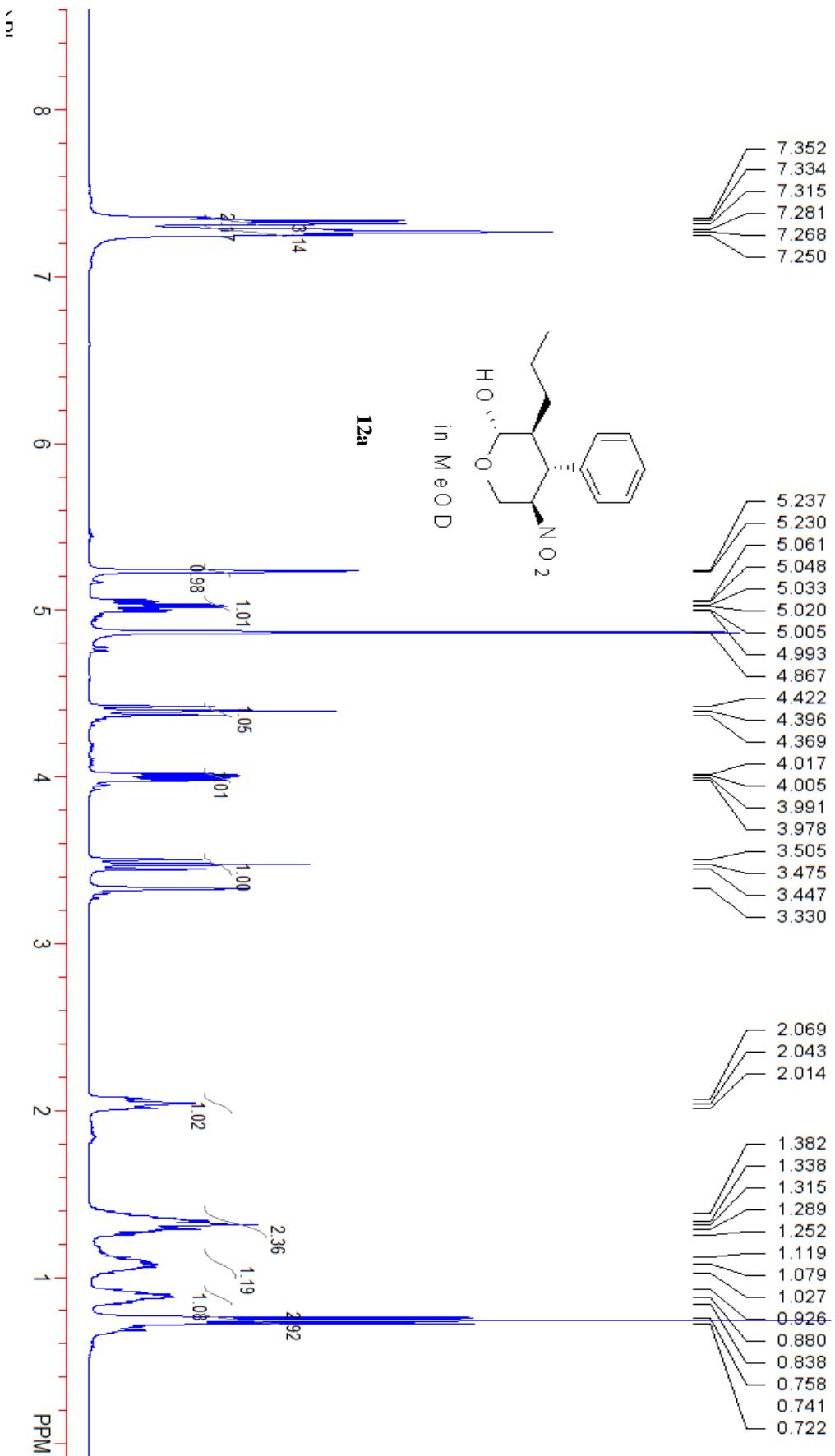


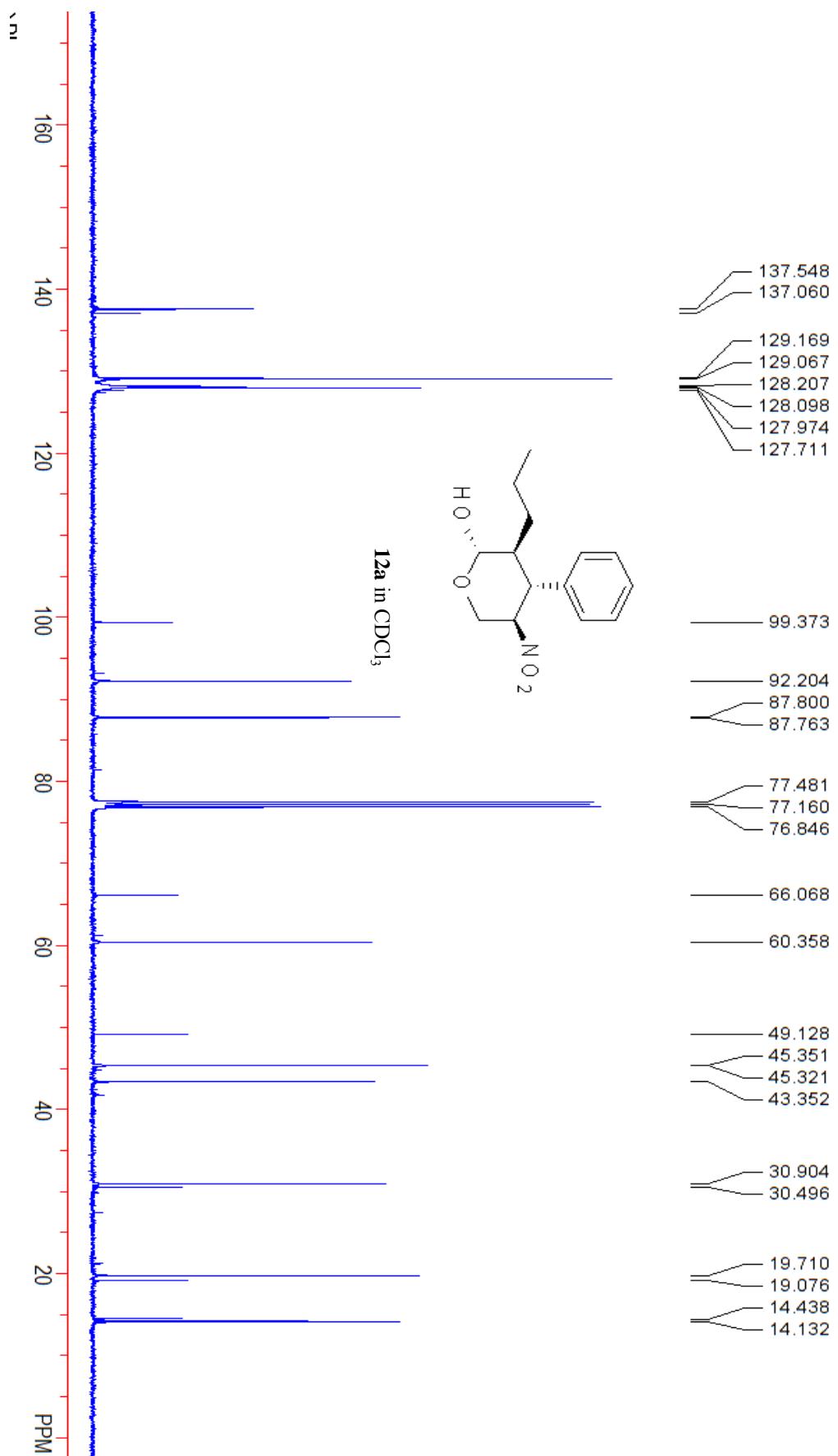


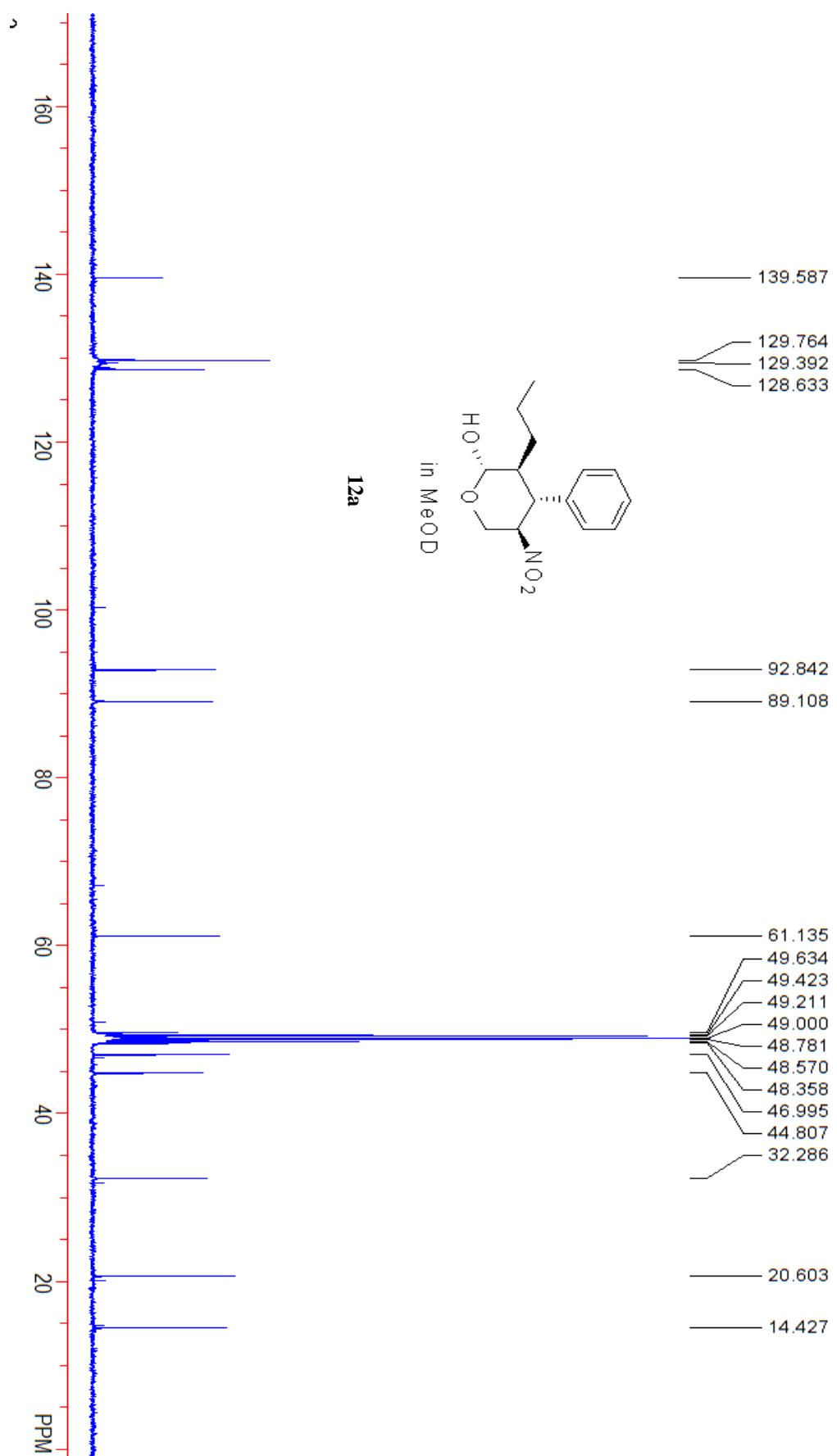












HPLC Report

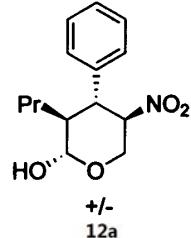
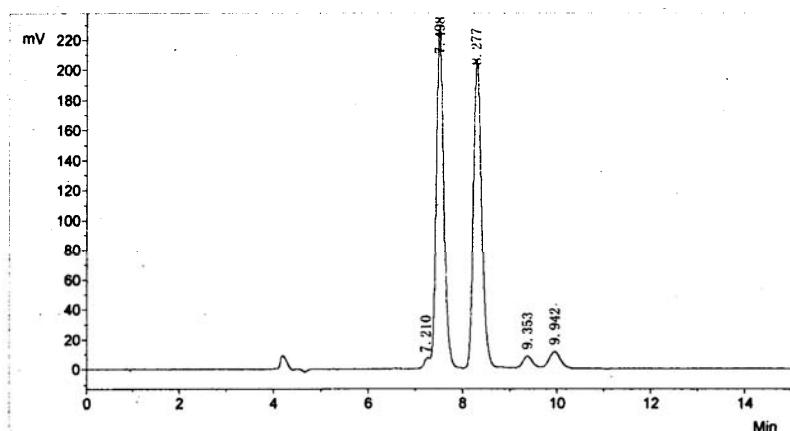
Sample Name:

Data File:ZSL-6-56+- IC 91 214 0.7.che

Operator:

Date:2010-09-09

Time:14:49



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	7.210	89.0	29.2	0.0006
2	2	7.498	217720.6	2319712.6	45.5464
3	3	8.277	198141.8	2522132.3	49.5208
4	4	9.353	7290.2	92011.1	1.8066
5	5	9.942	10316.2	159188.7	3.1256
Total			433557.8	5093073.9	100.0000

HPLC Report

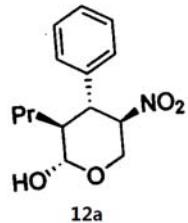
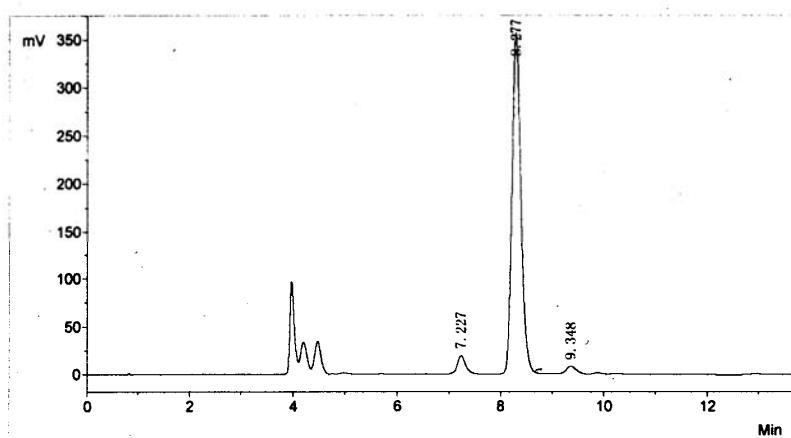
Sample Name:

Data File:ZSL-6-54.che

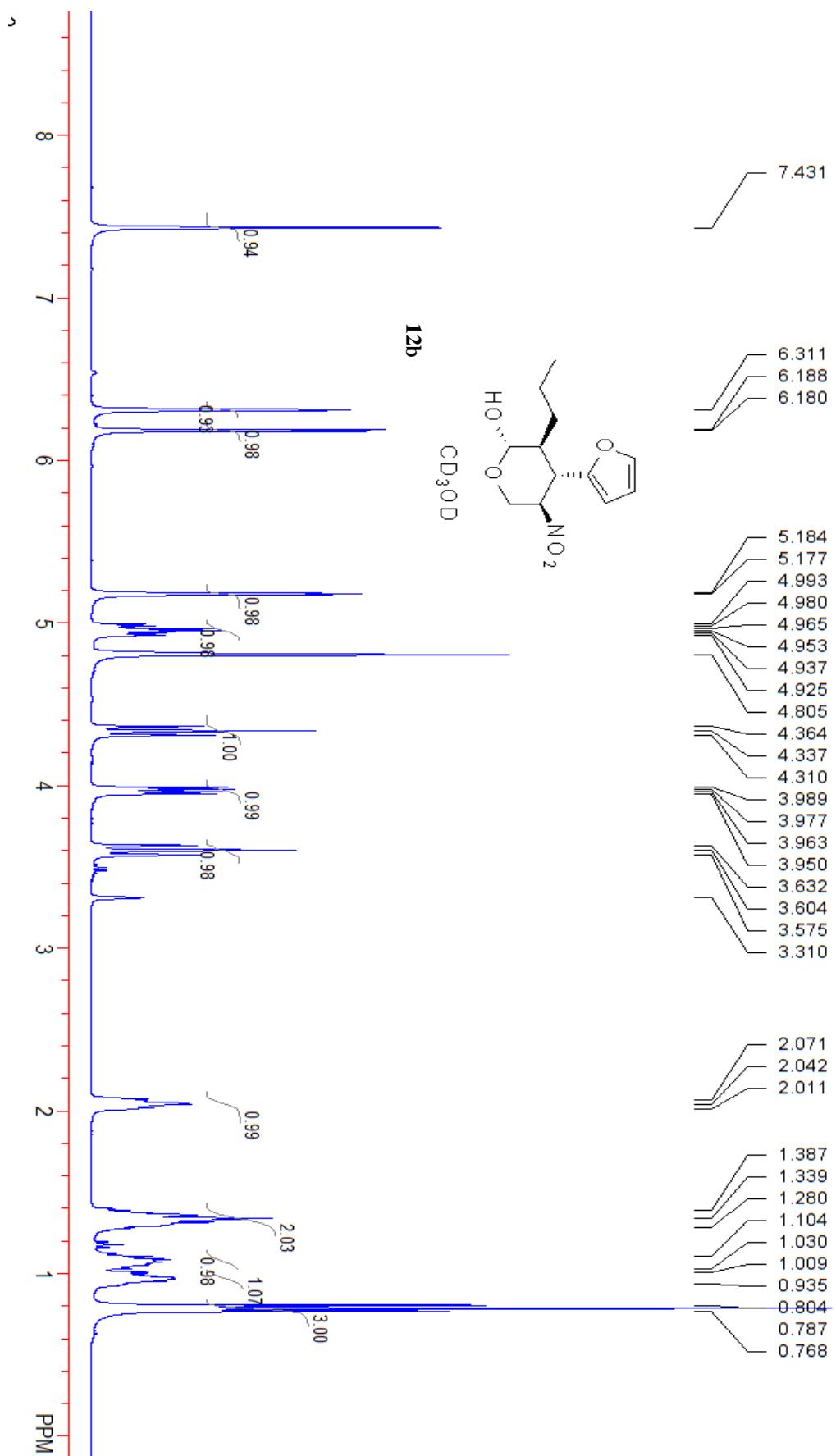
Operator:

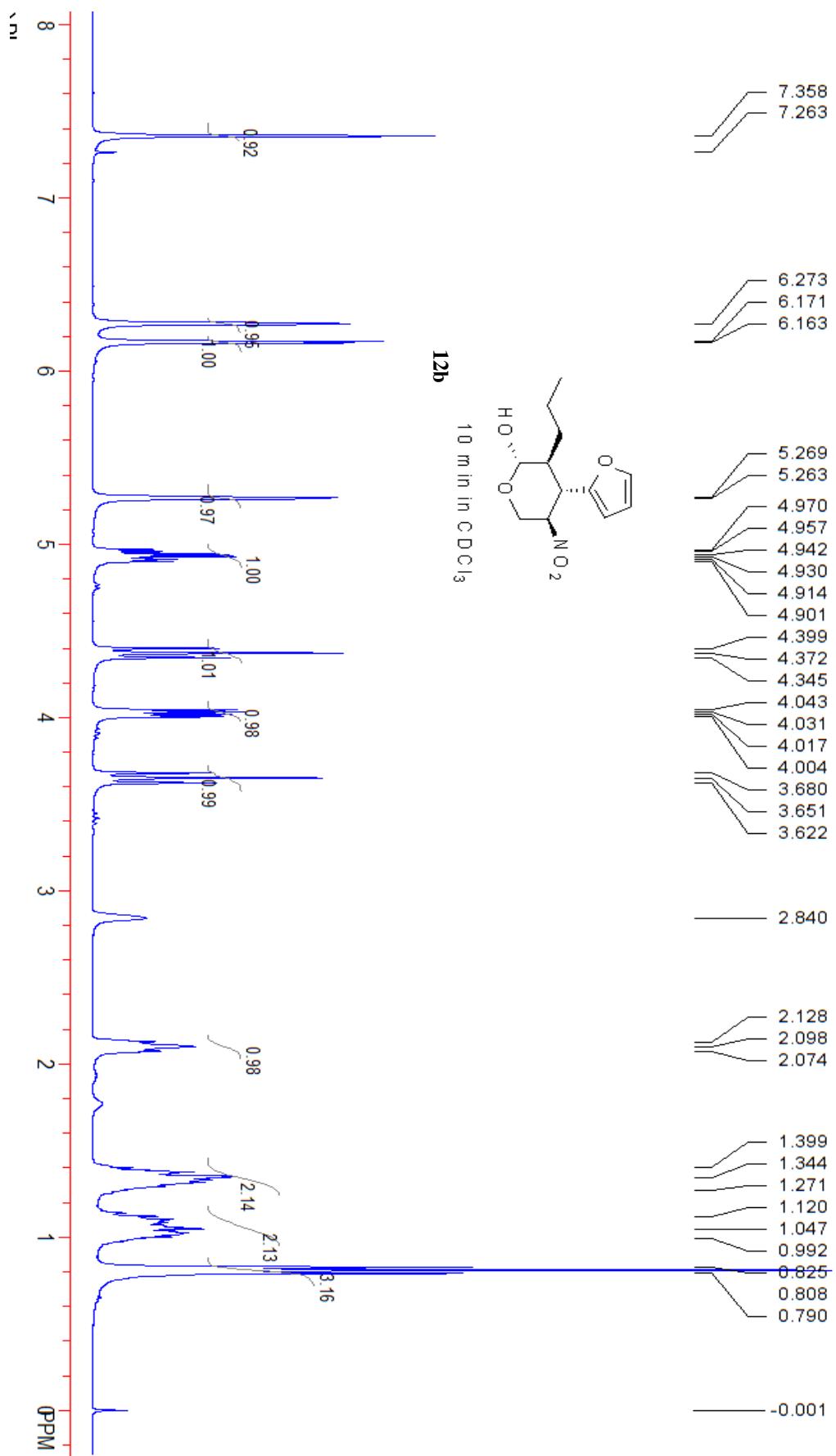
Date:2010-09-09

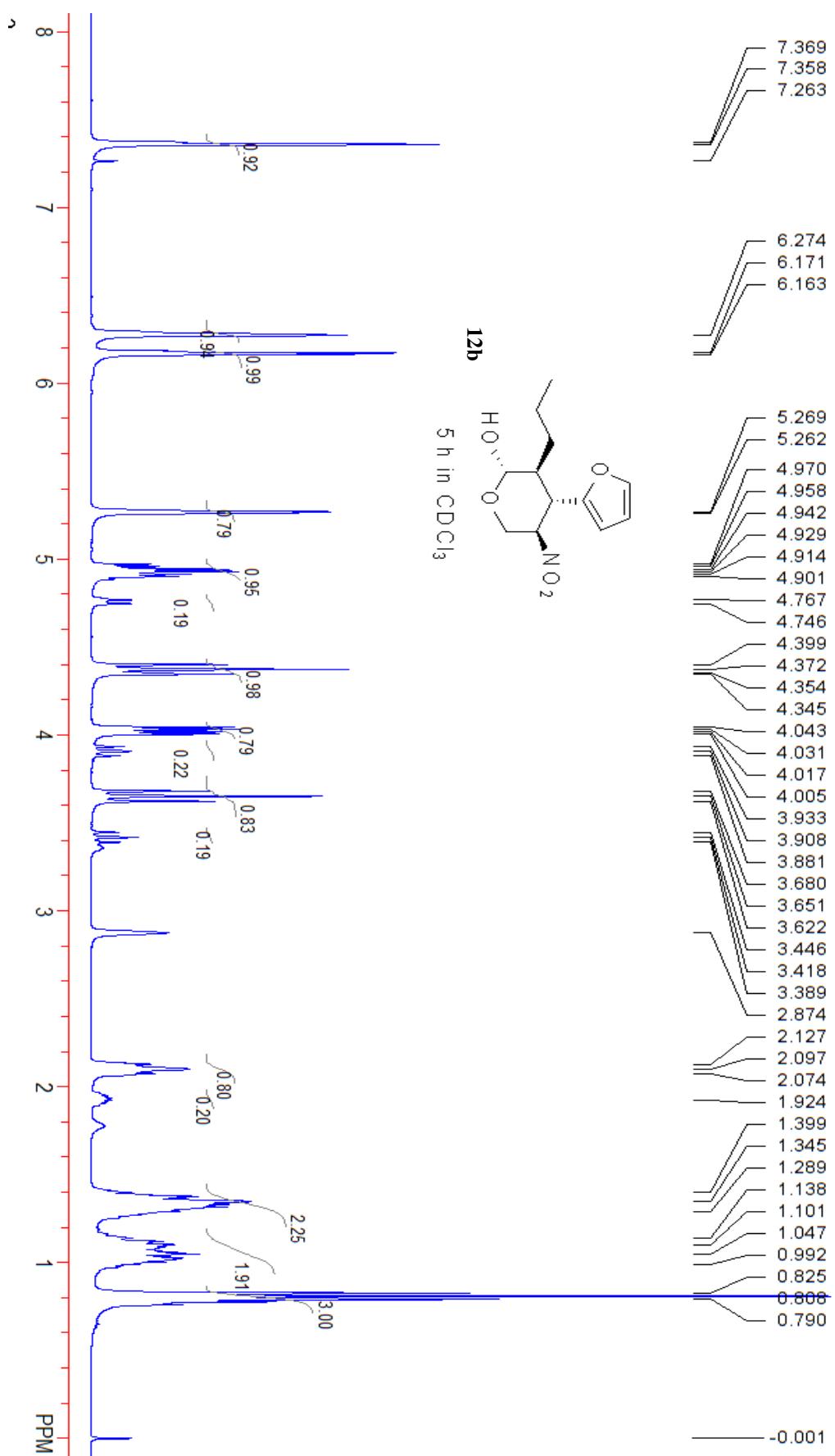
Time:15:11

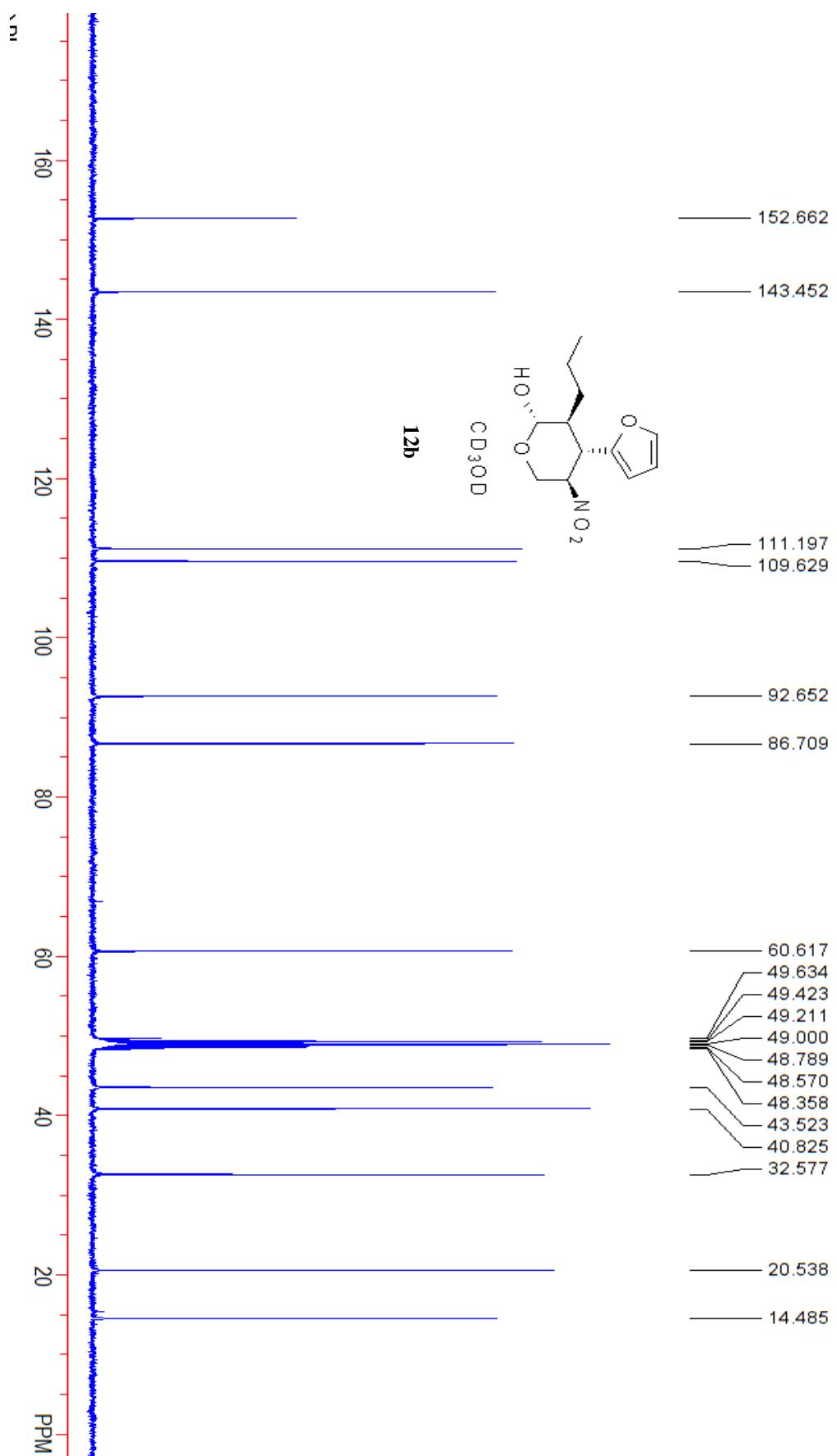


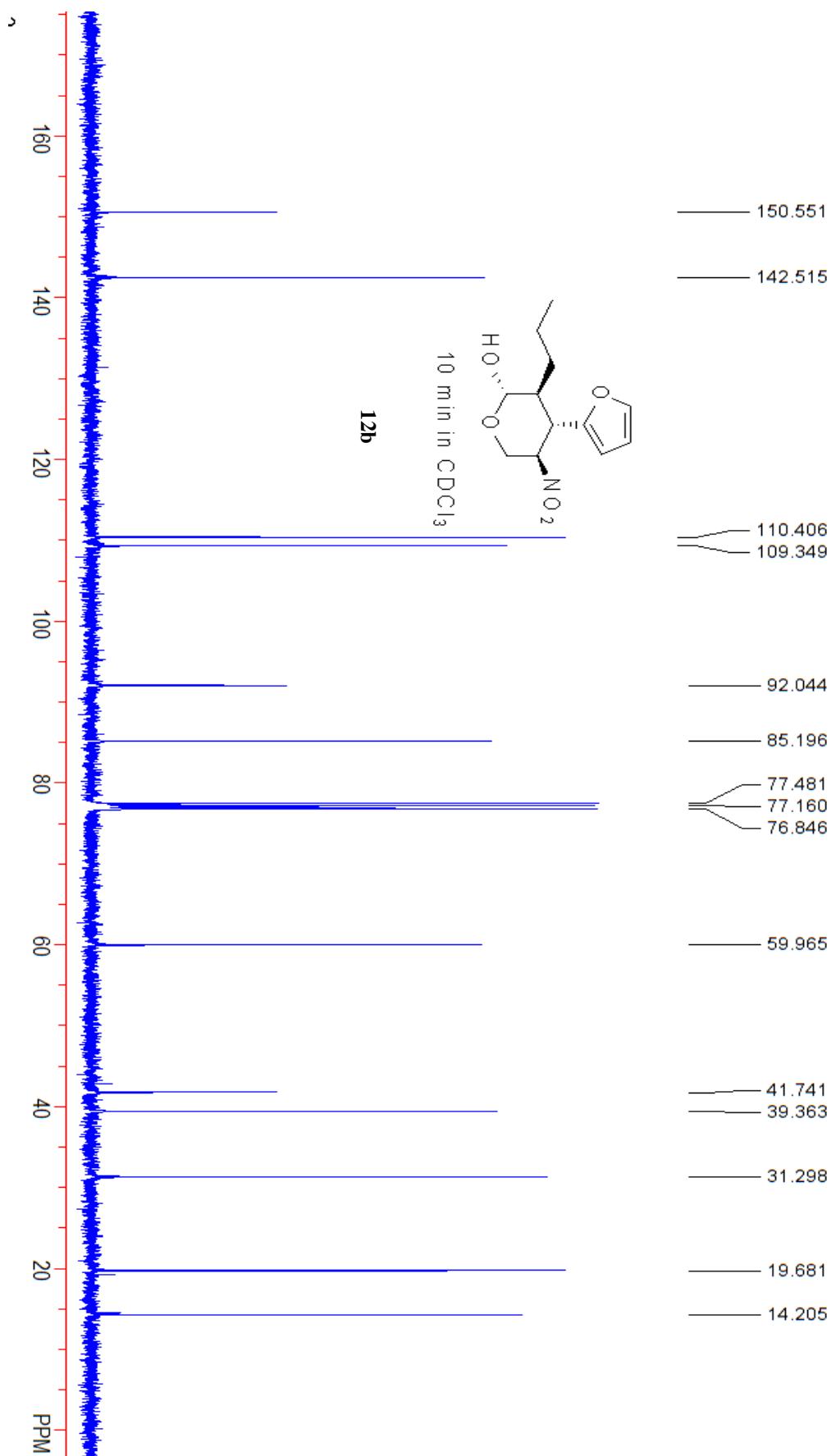
No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	7. 227	19627. 9	222910. 9	4. 8456
2	2	8. 277	353362. 9	4282969. 2	93. 1032
3	3	9. 348	7533. 0	94360. 6	2. 0512
	Total		380523. 8	4600240. 7	100. 0000

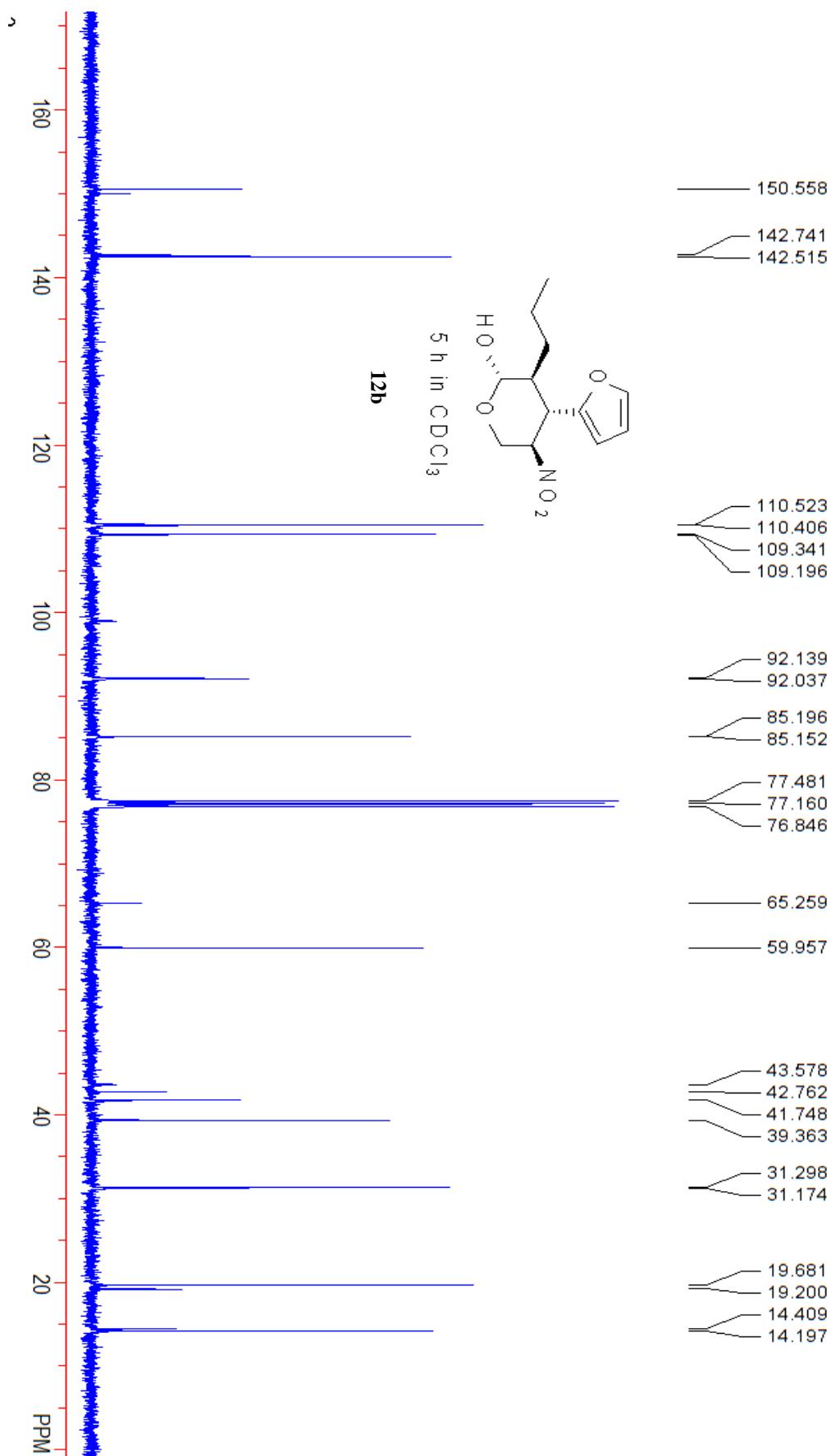






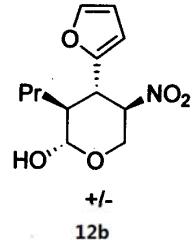
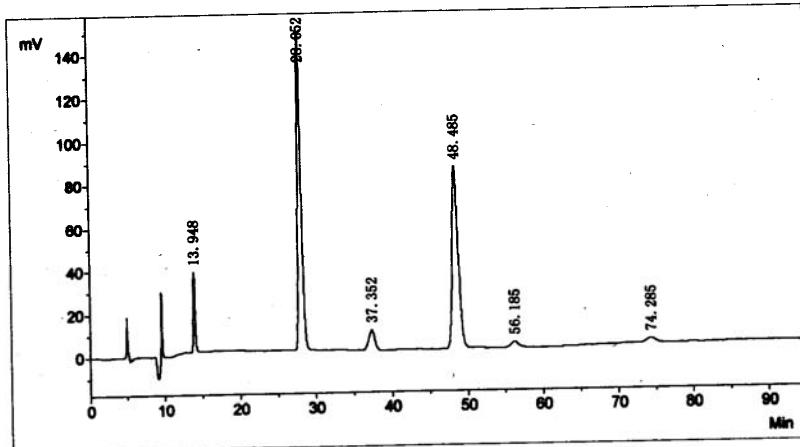






HPLC REPORT

Data File: ZSL-590+- AD-H 95EtOH5 214 0.7.che Sample Name:
 Date: 2009-09-27 Time: 14:54
 Operator:



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		13.948	37192.9	627017.1	5.6261
2	2		28.052	147968.1	4884582.5	43.8285
3	3		37.352	9507.6	515955.4	4.6296
4	4		48.485	84533.0	4735838.7	42.4939
5	5		56.185	2813.2	184939.2	1.6594
6	6		74.285	2300.2	196428.4	1.7625
Total				284315.0	11144761.3	100.0000

HPLC REPORT

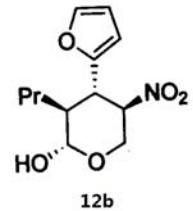
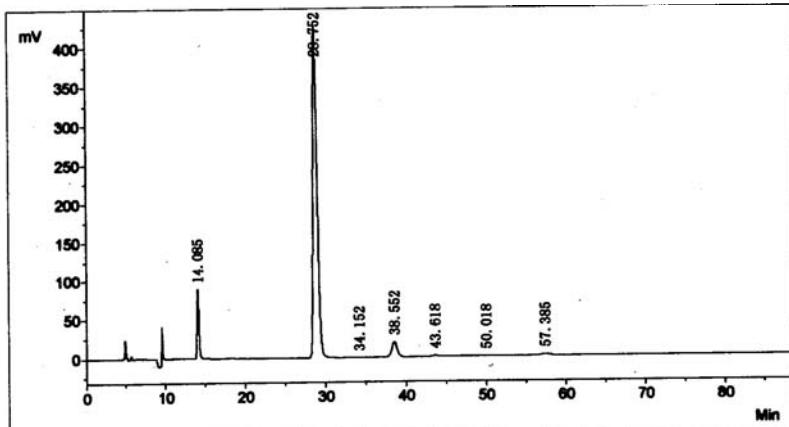
Data File:ZSL-13-85-2.che

Date:2009-09-27

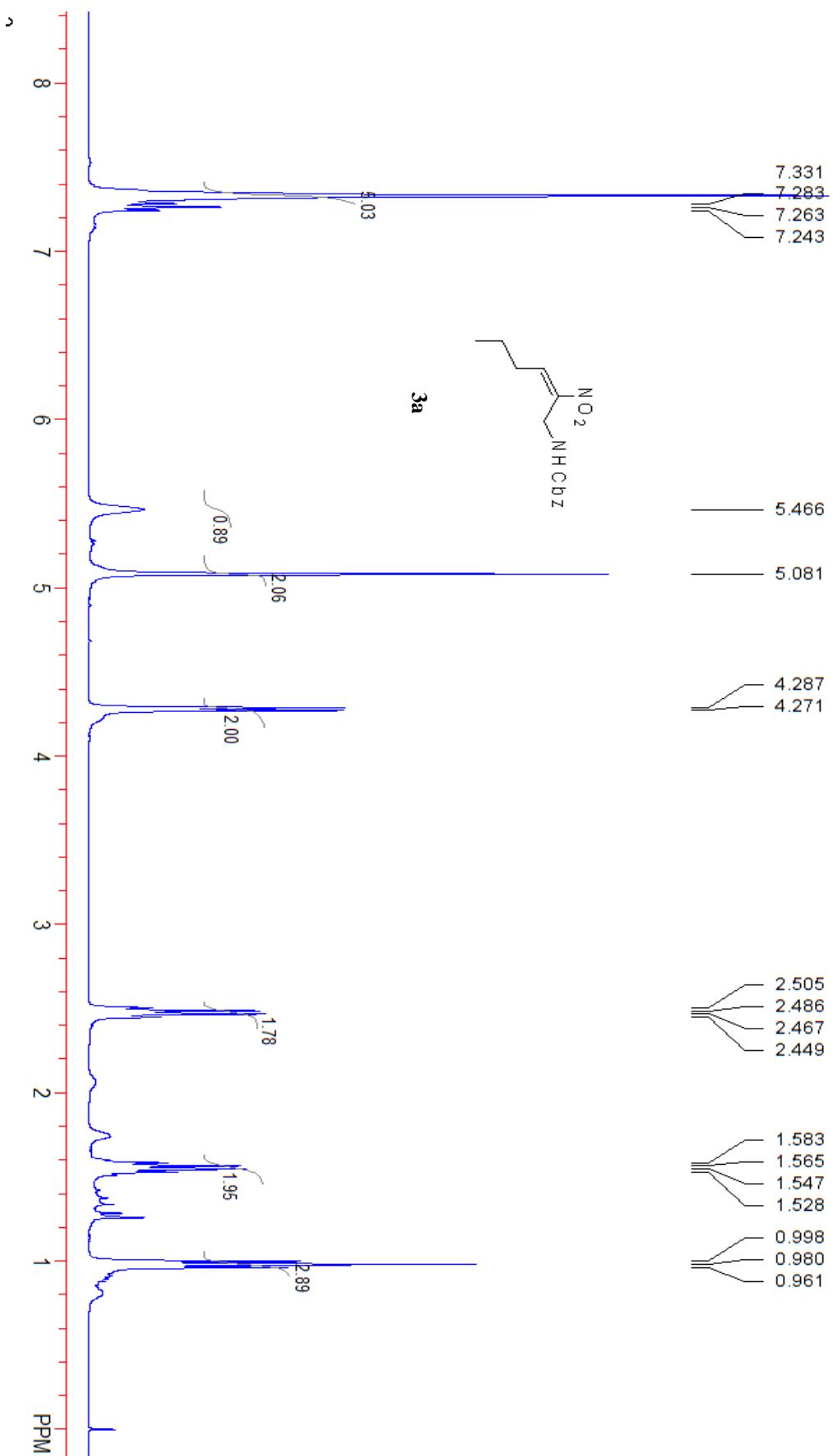
Operator:

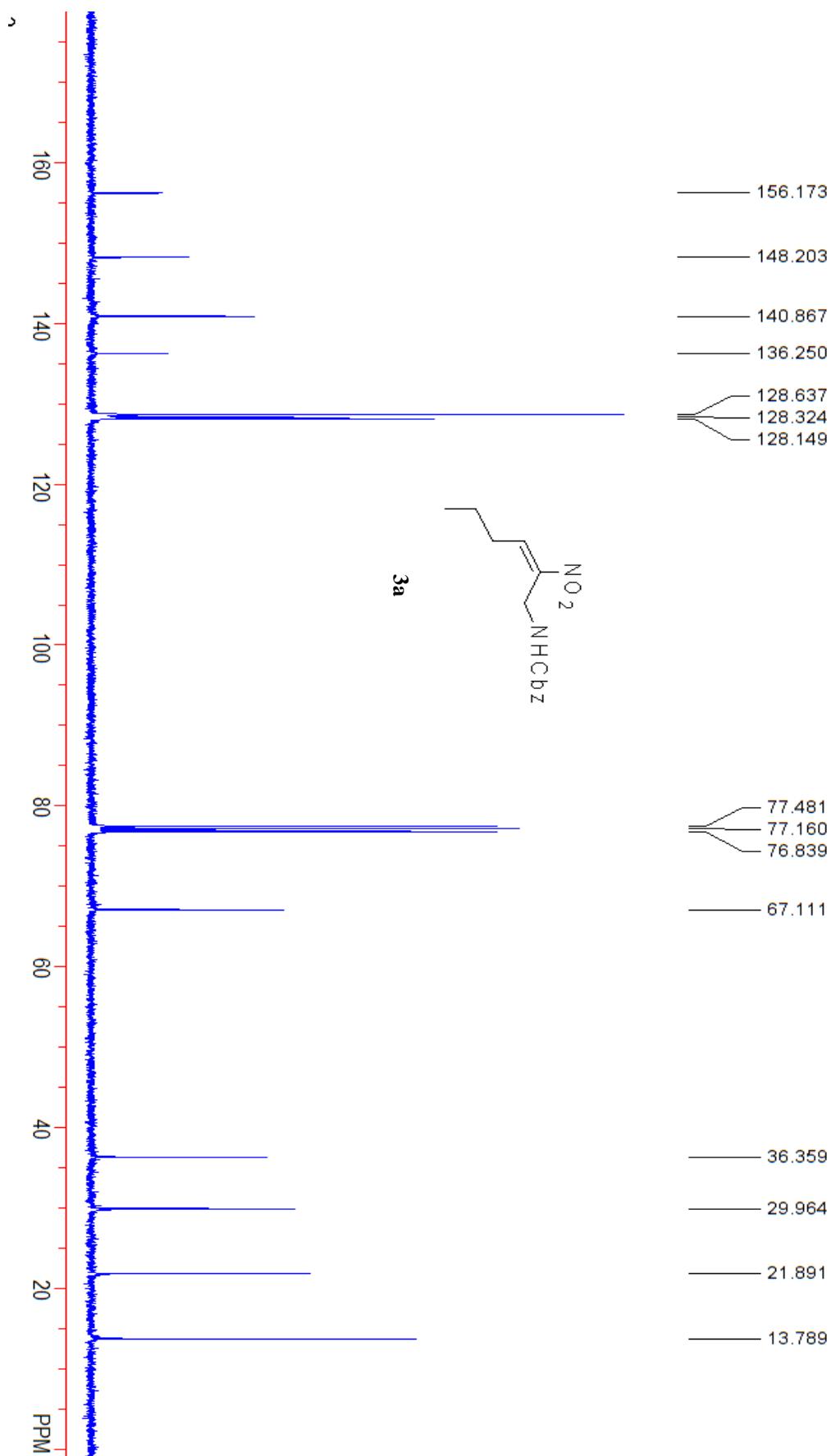
Sample Name:

Time:16:39



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		14.085	89367.0	1549891.6	8.5802
2	2		28.752	426646.6	15416639.9	85.3469
3	3		34.152	164.6	5826.5	0.0323
4	4		38.552	19655.5	905280.1	5.0117
5	5		43.618	1106.2	53651.2	0.2970
6	6		50.018	260.5	14094.0	0.0780
7	7		57.385	1781.9	118127.1	0.6540
Total				538982.3	18063510.3	100.0000





Std proton

File: x9

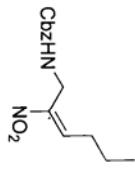
Pulse Sequence: NOESY

Solvent: cdcl3

Temp. 25.0 C / 298.1 K

Operator: vnmr1

VMRS-400 "400MR"



Relax. delay 2.000 sec

Mixing 1.000 sec

Acq. time 0.242 sec

Width 3811.0 Hz

2D Width 3811.0 Hz

4 repetition

2 x 128 increments

Observe H1, 399.622490 MHz

Data Processing

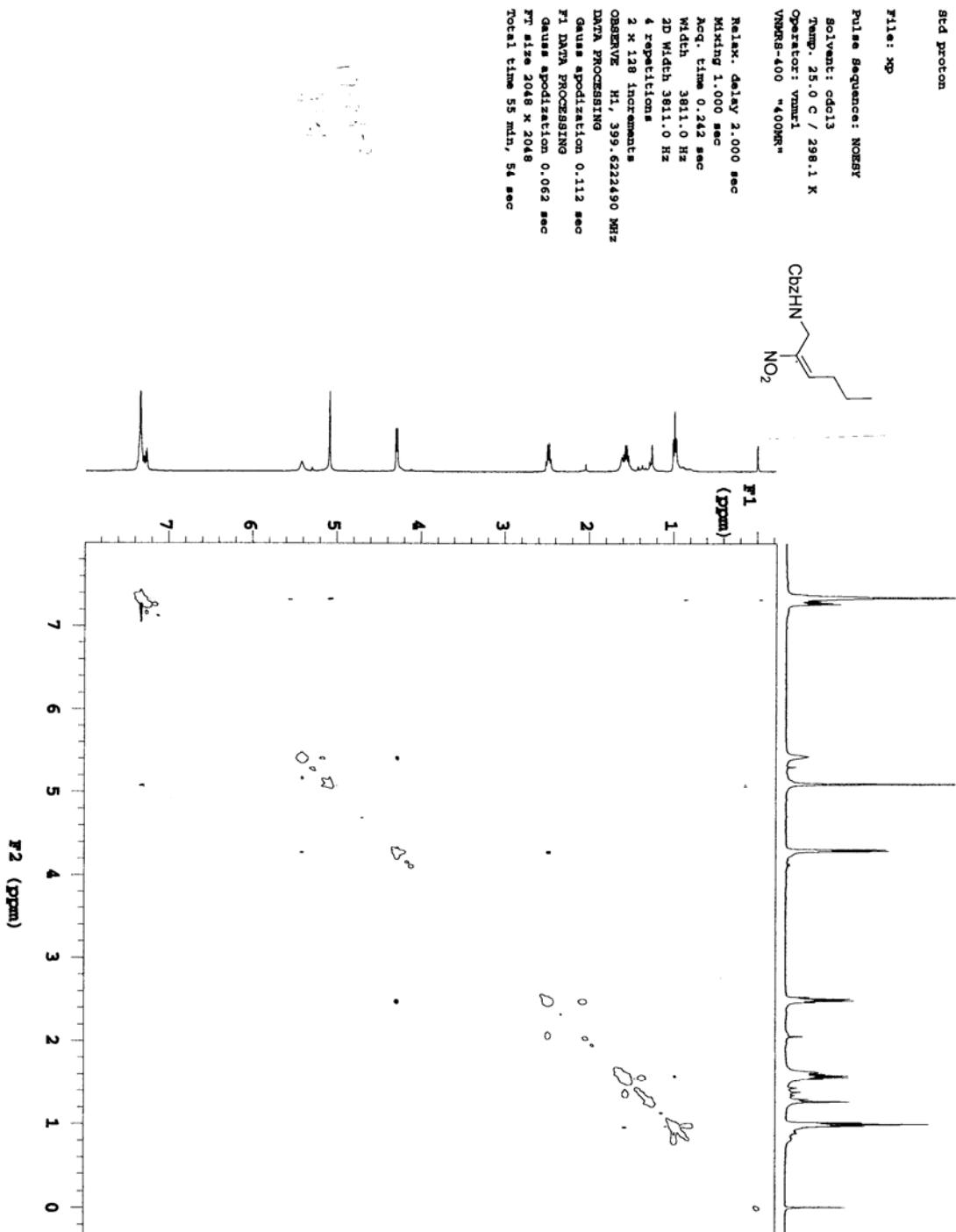
gauss apodization 0.112 sec

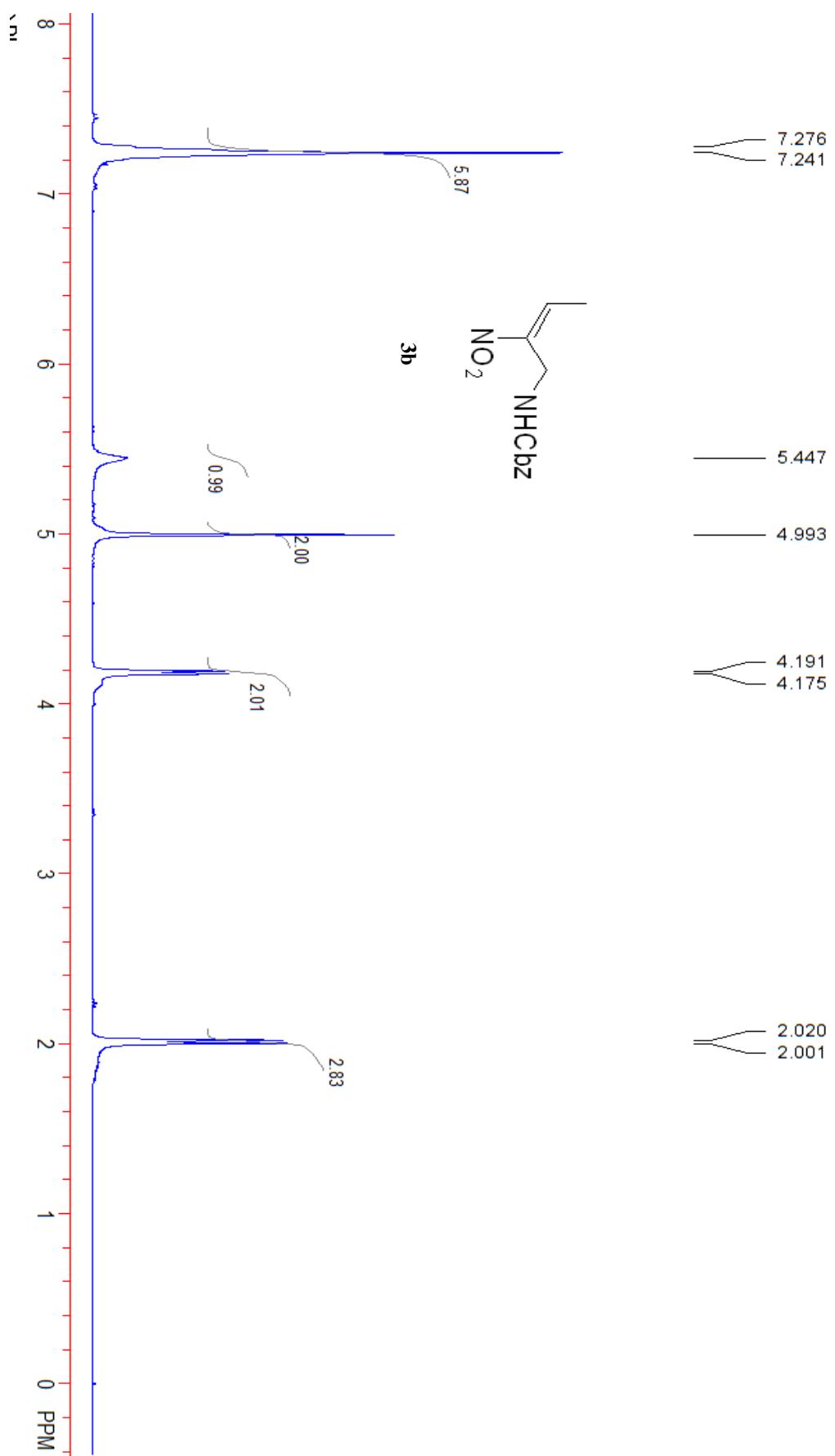
Fit DATA PROCESSING

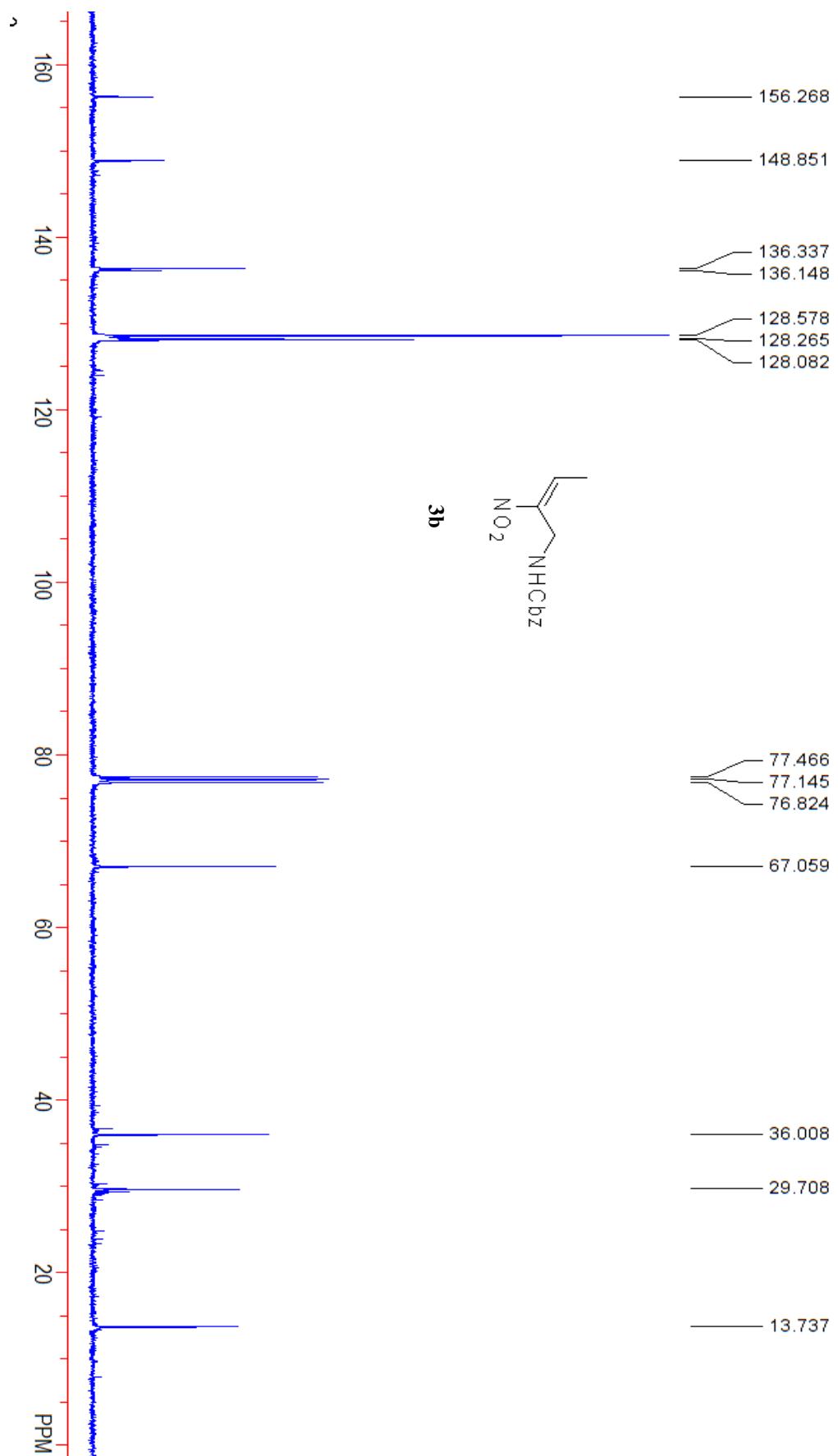
gauss apodization 0.062 sec

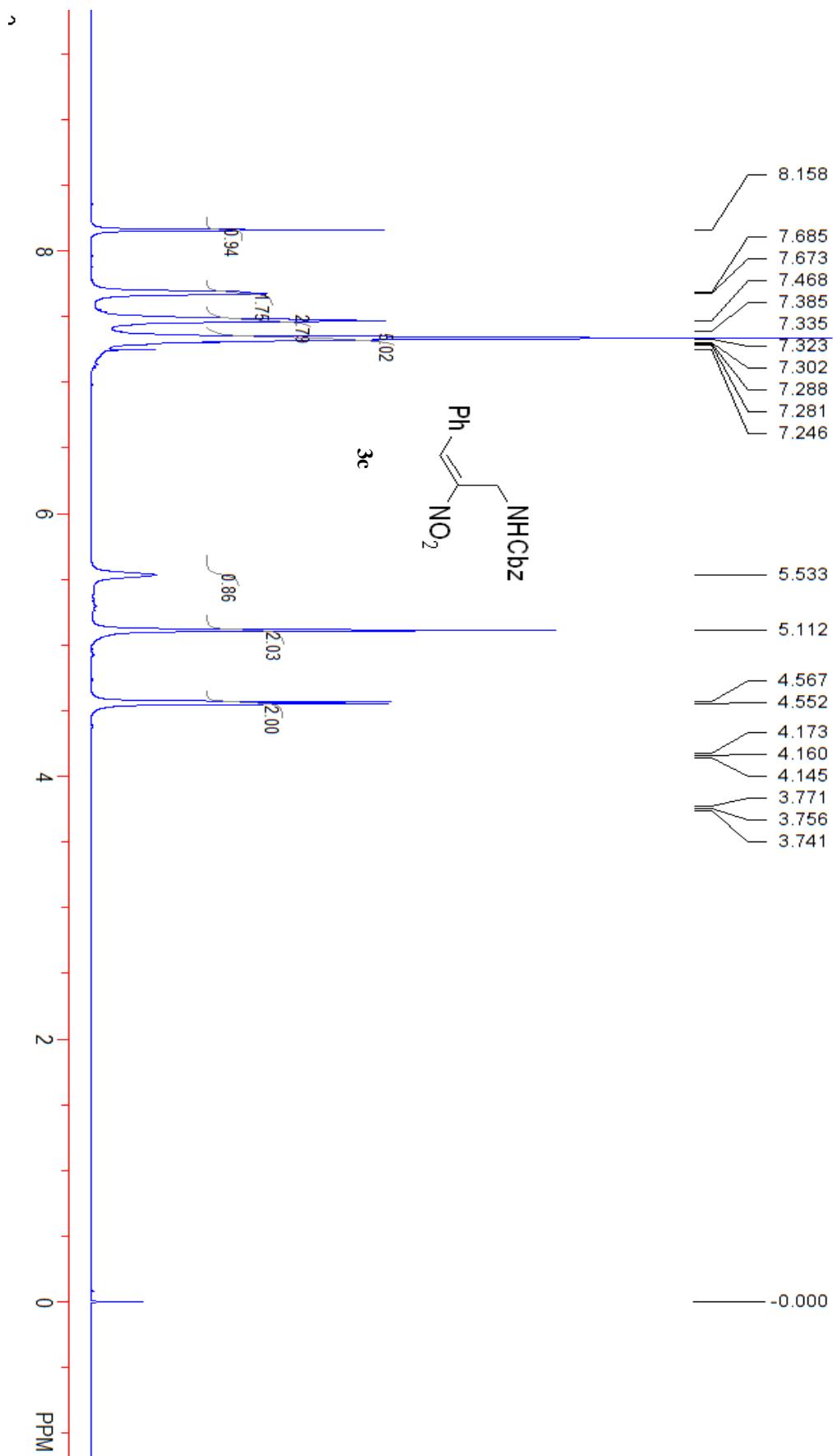
FT size 2048 x 2048

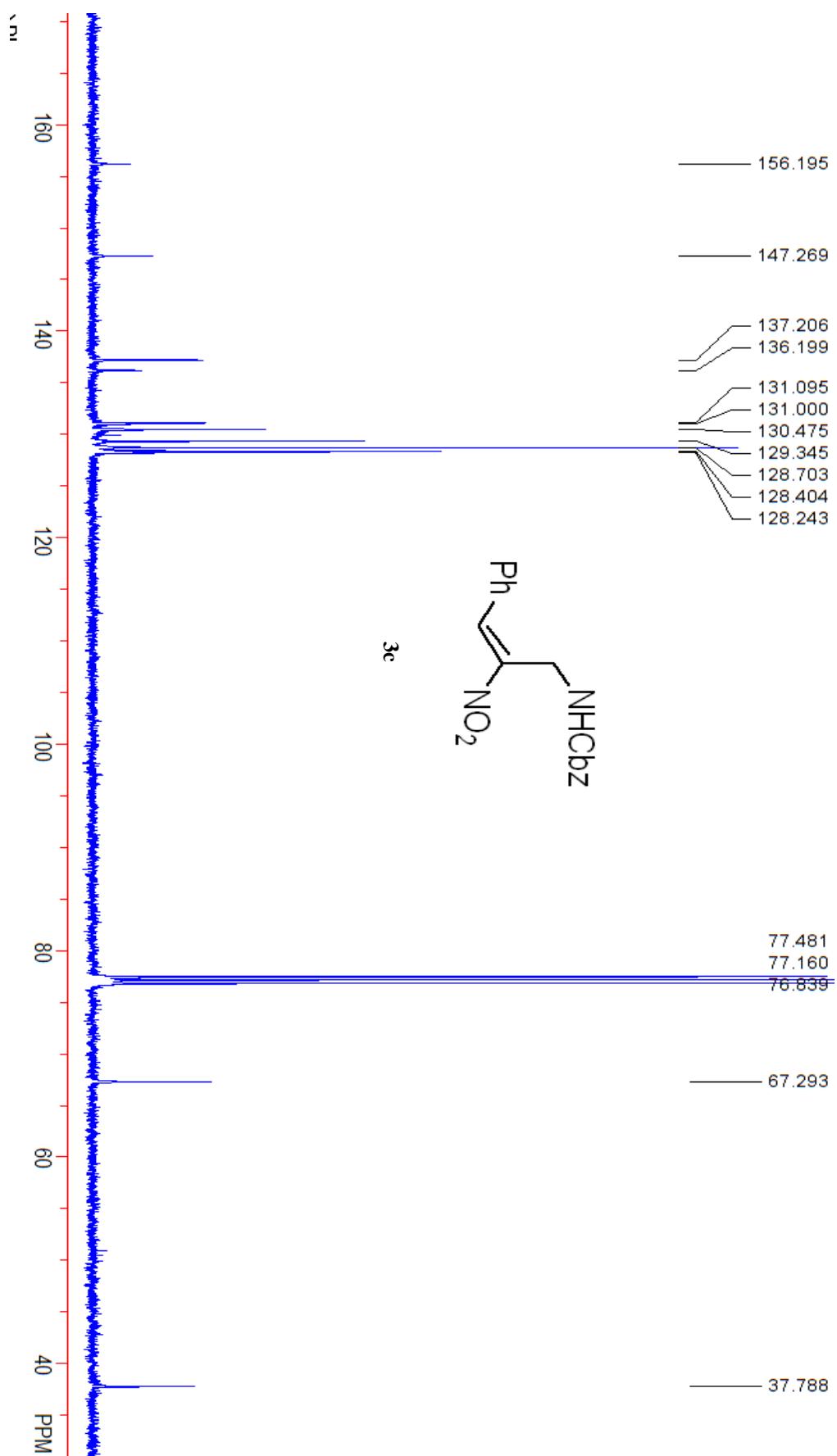
Total time 55 min, 54 sec

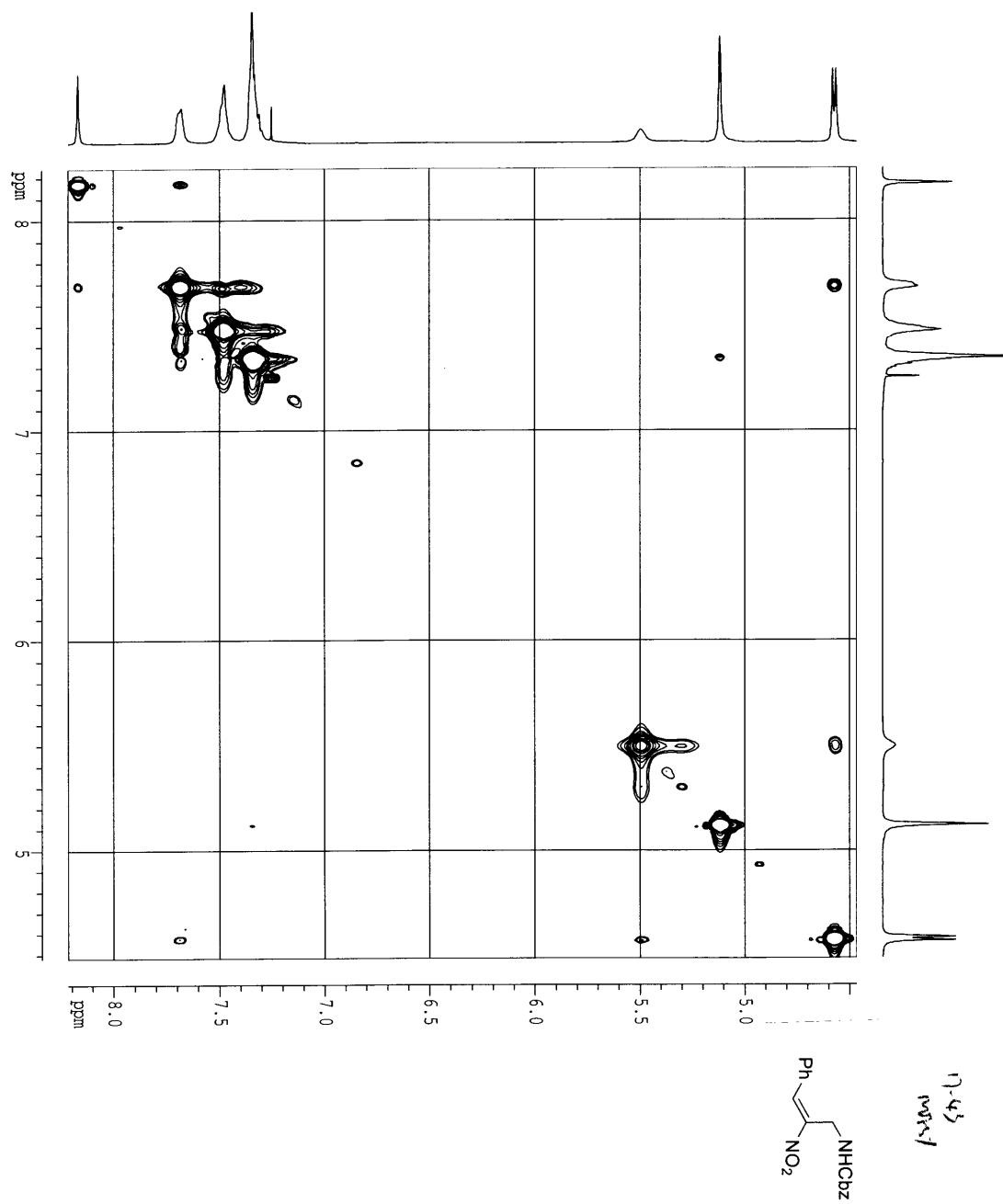


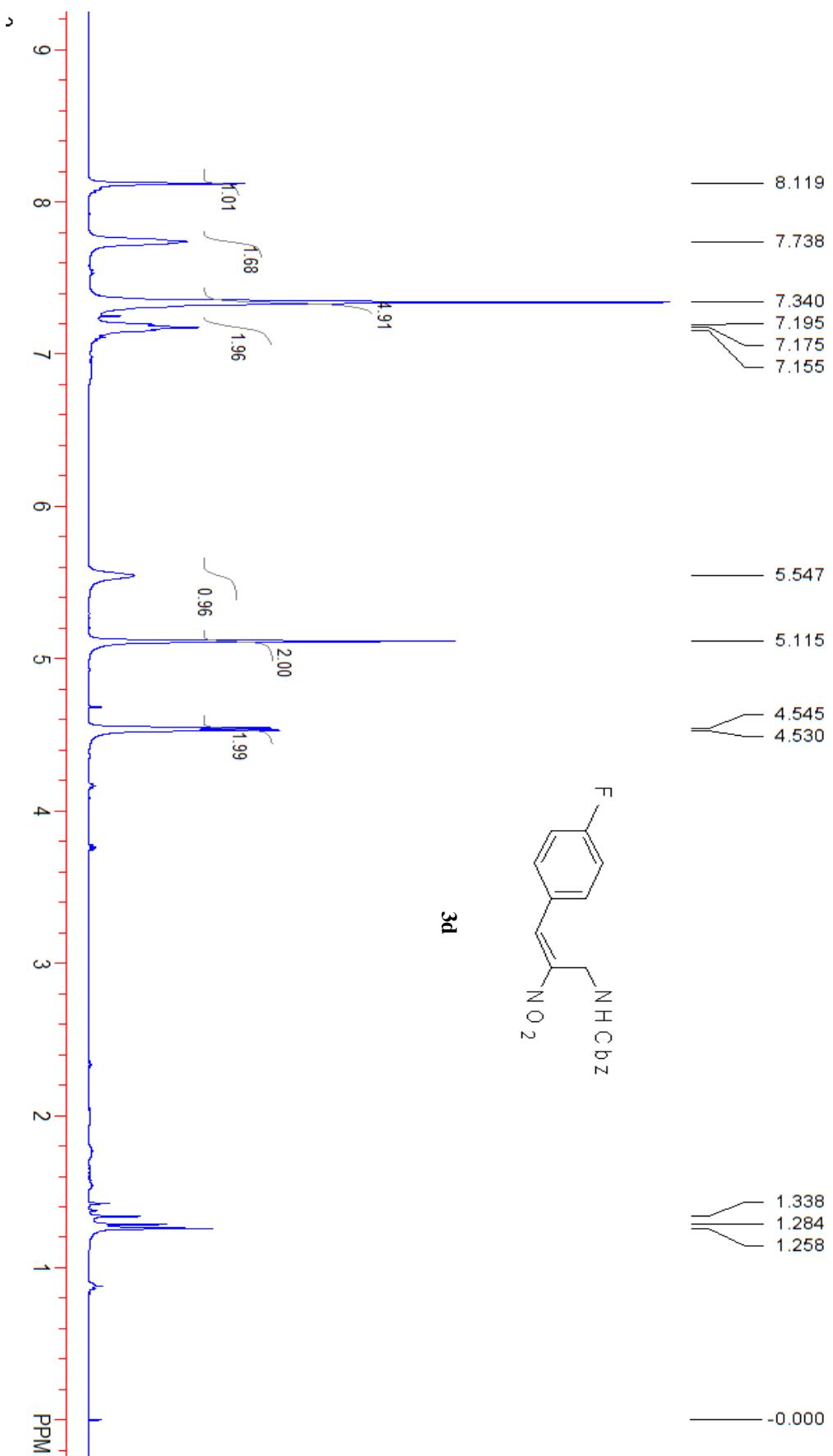


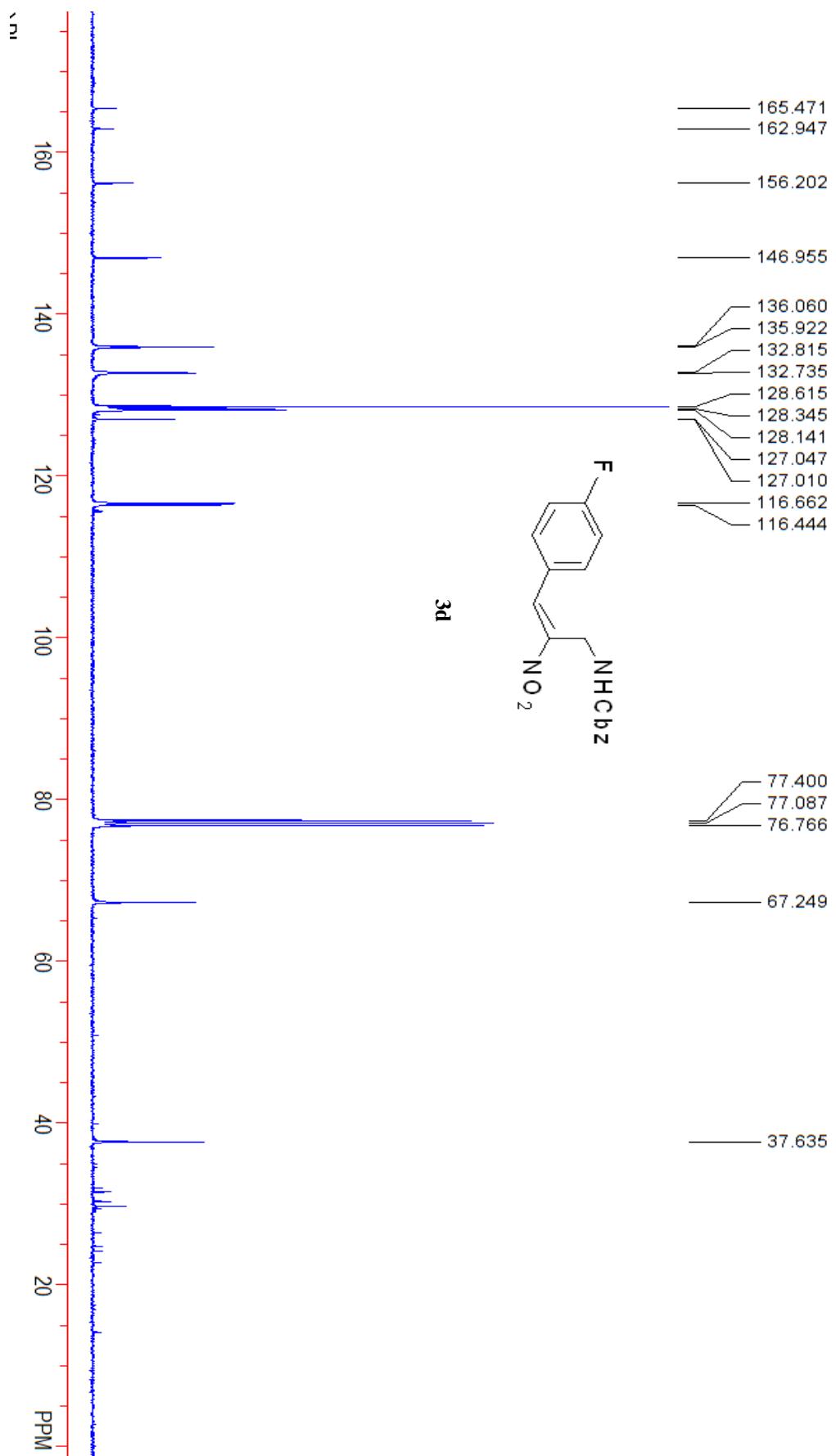


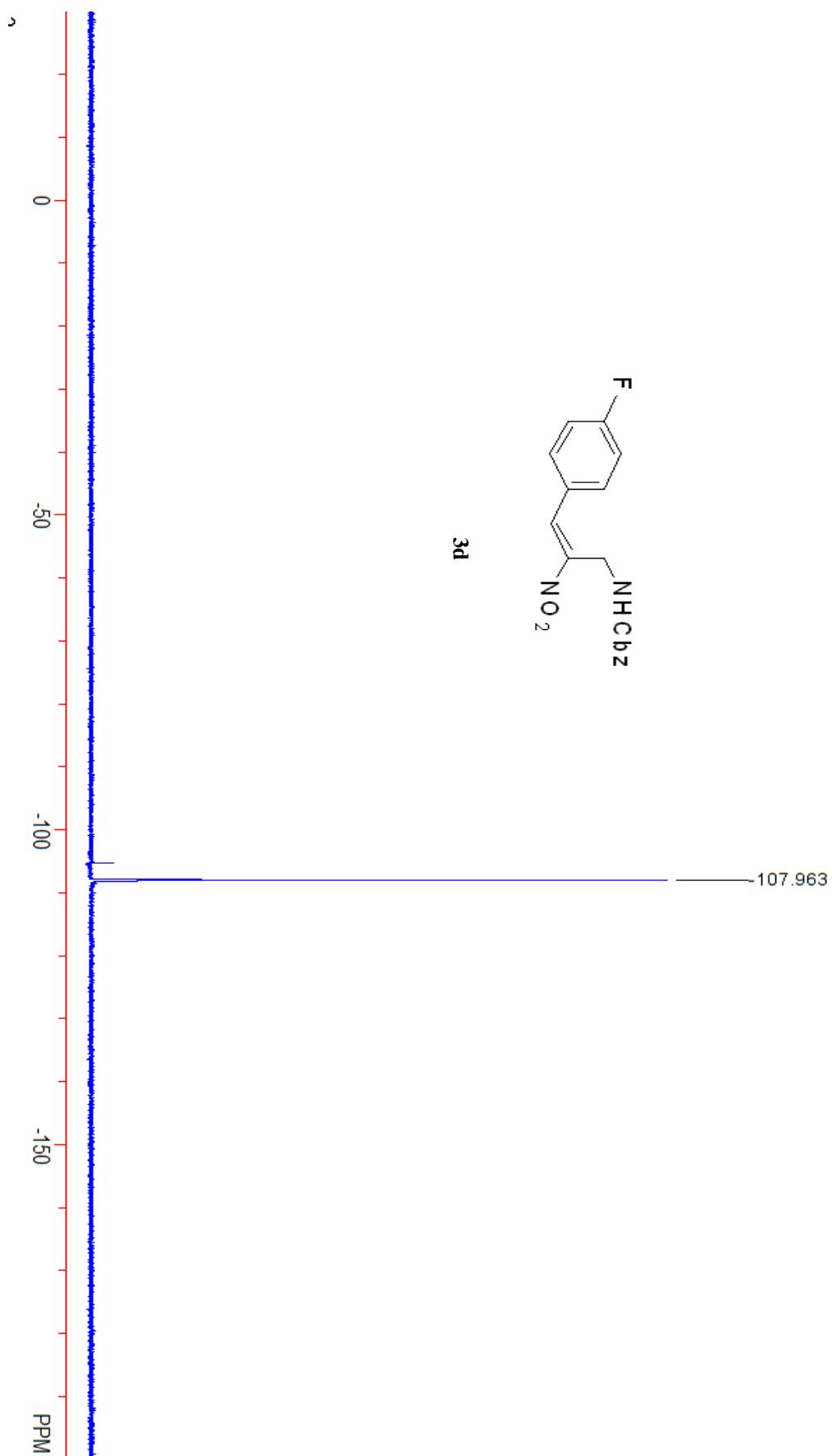


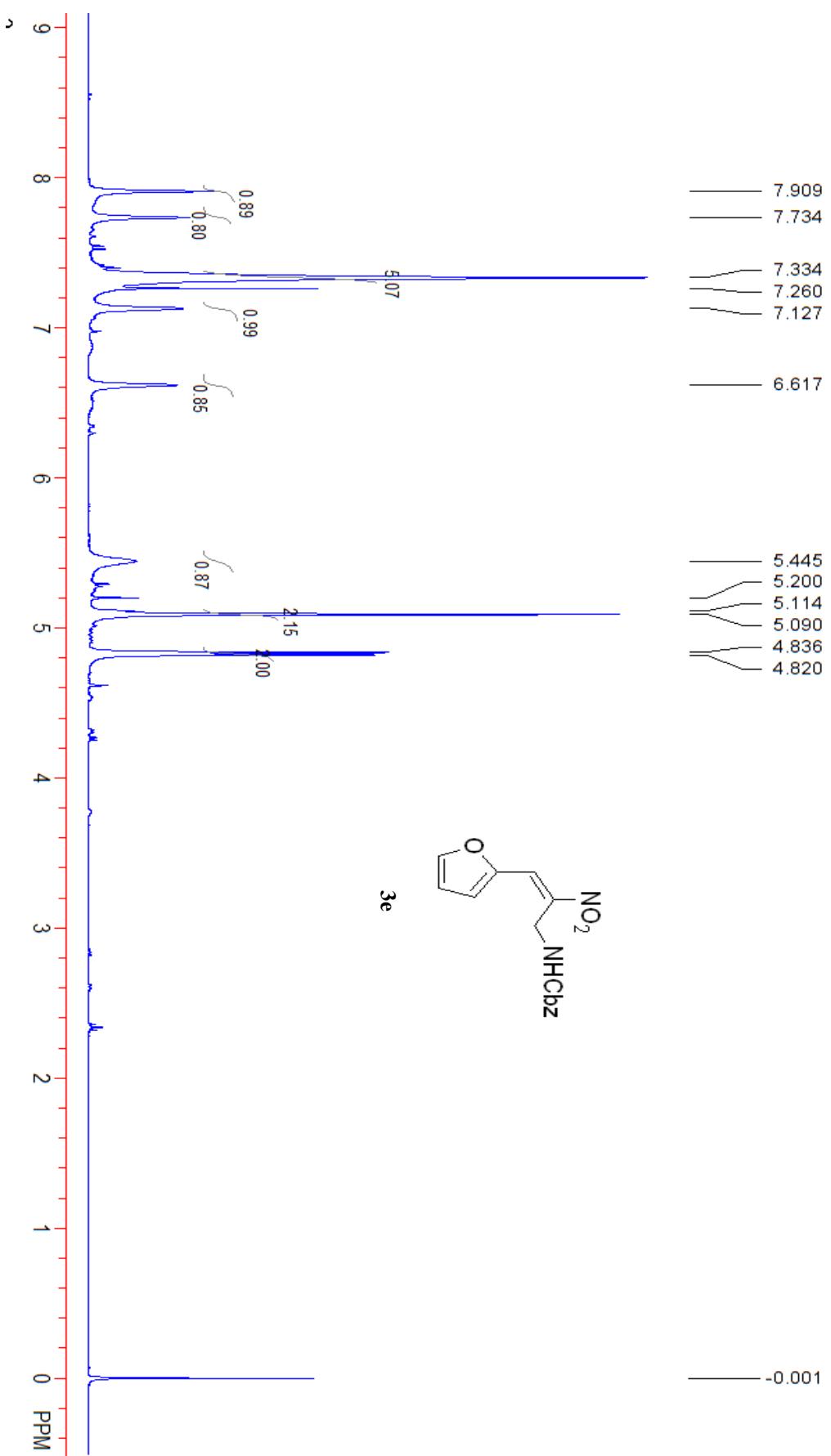


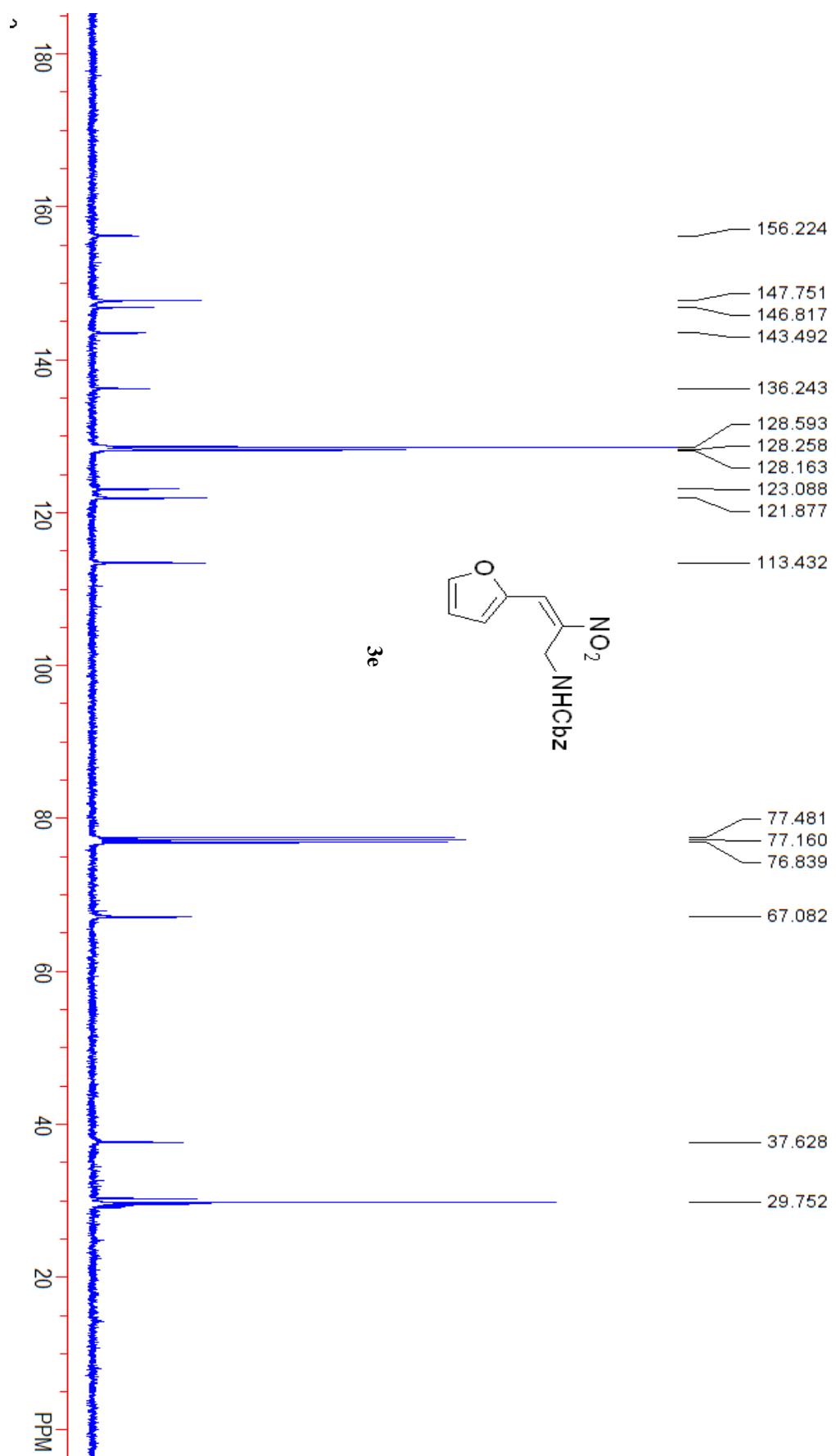


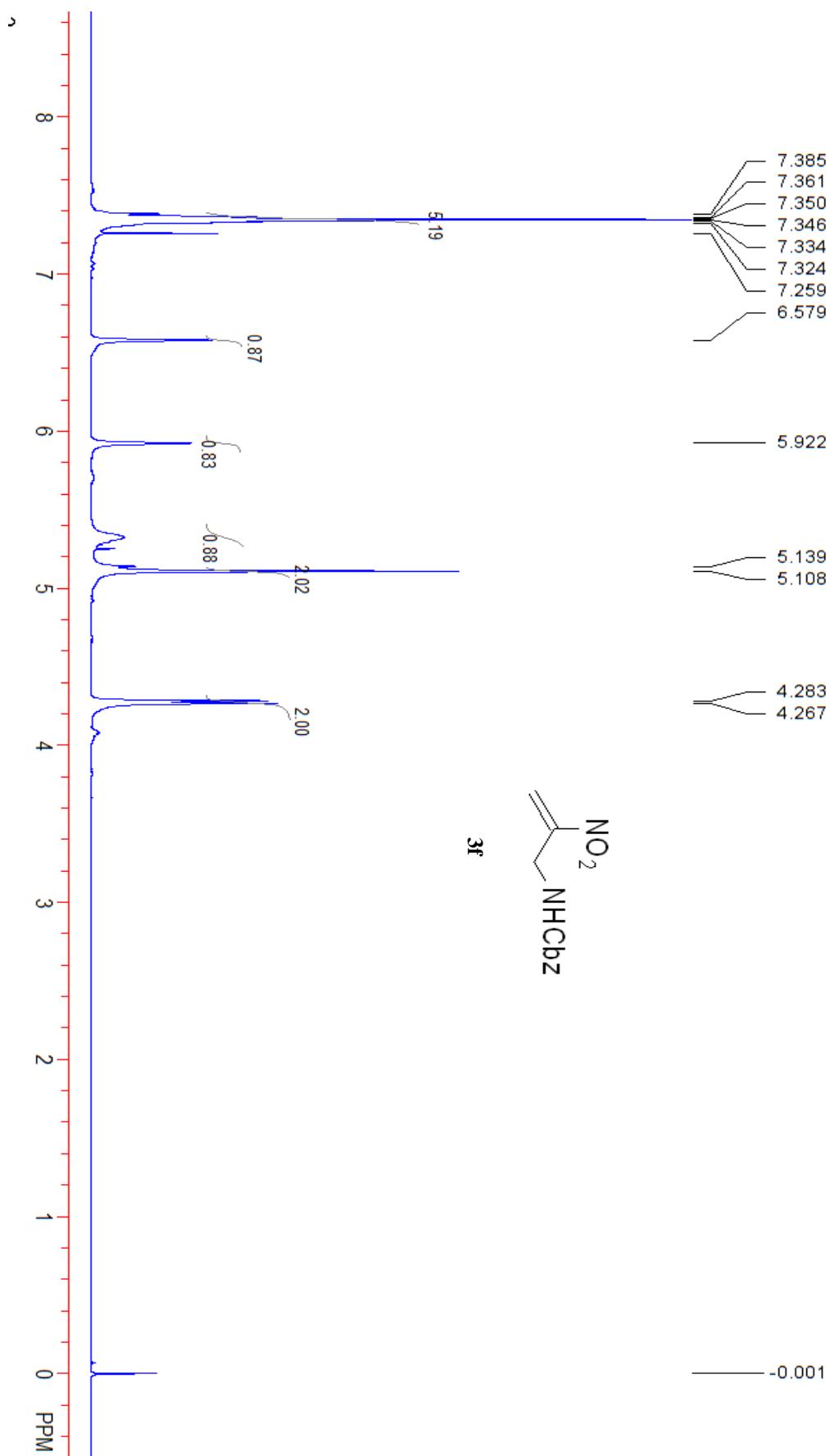


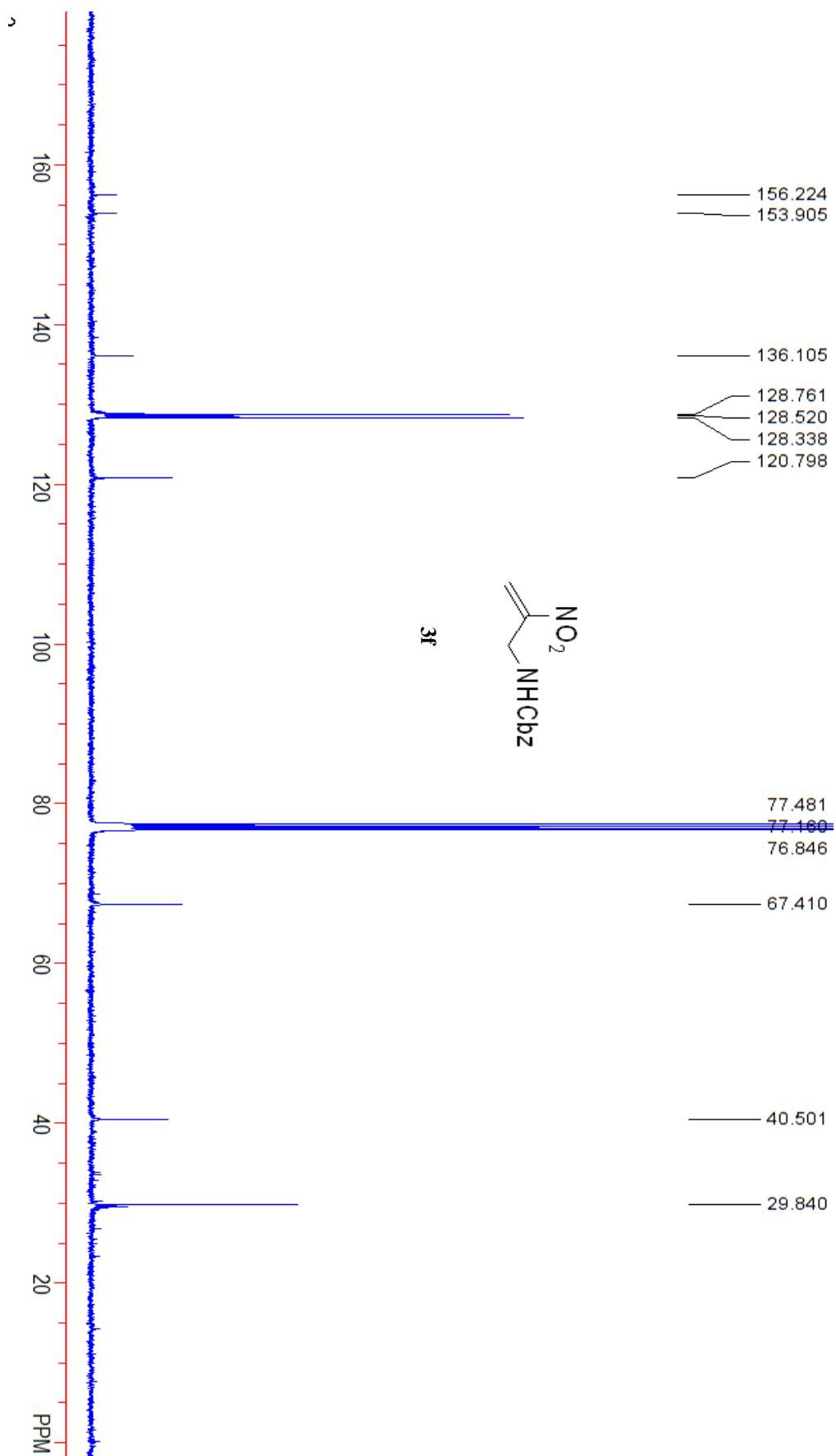


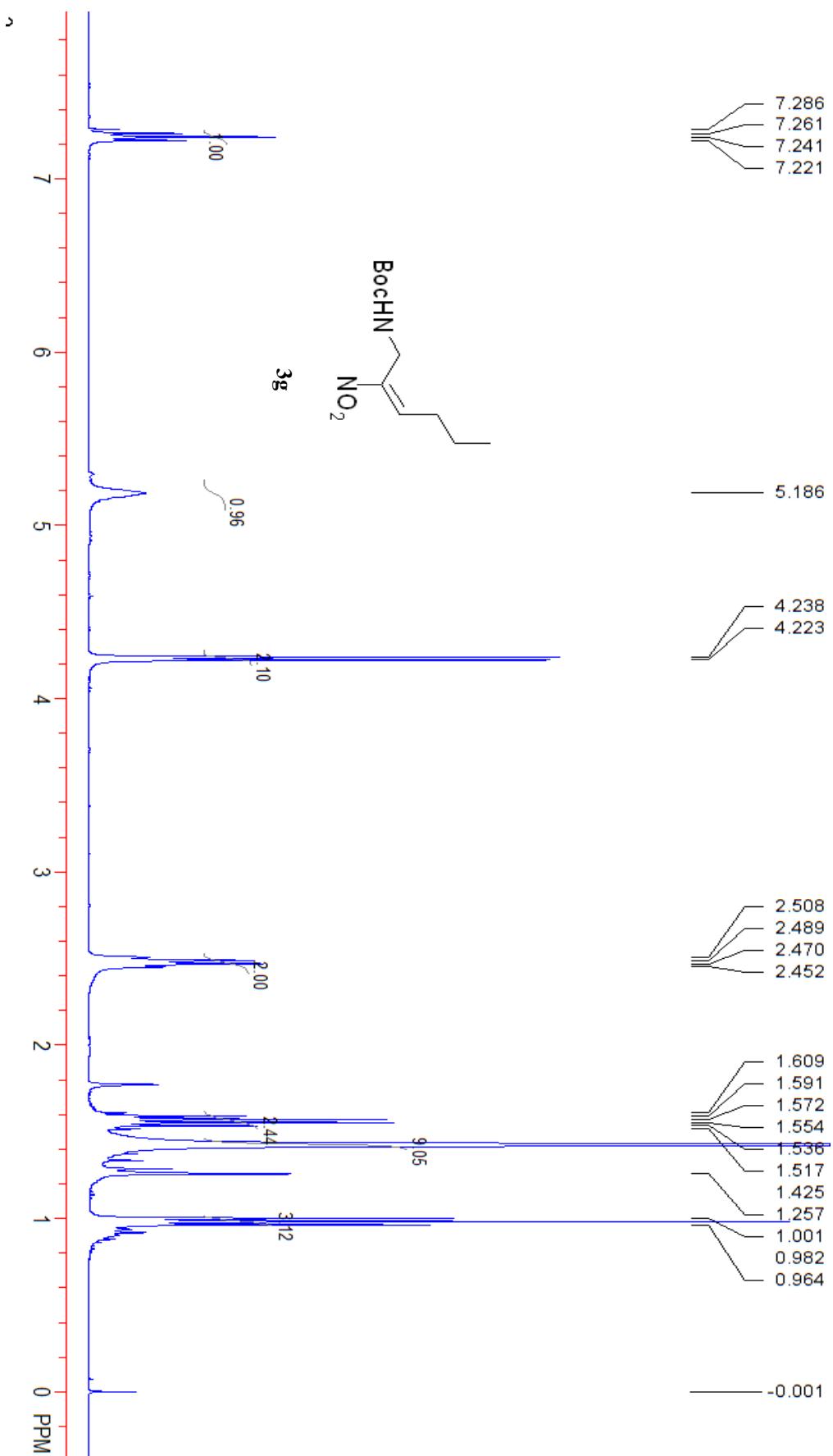


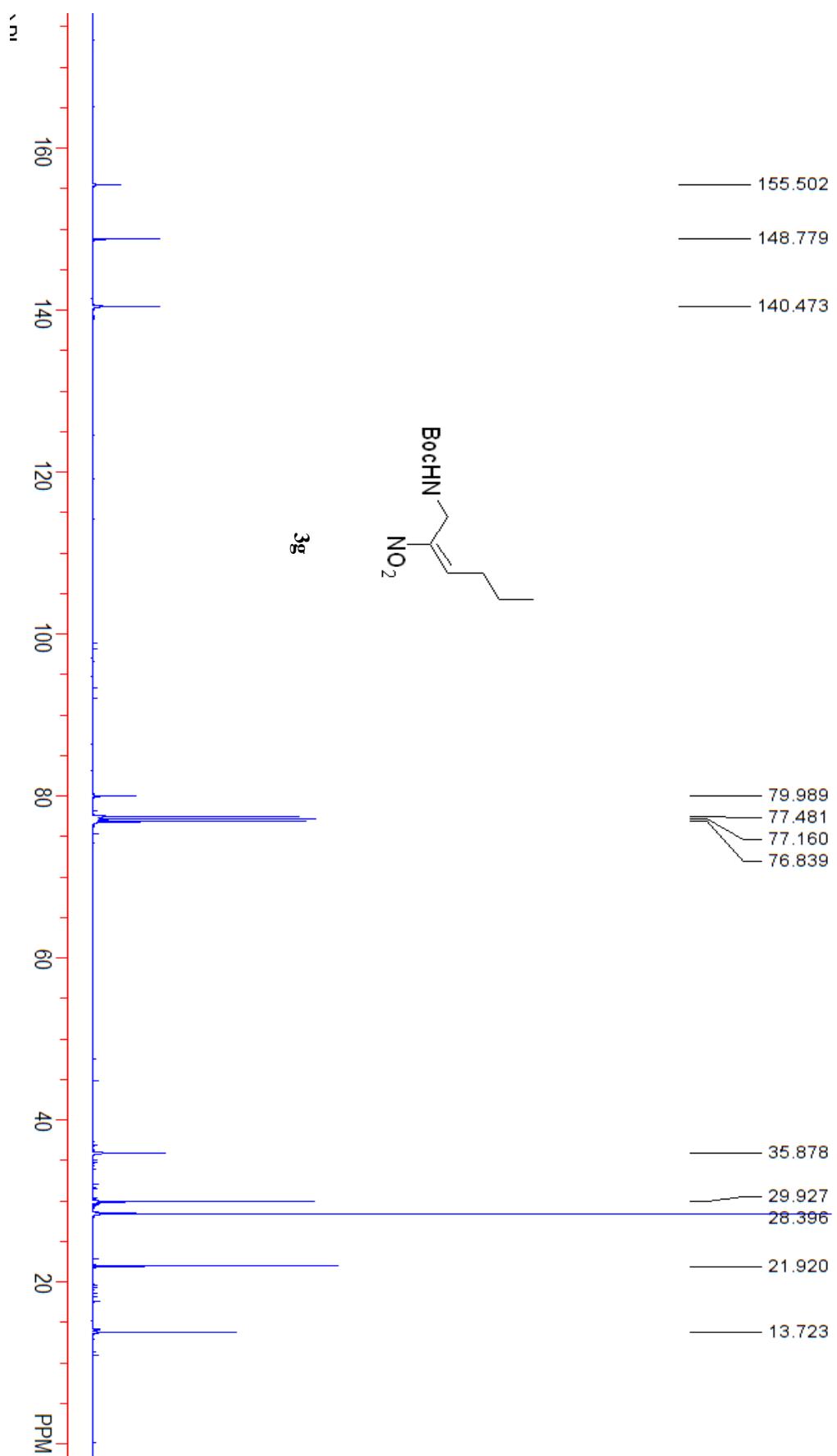


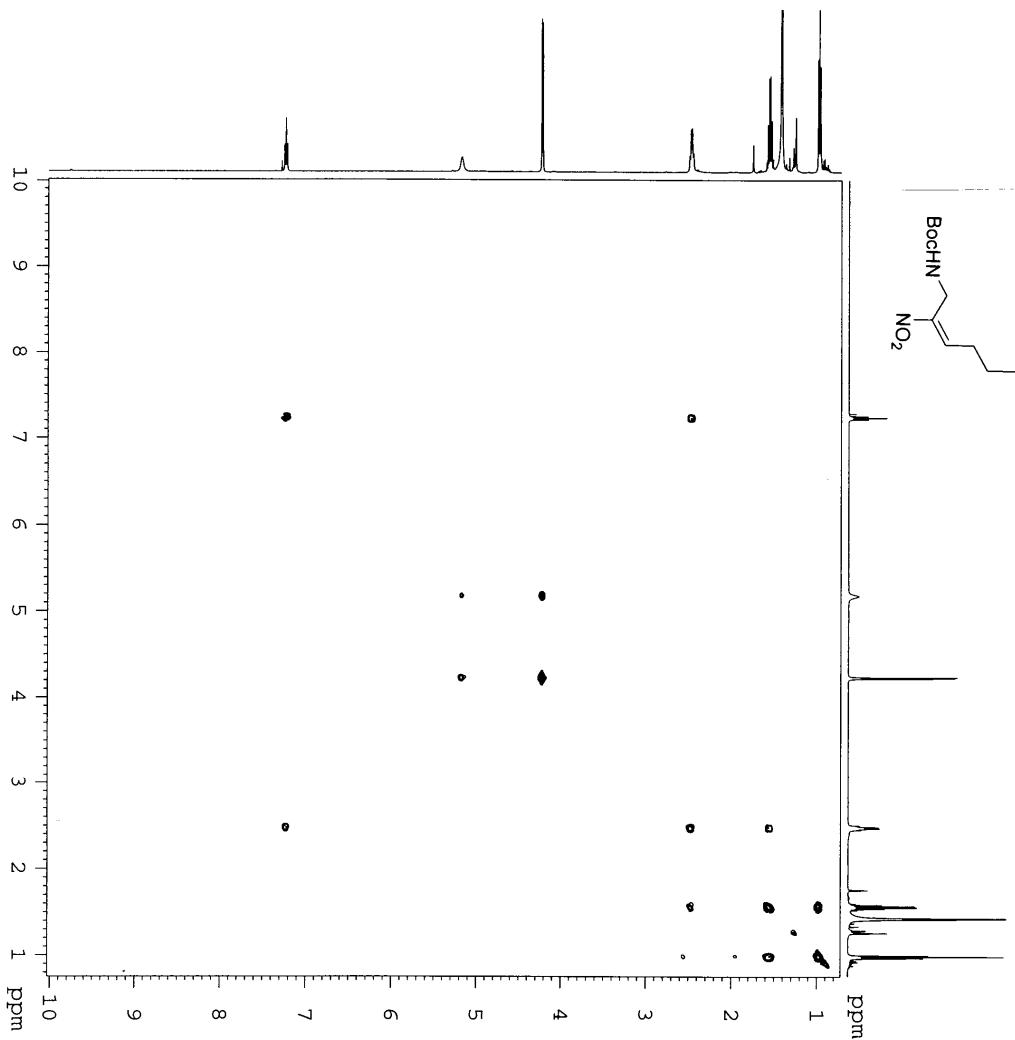












Current Data Parameters

NAME 20051025crys_1

ENZO PROCMD

F2 - Acquisition Parameters

DATE 20100824

TIME 12:56

INSTRUM spect

PROBHD 5 mm QNP 1H/13

PULPROG cosyoutgrf

TD 1024

SOLVENT CDCl3

NS 1

DS 16

TE 5482.456 Hz

SPHES 5.353.69 Hz

FTRES 0.0393320 sec

RG 1.04 sec

AD 6.50 usec

DW 91.200 usec

DE 300.0 K

TE 0.0000300 sec

DO 2.0000000 sec

D1 0.00000400 sec

d13 0.0002000 sec

D16 0.00018240 sec

INO 0.00018240 sec

===== CHANNEL f1 =====

NUCL 1H usec

T1 10.0 usec

PR1 4.00 dB

SW1 500.1324200 MHz

===== GRADIENT CHANNEL =====

GRADNAM1 SINE,100

GRADNAM2 SINE,100

GRADNAM3 SINE,100

GR21 16.00 %

GR22 12.00 %

GR23 40.00 %

P16 1000.00 usec

PL - Acquisition parameters

ND 1

TD 256

SW1 500.1324 MHz

SP1 21.41584 Hz

SW2 10.962 Ppm

FMODE Of

F2 - Processing parameters

SI 1024

SF 500.1300016 MHz

NOM SINE

SSB 0

LB 0.00 Hz

GB 0

PC 1.40

PL - Processing parameters

SI 1024

SF 500.1300016 MHz

NOM SINE

SSB 0

LB 0.00 Hz

GB 0

