

Organocatalytic Approach to Polysubstituted Piperidines and Tetrahydropyrans

You Wang,¹ Shaolin Zhu,² and Dawei Ma^{2*}

¹Department of Chemistry, Fudan University, Shanghai 200433, China

*²State Key Laboratory of Bioorganic and Natural Products Chemistry, Shanghai
Institute of Organic Chemistry, Chinese Academy of Sciences, 354 Fenglin Lu,
Shanghai 200032, China*

madw@mail.sioc.ac.cn

Supporting Information

Table of contents

1	Experimental-----	S2
2	Copies of NMR and HPLC spectra for new compounds-----	S22

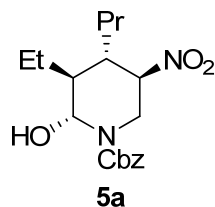
Experimental

General information. For thin-layer chromatography (TLC), Silica gel plates GF254 were used and compounds were visualized by irradiation with UV light, I₂, or by treatment with a solution of phosphomolybdic acid in ethanol followed by heating. Optical rotations were measured by used a Perkin–Elmer 241MC polarimeter in the solvent indicated. FT-IR spectra were recorded on an AVATAR-360 spectrophotometer. ¹H and ¹³C NMR spectra were recorded on MERCURY300, Bruker DRX-400, and Bruker AV-500 spectrometers with TMS as the internal standard. HRMS were recorded by using either FTMS-7 or IonSpec 4.7 spectrometers. HPLC was carried out using a Water 515 pump, waters 2487 UV detector, Millennium workstation, Phenomenex (Lux Amylose-2, Lux Cellulose-1, Lux Cellulose-2), Daicel Chiralpak HPLC column. Yields refer to pure compounds, unless otherwise indicated.

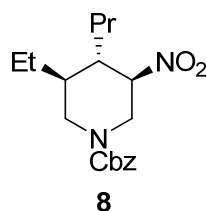
General procedure for cascade Michael-aminalization process of aldehydes with substituted nitroalkenes: Aldehyde **2** (0.40 mmol) was added to a suspension of catalyst **1** (0.02-0.03 mmol), nitroalkenes **3** (0.20 mmol) and benzoic acid (0.06-0.09 mmol) in water (0.40 mL) at room temperature. The stirring was continued until complete conversion of **3** (monitored by TLC). The reaction mixture was diluted with ethyl acetate, dried over Na₂SO₄ and concentrated. Purification by flash column chromatography (silica gel, petroleum ether/AcOEt) afforded the the corresponding substituted piperidine. The enantiomeric excess (ee) was determined by HPLC on a chiral phase. *Noteworthy is that these substituted piperidines could isomerize at the chiral center of hemiaminal part during standing in CDCl₃, and therefore some additional peaks (for isomer) were observed in ¹³C NMR spectra if ¹³C NMR determination took longer time.*

Reduction of **3 with triethylsilane and BF₃·OEt₂.** To a mixture of the above crude product (0.15 mmol) and triethylsilane (72 μL, 0.45 mmol) in CH₂Cl₂ (1.5 mL) was added BF₃·OEt₂ (23 μL, 0.3 mmol) at 0 °C. The reaction mixture was stirred for 1-3 h.

Purification by flash column chromatography (silica gel, petroleum ether/AcOEt) afforded the reduced product. The enantiomeric excess (ee) was determined by HPLC on a chiral phase.

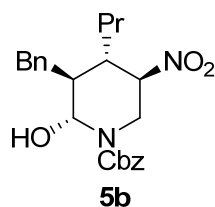


(2R,3S,4S,5R)-3-Ethyl-2-hydroxy-5-nitro-4-propyl-piperidine-1-carboxylic acid benzyl ester 5a. $[\alpha]_D^{28} = -8.4$ (c 1.00, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.39-7.32 (m, 5H), 5.74 (s, 1H), 5.15 (s, 2H), 4.47 (dt, $J = 4.4, 10.4$ Hz, 1H), 4.18 (d, $J = 10.0$ Hz, 1H), 3.78 (t, $J = 10.8$ Hz, 1H), 3.00-2.80 (br, 1H), 2.46-2.39 (m, 1H), 1.63-1.56 (m, 1H), 1.48-1.37 (m, 3H), 1.33-1.21 (m, 3H), 1.00 (t, $J = 6.8$ Hz, 3H), 0.89 (t, $J = 6.8$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 155.12, 135.89, 128.81, 128.60, 128.27, 85.39, 74.97, 68.07, 42.79, 41.22, 37.59, 30.28, 21.27, 17.17, 14.49, 11.38; IR (neat): $\nu = 3432$ br, 2962 m, 2934 m, 1690 s, 1550 s, 1431 m, 1145 m, 963 m, 748 m, 698 cm^{-1} ; ESI-MS m/z 373.4 ($\text{M} + \text{Na}$) $^+$, HRMS (MALDI) calcd for $\text{C}_{18}\text{H}_{26}\text{N}_2\text{O}_5\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 373.1734, found 373.1733; HPLC: PHENOMENEX LUX AMYLOSE-2 (PA-2) column, hexane/*i*-PrOH = 70:30, flow rate 0.7 mL/min, 214 nm, major isomer: $t_R = 8.5$ min (major), $t_R = 7.7$ min (minor), $ee=98\%$. (Noteworthy is that some additional peaks (for minor isomer) were observed in ^{13}C NMR spectra due to isomerization)

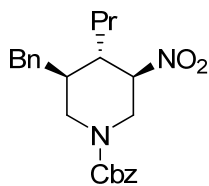


(3S,4S,5R)-3-Ethyl-5-nitro-4-propyl-piperidine-1-carboxylic acid benzyl ester 8. $[\alpha]_D^{28} = -40.6$ (c 1.00, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.39-7.31 (m, 5H), 5.20-5.07 (m, 2H), 4.65-4.38 (m, 2H), 4.30-4.10 (m, 1H), 3.30-3.10 (m, 1H),

2.63-2.45 (m, 1H), 1.97 (tt, $J = 3.2, 10.8$ Hz, 1H), 1.70-1.60 (m, 1H), 1.60-1.46 (m, 1H), 1.45-1.35 (m, 1H), 1.35-1.15 (m, 4H), 0.93 (t, $J = 7.2$ Hz, 3H), 0.88 (t, $J = 6.8$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 154.88, 136.35, 128.72, 128.39, 128.14, 85.57, 67.78, 47.80, 47.16, 43.73, 38.43, 30.37, 22.71, 17.65, 14.56, 10.75; IR (neat): $\nu = 2962$ m, 2875 m, 1694 s, 1546 s, 1440 s, 1222 m, 1146 m, 1098 w, 754 w, 699 m cm^{-1} ; ESI-MS m/z 335.2 ($\text{M} + \text{H}$) $^+$, 357.3 ($\text{M} + \text{Na}$) $^+$, HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{26}\text{N}_2\text{O}_4\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 357.1785, found 357.1793; HPLC: PHENOMENEX LUX AMYLOSE-2 (PA-2) column, hexane/*i*-PrOH = 70:30, flow rate 0.7 mL/min, 214 nm, $t_R = 12.5$ min (major), $t_R = 11.0$ min (minor), $ee = 98\%$.

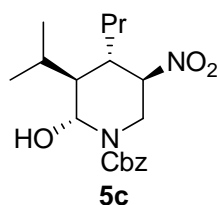


(2R,3S,4S,5R)-3-Benzyl-2-hydroxy-5-nitro-4-propyl-piperidine-1-carboxylic acid benzyl ester 5b. $[\alpha]_D^{27} = -8.7$ (c 1.02, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ major isomer: 7.31-7.19 (m, 10H), 5.26 (d, $J = 2.0$ Hz, 1H), 5.08-4.98 (m, 2H), 4.50 (dt, $J = 4.4, 10.4$ Hz, 1H), 4.05-4.22 (m, 1H), 3.80 (t, $J = 10.8$ Hz, 1H), 2.91 (dd, $J = 4.0, 13.6$ Hz, 1H), 2.65-2.57 (m, 2H), 1.78 (tt, $J = 3.6, 14.8$ Hz, 1H), 1.61-1.55 (m, 1H), 1.45-1.36 (m, 3H), 0.93 (t, $J = 6.8$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ major isomer: 154.67, 138.93, 135.76, 129.09, 128.78, 128.72, 128.43, 127.92, 126.66, 85.17, 74.73, 67.85, 43.51, 41.17, 37.49, 34.66, 30.39, 17.24, 14.50; IR (neat): $\nu = 3419$ br, 2959 m, 2932 m, 1705 s, 1550 s, 1432 m, 1305 m, 1143 m, 954 w, 737 w, 699 m cm^{-1} ; ESI-MS m/z 435.5 ($\text{M} + \text{Na}$) $^+$, HRMS (MALDI) calcd for $\text{C}_{23}\text{H}_{28}\text{N}_2\text{O}_5\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 435.1890, found 435.1889. (*Noteworthy is that some additional peaks (for minor isomer) were observed in ^{13}C NMR spectra due to isomerization*)



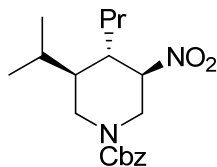
reduction product of **5b**

(3S,4S,5R)-3-Benzyl-5-nitro-4-propyl-piperidine-1-carboxylic acid benzyl ester (reduction preduct of 5b). $[\alpha]_D^{28} = -16.8$ (*c* 1.20, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.35-7.21 (m, 7H), 7.12-7.06 (m, 3H), 5.10 (d, *J* = 11.2 Hz, 1H), 4.99 (d, *J* = 11.2 Hz, 1H), 4.55-4.45 (m, 2H), 3.95 (d, *J* = 7.2 Hz, 1H), 3.20-3.30 (m, 1H), 3.02 (d, *J* = 11.6 Hz, 1H), 2.54 (t, *J* = 11.6 Hz, 1H), 2.20-2.35 (m, 1H), 2.10-2.04 (m, 1H), 1.76-1.61 (m, 2H), 1.42-1.32 (m, 3H), 0.92 (t, *J* = 7.0 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 154.65, 138.58, 136.18, 128.91, 128.81, 128.64, 128.18, 127.58, 126.72, 85.23, 67.61, 48.06, 47.06, 44.29, 39.28, 36.76, 30.58, 17.71, 14.56; IR (neat): ν = 2959 m, 2932 m, 2873 m, 1708 s, 1538 s, 1470 m, 1455 m, 1374 m, 1254 m, 1222 s, 1139 s, 736 m, 699 m cm⁻¹; ESI-MS *m/z* 419.4 (M + Na)⁺, HRMS (ESI) calcd for C₂₃H₂₈N₂O₄Na (M + Na)⁺ *m/z* 419.1941, found 419.1950; HPLC: PHENOMENEX LUX CELLULOSE-2 (PC-2) column, hexane/*i*-PrOH = 70:30, flow rate 0.7 mL/min, 214 nm, *t*_R = 14.5 min (major), *t*_R = 10.6 min (minor), *ee*=99%.



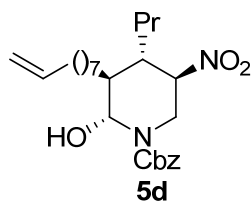
(2R,3S,4S,5R)-2-Hydroxy-3-isopropyl-5-nitro-4-propyl-piperidine-1-carboxylic acid benzyl ester 5c. $[\alpha]_D^{26} = -8.6$ (*c* 0.97, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ major isomer: 7.32-7.30 (m, 5H), 5.82 (s, 1H), 5.12 (s, 2H), 4.51 (dt, *J* = 4.4, 10.0 Hz, 1H), 4.16 (d, *J* = 8.8, Hz, 1H), 3.82 (t, *J* = 11.4, Hz, 1H), 2.73-2.67 (m, 1H), 1.98-1.90 (m, 1H), 1.46-1.43 (m, 1H), 1.42-1.40 (m, 1H), 1.37-1.28 (m, 2H), 1.24-1.16 (m, 1H), 1.09 (d, *J* = 7.2, Hz, 3H), 1.02 (d, *J* = 7.2 Hz, 3H), 0.90 (t, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ major isomer: 154.90, 135.89, 128.78, 128.55, 128.19, 85.34, 75.46, 67.98, 44.90, 40.97, 35.31, 29.80, 26.62, 20.88, 17.70, 16.39, 14.57; IR (neat): ν = 3437 br, 2962 s, 2934 m, 1702 s, 1552 s, 1429 m, 1266 m, 985 m, 739 s, 702 m

cm⁻¹; ESI-MS m/z 387.3 (M + Na)⁺, HRMS (ESI) calcd for C₁₉H₂₈N₂O₅Na (M + Na)⁺ m/z 387.1890, found 387.1883. (Noteworthy is that some additional peaks (for isomer) were observed in ¹³C NMR spectradue to isomerization)



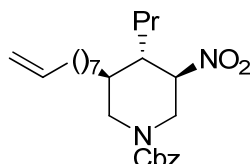
reduction product of **5c**

(3S,4S,5R)-3-Isopropyl-5-nitro-4-propyl-piperidine-1-carboxylic acid benzyl ester (reduction product of 5c). [α]_D²⁵ = -34.4 (*c* 0.97, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.39-7.31 (m, 5H), 5.19-5.09 (m, 2H), 4.54-4.42 (m, 2H), 4.20-4.08 (m, 1H), 3.26-3.14 (m, 1H), 2.68-2.56 (m, 1H), 2.15 (tt, *J* = 3.2, 10.8 Hz, 1H), 2.00-1.92 (m, 1H), 1.54-1.48 (m, 1H), 1.38-1.16 (m, 4H), 1.06-0.98 (m, 3H), 0.88 (t, *J* = 6.8 Hz, 3H), 0.86 (t, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 155.00, 136.37, 128.72, 128.39, 128.11, 85.67, 67.78, 47.28, 43.45, 41.87, 29.85, 25.76, 21.42, 17.12, 15.71, 14.60; IR (neat): ν = 2961 s, 2933 m, 1709 s, 1549 s, 1470 m, 1435 m, 1374 m, 1292 m, 1223 s, 1147 m, 1098 m, 970 w, 745 m, 698 w cm⁻¹; ESI-MS m/z 349.3 (M + H)⁺, 371.3 (M + Na)⁺, HRMS (ESI) calcd for C₁₉H₂₈N₂O₄Na (M + Na)⁺ m/z 397.1734, found 397.1741; HPLC: PHENOMENEX LUX AMYLOSE-2 (PA-2) column, hexane/*i*-PrOH = 50:50, flow rate 0.7 mL/min, 214 nm, *t*_R = 9.3 min (major), *t*_R = 8.7 min (minor), *ee* > 99%.



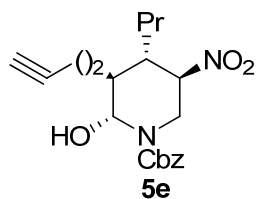
(2R,3S,4S,5R)-2-Hydroxy-5-nitro-3-non-8-enyl-4-propyl-piperidine-1-carboxylic acid benzyl ester 5d. [α]_D²⁵ = -13.5 (*c* 1.00, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.37-7.35 (m, 5H), 5.84-5.76 (m, 1H), 5.70 (s, 1H), 5.15 (s, 2H), 4.99 (d, *J* = 16.8 Hz, 1H), 4.93 (d, *J* = 10.0 Hz, 1H), 4.46 (dt, *J* = 4.4, 10.4 Hz, 1H), 4.17 (d, *J* = 8.4 Hz, 1H), 3.78 (t, *J* = 11.2 Hz, 1H), 2.42 (m, 1H), 2.33 (t, *J* = 7.6 Hz, 1H), 2.06-2.01 (m, 2H), 1.64-1.61 (m, 1H), 1.45-1.26 (m, 16H), 0.90 (t, *J* = 6.4 Hz, 3H); ¹³C NMR (100 MHz,

CDCl₃) δ 155.15, 139.27, 135.88, 128.81, 128.61, 128.28, 114.34, 85.37, 75.24, 68.09, 41.24, 37.61, 33.90, 30.33, 29.82, 29.53, 29.18, 29.03, 28.30, 26.70, 24.84, 17.21, 14.50; IR (neat): ν = 3448 br, 2928 s, 1686 s, 1551 s, 1336 w, 1143 m, 968 m, 910 m, 697 m cm⁻¹; ESI-MS m/z 469.5 (M + Na)⁺, HRMS (ESI) calcd for C₂₅H₃₈N₂O₅Na (M + Na)⁺ m/z 469.2673, found 469.2682. (Noteworthy is that some additional peaks (for isomer) were observed in ¹³C NMR spectradue to isomerization)



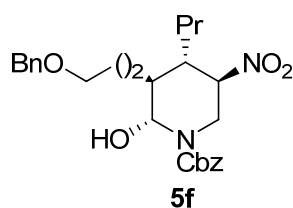
reduction product of **5d**

(3R,4S,5S)-3-Nitro-5-non-8-enyl-4-propyl-piperidine-1-carboxylic acid benzyl ester (reduction product of 5d). [α]_D²⁷ = -33.7 (*c* 1.00, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.39-7.32 (m, 5H), 5.86-5.76 (m, 1H), 5.18-5.10 (m, 2H), 5.00 (dd, *J* = 1.6, 17.2 Hz, 1H), 4.94 (d, *J* = 10.4 Hz, 1H), 4.54-4.38 (m, 2H), 4.27-4.10 (m, 1H), 3.27-3.15 (m, 1H), 2.60-2.45 (m, 1H), 2.04 (q, *J* = 6.8 Hz, 2H), 1.97-1.92 (m, 1H), 1.56-1.50 (m, 2H), 1.39-1.12 (m, 15H), 0.88 (t, *J* = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 154.83, 139.23, 136.35, 128.72, 128.40, 128.13, 114.35, 85.48, 67.79, 48.23, 47.16, 44.11, 37.16, 33.87, 30.42, 29.82, 29.44, 29.12, 29.00, 26.32, 17.68, 14.57; IR (neat): ν = 2928 s, 2855 s, 1710 s, 1550 s, 1467 s, 1433 s, 1376 m, 1290 m, 1221 s, 1142 m, 970 m, 911 m, 754 m, 698 m cm⁻¹; ESI-MS m/z 431.5 (M + H)⁺, 453.5 (M + Na)⁺, HRMS (MALDI) calcd for C₂₅H₃₈N₂O₄Na (M + Na)⁺ m/z 453.2724, found 453.2719; HPLC: PHENOMENEX LUX AMYLOSE-2 (PA-2) column, hexane/*i*-PrOH = 50:50, flow rate 0.7 mL/min, 214 nm, *t*_R = 9.2 min (major), *t*_R = 7.8 min (minor), *ee* > 99%.



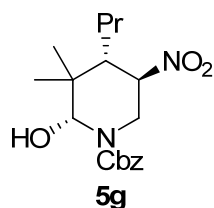
(2R,3S,4S,5R)-3-But-3-ynyl-2-hydroxy-5-nitro-4-propyl-piperidine-1-carboxylic acid benzyl ester 5e. [α]_D²⁵ = -26.2 (*c* 1.05, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ

7.38-7.31 (m, 5H), 5.75 (s, 1H), 5.18-5.10 (m, 2H), 4.50 (dt, $J = 4.0, 10.0$ Hz, 1H), 4.11-4.24 (m, 1H), 3.77 (t, $J = 11.2$ Hz, 1H), 2.45 (t, $J = 10.4$ Hz, 1H), 2.35-2.31 (m, 2H), 1.97 (brs, 1H), 1.76-1.66 (m, 3H), 1.48-1.42 (m, 1H), 1.34-1.26 (m, 4H), 0.90 (t, $J = 6.8$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 155.13, 135.78, 128.80, 128.61, 128.26, 85.15, 83.27, 74.64, 69.58, 68.13, 41.11, 39.72, 37.30, 30.20, 26.79, 17.05, 15.68, 14.46; IR (neat): $\nu = 3435$ br, 3297 s, 2959 s, 2932 s, 1694 s, 1549 m, 1430 m, 1337 m, 1144 m, 1095 m, 981 s, 698 m, 646 s cm^{-1} ; ESI-MS m/z 397.3 ($\text{M} + \text{Na}$) $^+$, HRMS (ESI) calcd for $\text{C}_{20}\text{H}_{26}\text{N}_2\text{O}_5\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 397.1734, found 397.1741; HPLC: sino-AD column, hexane/*i*-PrOH = 90:10, flow rate 0.7 mL/min, 214 nm, major isomer: $t_R = 10.2$ min (major), $t_R = 12.6$ min (minor), $ee=99\%$. (Noteworthy is that some additional peaks (for isomer) were observed in ^{13}C NMR spectradue to isomerization)



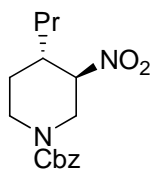
(2R,3S,4S,5R)-3-(3-Benzyloxy-propyl)-2-hydroxy-5-nitro-4-propyl-piperidine-1-carboxylic acid benzyl ester 5f. $[\alpha]_D^{25} = -11.7$ (c 1.22, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ major isomer: 7.35-7.29 (m, 10H), 5.71 (s, 1H), 5.12 (s, 2H), 4.47 (s, 2H), 4.43 (dd, $J = 4.0, 10.0$ Hz, 1H), 4.21-4.11 (m, 1H), 3.73 (t, $J = 11.6$ Hz, 1H), 3.47 (t, $J = 6.0$ Hz, 2H), 2.46-2.37 (m, 1H), 1.86-1.75 (m, 1H), 1.61-1.59 (m, 2H), 1.47-1.42 (m, 2H), 1.28-1.24 (m, 4H), 0.88 (t, $J = 6.4$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ major isomer: 154.98, 138.38, 135.87, 128.76, 128.54, 128.49, 128.24, 127.78, 127.74, 85.33, 75.04, 73.05, 70.18, 68.00, 41.13, 37.58, 30.21, 29.78, 26.59, 24.94, 17.15, 14.44; IR (neat): $\nu = 3418$ br, 2958 s, 2931 s, 2872 s, 1705 s, 1549 s, 1455 m, 1337 w, 1102 m, 968 m, 738 s, 698 m cm^{-1} ; ESI-MS m/z 493.5 ($\text{M} + \text{Na}$) $^+$, HRMS (MALDI) calcd for $\text{C}_{26}\text{H}_{34}\text{N}_2\text{O}_6\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 493.2309, found 493.2311; HPLC: PHENOMENEX LUX AMYLOSE-2 (PA-2) column, hexane/*i*-PrOH = 70:30, flow rate 0.7 mL/min, 214 nm, major isomer: $t_R = 18.3$ min (major), $t_R = 10.5$ min (minor),

ee > 99%. (Noteworthy is that some additional peaks (for isomer) were observed in ^{13}C NMR spectradue to isomerization).



(2R,4S,5R)-2-Hydroxy-3,3-dimethyl-5-nitro-4-propyl-piperidine-1-carboxylic

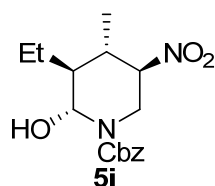
acid benzyl ester 5g. $[\alpha]_{\text{D}}^{28} = -8.7$ (*c* 0.67, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ major isomer: 7.40-7.32 (m, 5H), 5.26 (s, 1H), 5.14 (d, *J* = 7.2 Hz, 2H), 4.48 (dt, *J* = 4.8, 11.2 Hz, 1H), 4.39-4.17 (m, 1H), 3.70 (t, *J* = 12.0 Hz, 1H), 2.36-2.32 (m, 1H), 1.82-1.56 (m, 1H), 1.49-1.42 (m, 1H), 1.24-1.18 (m, 2H), 1.10 (s, 3H), 0.85 (t, *J* = 6.8 Hz, 3H), 0.82 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ major isomer: 155.30, 135.89, 128.82, 128.62, 128.26, 86.06, 82.44, 68.09, 41.62, 39.25, 30.20, 24.75, 23.24, 19.67, 14.47; IR (neat): ν = 3428 br, 2962 m, 2918 m, 1690 s, 1552 s, 1431 s, 1331 m, 1213 m, 1055 m, 968 m, 739 m, 698 m cm^{-1} ; ESI-MS *m/z* 373.4 ($\text{M} + \text{Na}$) $^+$, HRMS (MALDI) calcd for $\text{C}_{18}\text{H}_{26}\text{N}_2\text{O}_5\text{Na}$ ($\text{M} + \text{Na}$) $^+$ *m/z* 373.1734, found 373.1741. HPLC: PHENOMENEX LUX CELLULOSE-2 (PC-2) column, hexane/*i*-PrOH = 90:10, flow rate 0.7 mL/min, 214 nm, major isomer: t_{R} = 16.1 min (major), t_{R} = 9.7 min (minor), *ee* = 90%, minor isomer: t_{R} = 8.4 min (major), t_{R} = 11.6 min (minor), *ee* = 92%. (Noteworthy is that some additional peaks (for isomer) were observed in ^{13}C NMR spectradue to isomerization)



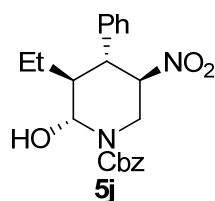
reduction product of **5h**

(3R,4S)-3-Nitro-4-propyl-piperidine-1-carboxylic acid benzyl ester (reduction product of 5h). $[\alpha]_{\text{D}}^{29} = -17.17$ (*c* 0.83, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.39-7.32 (m, 5H), 5.14-5.13 (m, 2H), 4.60-4.47 (m, 1H), 4.27-4.23 (m, 1H), 4.20-4.15 (m, 1H), 3.30-3.15 (m, 1H), 2.86 (t, *J* = 11.2 Hz, 1H), 2.14-2.12 (m, 1H), 1.91 (d, *J* = 13.2 Hz, 1H), 1.43-1.34 (m, 2H), 1.25-1.18 (m, 3H), 0.89 (t, *J* = 6.8 Hz,

3H); ^{13}C NMR (100 MHz, CDCl_3) δ 154.95, 136.31, 128.75, 128.45, 128.24, 87.06, 67.86, 47.01, 43.65, 39.73, 34.22, 28.61, 19.03, 14.06; IR (neat): ν = 2959 s, 2929 s, 1705 s, 1548 s, 1431 s, 1212 s, 1155 m, 1091 m, 973 m, 751 m, 698 cm^{-1} ; ESI-MS m/z 307.3 ($\text{M} + \text{H}$) $^+$, 329.3 ($\text{M} + \text{Na}$) $^+$, HRMS (ESI) calcd for $\text{C}_{16}\text{H}_{22}\text{N}_2\text{O}_4\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 329.1472, found 329.1472; HPLC: PHENOMENEX LUX AMYLOSE-2 (PA-2) column, hexane/*i*-PrOH = 80:20, flow rate 0.7 mL/min, 214 nm, major isomer: t_R = 20.1 min (major), t_R = 21.7 min (minor), *ee* = 86%.

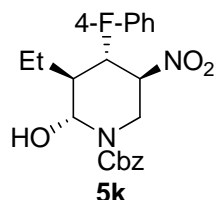


(2R,3S,4S,5R)-3-Ethyl-2-hydroxy-4-methyl-5-nitro-piperidine-1-carboxylic acid benzyl ester 5i. $[\alpha]_D^{29}$ = -26.8 (*c* 0.75, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.36-7.32 (m, 5H), 5.74 (d, J = 2.0 Hz, 1H), 5.14 (s, 2H), 4.27-4.21 (m, 2H), 3.67 (t, J = 12.0 Hz, 1H), 2.45-2.38 (m, 1H), 1.70-1.63 (m, 1H), 1.46-1.38 (m, 1H), 1.26-1.20 (m, 1H), 1.00 (t, J = 7.2 Hz, 3H), 0.98 (d, J = 6.8 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 155.06, 135.83, 128.81, 128.62, 128.24, 88.71, 74.91, 68.10, 46.04, 41.27, 33.82, 21.50, 15.25, 11.34; IR (neat): ν = 3466 br, 2962 m, 2929 s, 1698 s, 1669 s, 1549 s, 1536 s, 1472 m, 1439 m, 1337 m, 1246 m, 1149 m, 1019 m, 729 cm^{-1} ; ESI-MS m/z 345.3 ($\text{M} + \text{Na}$) $^+$, HRMS (ESI) calcd for $\text{C}_{16}\text{H}_{22}\text{N}_2\text{O}_5\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 345.1421, found 345.1424; HPLC: PHENOMENEX LUX AMYLOSE-2 (PA-2) column, hexane/*i*-PrOH = 70:30, flow rate 0.7 mL/min, 214 nm, major isomer: t_R = 10.3 min (major), t_R = 8.2 min (minor), *ee* = 98%. (Noteworthy is that some additional peaks (for isomer) were observed in ^{13}C NMR spectra due to isomerization)

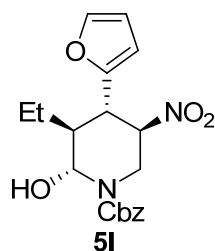


(2R,3S,4R,5R)-3-Ethyl-2-hydroxy-5-nitro-4-phenylpiperidine-1-carboxylic acid benzyl ester 5j. $[\alpha]_D^{25}$ = +33.0 (*c* 1.32, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.37-7.32 (m, 5H), 7.30-7.23 (m, 3H), 7.15-7.13 (m, 2H), 5.90 (d, J = 1.6 Hz, 1H),

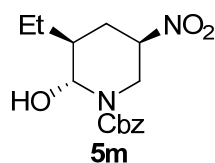
5.17 (s, 2H), 4.76 (dt, $J = 4.8, 11.2$ Hz, 1H), 4.42 (d, $J = 8.0$ Hz, 1H), 3.79 (t, $J = 12.0$ Hz, 1H), 3.48 (t, $J = 11.6$ Hz, 1H), 1.80-1.74 (m, 1H), 1.38-1.29 (m, 1H), 1.09-1.03 (m, 1H), 0.83 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 155.10, 137.49, 135.72, 129.08, 128.83, 128.67, 128.28, 128.01(2C), 87.86, 74.95, 68.23, 46.19, 45.65, 41.56, 22.04, 11.22; IR (neat): $\nu = 3396$ br, 2962 w, 1675 s, 1434 s, 1357 m, 1336 m, 1288 m, 1193 m, 1151 m, 1109 m, 965 m, 765 w, 701 w cm^{-1} ; ESI-MS m/z 407.4 ($\text{M} + \text{Na}$) $^+$, HRMS (ESI) calcd for $\text{C}_{21}\text{H}_{24}\text{N}_2\text{O}_5\text{Na}$ ($\text{M} + \text{Na}$) $^+$ m/z 407.1577, found 407.1578; HPLC: PHENOMENEX LUX CELLULOSE-2 (PC-2) column, hexane/*i*-PrOH = 70:30, flow rate 0.7 mL/min, 214 nm, major isomer: $t_R = 11.1$ min (major), $t_R = 6.7$ min (minor), $ee > 99\%$. (Noteworthy is that some additional peaks (for isomer) were observed in ^{13}C NMR spectra due to isomerization)



(3S,4R,5R)-3-Ethyl-4-(4-fluorophenyl)-5-nitro-piperidine-1-carboxylic acid benzyl ester 5k. $[\alpha]_D^{23} = +27.6$ (c 1.43, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.39-7.34 (m, 5H), 7.13-7.10 (m, 2H), 7.02-6.97 (m, 2H), 5.90 (s, 1H), 5.17 (s, 2H), 4.70 (dt, $J = 4.4, 11.2$ Hz, 1H), 4.43 (d, $J = 7.6$ Hz, 1H), 3.79 (t, $J = 11.6$ Hz, 1H), 3.48 (t, $J = 11.6$ Hz, 1H), 1.72 (tt, $J = 2.8, 11.2$ Hz, 1H), 1.37-1.29 (m, 1H), 1.07-1.01 (m, 1H), 0.83 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 162.35 ($J = 245.7$ Hz), 155.10, 135.63, 133.21 ($J = 3.6$ Hz), 129.55, 128.83, 128.69, 128.28, 116.07 ($J = 21.1$ Hz), 87.84, 74.86, 68.28, 45.74, 45.49, 41.49, 21.98, 11.19; ^{19}F NMR (300 MHz, CDCl_3) δ -114.50; IR (neat): $\nu = 3418$ br, 1683 s, 1549 s, 1513 m, 1435 m, 1335 m, 1226 m, 833 m, 733 m cm^{-1} ; ESI-MS m/z 425.2 ($\text{M} + \text{Na}$) $^+$, HRMS (ESI) calcd for $\text{C}_{21}\text{H}_{23}\text{N}_2\text{O}_5\text{FNa}$ ($\text{M} + \text{Na}$) $^+$ m/z 425.1483, found 425.1501; HPLC: PHENOMENEX LUX CELLULOSE-2 (PC-2) column, hexane/*i*-PrOH = 80:20, flow rate 0.7 mL/min, 214 nm, major isomer: $t_R = 16.7$ min (major), $t_R = 8.6$ min (minor), $ee > 99\%$. (Noteworthy is that some additional peaks (for isomer) were observed in ^{13}C NMR spectra due to isomerization)

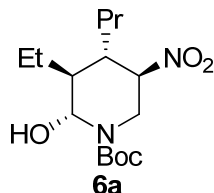


(2R,3S,4S,5R)-3-Ethyl-4-furan-2-yl-2-hydroxy-5-nitro-piperidine-1-carboxylic acid benzyl ester 5l. $[\alpha]_D^{26} = +40.0$ (*c* 1.16, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.40-7.34 (m, 6H), 6.26 (dd, *J* = 2.0, 2.8 Hz, 1H), 6.15 (d, *J* = 2.8 Hz, 1H), 5.86 (d, *J* = 2.0 Hz, 1H), 5.17 (s, 2H), 4.81 (dt, *J* = 4.8, 11.2 Hz, 1H), 4.40 (d, *J* = 11.6 Hz, 1H), 3.76 (t, *J* = 12.0 Hz, 1H), 3.62 (t, *J* = 11.6 Hz, 1H), 1.92-1.85 (m, 1H), 1.42-1.36 (m, 1H), 1.17-1.10 (m, 1H), 0.86 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 155.03, 150.48, 142.62, 135.74, 128.85, 128.68, 128.30, 110.41, 109.54, 85.36, 74.93, 68.25, 43.98, 41.14, 40.14, 22.37, 11.18; IR (neat): ν = 3419 s, 2965 m, 1678 s, 1556 s, 1435 w, 1335 w, 1192 m, 1151 m, 965 m, 883 m, 758 s cm⁻¹; ESI-MS *m/z* 397.3 (M + Na)⁺, HRMS (ESI) calcd for C₁₉H₂₂N₂O₆Na (M + Na)⁺ *m/z* 397.1370, found 397.1370; HPLC: PHENOMENEX LUX CELLULOSE-2 (PC-2) column, hexane/*i*-PrOH = 70:30, flow rate 0.7 mL/min, 214 nm, major isomer: *t_R* = 8.3 min (major), *t_R* = 6.6 min (minor), *ee* > 99%. (Noteworthy is that some additional peaks (for isomer) were observed in ¹³C NMR spectra due to isomerization)

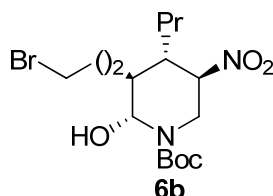


(2R,3S,5R)-3-Ethyl-2-hydroxy-5-nitro-piperidine-1-carboxylic acid benzyl ester 5m. $[\alpha]_D^{29} = +0.71$ (*c* 0.38, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.40-7.33 (m, 5H), 5.87 (s, 1H), 5.16 (s, 2H), 4.43 (tt, *J* = 4.0, 12.0 Hz, 2H), 3.56 (t, *J* = 12.8 Hz, 1H), 2.80-2.60 (br s, 1H), 2.29-2.25 (m, 1H), 2.06 (q, *J* = 12.4 Hz, 1H), 1.58-1.55 (m, 2H), 1.42-1.38 (m, 1H), 0.98 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 155.26, 135.92, 128.84, 128.64, 128.35, 80.25, 75.69, 68.11, 40.92, 40.59, 29.31, 24.74, 11.38; IR (neat): ν = 3418 br, 2963 m, 2924 m, 1701s, 1545 s, 1328 m, 1268 m, 1164 m, 1004 m, 958 m, 901 m, 751 m, 698 m cm⁻¹; ESI-MS *m/z* 331.3 (M + Na)⁺, HRMS

(ESI) calcd for $C_{15}H_{20}N_2O_5Na$ ($M + Na$)⁺ m/z 331.1264, found 331.1270; HPLC: PHENOMENEX LUX AMYLOSE-2 (PA-2) column, hexane/*i*-PrOH = 70:30, flow rate 0.5 mL/min, 214 nm, major isomer: t_R = 10.0 min (major), t_R = 12.0 min (minor), ee = 79%. (Noteworthy is that some additional peaks (for isomer) were observed in ^{13}C NMR spectradue to isomerization)

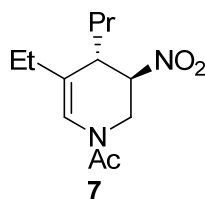


(2R,3S,4S,5R)-3-Ethyl-2-hydroxy-5-nitro-4-propyl-piperidine-1-carboxylic acid *tert*-butyl ester 6a. $[\alpha]_D^{27} = -18.4$ (c 1.88, $CHCl_3$); 1H NMR (400 MHz, $CDCl_3$) δ 5.69 (s, 1H), 4.46 (dt, J = 4.8, 10.8 Hz, 1H), 4.10 (d, J = 10.4 Hz, 1H), 3.70 (t, J = 10.8 Hz, 1H), 3.27 (brs, 1H), 2.44-2.39 (m, 1H), 1.62-1.56 (m, 1H), 1.47 (s, 9H), 1.43-1.22 (m, 6H), 1.01 (t, J = 6.8 Hz, 3H), 0.89 (t, J = 6.8 Hz, 3H); ^{13}C NMR (100 MHz, $CDCl_3$) δ 154.43, 85.69, 81.72, 74.35, 42.75, 41.37, 37.66, 30.26, 28.39, 21.28, 17.19, 14.49, 11.32; IR (neat): ν = 3419 s, 2962 m, 2932 m, 1683 s, 1557 m, 1540 m, 1430 m, 1387 w, 1162 s, 1007 w, 884 w cm^{-1} ; ESI-MS m/z 339.3 ($M + Na$)⁺, HRMS (ESI) calcd for $C_{15}H_{28}N_2O_3Na$ ($M + Na$)⁺ m/z 339.1890, found 339.1891; HPLC: sino-AD column, hexane/*i*-PrOH = 90:10, flow rate 0.7 mL/min, 214 nm, major isomer: t_R = 7.9 min (major), t_R = 6.5 min (minor), ee > 99%. (Noteworthy is that some additional peaks (for isomer) were observed in ^{13}C NMR spectradue to isomerization)



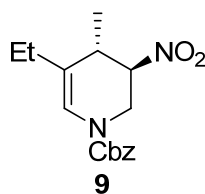
(2R,3S,4S,5R)-3-(3-Bromo-propyl)-2-hydroxy-5-nitro-4-propyl-piperidine-1-carboxylic acid *tert*-butyl ester 6b. $[\alpha]_D^{29} = -27.7$ (c 1.0, $CHCl_3$); 1H NMR (400 MHz, $CDCl_3$) δ 5.61 (s, 1H), 4.47 (dt, J = 4.4, 10.4 Hz, 1H), 4.09 (d, J = 10.0 Hz, 1H), 3.69 (t, J = 12.4 Hz, 1H), 3.46-3.42 (br, 1H), 3.44 (t, J = 6.4 Hz, 2H), 2.47-2.42 (m, 1H), 2.11-2.03 (m, 1H), 1.91-1.83 (m, 1H), 1.72-1.66 (m, 1H), 1.62-1.54 (m, 1H), 1.51-1.46 (m, 1H), 1.46 (s, 9H), 1.37-1.23 (m, 4H), 0.91 (t, J = 6.8 Hz, 3H); ^{13}C NMR

(100 MHz, CDCl₃) δ 154.37, 85.40, 81.94, 74.48, 41.23, 40.50, 37.45, 33.64, 30.21, 29.70, 28.37, 26.93, 17.14, 14.47; IR (neat): ν = 3445 br, 2959 m, 2932 m, 1678 s, 1557 m, 1538 m, 1472 m, 1389 m, 1165 m, 1006 w, 917 w cm⁻¹; ESI-MS m/z 431.4 (M + Na)⁺, HRMS (ESI) calcd for C₁₆H₂₉BrN₂O₅Na (M + Na)⁺ m/z 431.1152, found 431.1156; HPLC: Chiralpak IC column, hexane/*i*-PrOH = 98:2, flow rate 0.7 mL/min, 214 nm, major isomer: t_R = 24.1 min (major), t_R = 19.3 min (minor), *ee* > 99%. (Noteworthy is that some additional peaks (for isomer) were observed in ¹³C NMR spectradue to isomerization)

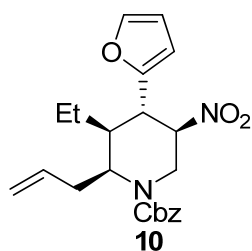


1-((3R,4S)-5-Ethyl-3-nitro-4-propyl-3,4-dihydro-2H-pyridin-1-yl)-ethanone 7.

$[\alpha]_D^{26} = +11.8$ (*c* 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 6.35 & 7.02 (both s, due to rotamers, 1H), 4.76 & 4.47 (ddd, *J* = 1.6, 3.2, 15.2 Hz, & dd, *J* = 3.2, 13.2 Hz, due to rotamers, 1H), 4.67 & 4.59 (q, *J* = 3.2 Hz & q, *J* = 2.8 Hz, due to rotamers, 1H), 3.51 & 3.67 (dd, *J* = 3.2, 14.8 Hz & dd, *J* = 3.2, 13.6 Hz, due to rotamers, 1H), 2.96 & 3.02 (t, *J* = 4.4 Hz & t, *J* = 4.4 Hz, due to rotamers, 1H), 2.19 & 2.23 (both s, due to rotamers, 3H), 2.09 (q, *J* = 7.6 Hz, 2H), 1.69-1.60 (m, 2H), 1.46-1.35 (m, 2H), 1.04 (t, *J* = 7.6 Hz, 3H), 0.98 (t, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 168.32 & 167.59 (due to rotamers), 121.27 & 121.70 (due to rotamers), 119.86 & 118.29 (due to rotamers), 80.83 & 80.89 (due to rotamers), 38.85 & 42.26 (due to rotamers), 38.44 & 37.58 (due to rotamers), 34.74 & 34.93 (due to rotamers), 26.03 & 25.81 (due to rotamers), 21.60 & 22.00 (due to rotamers), 19.98 & 20.04 (due to rotamers), 14.05, 12.57 & 12.55 (due to rotamers); IR (neat): ν = 3418 m, 2964 s, 2934 s, 2875 m, 1652 s, 1549 s, 1410 m, 1316 m, 1210 m, 1179 m, 1017 m, 740 m, 611 m cm⁻¹; ESI-MS m/z 241.2 (M + H)⁺, HRMS (ESI) calcd for C₁₂H₂₀N₂O₃Na (M + Na)⁺ m/z 263.1366, found 263.1372; HPLC: PHENOMENEX LUX AMYLOSE-2 (PA-2) column, hexane/*i*-PrOH = 70:30, flow rate 0.7 mL/min, 214 nm, t_R = 15.2 min (major), t_R = 21.8 min (minor), *ee* = 93%.



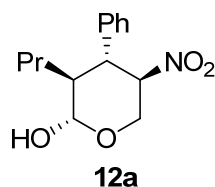
(3R,4S)-5-Ethyl-4-methyl-3-nitro-3,4-dihydro-2H-pyridine-1-carboxylic acid benzyl ester 9. To a solution of **4i** (32 mg, 0.10 mol) in 2 mL CH₂Cl₂ was added several drops of TFA at rt. The reaction mixture was stirred at this temperature until **4i** was consumed as monitored by TLC. Direct concentration of the reaction mixture gave **9** (30 mg, 100%), which is pure enough for analysis. $[\alpha]_D^{23} = +26.5$ (*c* 0.83, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.38-7.33 (m, 5H), 6.69 & 6.58 (both s, due to rotamers, 1H), 5.26-5.16 (m, 2H), 4.50-4.39 (m, 2H), 3.81-3.76 (m, 1H), 3.04 (s, 1H), 2.10-2.02 (m, 2H), 1.17 (d, *J* = 7.2 Hz, 3H), 1.04 & 1.02 (both t, *J* = 7.6 Hz, due to rotamers, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 153.00 & 152.95 (due to rotamers), 136.05, 128.74, 128.47, 128.26, 120.93 & 120.51, 119.00 & 118.65 (due to rotamers), 83.82 & 83.86 (due to rotamers), 68.07 & 68.12 (due to rotamers), 40.98 & 40.88 (due to rotamers), 33.09 & 33.27 (due to rotamers), 25.37, 18.31 & 18.23 (due to rotamers), 12.54 & 12.67 (due to rotamers); IR (neat): ν = 2918 m, 1708 s, 1546 m, 1215 m, 1139 m, 1029 m, 977 m, 936 m, 692 m, 600 m cm⁻¹; ESI-MS *m/z* 327.2 (M + Na)⁺, HRMS (ESI) calcd for C₁₆H₂₀N₂O₄Na (M + Na)⁺ *m/z* 327.1315, found 327.1322.



(2S,3S,4S,5R)-2-Allyl-3-ethyl-4-furan-2-yl-5-nitro-piperidine-1-carboxylic acid benzyl ester 10. To a solution of **4i** (37 mg, 0.10 mmol) and allyl trimethylsilane (60 μ L, 0.5 mmol) in CH₂Cl₂ (2 ml) was added BF₃·OEt₂ (25 μ L, 0.2 mmol) at -78 °C. The reaction mixture was stirred at this temperature until **4i** was consumed as monitored by TLC. Purification by flash column chromatography (silica gel,

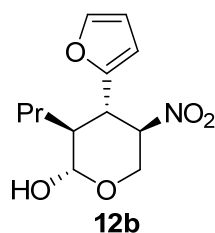
petroleum ether/AcOEt 20:1 to 15:1) afforded 37 mg (93%) of **10**. $[\alpha]_{\text{D}}^{23} = +31.6$ (*c* 0.93, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.39-7.32 (m, 6H), 6.29-6.28 (m, 1H), 6.17 (d, *J* = 2.8 Hz, 1H), 5.82-5.72 (m, 1H), 5.17-5.09 (m, 2H), 5.05 (d, *J* = 3.6 Hz, 1H), 5.02 (s, 1H), 4.91-4.88 (m, 1H), 4.60 (d, *J* = 15.6 Hz, 1H), 4.15 (m, 1H), 3.55 (dd, *J* = 6.4, 15.6 Hz, 1H), 3.47 (dd, *J* = 9.2, 11.6 Hz, 1H), 2.46-2.42 (m, 1H), 2.29-2.21 (m, 1H), 1.86-1.79 (m, 1H), 1.44-1.35 (m, 1H), 1.27-1.21 (m, 1H), 0.72 (t, *J* = 7.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 155.74, 151.28, 142.57, 136.34, 133.86, 128.64, 128.27, 128.19, 118.40, 110.64, 109.03, 86.06, 67.82, 54.96, 42.19, 39.77, 39.69, 36.95, 23.53, 9.60; IR (neat): ν = 2964 m, 2933 m, 1703 s, 1552 s, 1424 s, 1243 s, 1114 m, 1011 m, 925 m, 737 s, 598 m cm⁻¹; ESI-MS *m/z* 399.2 (M + H)⁺, 421.3 (M + Na)⁺, HRMS (ESI) calcd for C₂₂H₂₆N₂O₅Na (M + Na)⁺ *m/z* 421.1734, found 421.1738.

General procedure for cascade Michael-acetalization process of aldehydes with hydroxymethyl substituted nitroolefins: Valeraldehyde (0.40 mmol) was added to a suspension of catalyst **1** (20 mol %), hydroxymethyl substituted nitroolefins **11** (0.20 mmol) and benzoic acid (60 mol %) in water (0.40 mL) at room temperature, and stirred until complete conversion of the starting materials (monitored by TLC). The reaction mixture was diluted with ethyl acetate, dried over Na₂SO₄ and concentrated. Purification by flash column chromatography afforded **12**. The enantiomeric excess (ee) was determined by HPLC on a chiral phase.



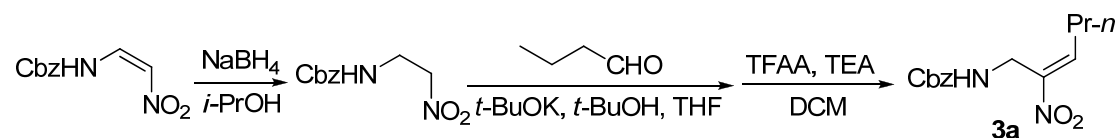
(2S,3S,4R,5R)-5-Nitro-4-phenyl-3-propyl-tetrahydro-pyran-2-ol 12a. $[\alpha]_{\text{D}}^{28} = -8.43$ (*c* 0.42, MeOH); ¹H NMR (400 MHz, CDCl₃) δ 7.34-7.24 (m, 3H), 7.20-7.18 (m, 2H), 5.31 (s, 1H), 4.87 (dq, *J* = 4.8, 11.2 Hz, 1H), 4.43 (t, *J* = 10.8 Hz, 1H), 4.04 (dd, *J* = 4.8, 10.4 Hz, 1H), 3.52 (t, *J* = 11.6 Hz, 1H), 2.68 (brs, 1H), 2.03-1.98 (m, 1H), 1.36-1.27 (m, 2H), 1.11-1.04 (m, 1H), 0.97-0.90 (m, 1H), 0.75 (t, *J* = 7.2 Hz, 3H); ¹H

NMR (400 MHz, MeOD) δ 7.35-7.32 (m, 2H), 7.28-7.25 (m, 3H), 5.23 (d, $J = 2.8$ Hz, 1H), 5.03 (dt, $J = 5.2, 11.2$ Hz, 1H), 4.40 (t, $J = 10.4$ Hz, 1H), 4.00 (dd, $J = 4.8, 10.4$ Hz, 1H), 3.48 (t, $J = 12.0$ Hz, 1H), 2.06-2.01 (m, 1H), 1.38-1.25 (m, 2H), 1.11-1.03 (m, 1H), 0.93-0.84 (m, 1H), 0.74 (t, $J = 7.6$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 137.55, 129.39, 129.07, 128.10, 127.97, 92.20, 87.76, 60.36, 45.35, 43.35, 30.90, 19.71, 14.13; ^{13}C NMR (100 MHz, MeOD) δ 139.59, 129.76, 128.63, 92.84, 89.11, 61.14, 47.00, 44.81, 32.29, 20.60, 14.43; IR (neat): $\nu = 3419$ br, 2950 m, 1542 s, 1455m, 1352 m, 1059 m, 1033 m, 930 m, 692 m cm^{-1} ; EI-MS m/z 265 M^+ , HRMS (EI) calcd for $\text{C}_{14}\text{H}_{19}\text{NO}_4$ M^+ m/z 265.1314, found 265.1319; HPLC: IC column, hexane/*i*-PrOH = 90:10, flow rate 0.7 mL/min, 214 nm, $t_R = 8.3$ min (major), $t_R = 7.5$ min (minor), $ee > 99\%$. (Noteworthy is that some additional peaks (for isomer) were observed in ^{13}C NMR spectra due to isomerization)



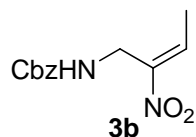
(2S,3S,4S,5R)-4-Furan-2-yl-5-nitro-3-propyl-tetrahydro-pyran-2-ol 12b. $[\alpha]_D^{28} = -0.01$ (c 1.96, MeOH); ^1H NMR (400 MHz, CDCl_3) δ 7.37-7.36 (m, 1H), 6.30-6.26 (m, 1H), 6.18-6.16 (m, 1H), 5.27 (d, $J = 2.8$ Hz, 1H), 4.94 (dt, $J = 4.8, 11.2$ Hz, 1H), 4.37 (t, $J = 10.8$ Hz, 1H), 4.02 (dd, $J = 4.8, 10.4$ Hz, 2H), 3.65 (t, $J = 11.6$ Hz, 1H), 2.13-2.07 (m, 1H), 1.40-1.25 (m, 2H), 1.14-0.97 (m, 2H), 0.81 (t, $J = 6.8$ Hz, 3H); ^1H NMR (400 MHz, MeOD) δ 7.44-7.42 (m, 1H), 6.32-6.30 (m, 1H), 6.19-6.18 (m, 1H), 5.18 (d, $J = 2.8$ Hz, 1H), 4.96 (dt, $J = 5.2, 11.2$ Hz, 1H), 4.34 (t, $J = 10.8$ Hz, 1H), 3.97 (dd, $J = 4.8, 10.4$ Hz, 1H), 3.60 (t, $J = 11.2$ Hz, 1H), 2.07-2.01 (m, 1H), 1.39-1.28 (m, 2H), 1.10-1.03 (m, 1H), 1.01-0.93 (m, 1H), 0.71 (t, $J = 6.8$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) 150.55, 142.51, 110.41, 109.35, 92.04, 85.20, 59.97, 41.74, 39.36, 31.30, 19.68, 14.20; ^{13}C NMR (100 MHz, MeOD) δ 152.66, 143.45, 111.20, 109.63, 92.65, 86.71, 60.62, 43.52, 40.82, 32.58, 20.54, 14.48; IR (neat): $\nu = 3396$ br, 2954 m, 2873 w, 1541 s, 1466 w, 1357 w, 1277 w, 1152 m, 1032 w, 1009 m, 928 m, 908 w, 726 m cm^{-1} ; EI-MS m/z 255 M^+ , HRMS (EI) calcd for $\text{C}_{12}\text{H}_{17}\text{NO}_5$ M^+ m/z

255.1107, found 255.1113; HPLC: AD-H column, hexane/*i*-PrOH = 95:5, flow rate 0.7 mL/min, 214 nm, major isomer: t_R = 28.0 min (major), t_R = 48.5 min (minor), *ee* > 99%. (Noteworthy is that some additional peaks (for isomer) were observed in ^{13}C NMR spectra due to isomerization)

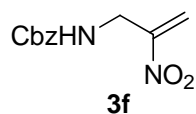


Typical procedure for assembly of 1-protected aminomethyl 2-alkyl substituted nitroalkenes **3a.** To a solution of ((*Z*)-2-nitrovinyl)-carbamic acid benzyl ester (2.24 g, 10 mmol) in 50 mL of *i*-PrOH were added NaBH_4 (760 mg, 2.0 eq, 20 mmol) at 0 °C. The resultant solution was warmed to rt. After stirring at rt for 2 h, 1 mL acetone and 20 mL saturate NH_4Cl aqueous solution was added slowly, the aqueous phase was extracted with ethyl acetate. The combined organic layers were washed with brine and dried over Na_2SO_4 and concentrated to give reduced product as light yellow solid, which was dissolved in 10 mL *t*-BuOH and 10 mL THF. To this solution was added *n*-butylaldehyde (15 mmol, 1.5 equiv.) and *t*-BuOK (112 mg, 1 mmol, 0.1 equiv.) at 0 °C. The resultant solution was warmed to rt. After stirring at rt for 5 h, the reaction mixture was diluted with ethyl ether and washed with water, the aqueous phase was extracted with ethyl ether three times. The combined organic phase was washed with brine and dried over Na_2SO_4 and concentrated to give condensation product, which was dissolved in 20 mL CH_2Cl_2 . To this solution was added TEA (20 mmol, 2 equiv.) and TFAA (15 mmol) slowly at -20 °C. After stirring at -20 °C for 1.5 h, the reaction mixture was diluted with CH_2Cl_2 and washed with water. The aqueous phase was extracted with CH_2Cl_2 three times. The combined organic phase was washed with brine and dried over Na_2SO_4 and concentrated. Purification of the residue by flash column chromatography (silica gel, 10:1 petroleum ether/ethyl acetate as eluent) afforded the corresponding (*E*)-2-nitro-hex-2-enyl carbamic acid benzyl ester **3a** (2.06 g, 74% for 3 steps). ^1H NMR (400 MHz, CDCl_3) δ 7.33-7.24 (m, 5H), 5.47 (brs, 1H), 5.08 (s, 2H), 4.28 (d, J = 6.4 Hz, 2H), 2.50-2.45 (m, 2H), 1.58-1.53 (m, 2H), 0.98 (t, J = 7.2 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 156.17, 148.20, 140.87,

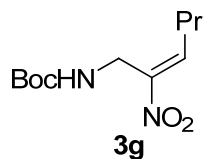
136.25, 128.64, 128.32, 128.15, 67.11, 36.36, 29.96, 21.89, 13.79; IR (neat): ν = 3334 br, 2962 m, 1714 s, 1520 s, 1332 m, 1238 s, 1136 s, 738 m, 698 m cm^{-1} ; ESI-MS m/z 279.2 ($\text{M} + \text{H}$)⁺, 301.2 ($\text{M} + \text{Na}$)⁺, HRMS (ESI) calcd for $\text{C}_{14}\text{H}_{18}\text{N}_2\text{O}_4\text{Na}$ ($\text{M} + \text{Na}$)⁺ m/z 301.1159, found 301.1164.



((E)-2-Nitrobut-2-enyl)carbamic acid benzyl ester 3b. 49% yield for three steps. ¹H NMR (400 MHz, CDCl_3) δ 7.28-7.24 (m, 6H), 5.45 (brs, 1H), 4.99 (s, 2H), 4.18 (d, J = 6.4 Hz, 2H), 2.01 (d, J = 7.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl_3) δ 156.27, 148.85, 136.34, 136.15, 128.58, 128.26, 128.08, 67.06, 36.01, 13.74; IR (neat): ν = 3333 br, 2956 m, 1714 s, 1515 s, 1341 m, 1241 s, 1137 m, 1080 m, 975 m, 846 m, 777 m, 698 m cm^{-1} ; ESI-MS m/z 273.3 ($\text{M} + \text{Na}$)⁺, HRMS (ESI) calcd for $\text{C}_{12}\text{H}_{14}\text{N}_2\text{O}_4\text{Na}$ ($\text{M} + \text{Na}$)⁺ m/z 273.0846, found 273.0846.

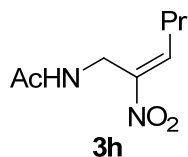


(2-Nitroallyl)carbamic acid benzyl ester 3f. 40% for three steps. ¹H NMR (400 MHz, CDCl_3) δ 7.38-7.32 (m, 5H), 6.58 (s, 1H), 5.92 (s, 1H), 5.38 (brs, 1H), 5.11 (s, 2H), 4.27 (d, J = 6.4 Hz, 2H); ¹³C NMR (100 MHz, CDCl_3) δ 156.22, 153.90, 136.10, 128.76, 128.52, 128.34, 120.80, 67.41, 40.50; ESI-MS m/z 236.1 ($\text{M} + \text{H}$)⁺, 259.1 ($\text{M} + \text{Na}$)⁺.

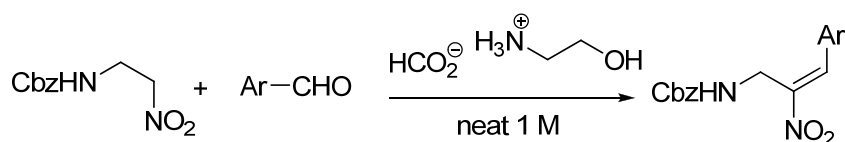


((E)-2-Nitrohex-2-enyl)carbamic acid *tert*-butyl ester 3g. Compound **3g** was prepared from ((*Z*)-2-nitrovinyl)carbamic acid *tert*-butyl ester and *n*-butylaldehyde in 59% overall yield according to typical procedure. ¹H NMR (400 MHz, CDCl_3) δ 7.24 (t, J = 8.0 Hz, 1H), 5.19 (brs, 1H), 4.23 (d, J = 6.0 Hz, 2H), 2.48 (q, J = 7.6 Hz, 2H), 1.61-1.52 (m, 2H), 1.42 (s, 9H), 0.98 (t, J = 7.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl_3) δ 155.50, 148.78, 140.47, 79.99, 35.88, 29.93, 28.40, 21.92, 13.72; IR (neat):

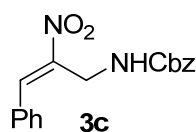
$\nu = 3419$ br, 2966 s, 2935 m, 1716 s, 1526 s, 1367 m, 1332 m, 1249 m, 1167 s, 932 w, 857 w cm^{-1} ; ESI-MS m/z 267.3 ($M + \text{Na}$)⁺, HRMS (ESI) calcd for $\text{C}_{11}\text{H}_{20}\text{N}_2\text{O}_4\text{Na}$ ($M + \text{Na}$)⁺ m/z 267.1315, found 267.1319.



***N*-((*E*)-2-Nitrohex-2-enyl)acetamide **3h**.** Compound **3h** was prepared from *N*-((*Z*)-2-nitrovinyl)acetamide and *n*-butylaldehyde in 50% overall yield according to typical procedure. ¹H NMR (400 MHz, CDCl_3) δ 7.26 (t, $J = 8.0$ Hz, 1H), 6.30 (br s, 1H), 4.35 (d, $J = 6.4$ Hz, 2H), 2.50 (q, $J = 7.6$ Hz, 2H), 1.98 (s, 3H), 1.61-1.52 (m, 2H), 0.99 (t, $J = 7.6$ Hz, 3H); ¹³C NMR (100 MHz, CDCl_3) δ 170.18, 148.03, 140.95, 34.65, 29.93, 22.93, 21.80, 13.73; IR (neat): $\nu = 3255$ s, 2962 m, 1668 m, 1644 m, 1516 s, 1338 m, 1288 m, 1095 w, 1075 w, 748 w cm^{-1} ; ESI-MS m/z 187.1 ($M + \text{H}$)⁺, 219.2 ($M + \text{Na}$)⁺, HRMS (MALDI) calcd for $\text{C}_8\text{H}_{14}\text{N}_2\text{O}_3\text{Na}$ ($M + \text{Na}$)⁺ m/z 209.0897, found 209.0897.

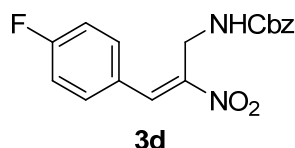


Typical procedure for assembly of 1-protected aminomethyl 2-aryl substituted nitroalkenes. A mixture of (2-nitroethyl)carbamate benzyl ester (1 mmol) and arylaldehyde (1 mmol) in 1 mL 2-HEAF was stirred overnight at rt until the starting material disappeared. The mixture was partitioned between water and ethyl acetate. The aqueous phase was extracted with ethyl acetate. The combined organic phase was washed with brine, dried over Na_2SO_4 and concentrated. Purification of the residue by flash column chromatography (silica gel, 15:1 to 10:1 petroleum ether/ethyl acetate as eluent) afforded the corresponding nitroalkene.

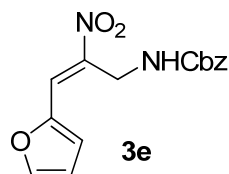


((*E*)-2-Nitro-3-phenylallyl)carbamate benzyl ester **3c.** 85% yield. ¹H NMR

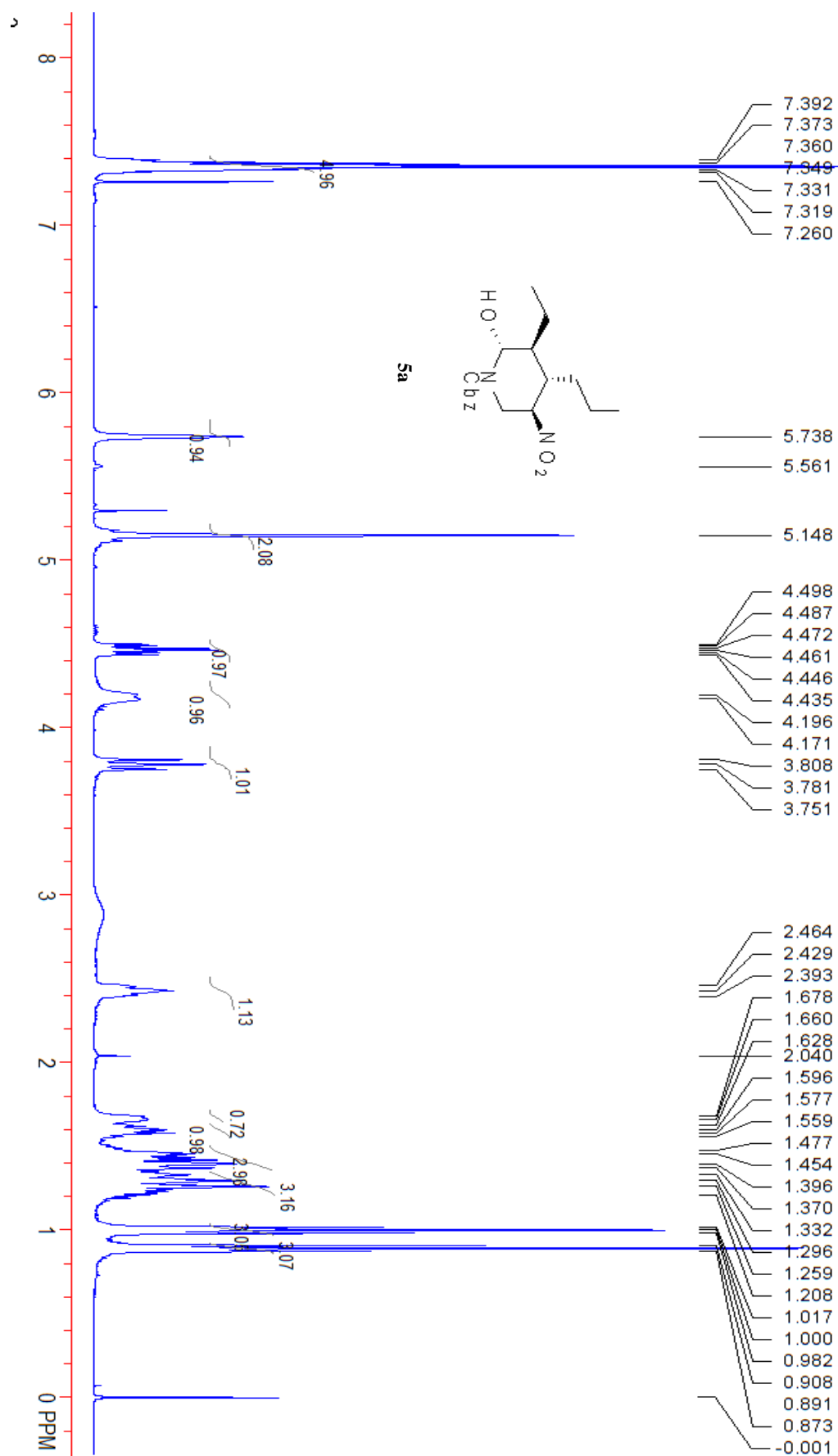
(400 MHz, CDCl₃) δ 8.17 (s, 1H), 7.72-7.66 (m, 2H), 7.52-7.45 (m, 3H), 7.40-7.30 (m, 5H), 5.50 (brs, 1H), 5.12 (s, 2H), 4.58 (d, J = 6.0 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 156.20, 147.27, 137.21, 136.20, 131.10, 131.00, 130.48, 129.34, 128.70, 128.40, 128.24, 67.29, 37.79; ESI-MS m/z 335.3 (M + Na)⁺, HRMS (MALDI) calcd for C₁₇H₁₆N₂O₄Na (M + Na)⁺ m/z 335.1002, found 335.1004.

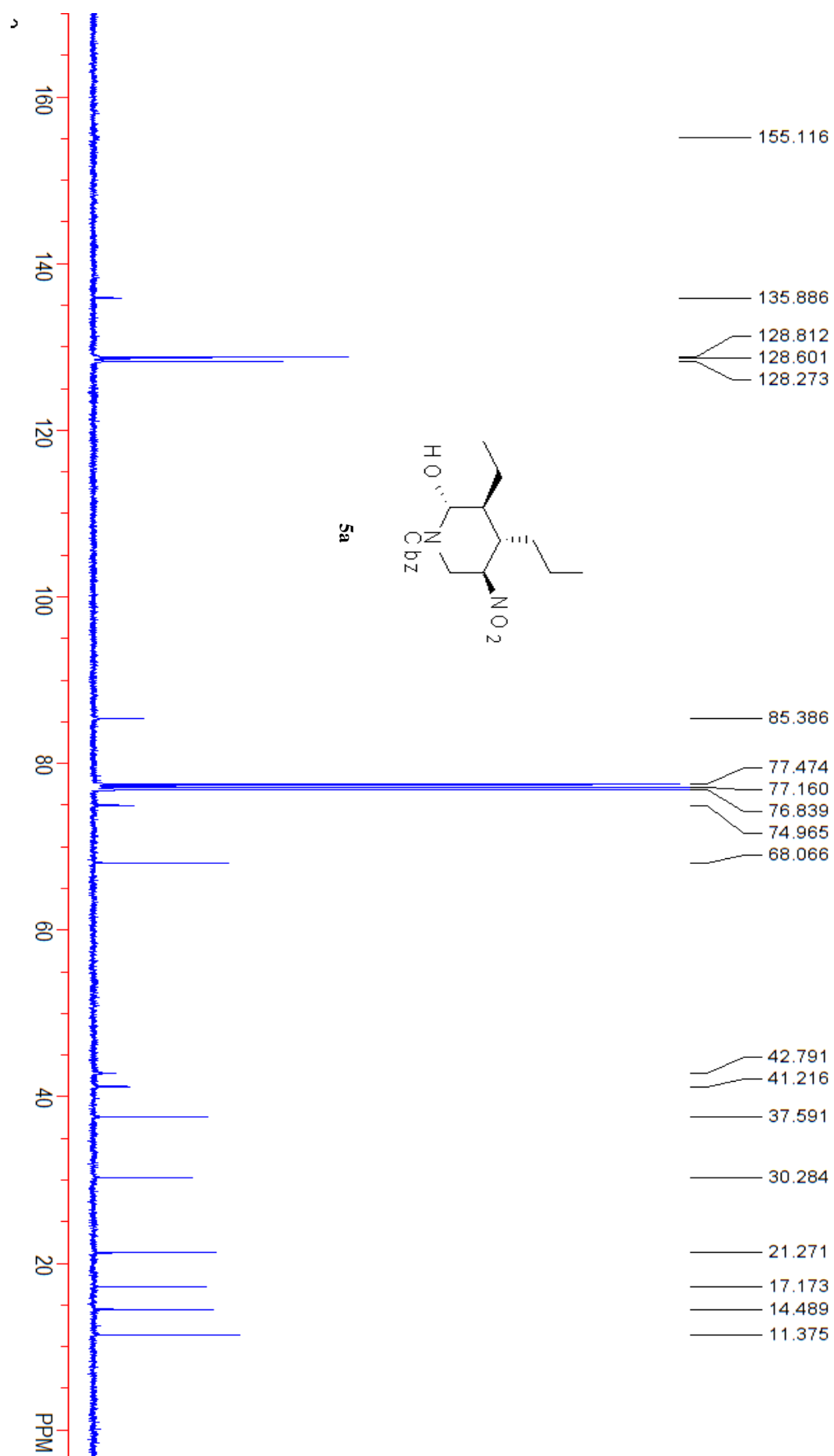


((E)-3-(4-Fluorophenyl)-2-nitroallyl)carbamic acid phenethyl ester 3d. 83% yield, ¹H NMR (400 MHz, CDCl₃) δ 8.12 (s, 1H), 7.78-7.70 (m, 2H), 7.38-7.30 (m, 5H), 7.19-7.15 (m, 2H), 5.55 (brs, 1H), 5.12 (s, 2H), 5.54 (d, J = 6.0 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 164.21 (d, J = 252.4 Hz), 156.20, 146.96, 136.06, 135.92, 132.78 (d, J = 8.0 Hz), 128.62, 128.24 (d, J = 20.0 Hz), 127.03 (d, J = 3.7 Hz), 116.55 (d, J = 21.8 Hz), 67.25, 37.64; IR (neat): ν = 3249 w, 3144 w, 1728 s, 1600 m, 1516 m, 1407 m, 1347 m, 1242 m, 1033 m, 858 w, 755 w, 696 w cm⁻¹; ESI-MS m/z 353.3 (M + Na)⁺, HRMS (ESI) calcd for C₁₇H₁₅FN₂O₄Na (M + Na)⁺ m/z 353.0908, found 353.0907.



((E)-3-Furan-2-yl-2-nitroallyl)carbamic acid benzyl ester 3e. 86% yield. ¹H NMR (400 MHz, CDCl₃) δ 7.91 (s, 1H), 7.73 (s, 1H), 7.40-7.27 (m, 5H), 7.13 (s, 1H), 6.62 (s, 1H), 5.44 (brs, 1H), 5.09 (s, 2H), 4.83 (d, J = 6.4 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 156.22, 147.75, 146.82, 143.49, 136.24, 128.59, 128.26, 128.16, 123.09, 121.88, 113.43, 67.08, 37.63; IR (neat): ν = 3318 s, 2922 s, 2851 m, 1686 s, 1498 m, 1311 m, 1269 m, 981 w, 770 m, 699 w cm⁻¹; ESI-MS m/z 325.3 (M + Na)⁺, HRMS (ESI) calcd for C₁₅H₁₄N₂O₅Na(M + Na)⁺ m/z 325.0795, found 325.0799.





HPLC REPORT

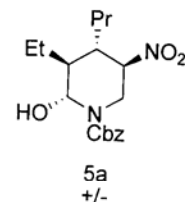
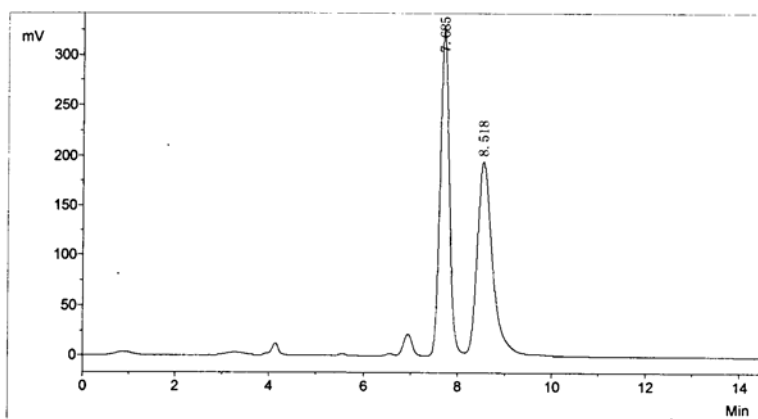
Data File:ZSL-17-41+- PA-2 73 214 0.7. che

Sample name:

Date:2010-06-01

Time :09:21

Operator:



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		7.685	324496.4	4449265.3	50.8309
2	2		8.518	192347.9	4303805.4	49.1691
Total				516844.3	8753070.6	100.0000

HPLC REPORT

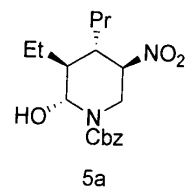
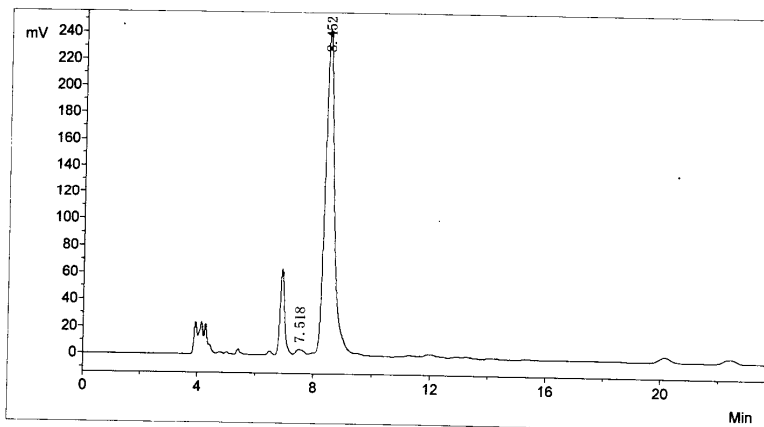
Data File: ZSL-17-40. che

Sample name:

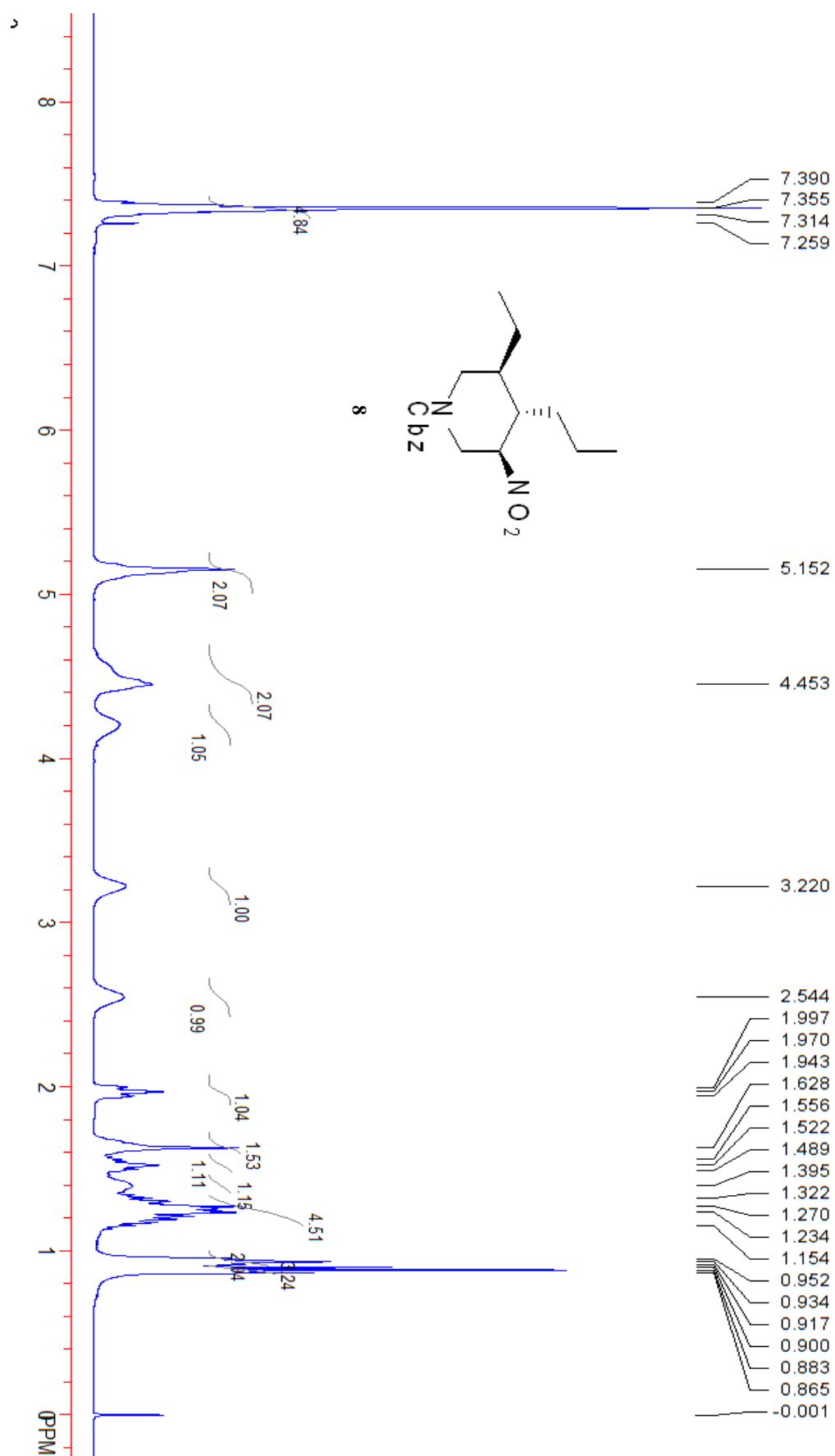
Date: 2010-06-01

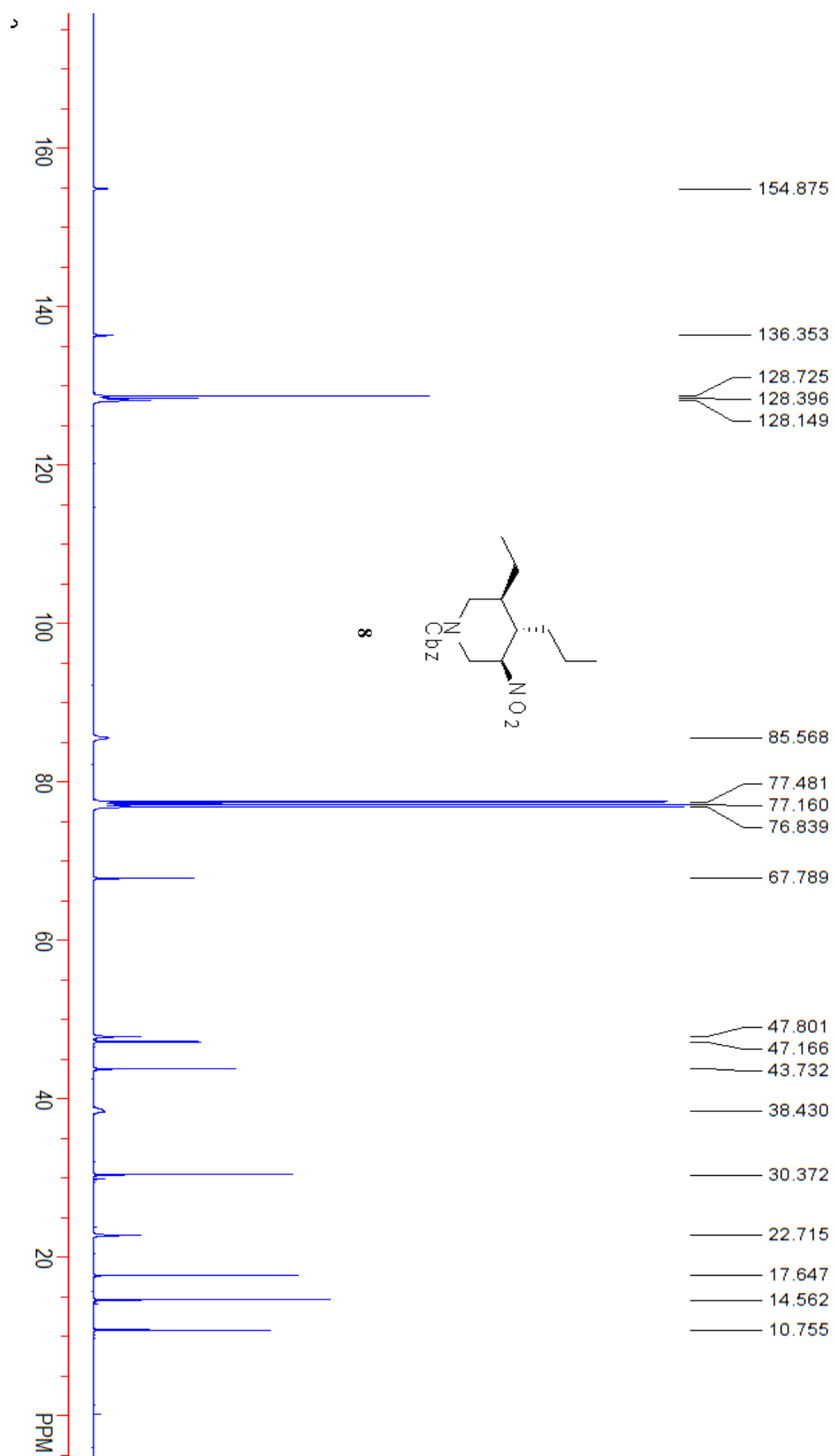
Time : 09:46

Operator:



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		7.518	3392.2	55822.7	1.0677
2	2		8.452	241699.8	5172392.5	98.9323





HPLC Report

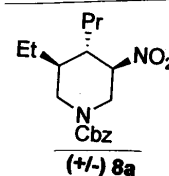
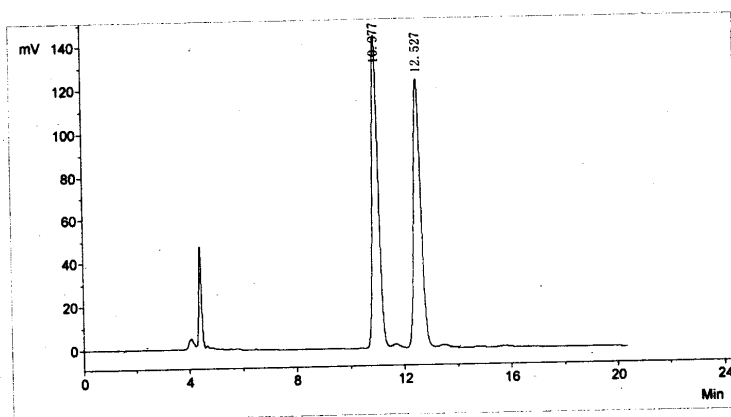
Sample Name:

Data File: ZSL6-25+- PA-2730.7214.che

Operator:

Date: 2010-07-07

Time: 13:28



No.	PeakNo	R.Time	PeakHeight	PeakArea	PerCent
1	1	10.977	144556.2	2412167.5	50.9185
2	2	12.527	123530.9	2325147.1	49.0815
Total			268087.1	4737314.6	100.0000

HPLC Report

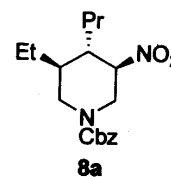
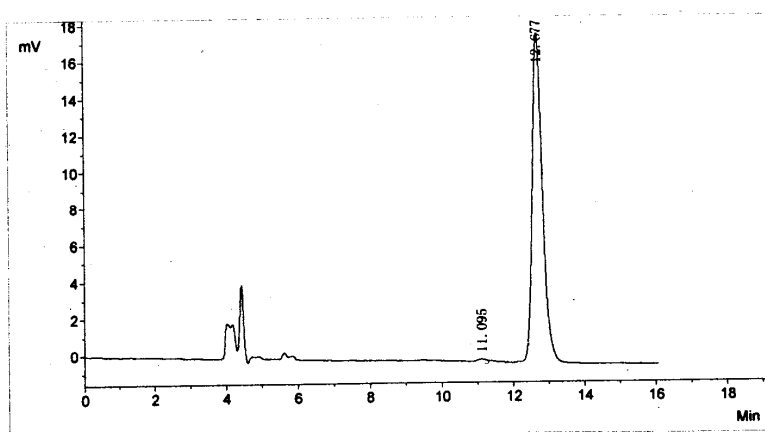
Sample Name:

Data File: ZSL6-24-1.che

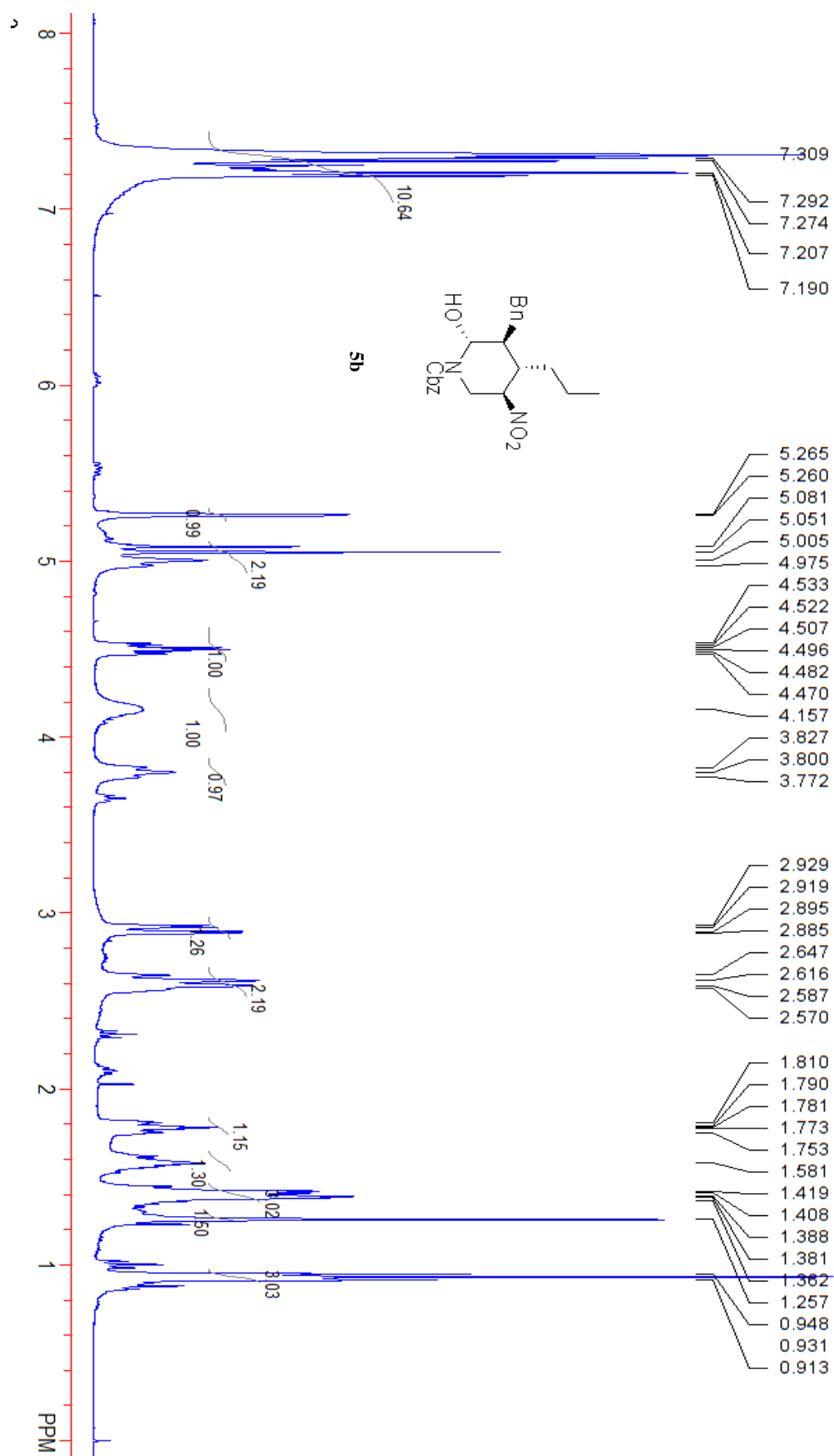
Operator:

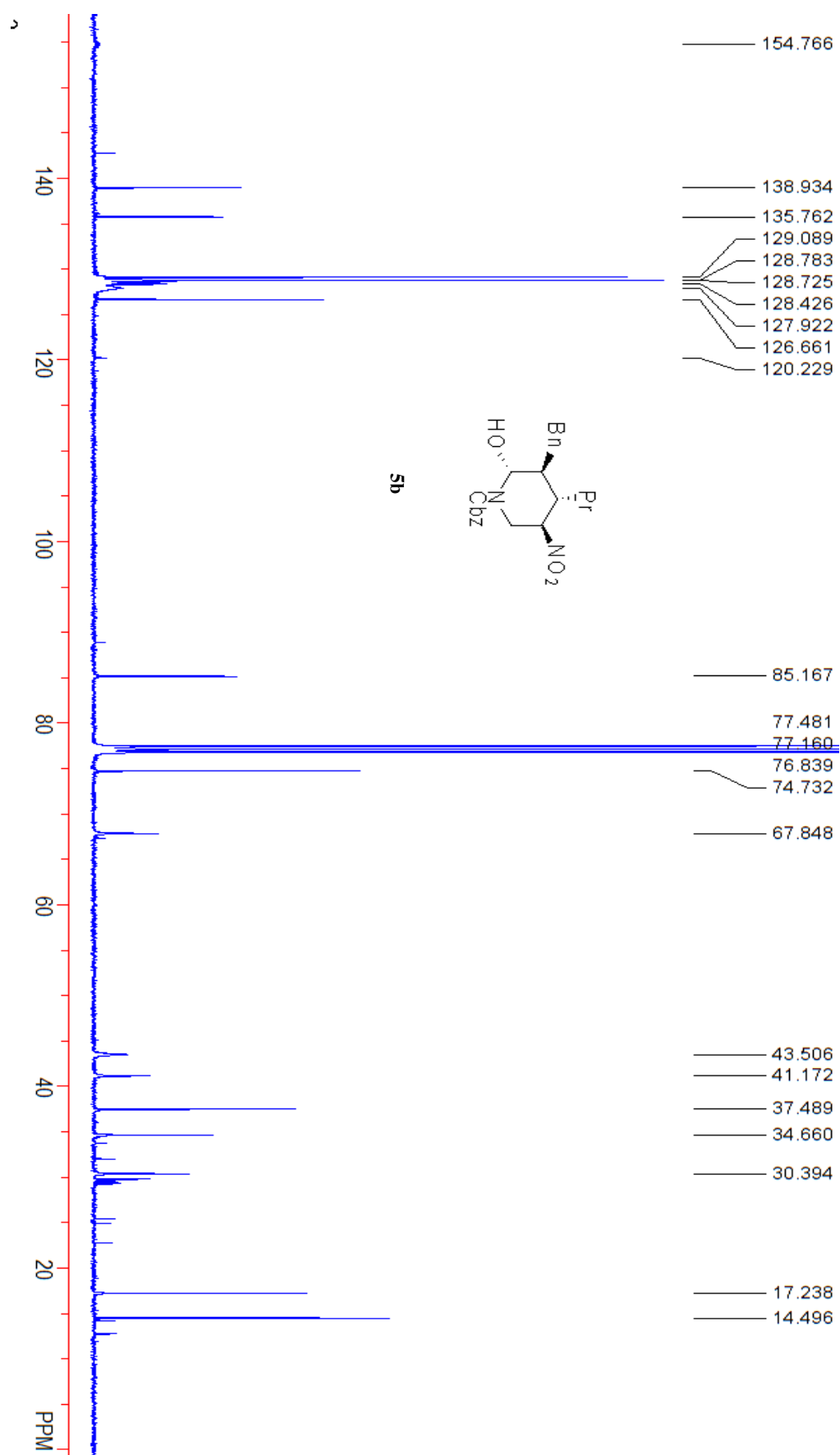
Date: 2010-07-07

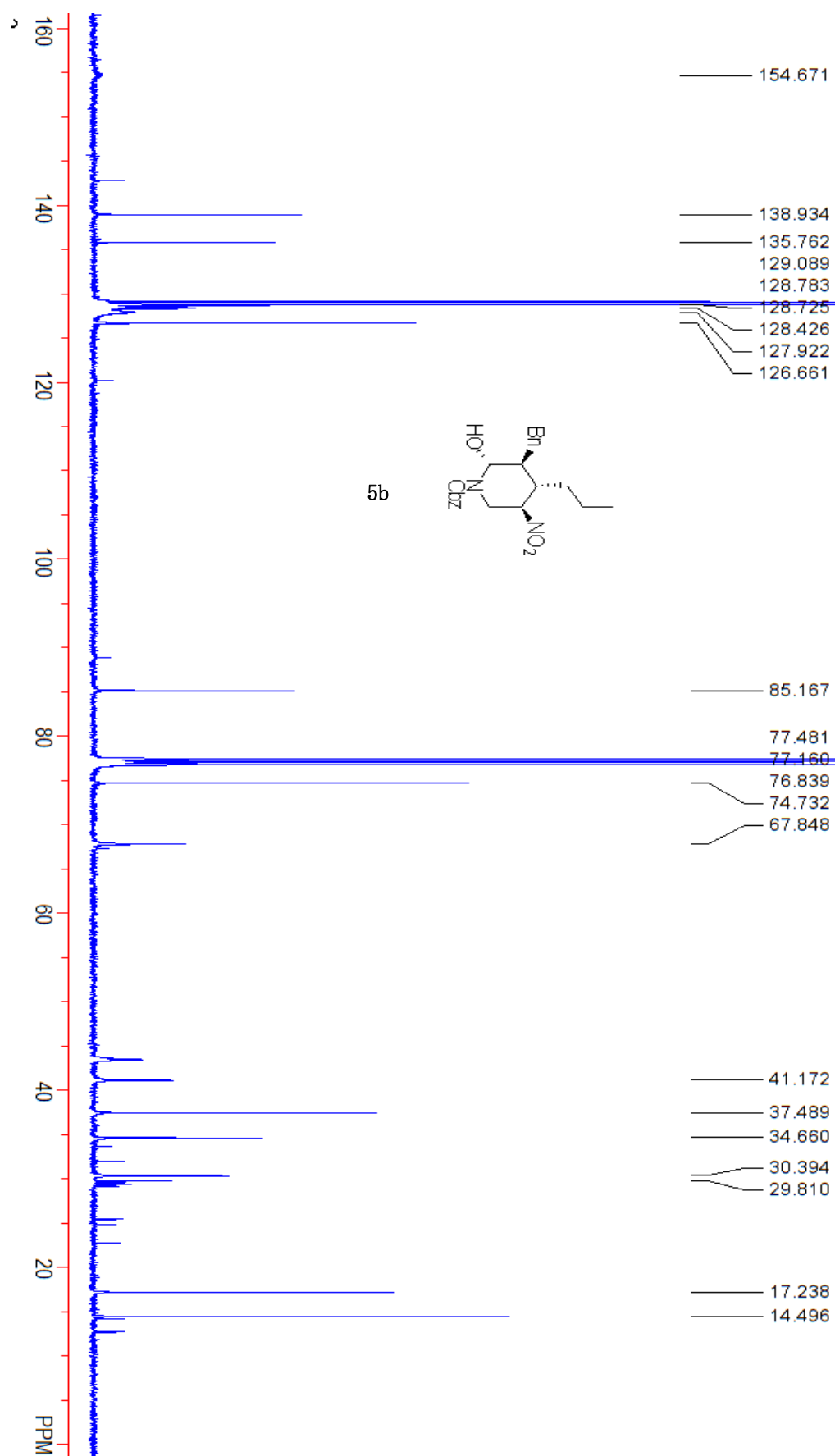
Time: 14:15

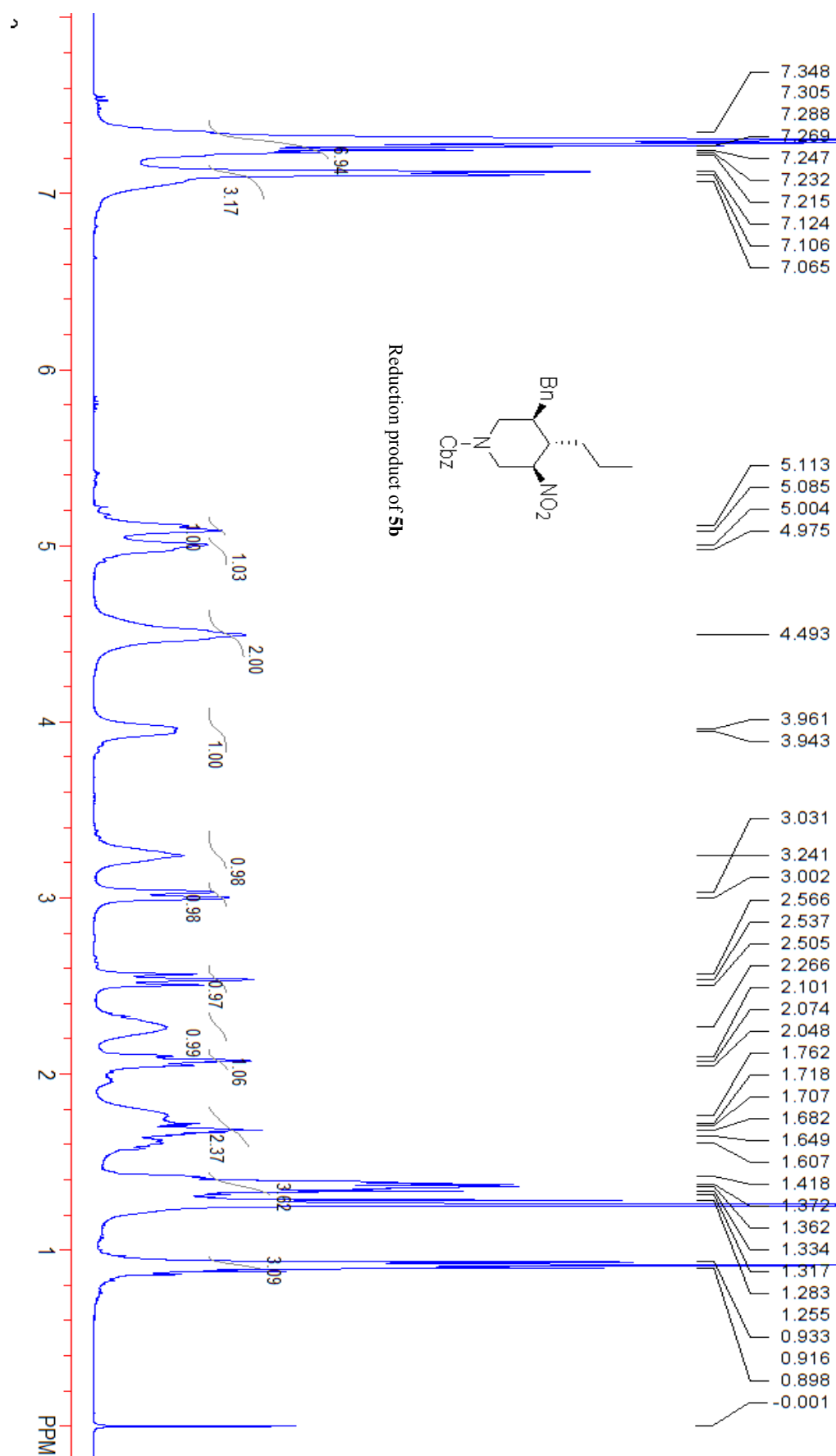


No.	PeakNo	R.Time	PeakHeight	PeakArea	PerCent
1	1	11.095	136.6	2559.5	0.7148
2	2	12.677	17767.1	355502.8	99.2852
Total			17903.7	358062.3	100.0000









HPLC Report

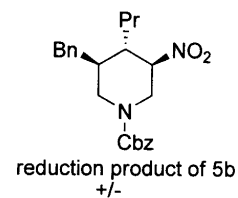
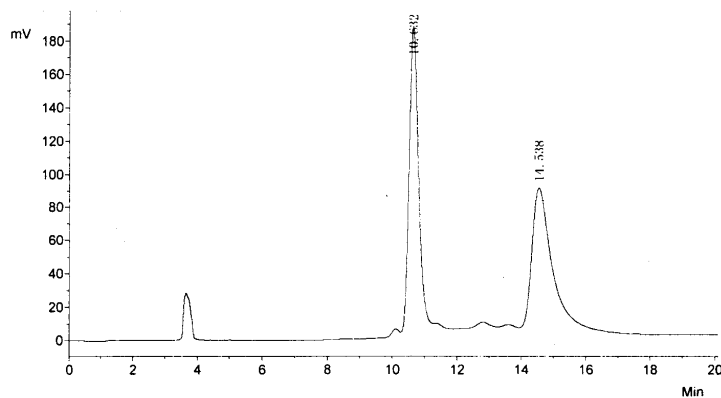
Sample Name:

Data File: ZSL17-29-2+-PC-2-730.7214.che

Operator:

Date: 2010-09-07

Time: 13:51



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	10.632	181398.8	3576778.5	49.7194
2	2	14.538	74437.0	3617148.2	50.2806
Total			255835.8	7193926.7	100.0000

HPLC Report

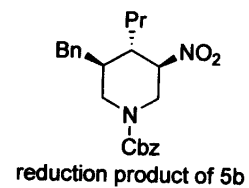
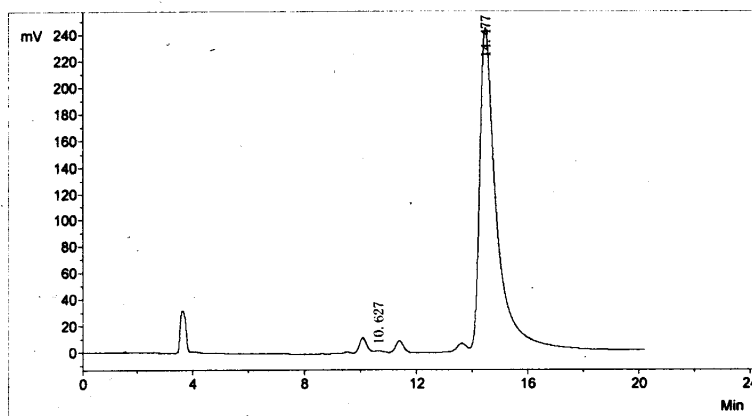
Sample Name:

Data File: ZSL6-33-2.che

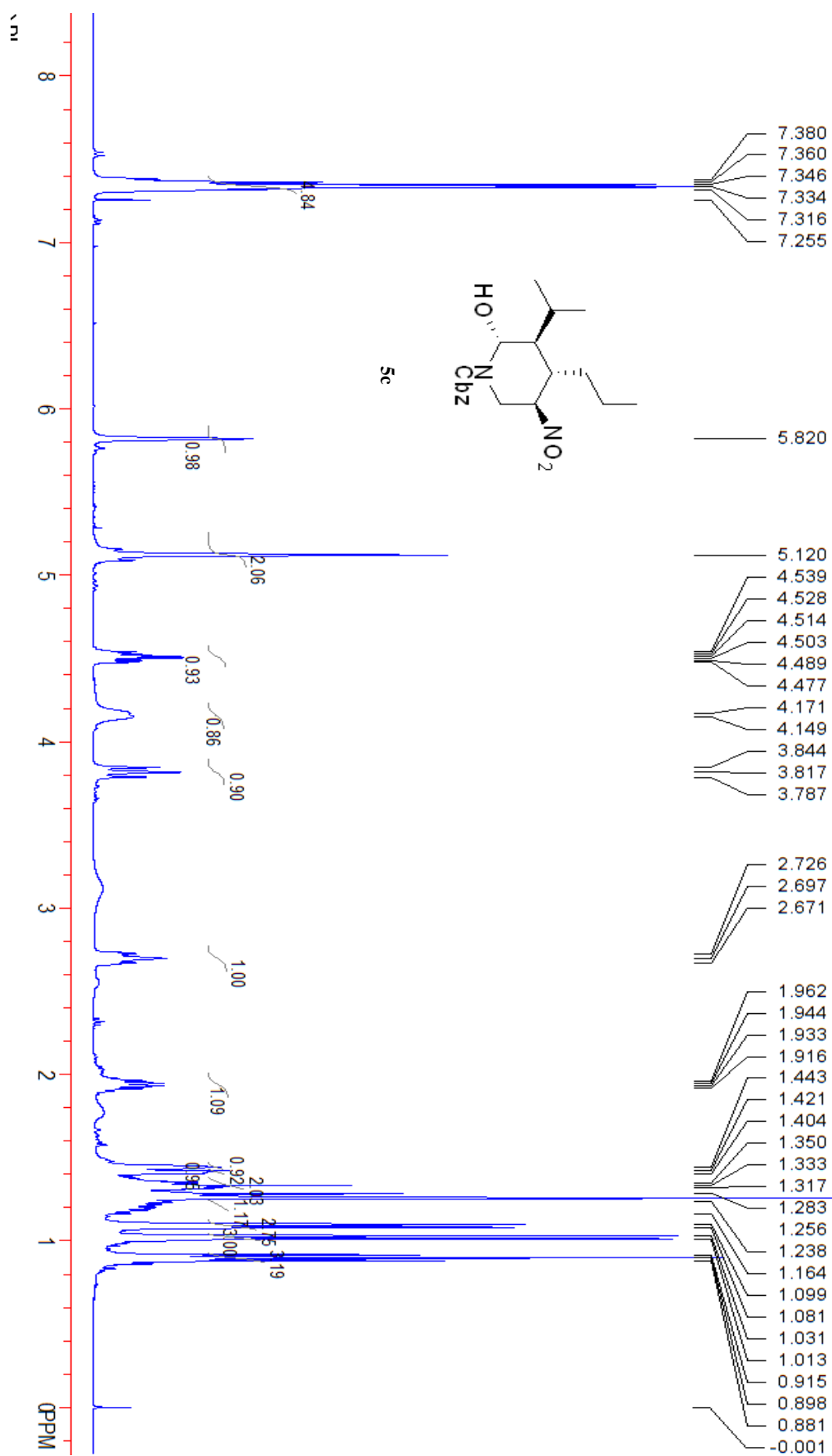
Operator:

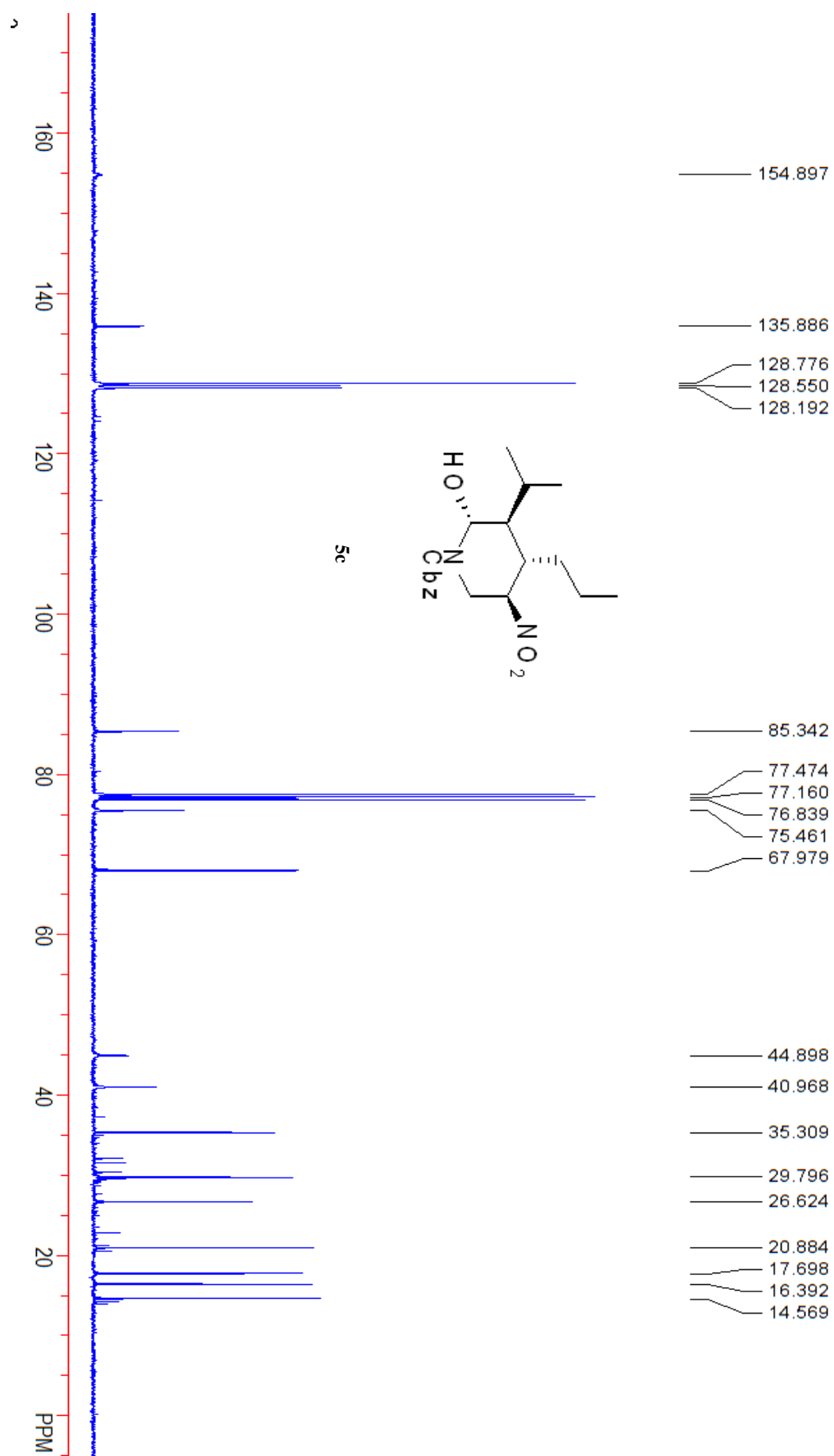
Date: 2010-09-07

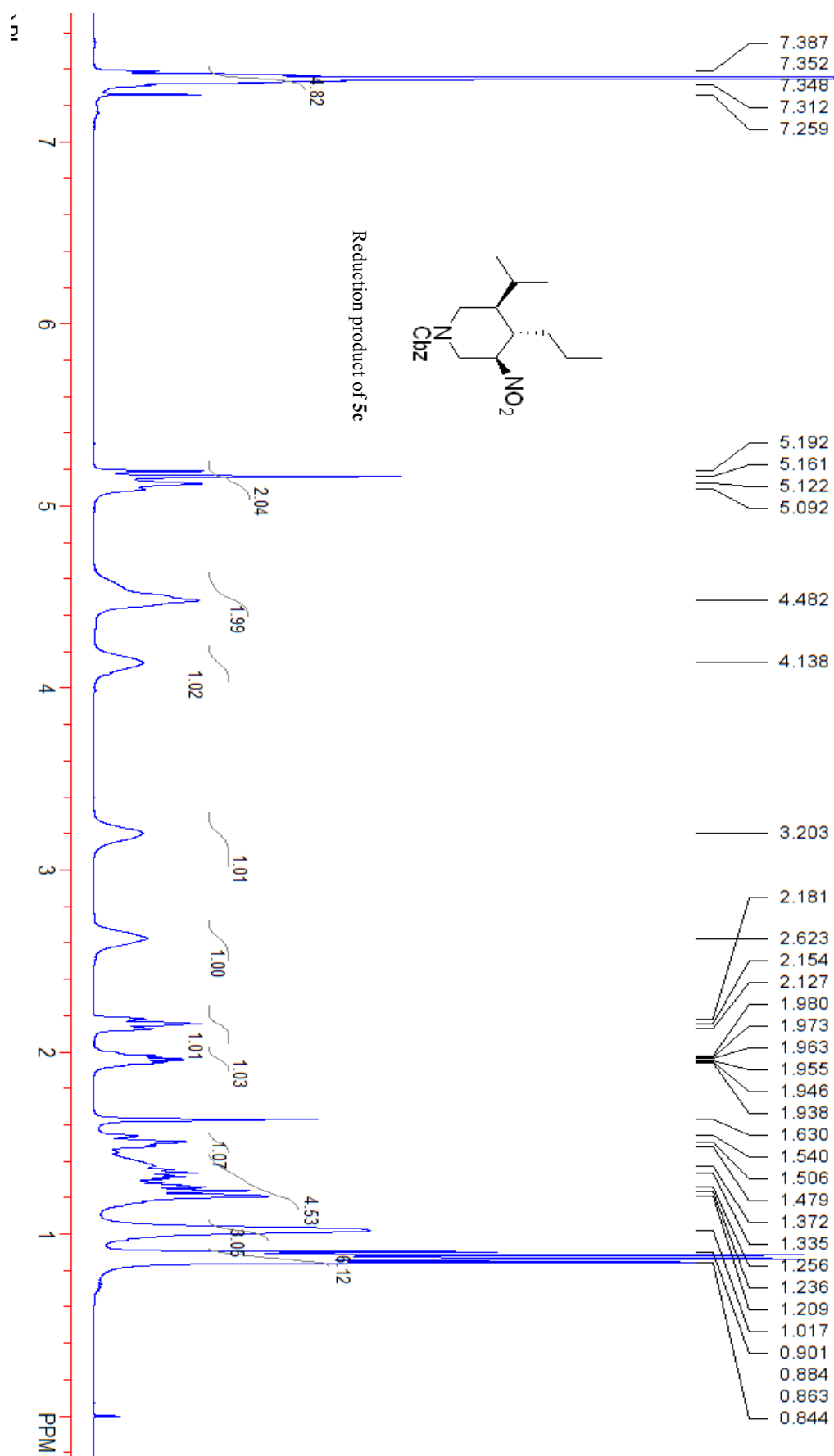
Time: 14:16

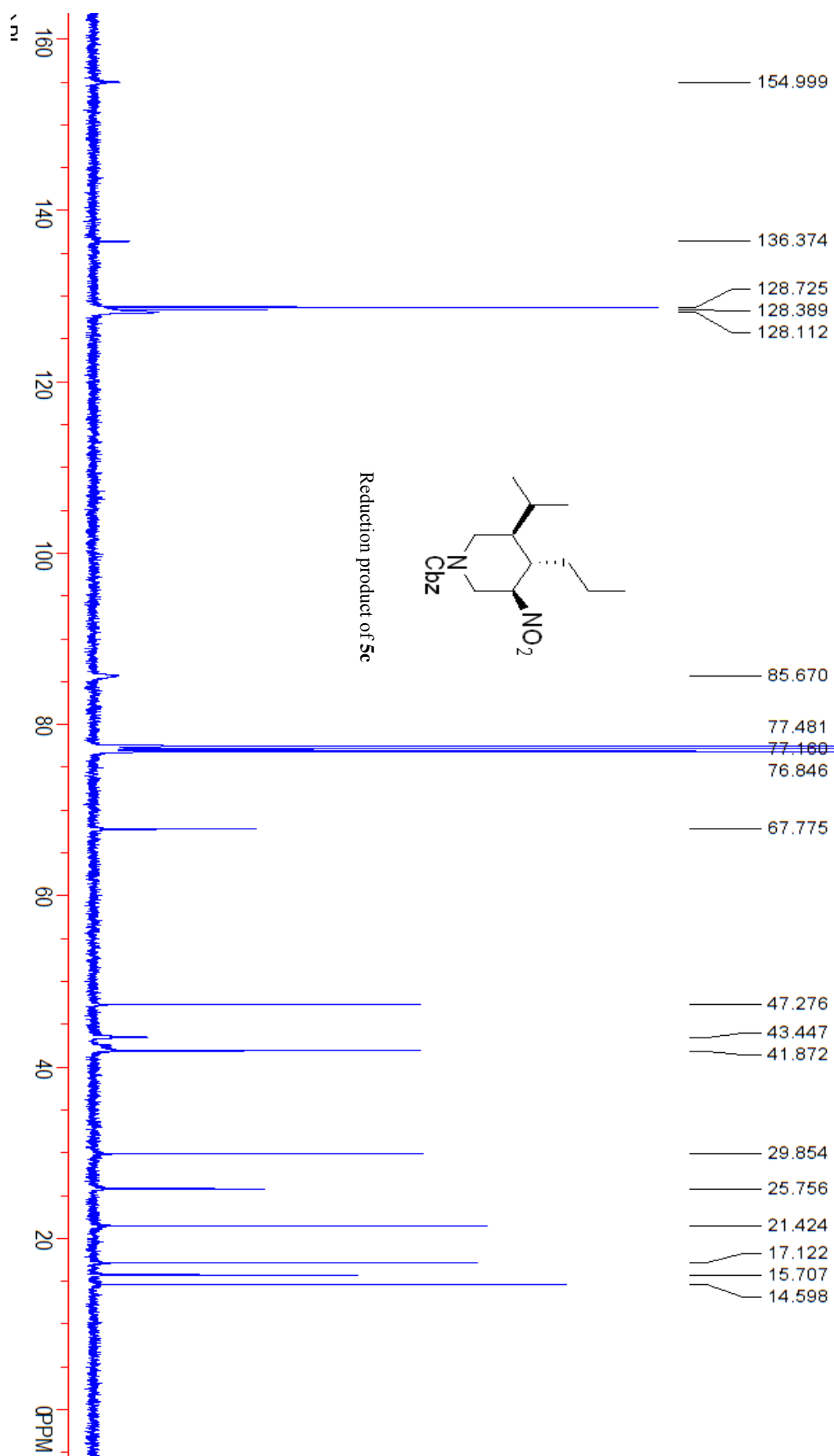


No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	10.627	621.7	8241.4	0.0831
2	2	14.477	241752.0	9907979.4	99.9169
Total			242373.7	9916220.9	100.0000









HPLC Report

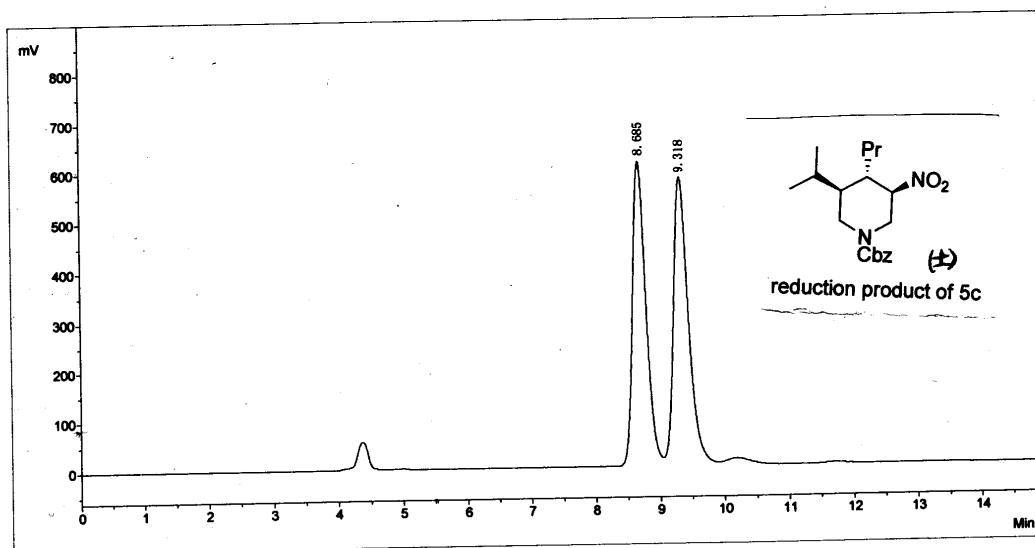
Sample Name:

Data File: ZSL-6-16-2+- PA-2 55 214 0.7. che

Operator:

Date: 2010-07-20

Time: 14:01



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		8.685	612982.1	8794990.6	49.1811
2	2		9.318	574153.1	9087883.3	50.8189
Total				1187135.2	17882873.9	100.0000

HPLC Report

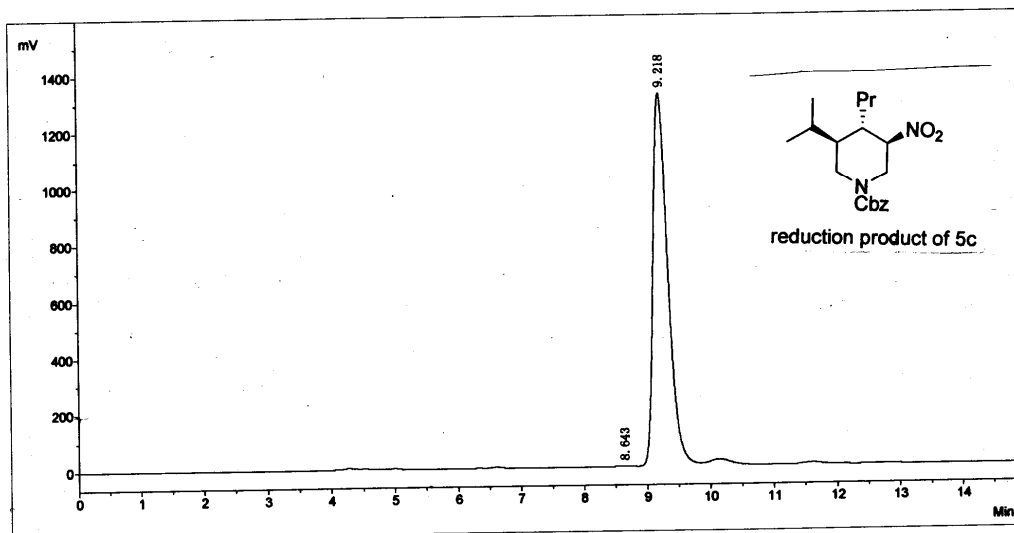
Sample Name:

Data File: ZSL-6-30-2. che

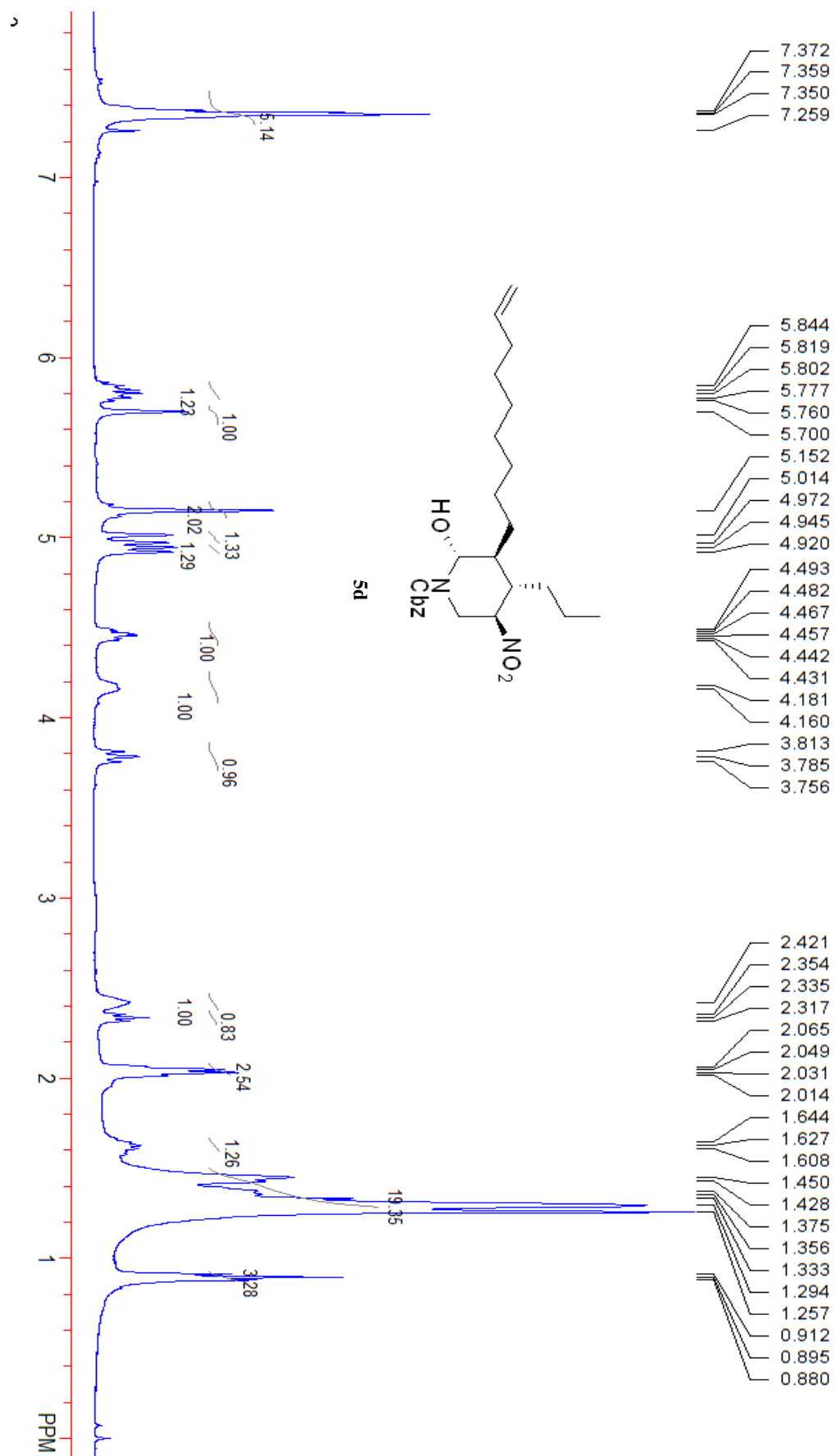
Operator:

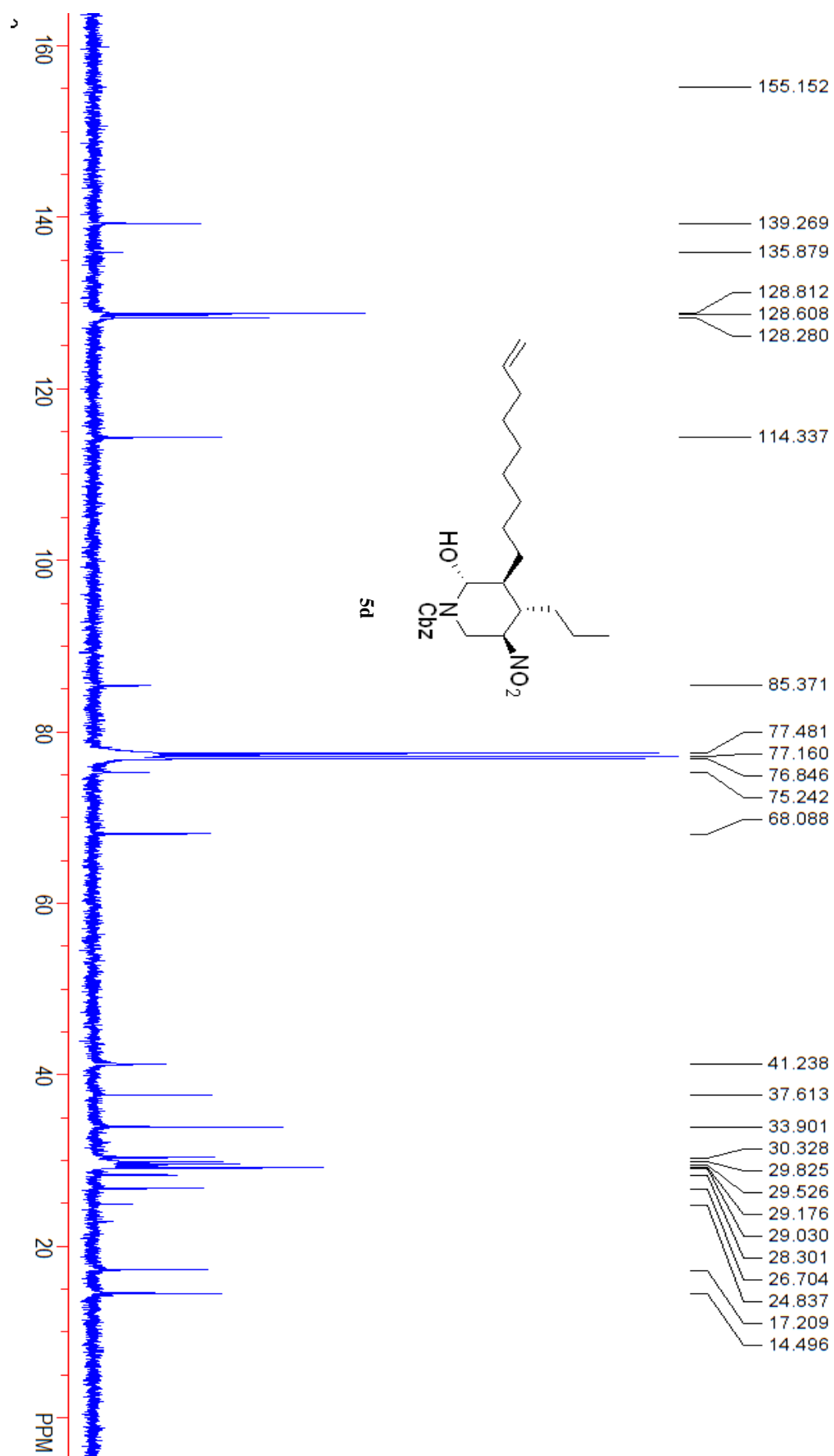
Date: 2010-07-20

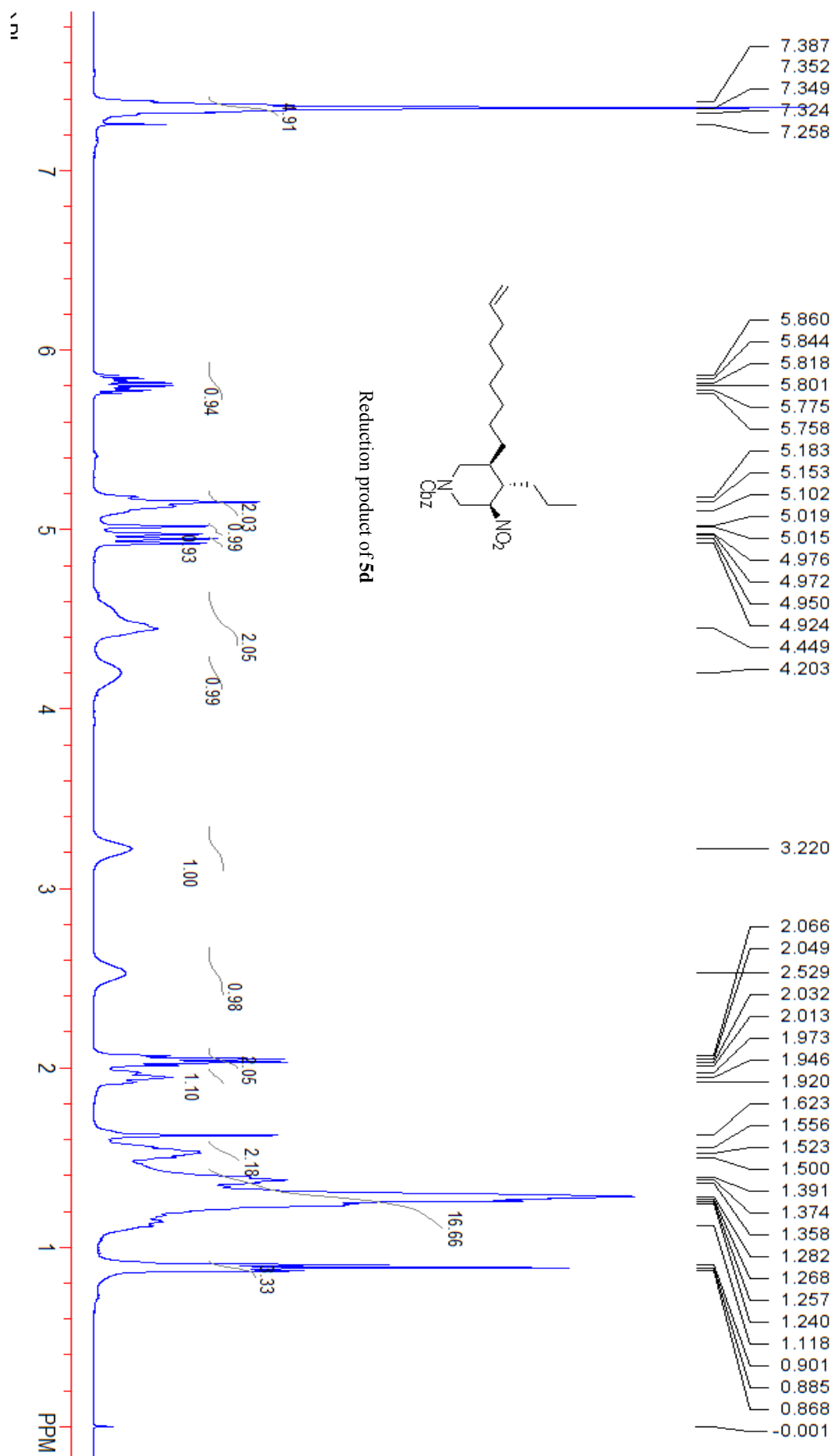
Time: 14:27

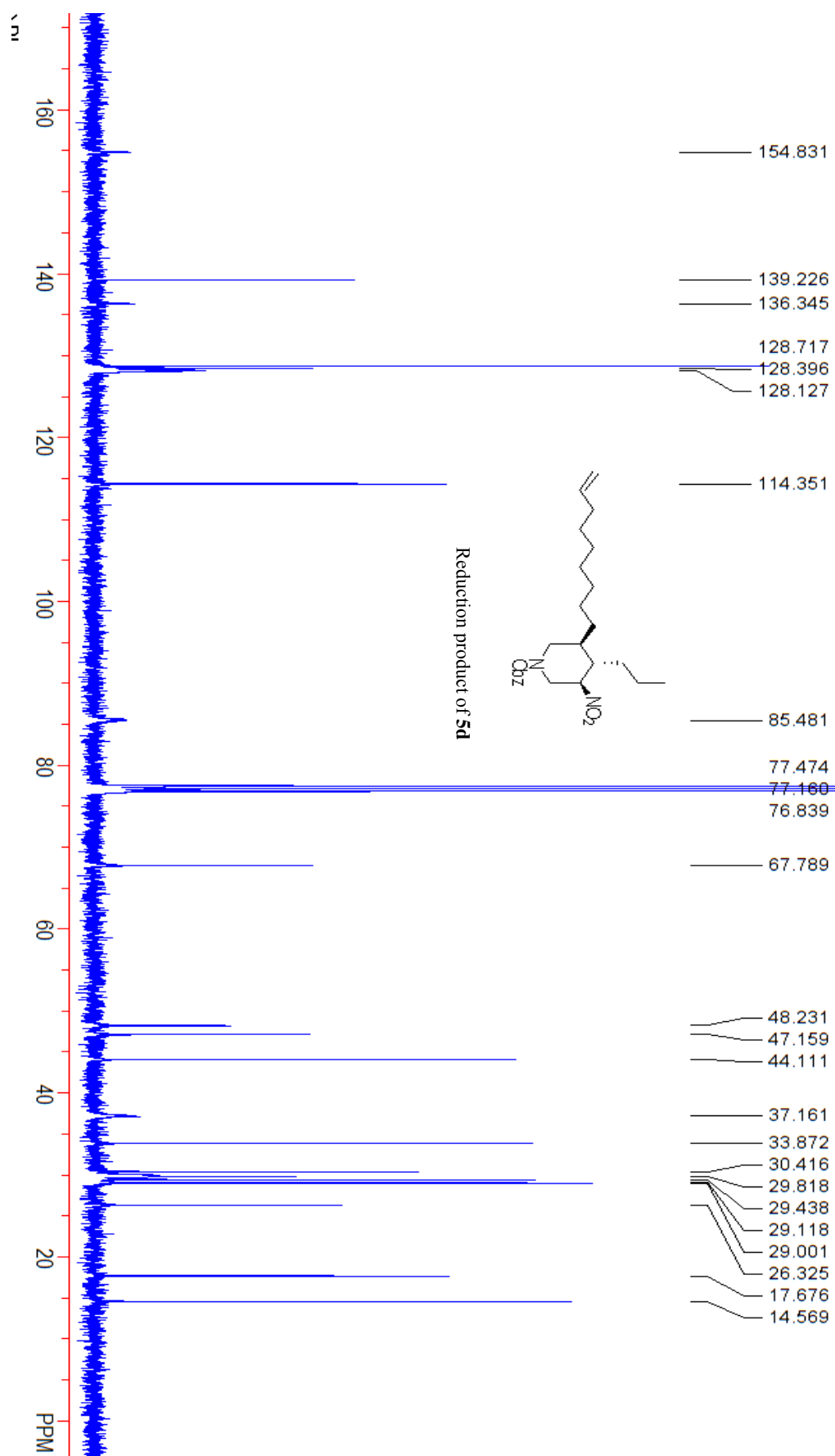


No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		8.643	2207.4	26436.7	0.1249
2	2		9.218	1318725.3	21136472.8	99.8751
Total				1320932.7	21162909.5	100.0000









HPLC Report

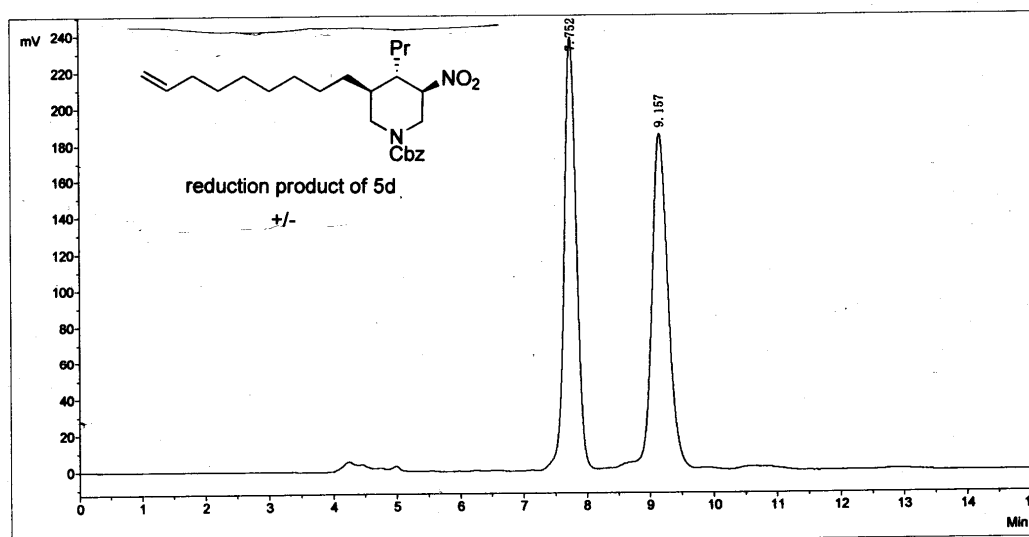
Sample Name:

Data File:ZSL-6-19-2+- PA-2 55 214 0.7.che

Operator:

Date:2010-07-20

Time:13:25



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		7.752	238116.7	3174916.5	50.4255
2	2		9.157	182479.9	3121331.3	49.5745
Total				420596.5	6296247.8	100.0000

HPLC Report

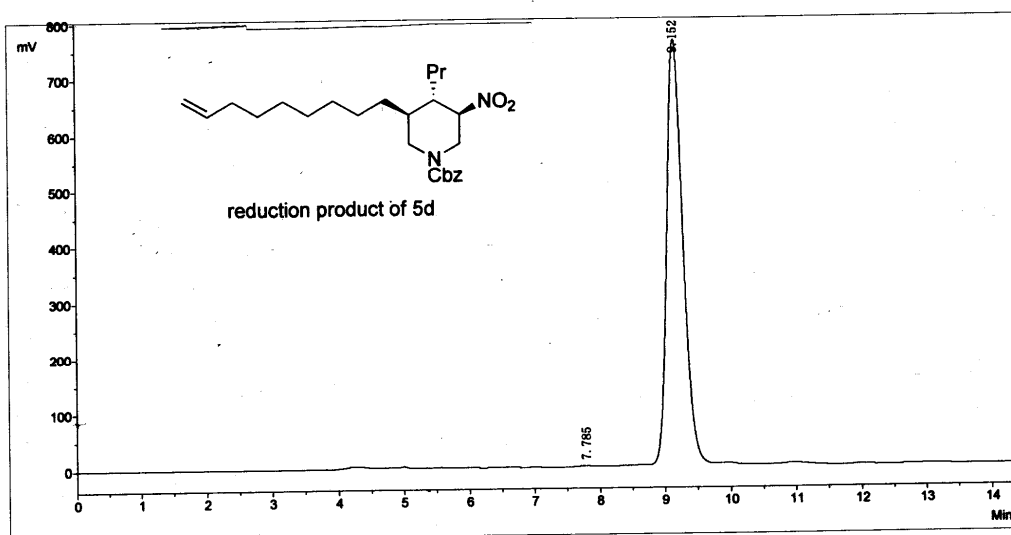
Sample Name:

Data File: ZSL-6-29.che

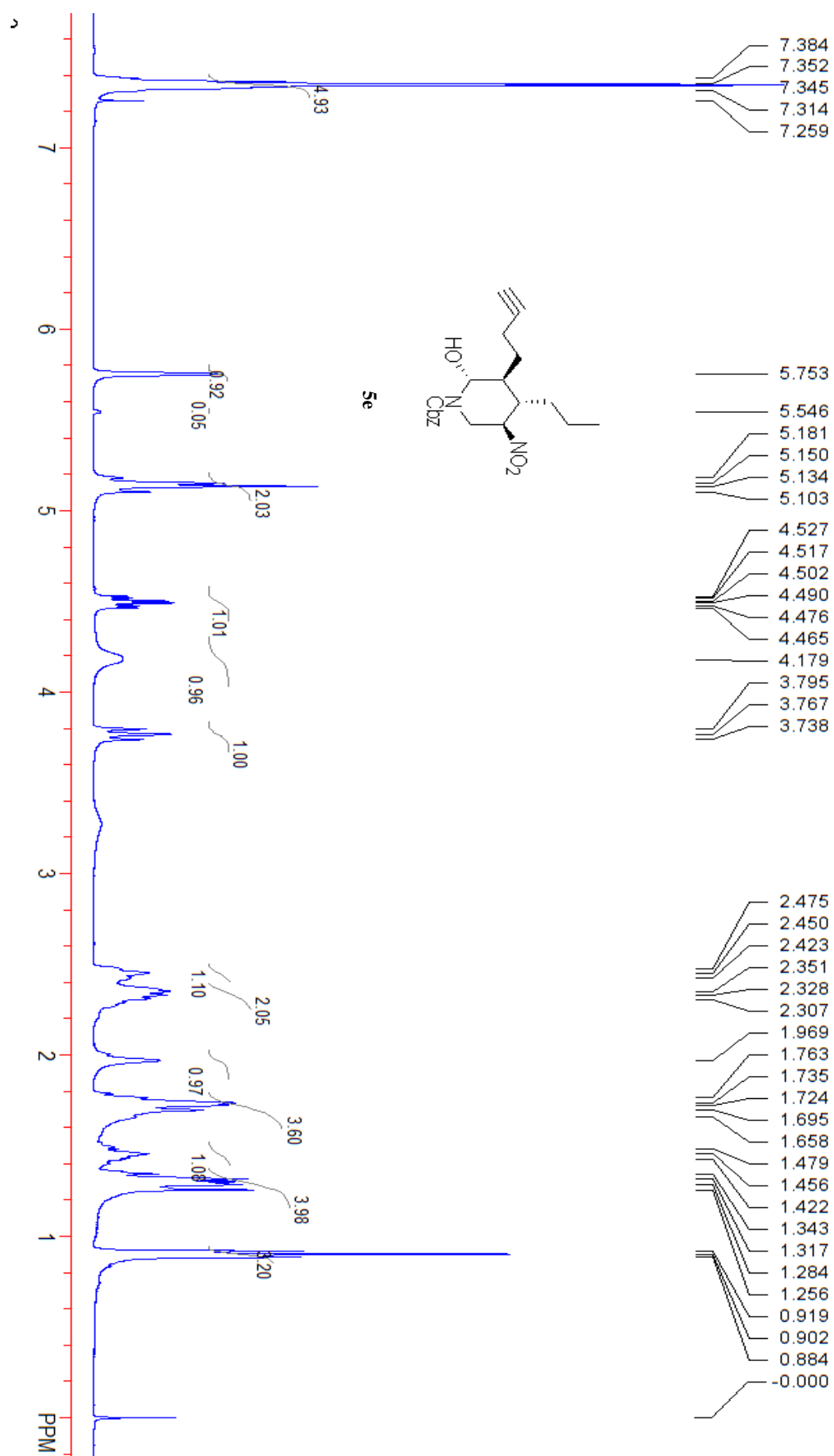
Operator:

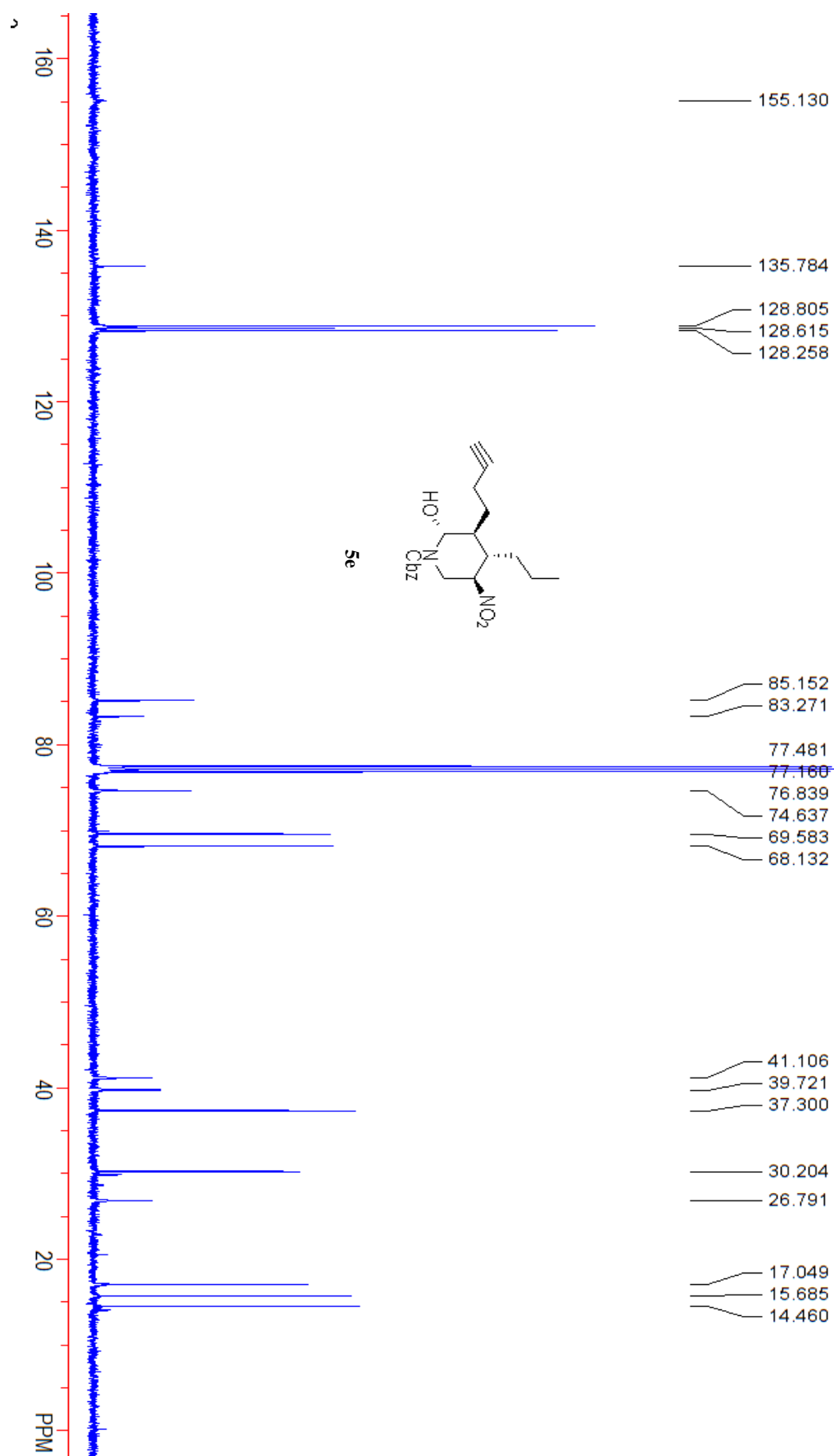
Date: 2010-07-20

Time: 13:43



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		7.785	1439.2	16491.4	0.1275
2	2		9.152	760472.5	12914286.9	99.8725
Total				761911.7	12930778.3	100.0000





HPLC Report

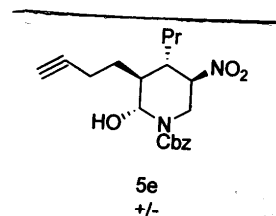
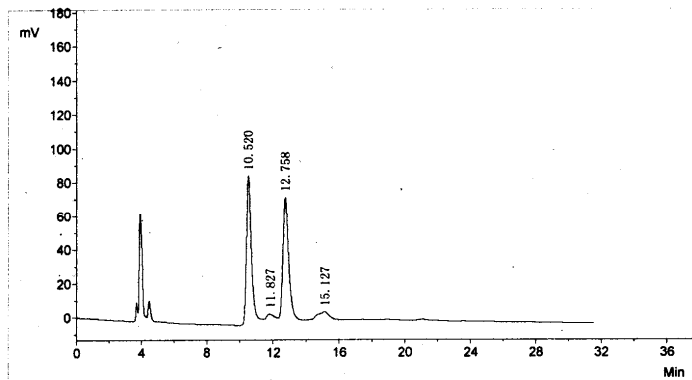
Sample Name:

Data File:ZSL6-20+-SINO-AD-910.7214.che

Operator:

Date:2010-08-19

Time:09:04



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	10.520	86975.5	1719927.1	46.1434
2	2	11.827	2685.0	69032.4	1.8521
3	3	12.758	71819.8	1698966.1	45.5811
4	4	15.127	4827.4	239425.0	6.4235
Total			166307.7	3727350.6	100.0000

HPLC Report

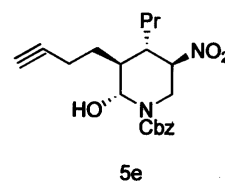
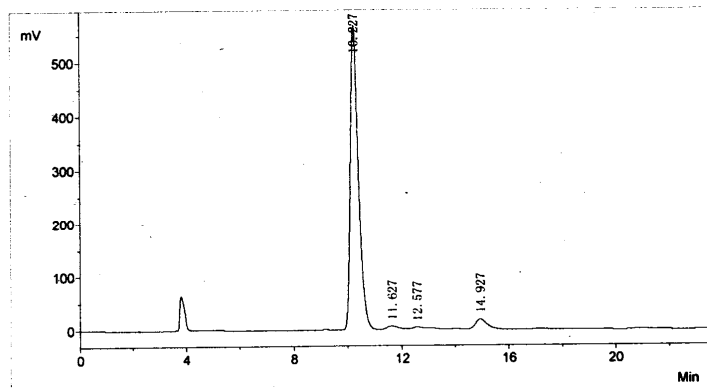
Sample Name:

Data File:ZSL6-28.che

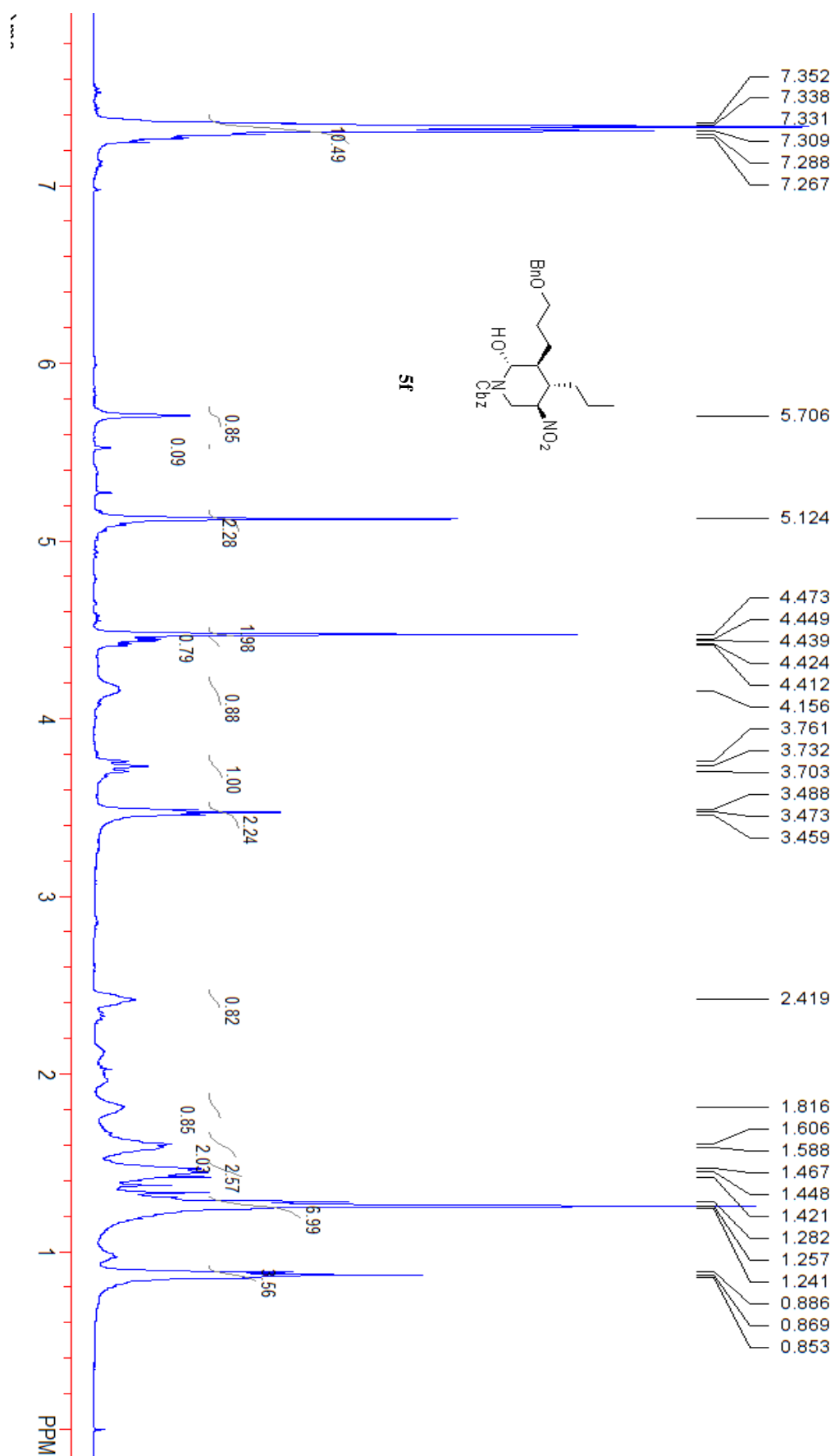
Operator:

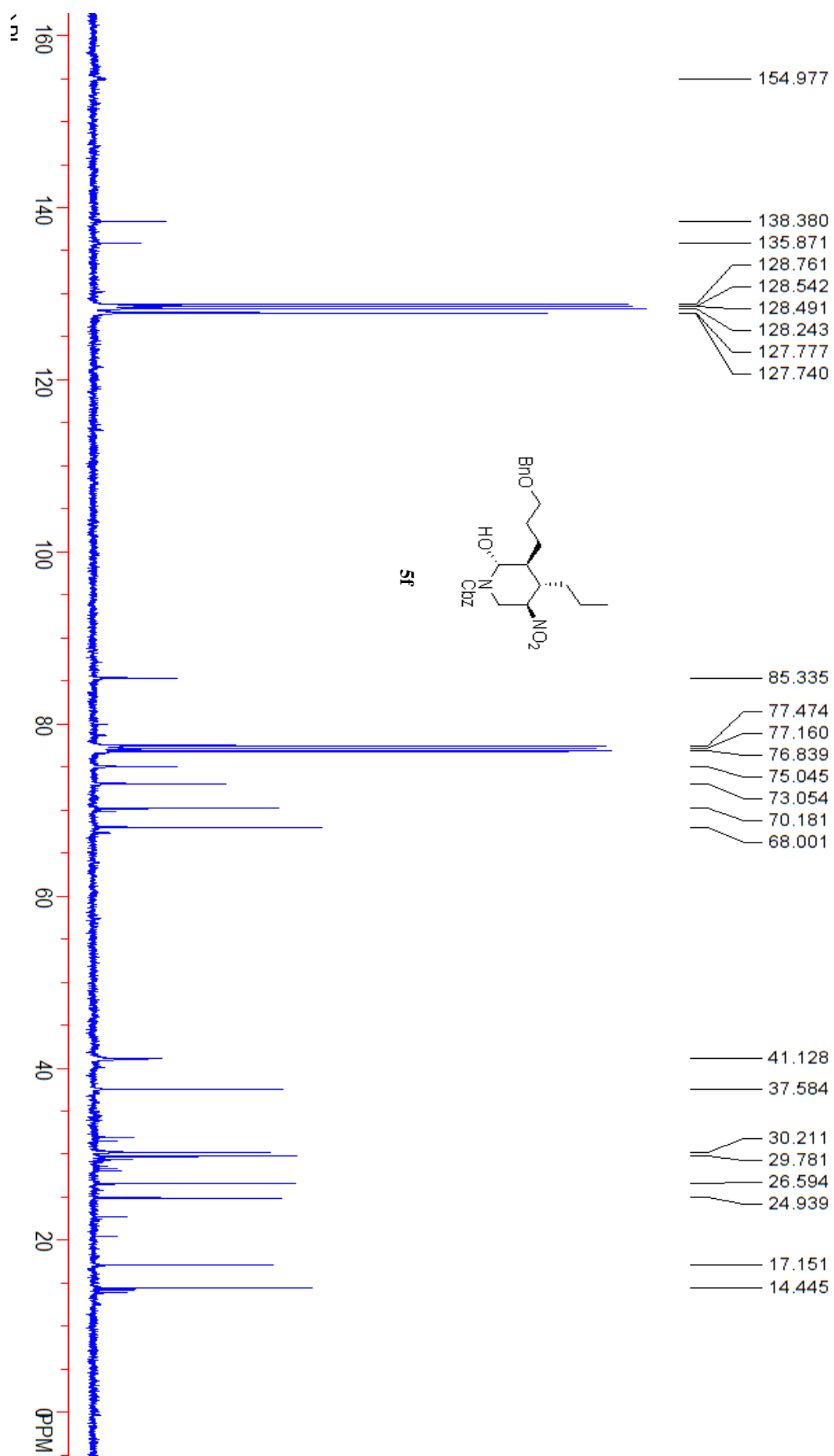
Date:2010-08-19

Time:09:41



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	10.227	564408.8	11776232.0	93.7569 ✓
2	2	11.627	5774.7	161741.2	1.2877
3	3	12.577	3127.3	59477.1	0.4735 ✓
4	4	14.927	19044.9	562943.1	4.4819
Total			592355.7	12560393.3	100.0000





HPLC Report

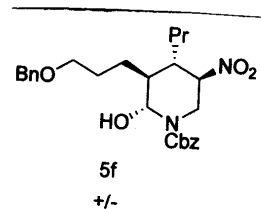
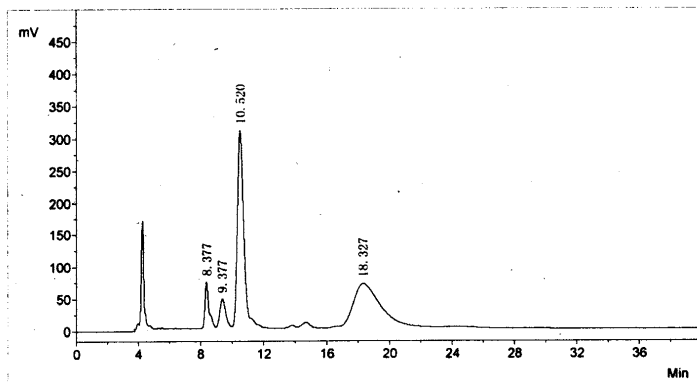
Sample Name:

Data File: ZSL-17-49+- PA-2 73 214 0.7. che

Operator:

Date: 2010-08-16

Time: 16:35



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	8.377	72153.0	1264820.3	7.0741
2	2	9.377	44074.8	1111479.7	6.2165
3	3	10.520	300193.5	7770978.7	43.4628
4	4	18.327	66426.7	7732317.3	43.2466
Total			482848.0	17879596.0	100.0000

HPLC Report

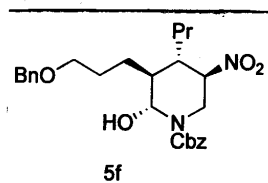
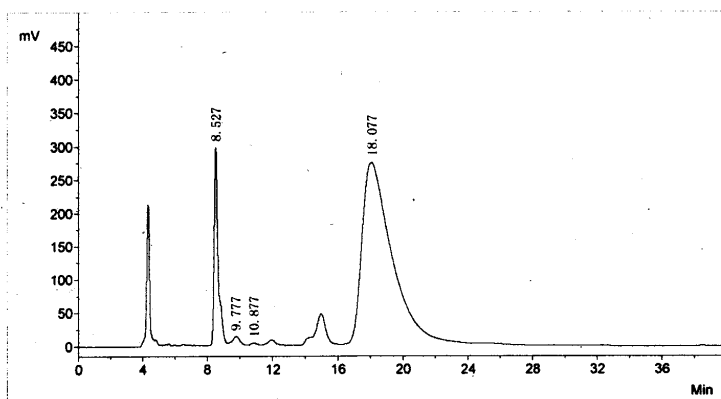
Sample Name:

Data File: ZSL-6-32. che

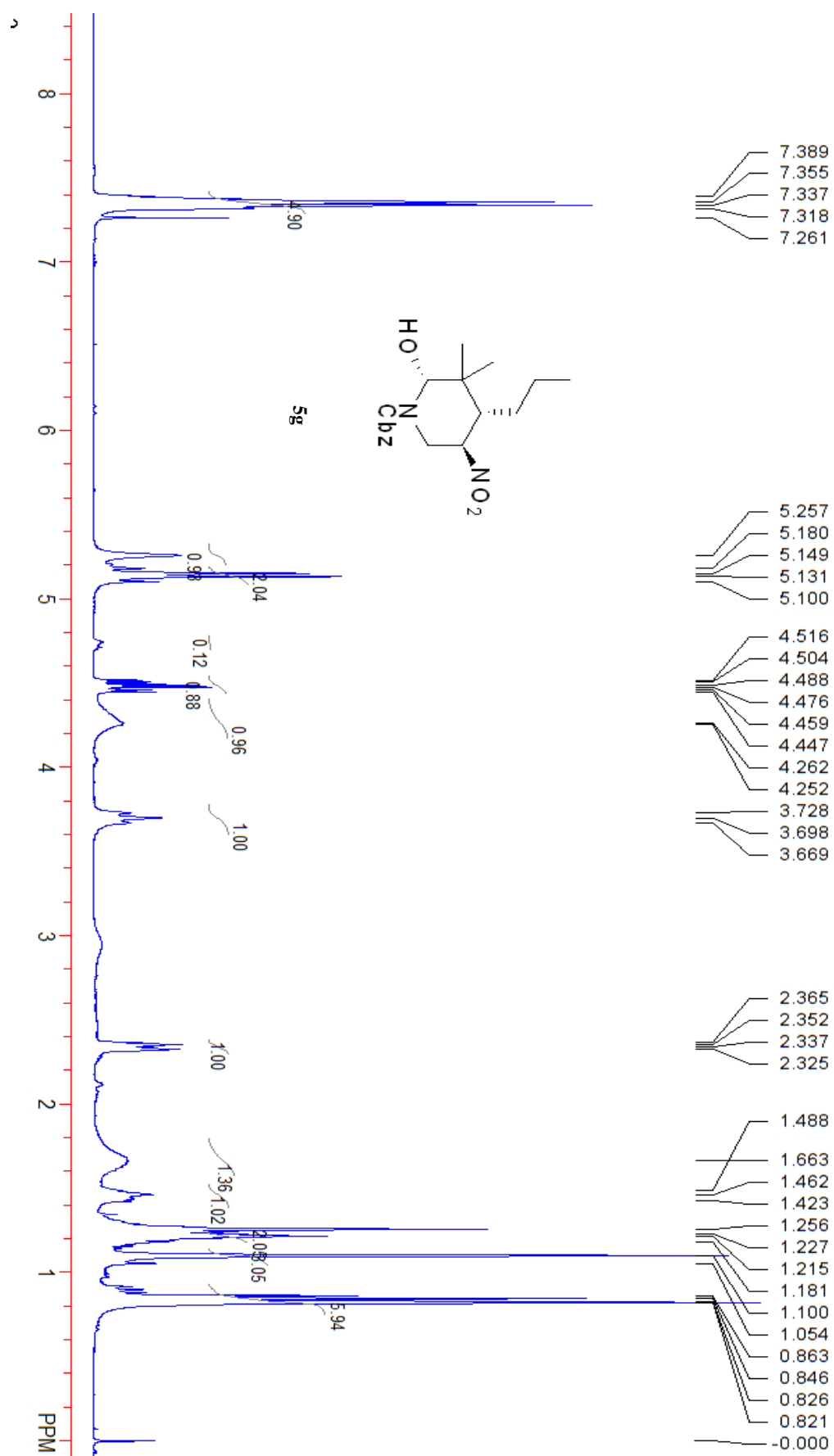
Operator:

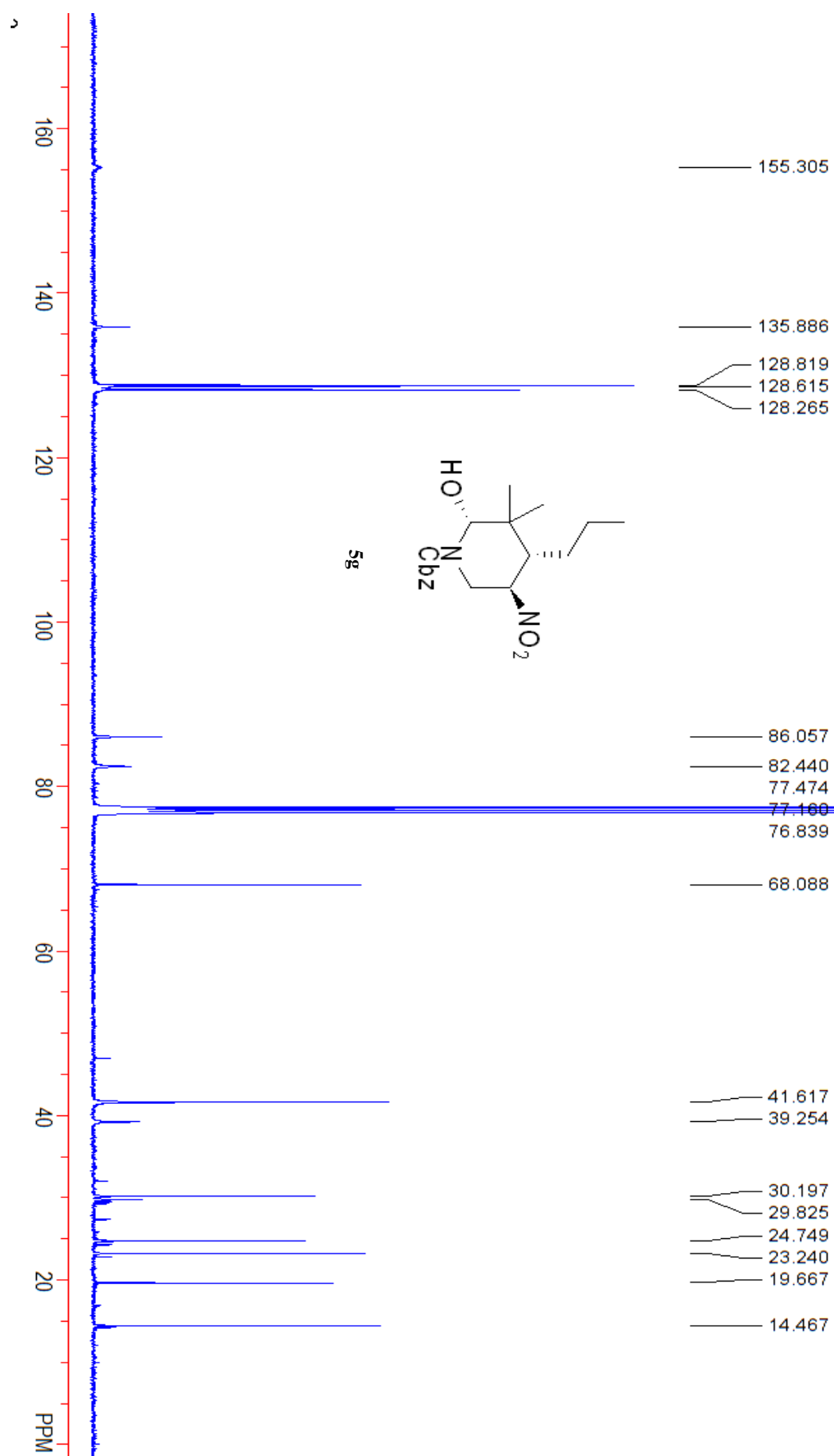
Date: 2010-08-16

Time: 17:33



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	8.527	297646.4	5204237.6	12.9060
2	2	9.777	13661.7	434562.6	1.0777
3	3	10.877	3312.1	82949.2	0.2057
4	4	18.077	273054.8	34602347.9	85.8106
Total			587675.0	40324097.2	100.0000





HPLC Report

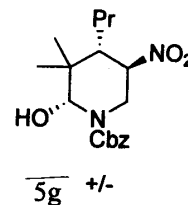
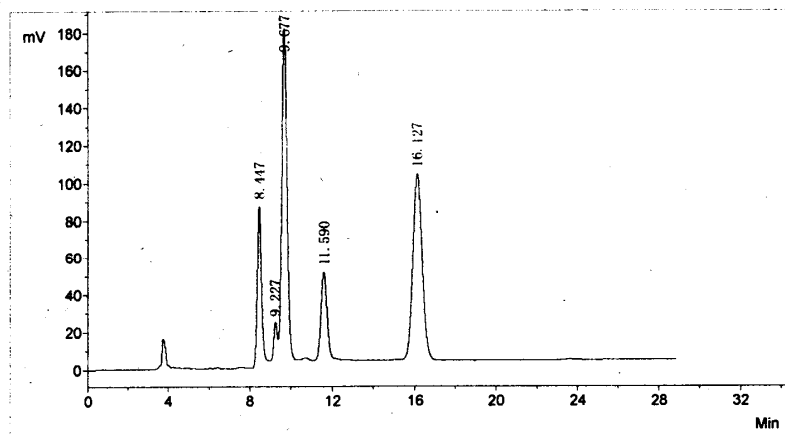
Sample Name:

Data File:ZSL-6-12+- PC-2 91 214 0.7. che

Operator:

Date:2010-09-01

Time:10:18



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	8.447	84949.7	1170842.7	13.4630
2	2	9.227	20003.7	240992.5	2.7711
3	3	9.677	177394.2	3136117.0	36.0608
4	4	11.590	46931.6	957040.2	11.0046
5	5	16.127	99869.3	3191744.0	36.7005
Total			429148.5	8696736.4	100.0000

HPLC Report

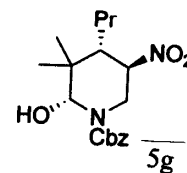
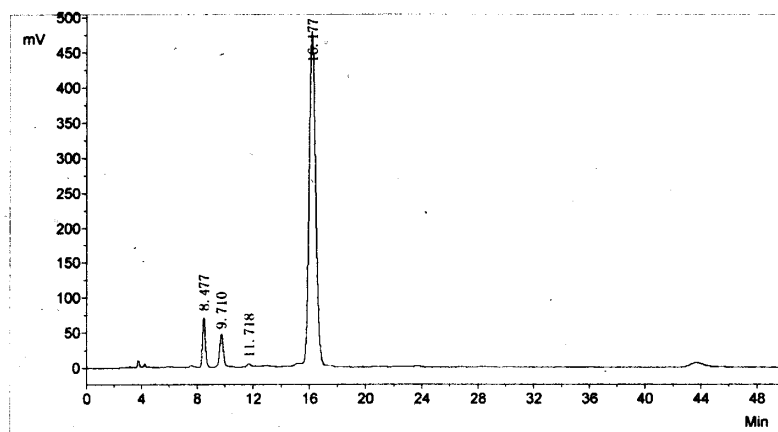
Sample Name:

Data File: ZSL-6-58-1.che

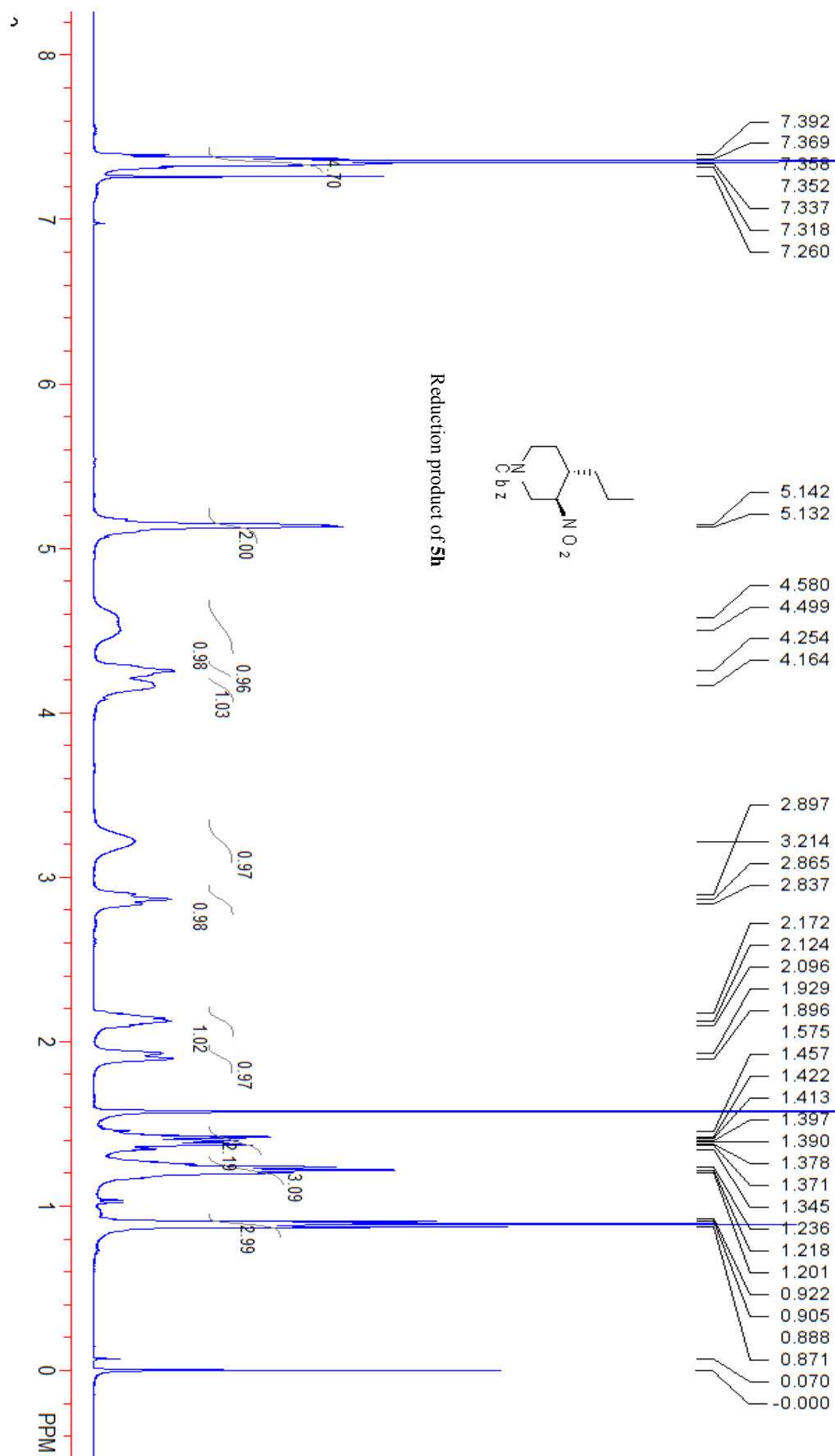
Operator:

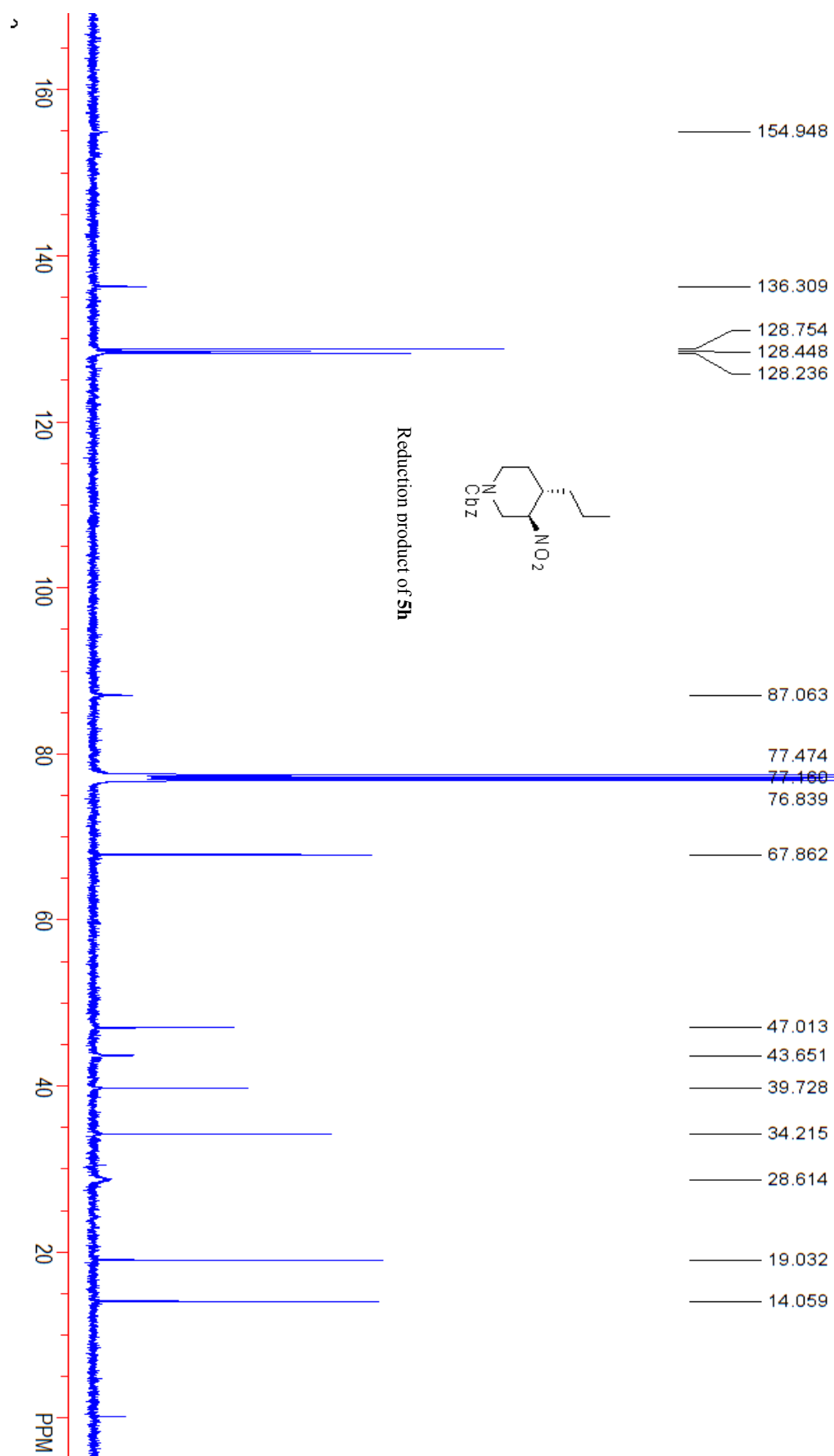
Date: 2010-09-01

Time: 10:56



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	8.477	71228.2	1025711.5	6.0447
2	2	9.710	43662.8	785706.3	4.6303
3	3	11.718	2788.2	41095.3	0.2422
4	4	16.177	475484.4	15116135.5	89.0827
Total			593163.6	16968648.5	100.0000





HPLC Report

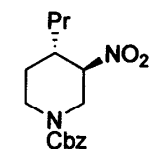
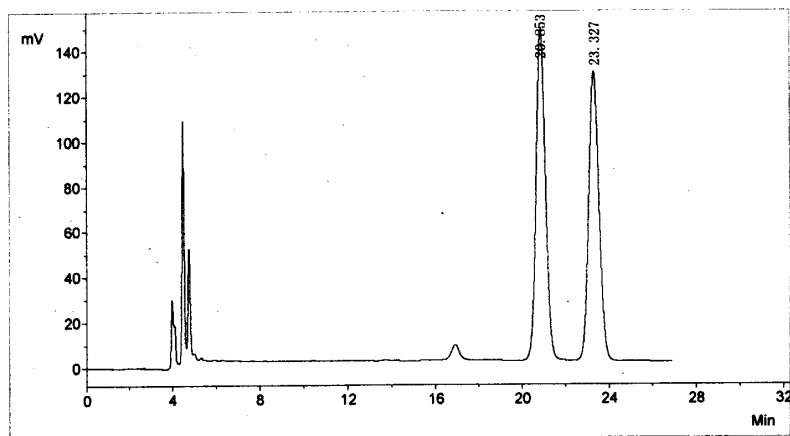
Sample Name:

Data File: ZSL6-82+-.che

Operator:

Date: 2010-10-18

Time: 08:49



reduction product of 5h

+/-

No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	20.853	145686.3	4283260.4	50.4646
2	2	23.327	127371.2	4204392.6	49.5354
Total			273057.6	8487652.9	100.0000

HPLC Report

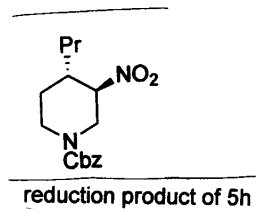
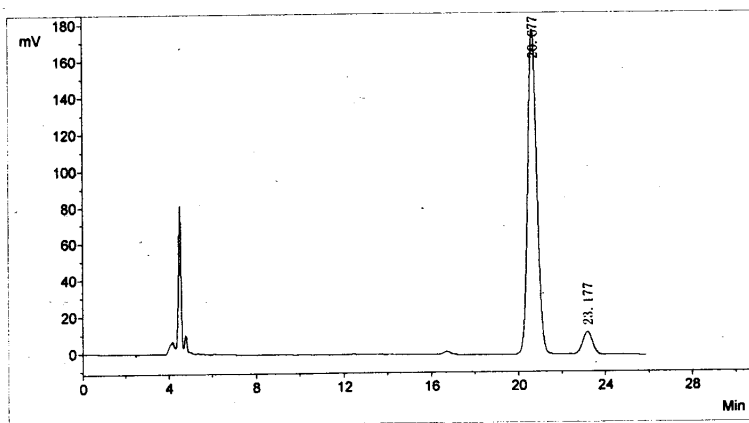
Sample Name:

Data File: ZSL6-89.che

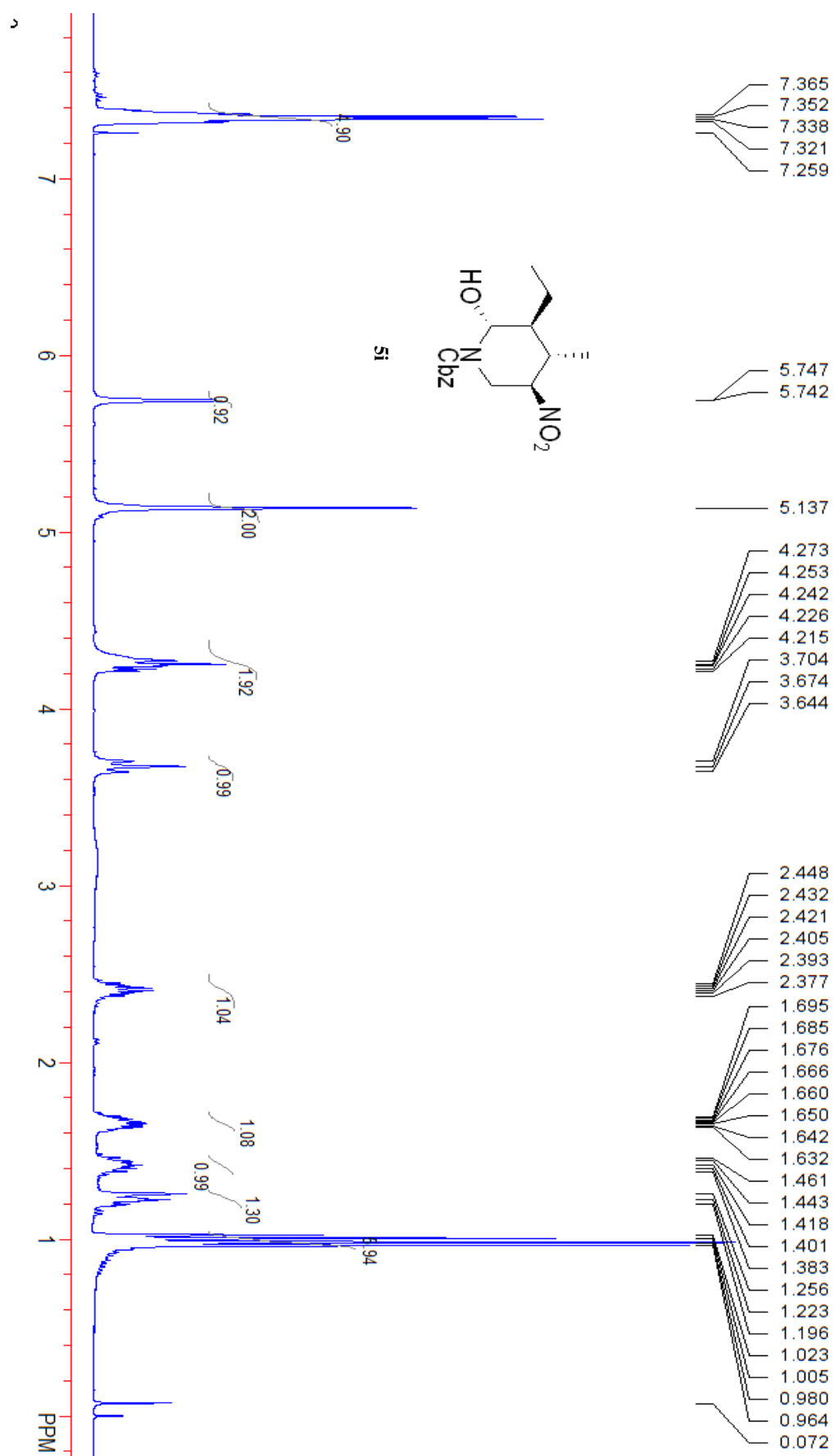
Operator:

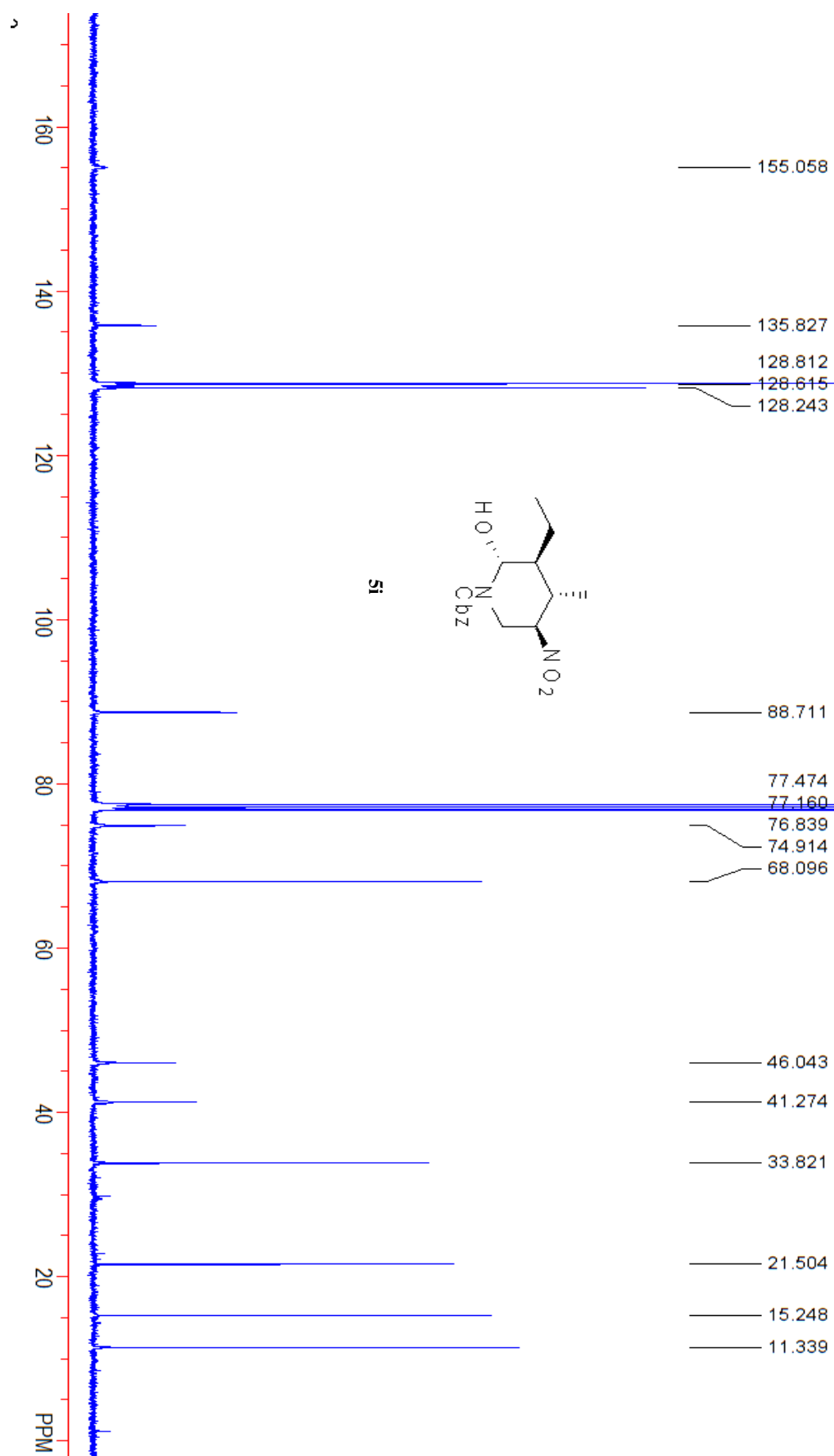
Date: 2010-10-18

Time: 09:52



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	20.677	177490.1	5171404.2	92.9367
2	2	23.177	12366.4	393031.5	7.0633
Total			189856.4	5564435.8	100.0000





HPLC Report

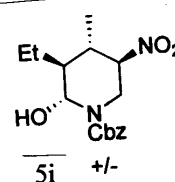
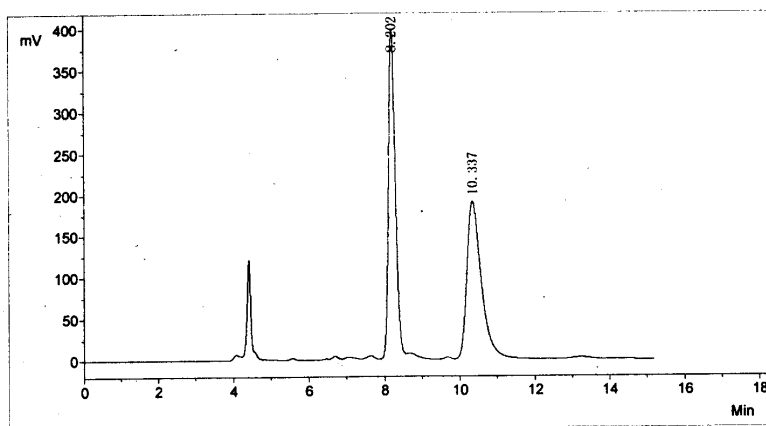
Sample Name:

Data File: ZSL6-88+-PA-2730.7214.che

Operator:

Date: 2010-10-20

Time: 13:45



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	8.202	390934.8	4789758.2	50.5280
2	2	10.337	187029.8	4689654.3	49.4720
Total			577964.5	9479412.5	100.0000

HPLC Report

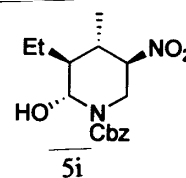
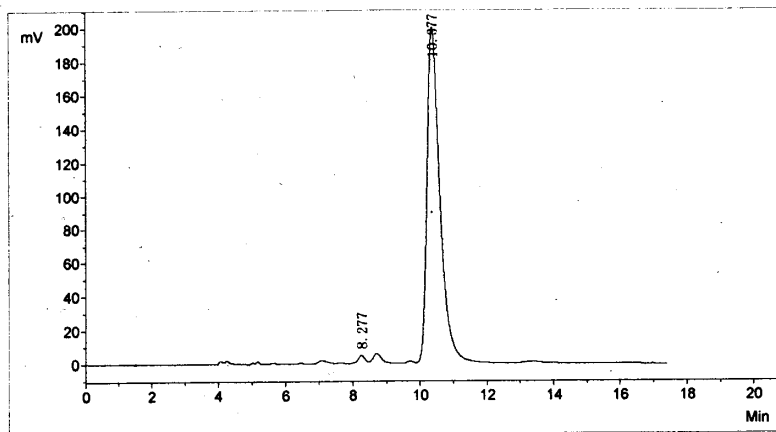
Sample Name:

Data File: ZSL6-90.che

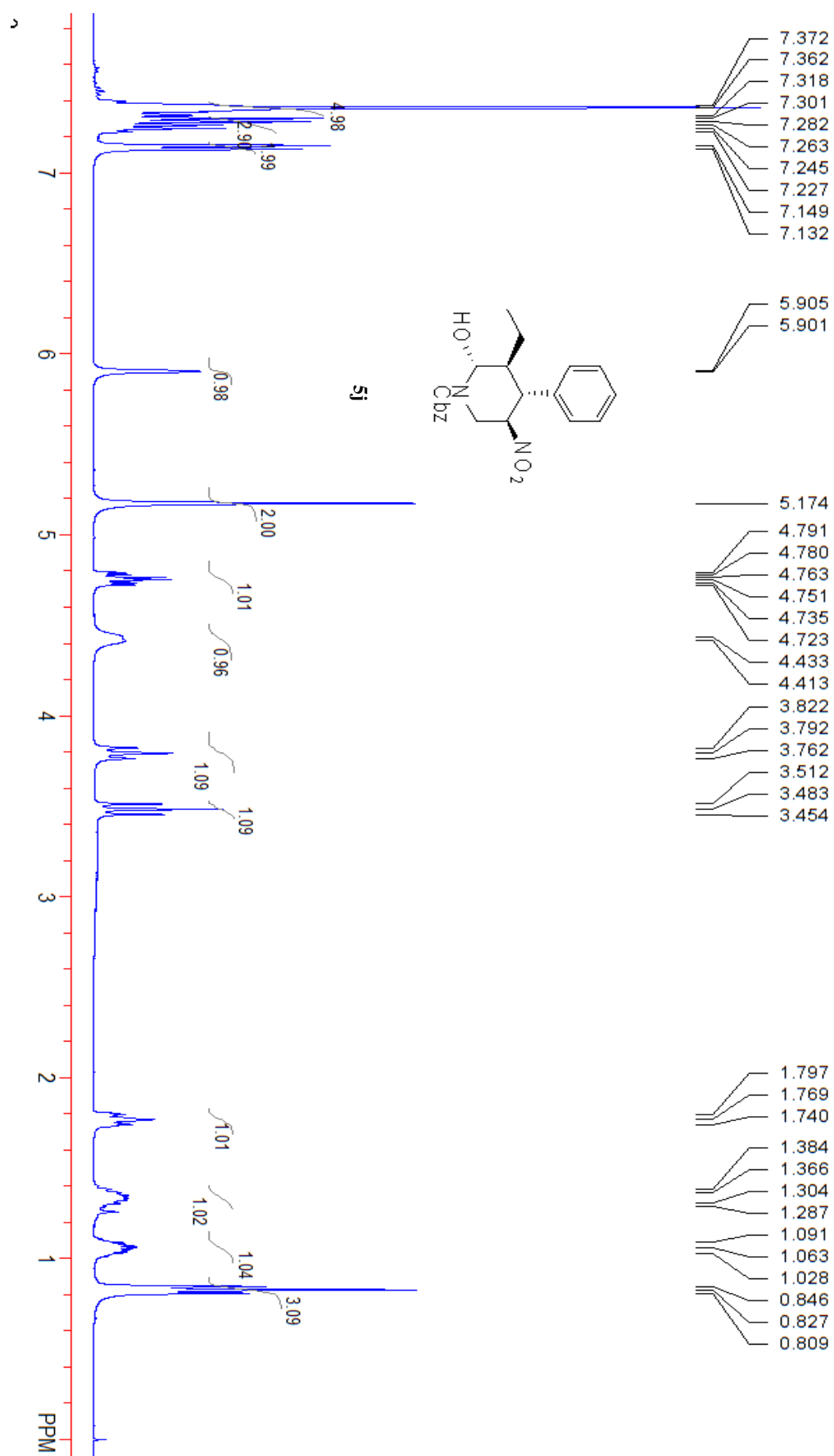
Operator:

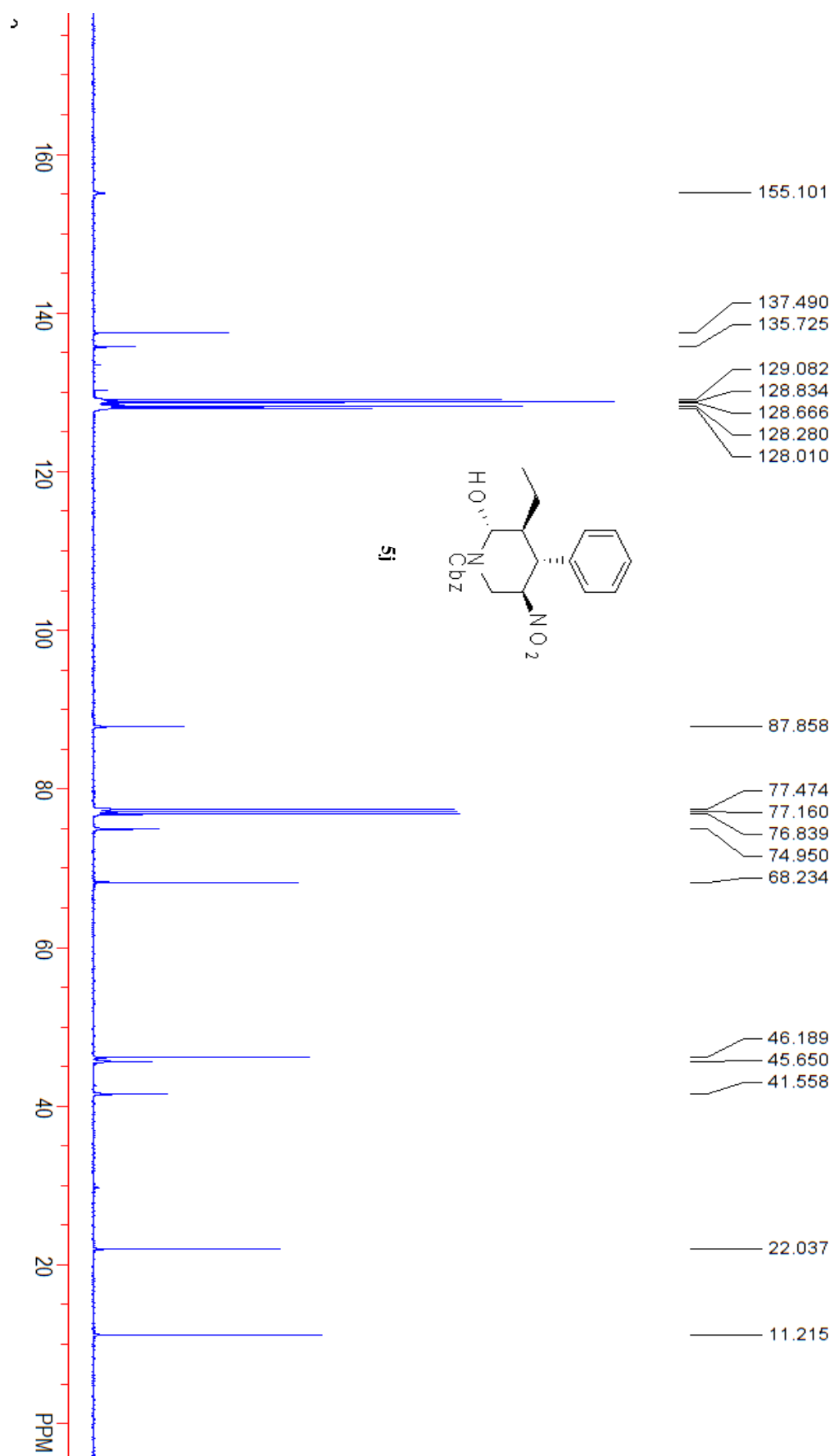
Date: 2010-10-20

Time: 14:07



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	8.277	4068.2	50828.0	0.9310
2	2	10.377	200075.3	5408880.0	99.0690
Total			204143.6	5459707.9	100.0000





HPLC Report

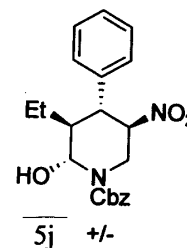
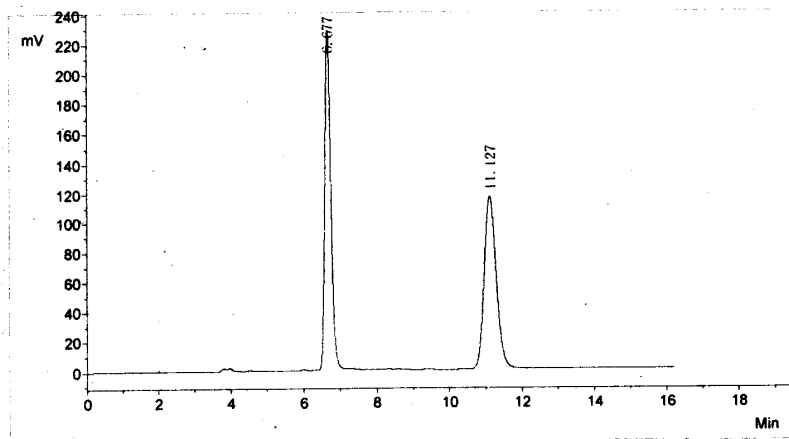
Sample Name:

Data File: ZSL6-17+-PC-2730.7214.che

Operator:

Date: 2010-08-25

Time: 09:09



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	6.677	227041.6	2713102.9	50.0982
2	2	11.127	116279.6	2702463.8	49.9018
Total			343321.3	5415566.6	100.0000

HPLC Report

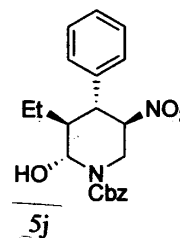
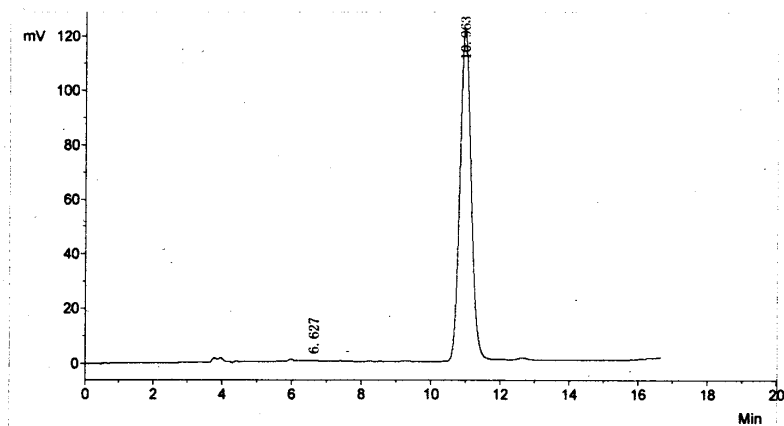
Sample Name:

Data File: ZSL6-60.che

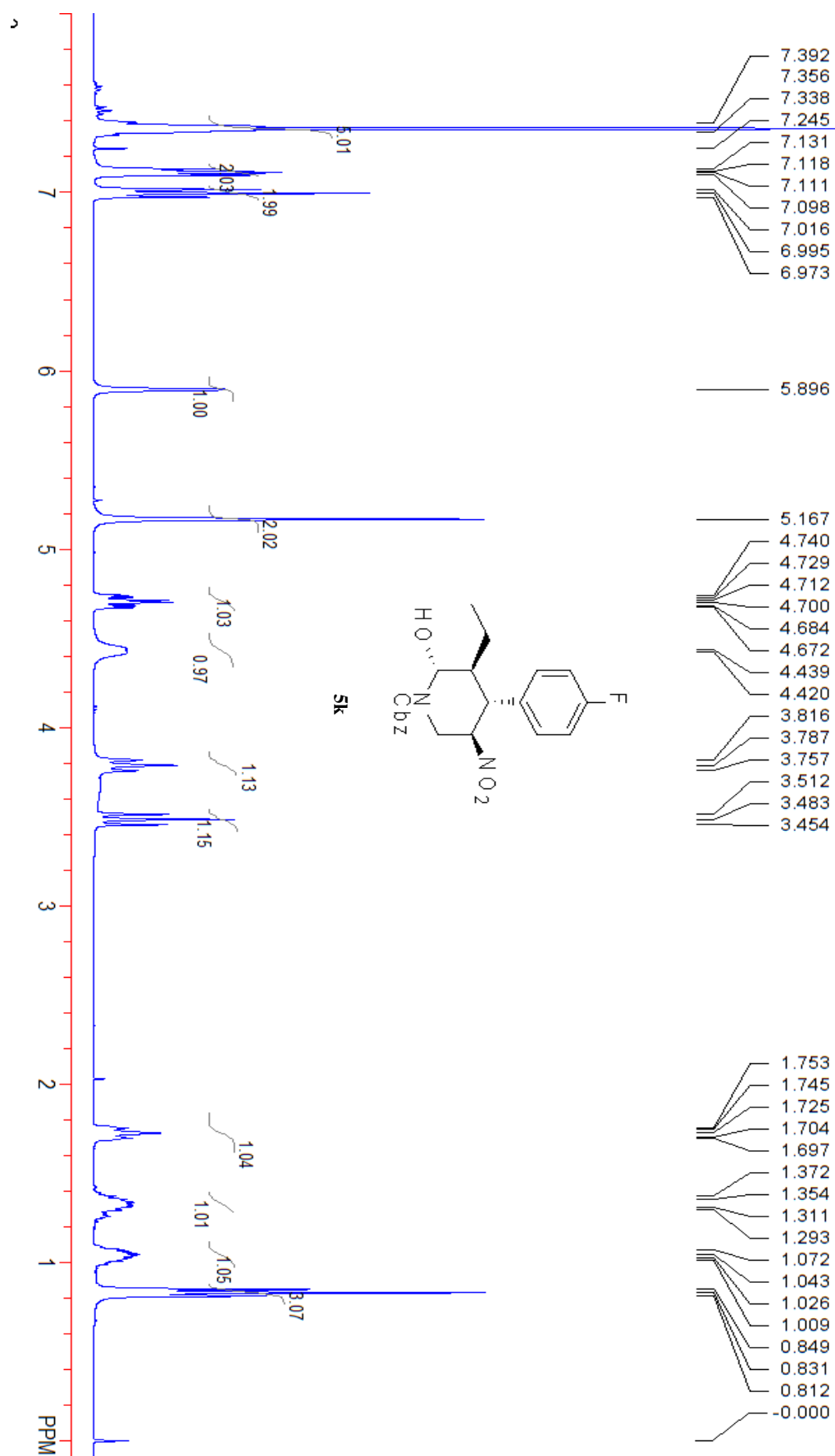
Operator:

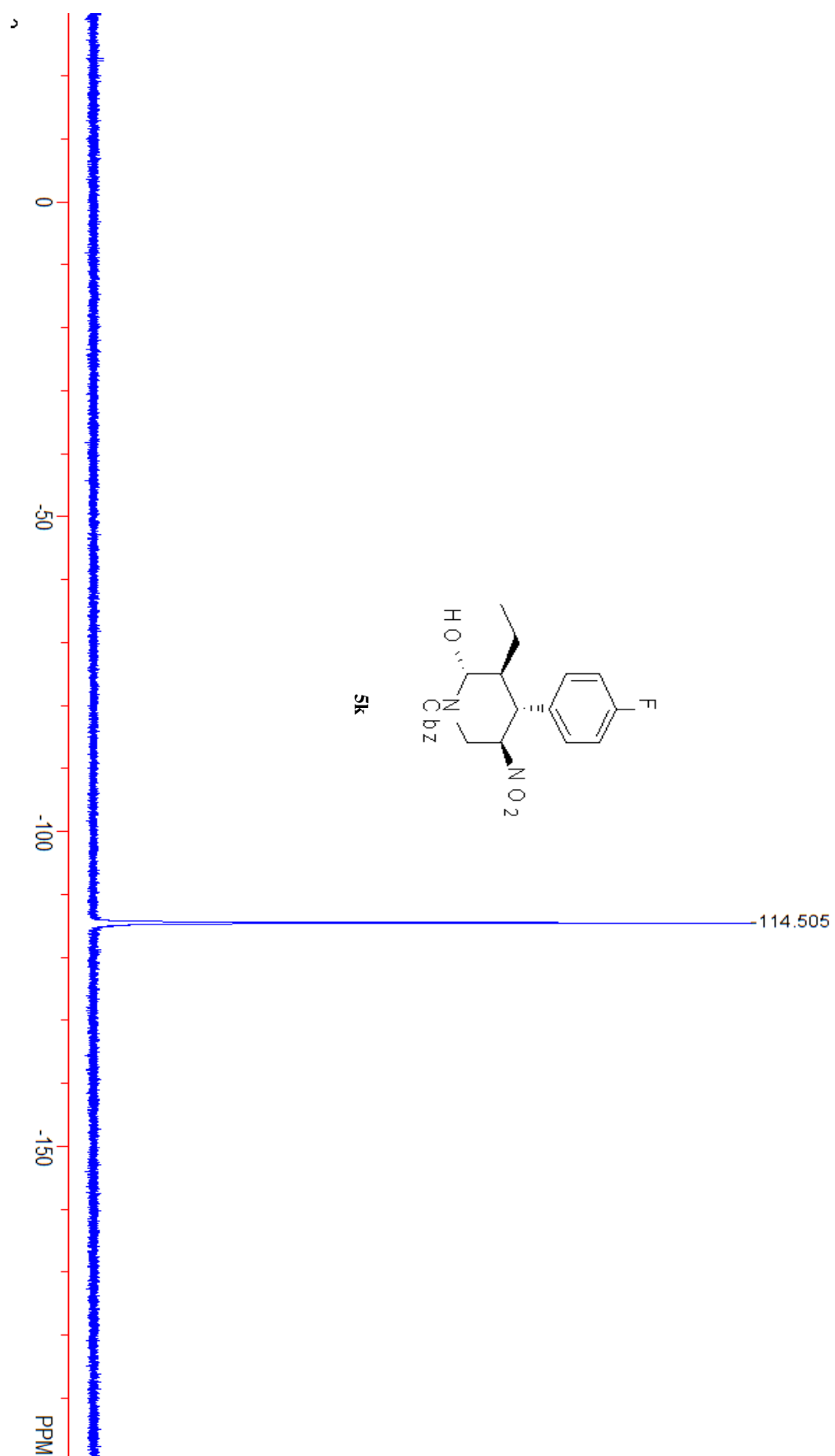
Date: 2010-08-25

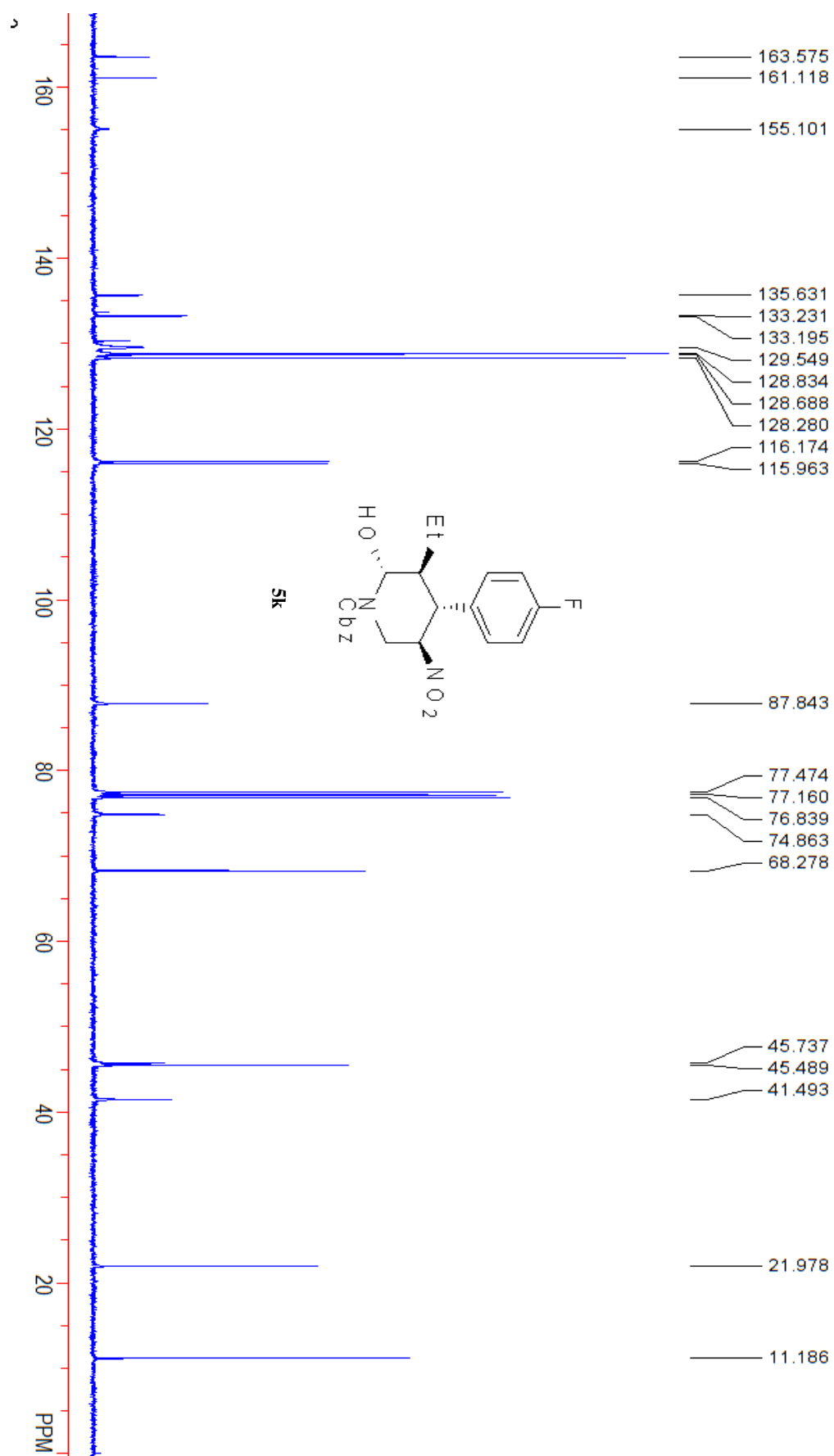
Time: 09:31



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	6.627	272.0	5650.9	0.2090
2	2	10.963	121082.4	2698644.4	99.7910
Total			121354.4	2704295.3	100.0000







HPLC Report

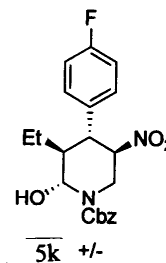
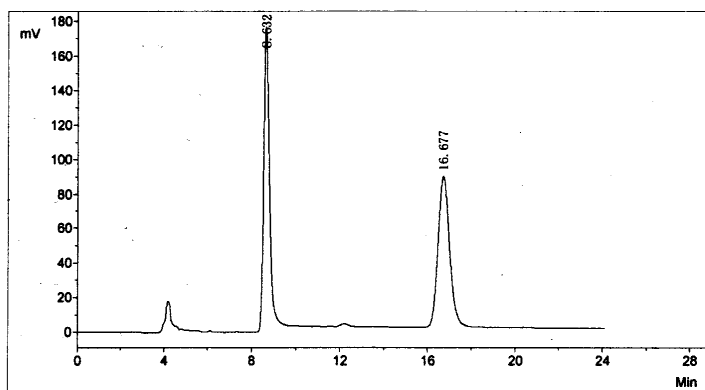
Sample Name:

Data File: ZSL6-97+-PC-2820.7214.che

Operator:

Date: 2010-11-23

Time: 13:29



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	8.632	174752.3	3111513.3	49.1262
2	2	16.677	87305.2	3222207.0	50.8738
Total			262057.5	6333720.3	100.0000

HPLC Report

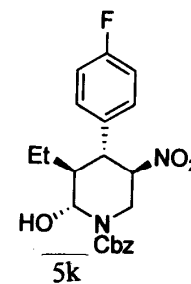
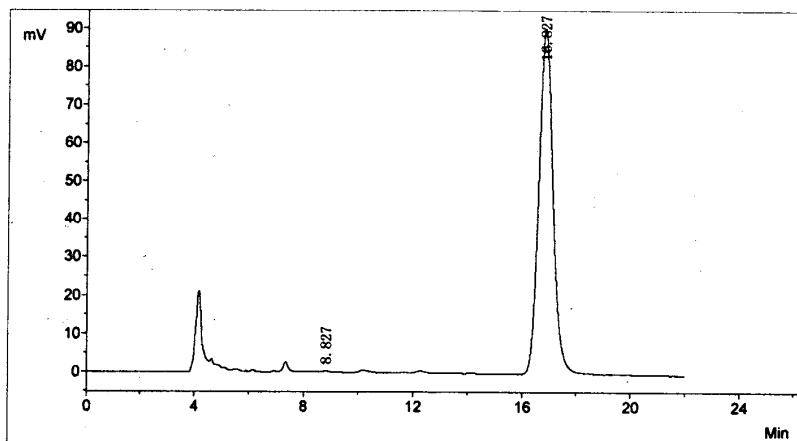
Sample Name:

Data File:ZSL6-98.che

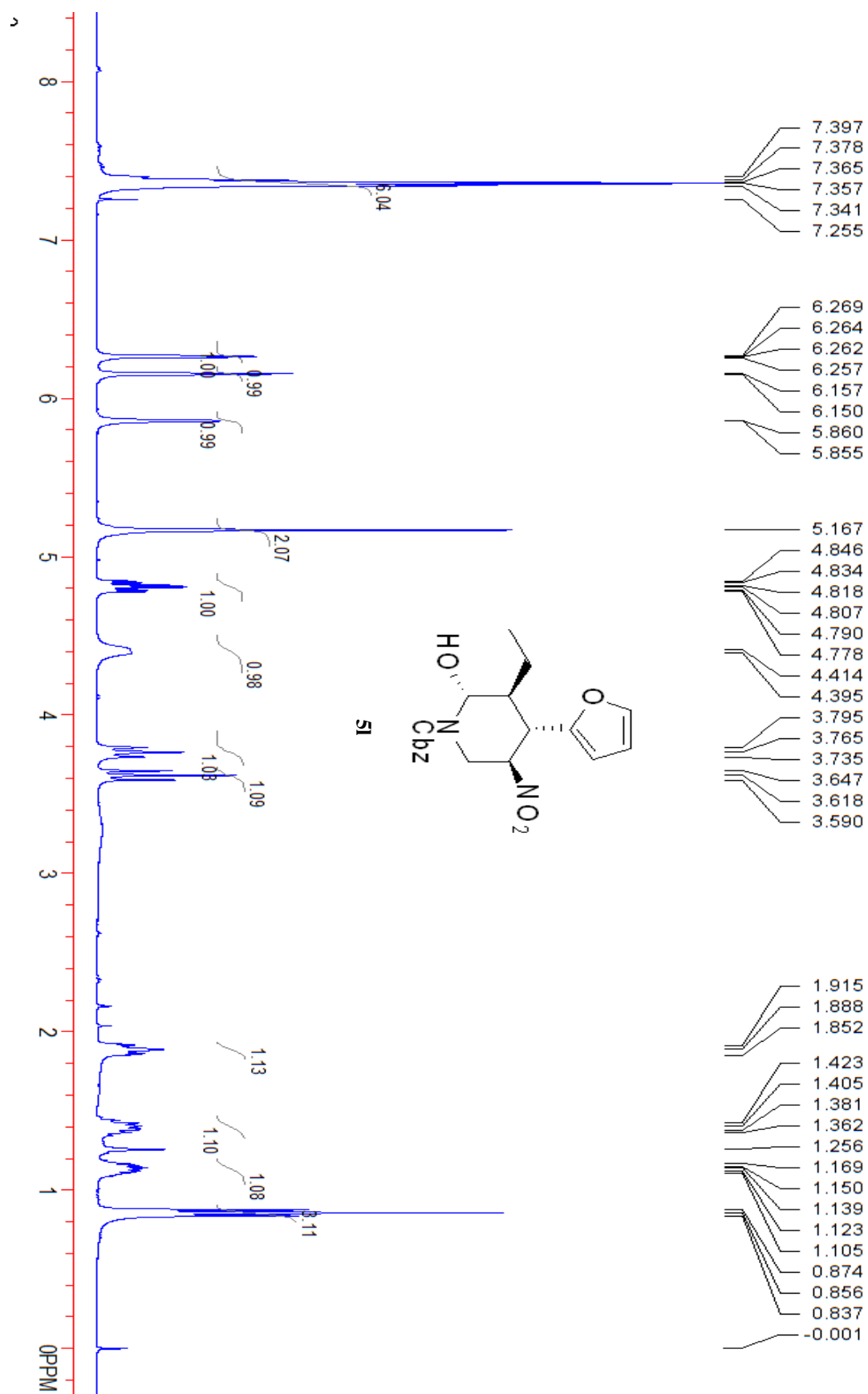
Operator:

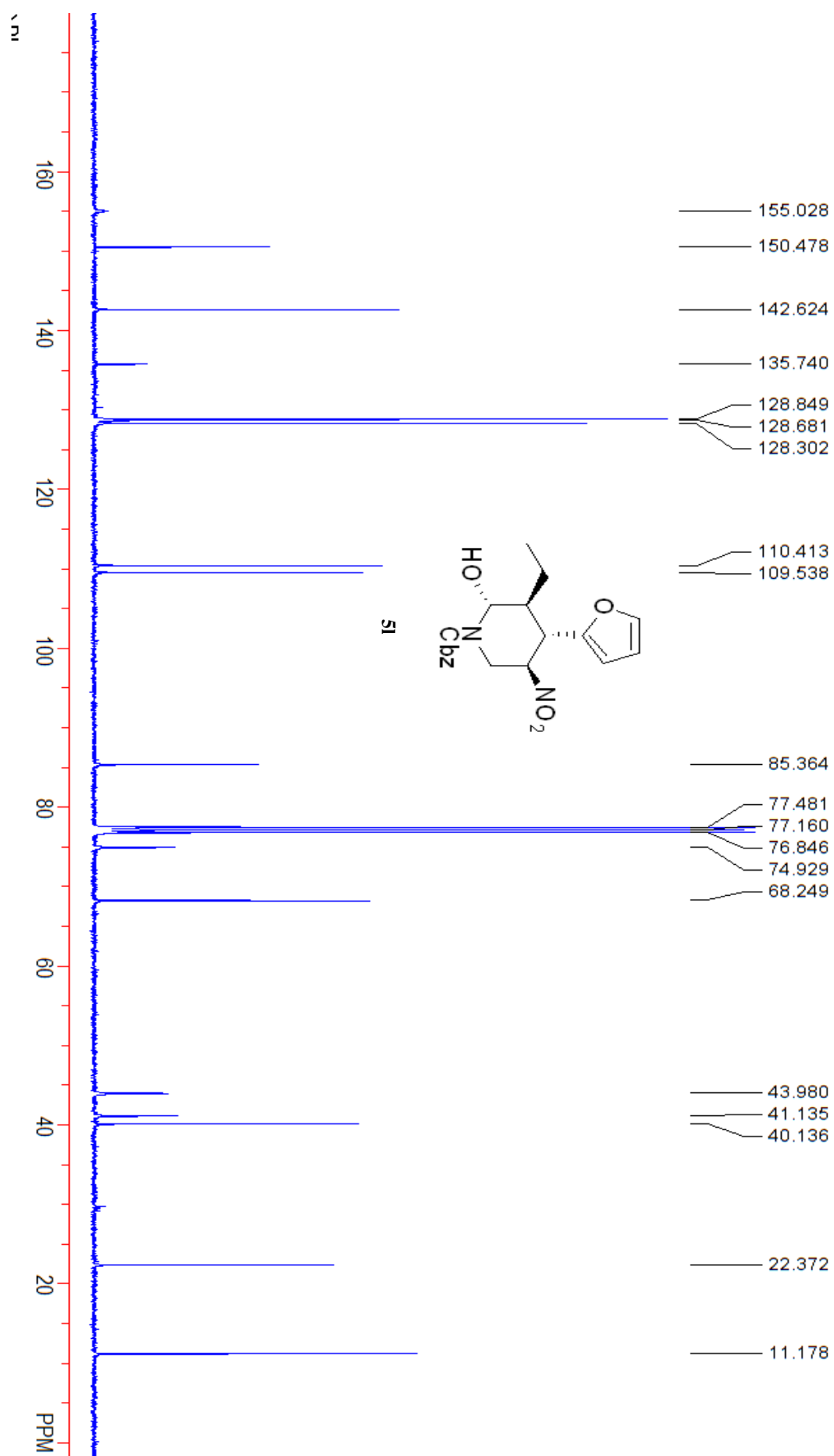
Date:2010-11-23

Time:13:57



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	8.827	246.4	1609.5	0.0489
2	2	16.827	90385.6	3292203.7	99.9511
Total			90632.0	3293813.2	100.0000





HPLC Report

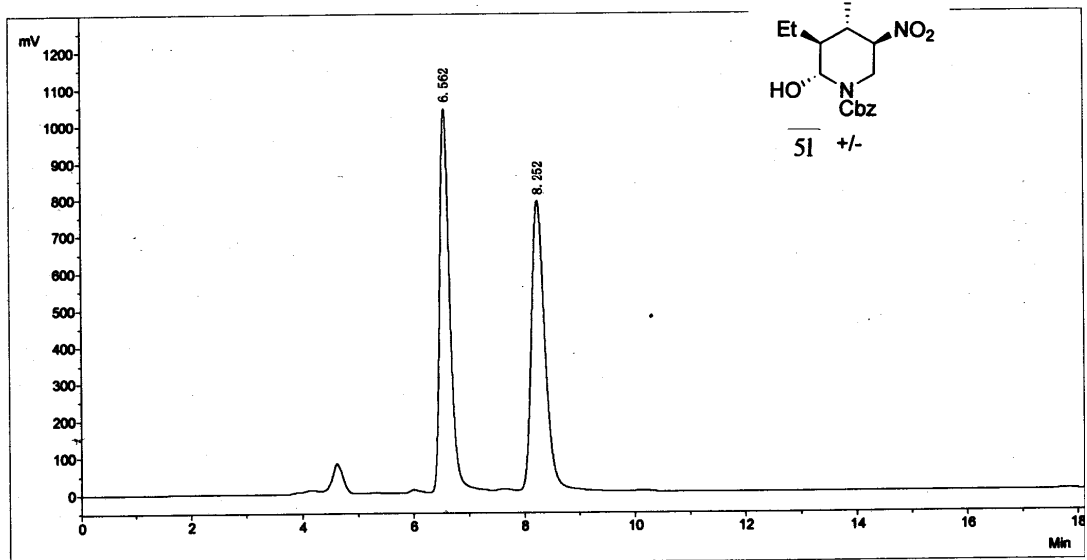
Sample Name:

Data File: ZSL-6-52+- PC-2 73 214 0.7. che

Operator:

Date: 2010-08-20

Time: 09:32



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		6.562	1039130.9	12968537.7	49.4847
2	2		8.252	783219.4	13238615.3	50.5153
Total				1822350.3	26207153.0	100.0000

HPLC Report

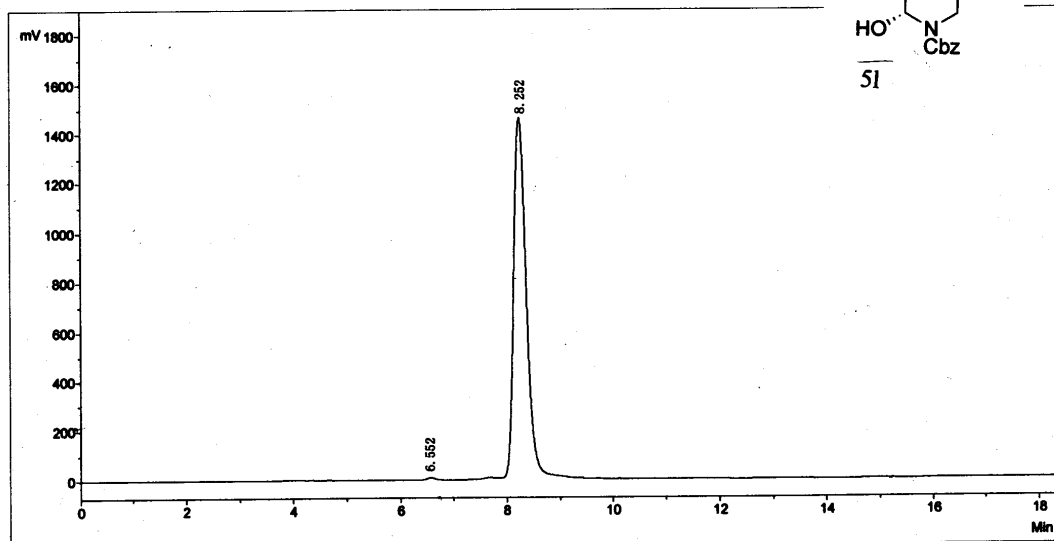
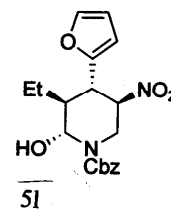
Sample Name:

Data File: ZSL-6-57.che

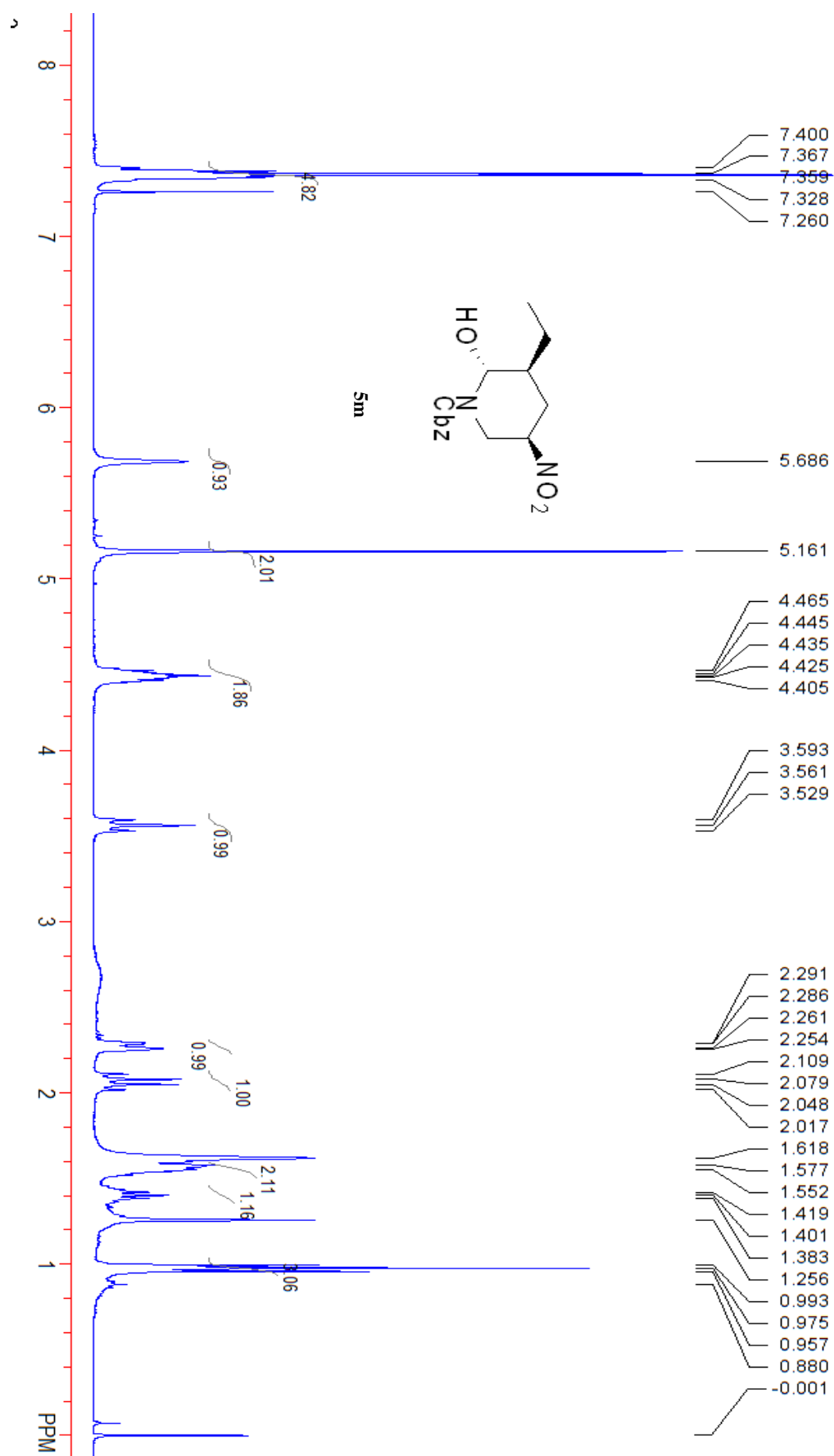
Operator:

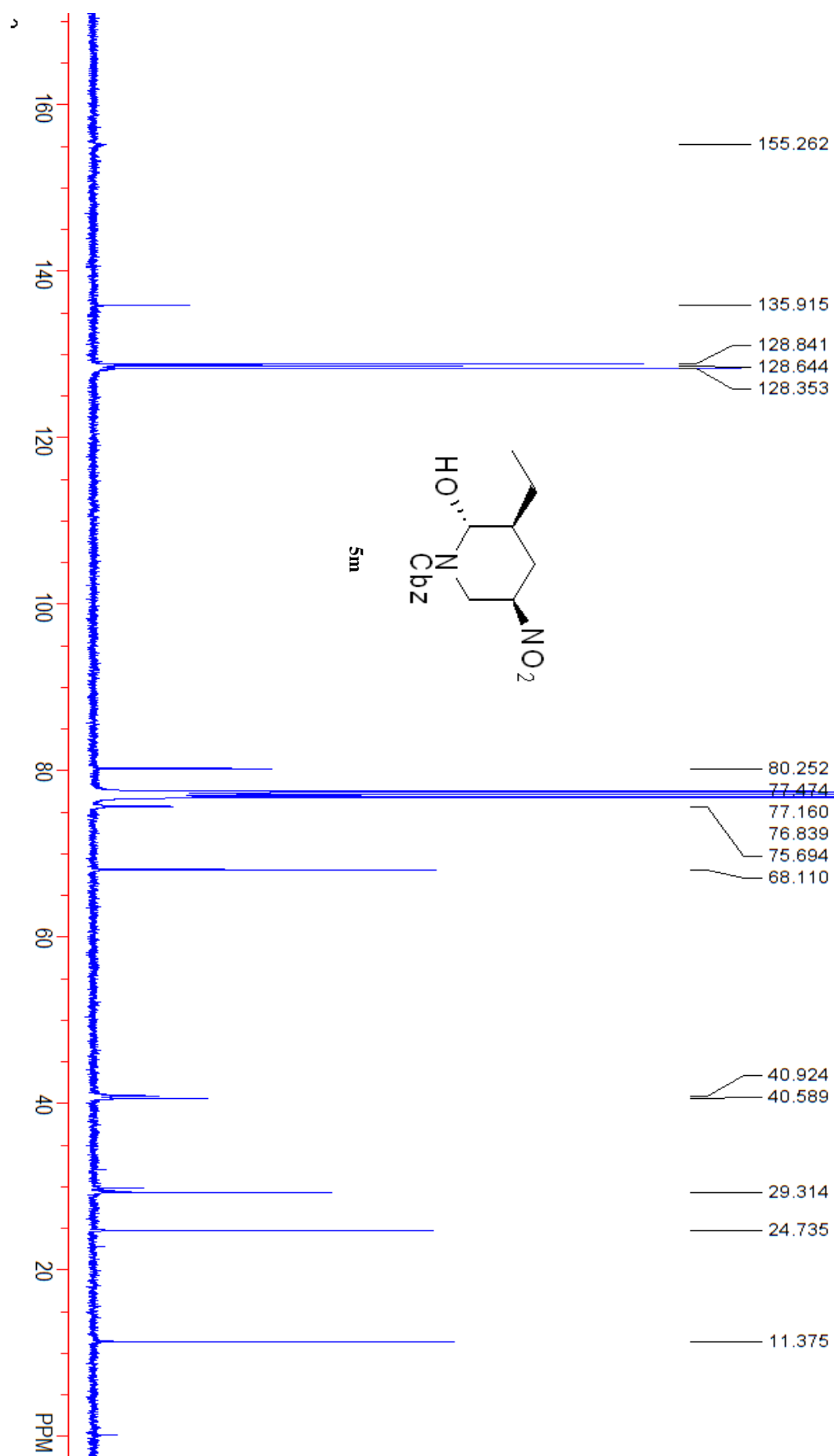
Date: 2010-08-20

Time: 09:53



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		6.552	9232.6	106097.7	0.4436
2	2		8.252	1448408.9	23811995.8	99.5564
Total				1457641.5	23918093.5	100.0000





HPLC Report

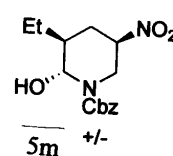
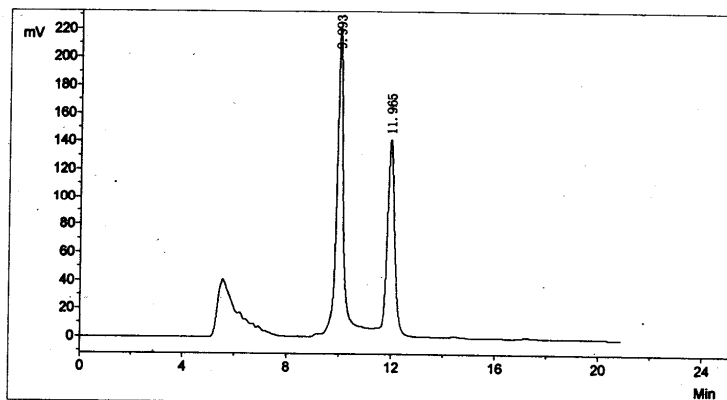
Sample Name:

Data File:ZSL6-75+-PA-2730.5214.che

Operator:

Date:2010-10-13

Time:10:01



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		9.993	215869.9	3477952.2	57.8622
2	2		11.965	136573.4	2532800.5	42.1378
Total				352443.2	6010752.6	100.0000

HPLC Report

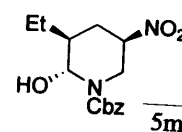
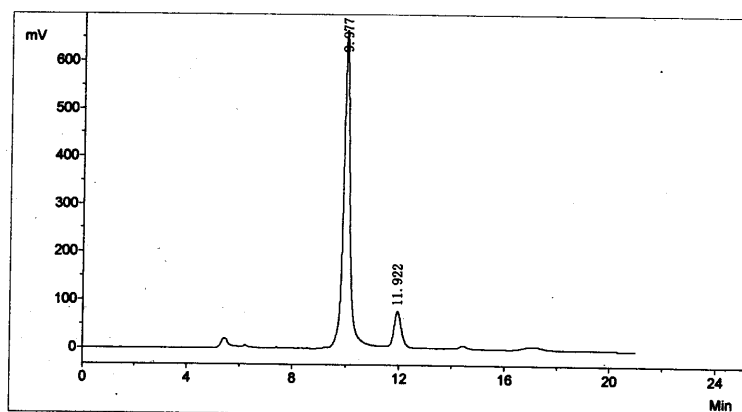
Sample Name:

Data File:ZSL6-78.che

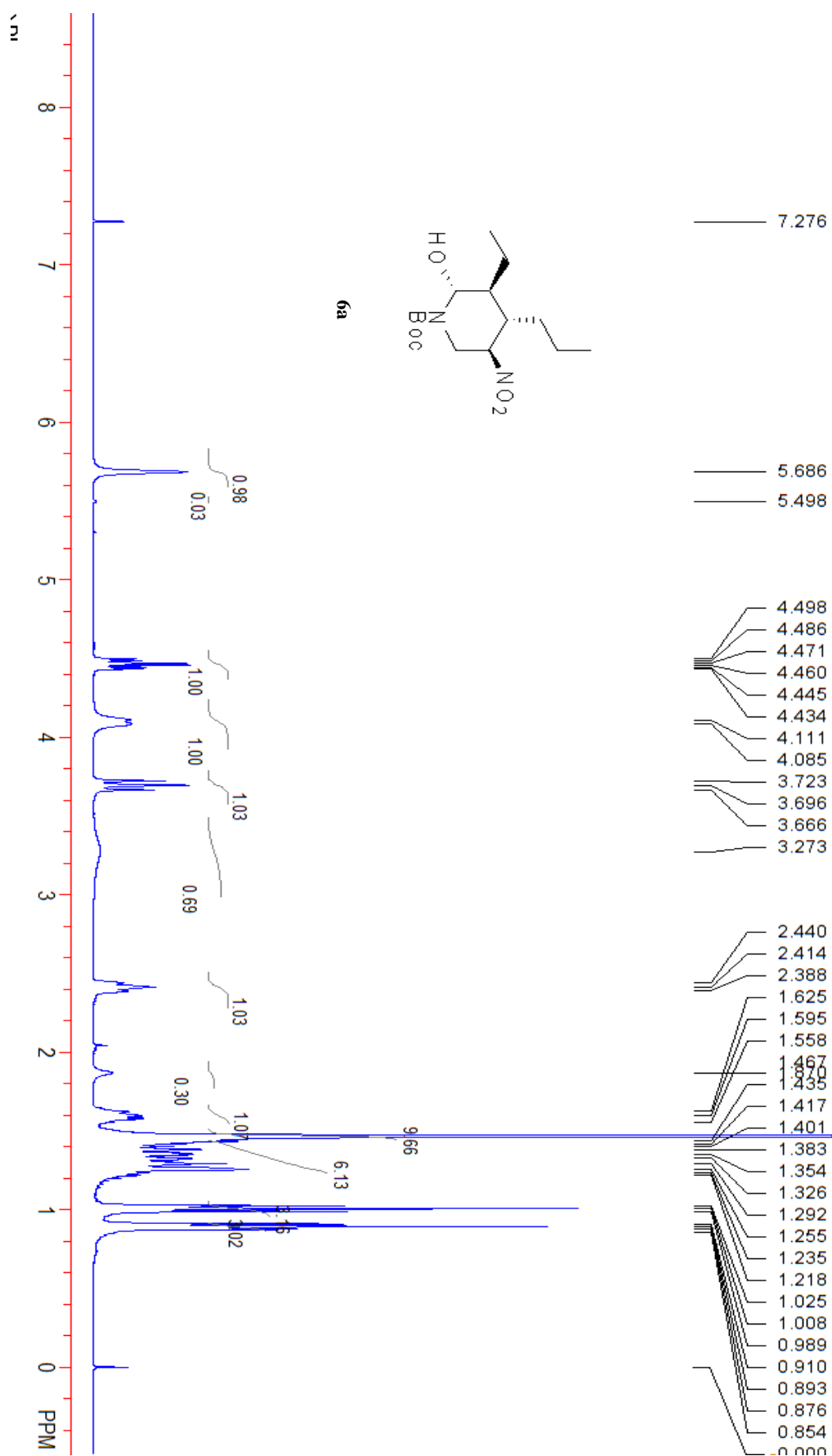
Operator:

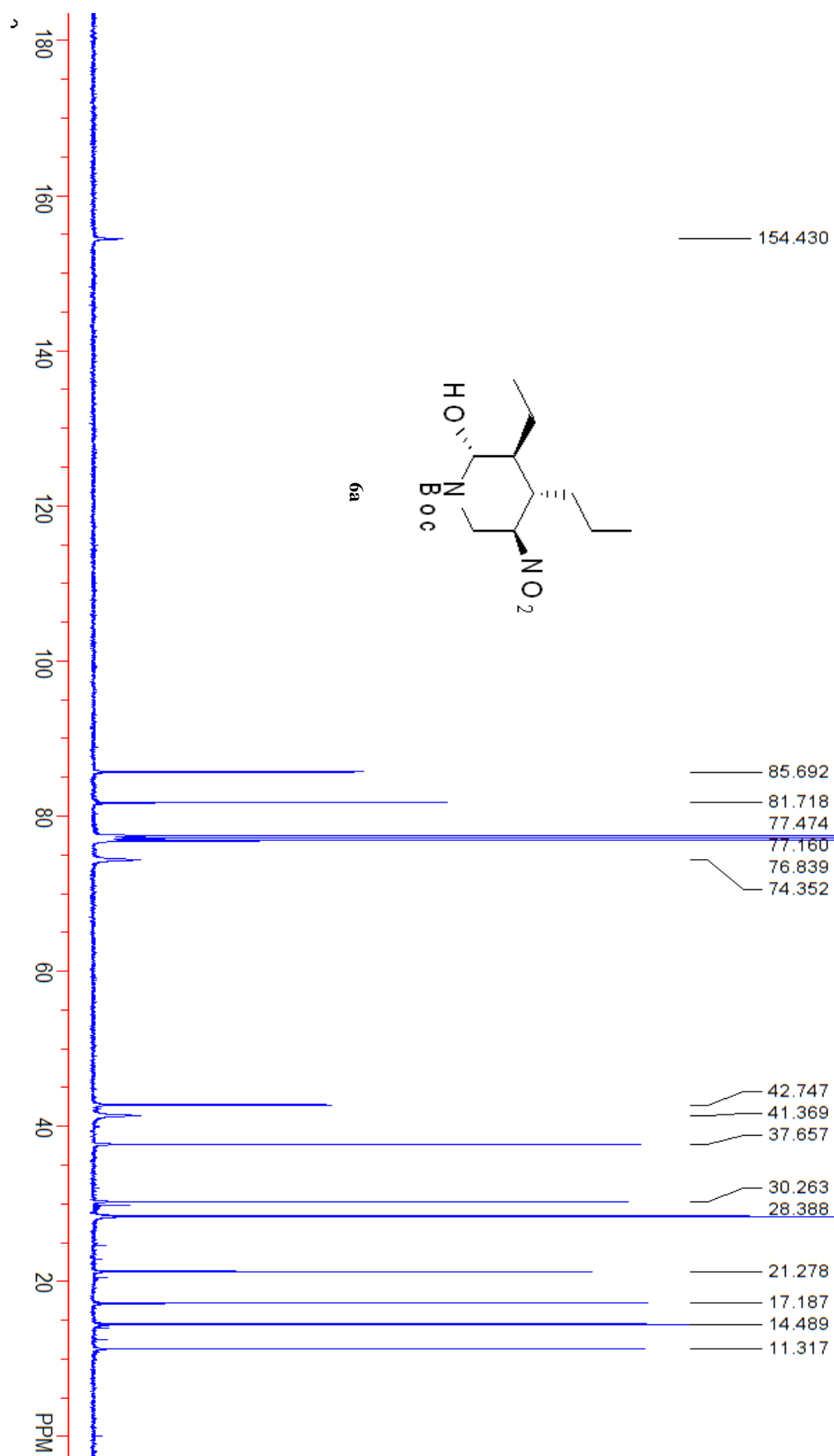
Date:2010-10-13

Time:10:26



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		9.977	659599.3	11319465.2	89.2951
2	2		11.922	71528.3	1357006.8	10.7049
Total				731127.6	12676472.0	100.0000





HPLC REPORT

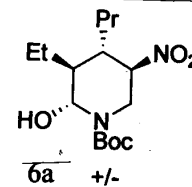
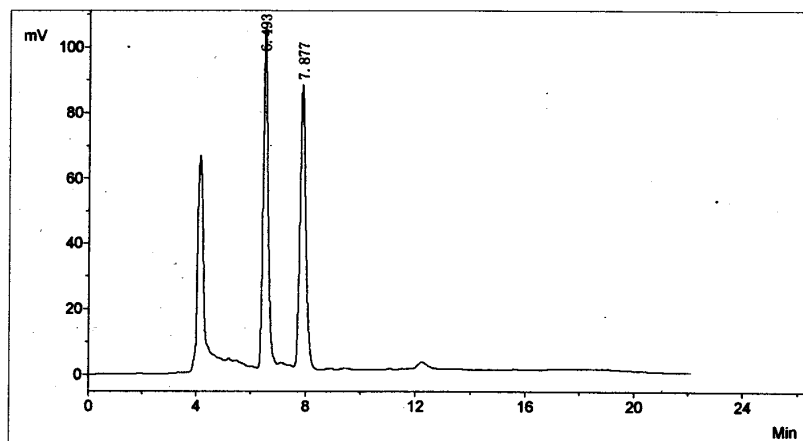
Data File: ZSL6-39-2+-SINO-AD910.7214. che

Sample name:

Date: 2010-08-26

Time : 08:58

Operator:



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		6.493	102897.1	1258809.9	50.0411
2	2		7.877	85796.0	1256741.0	49.9589
Total				188693.0	2515550.9	100.0000

HPLC REPORT

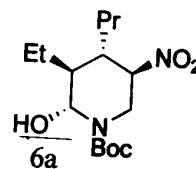
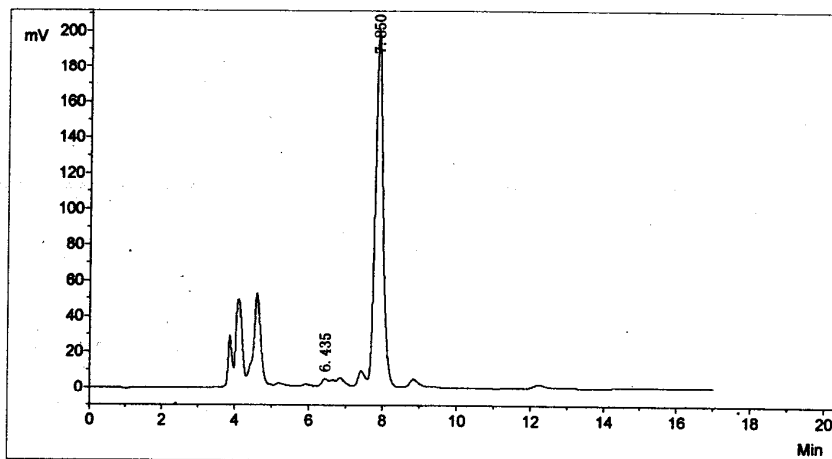
Data File: 6-41-2. che

Sample name:

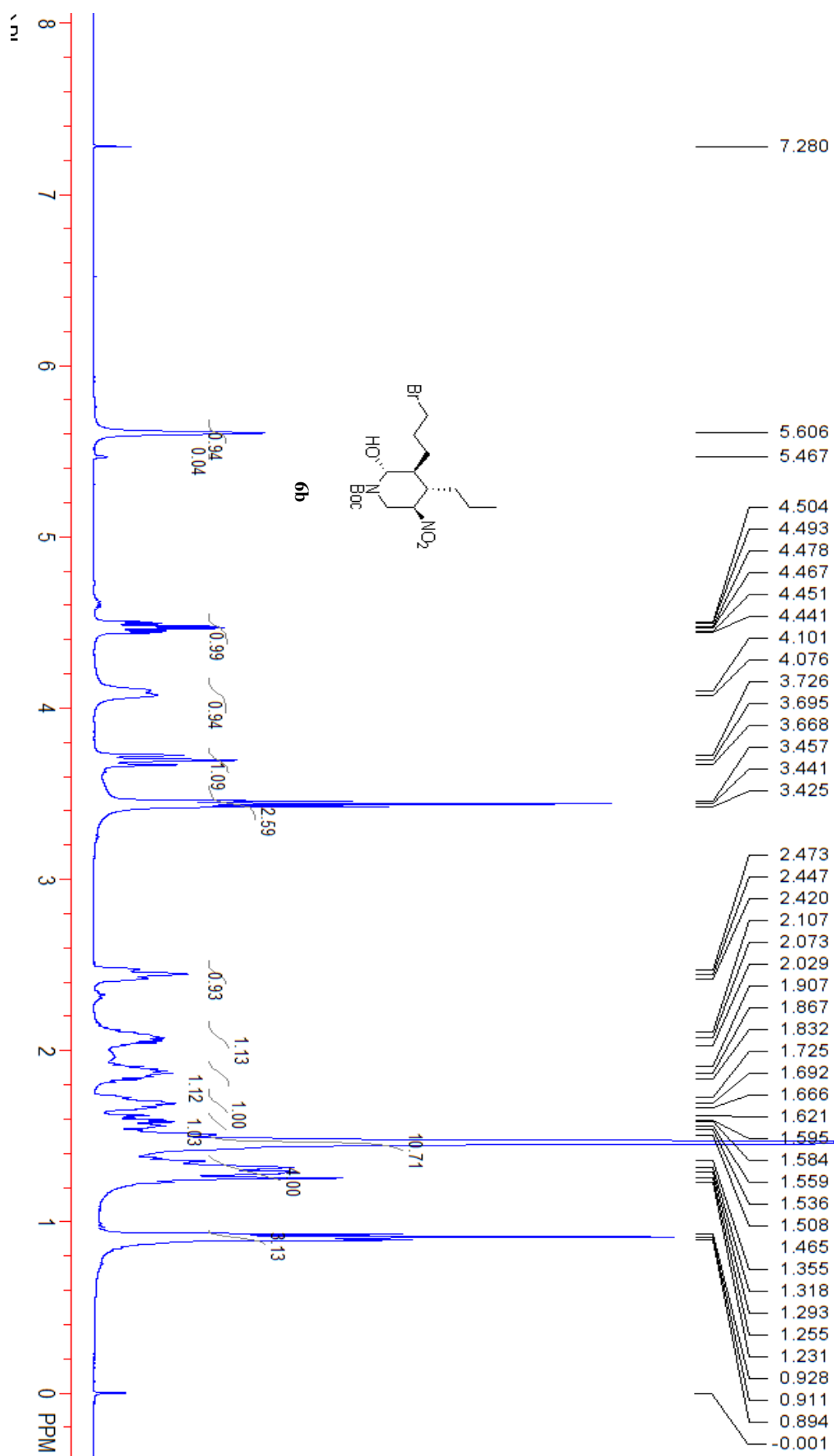
Date: 2010-08-26

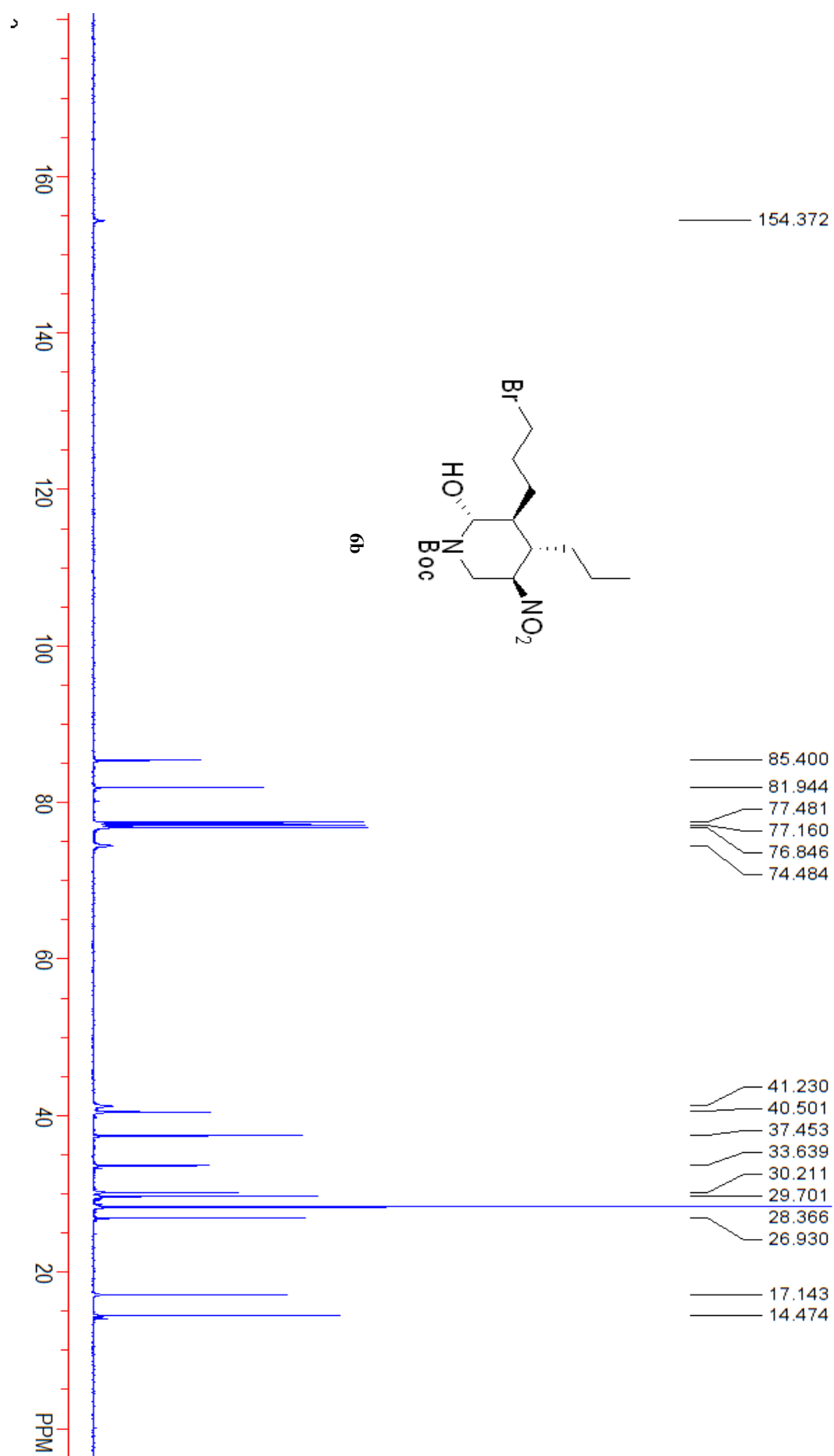
Time : 09:27

Operator:



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		6.435	807.1	3981.3	0.1446
2	2		7.850	196129.3	2748652.6	99.8554
Total				196936.4	2752633.9	100.0000





HPLC REPORT

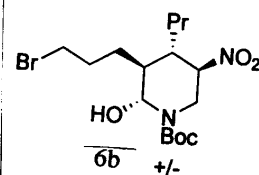
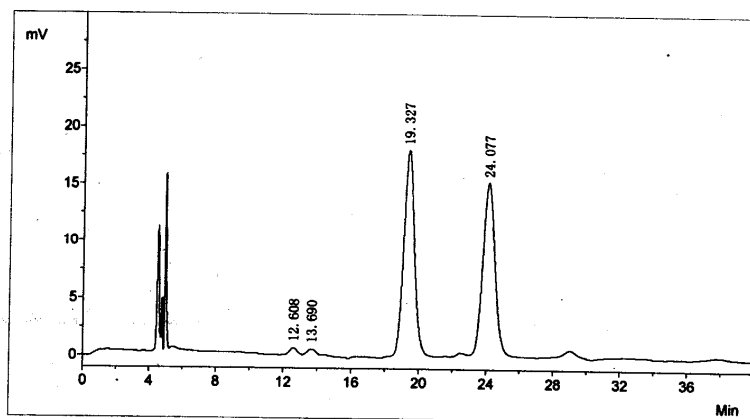
Data File: ZSL-6-44+- IC 982 214 0.7. che

Sample name:

Date: 2010-08-16

Time : 15:56

Operator:



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		12.608	579.8	17664.9	1.1128
2	2		13.690	504.5	16716.8	1.0531
3	3		19.327	18018.8	799430.5	50.3596
4	4		24.077	15054.6	753631.6	47.4745
Total				34157.7	1587443.7	100.0000

HPLC REPORT

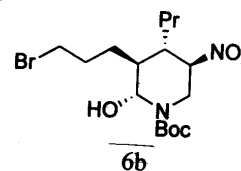
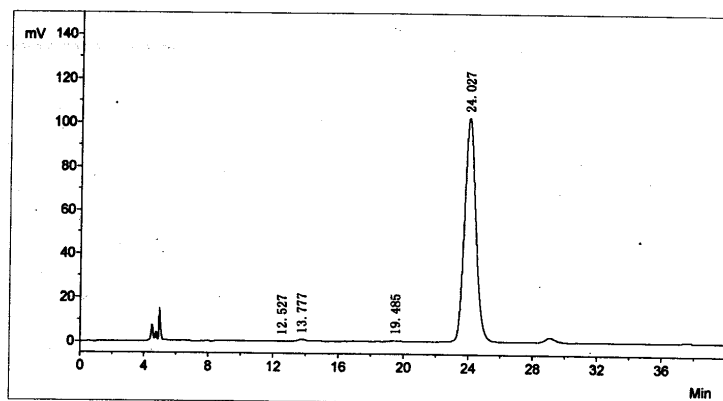
Data File: ZSL-6-42. che

Sample name:

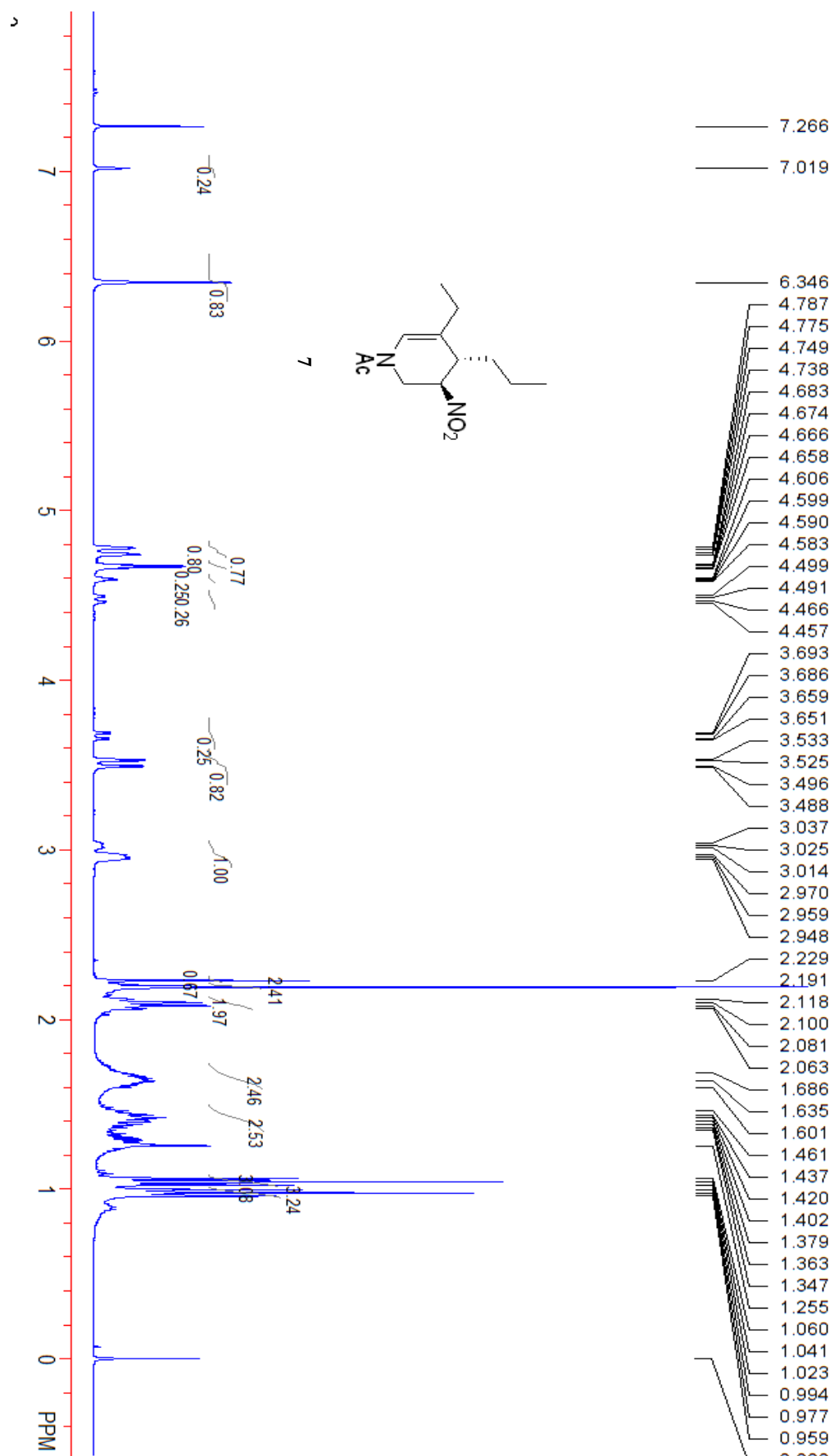
Date: 2010-08-16

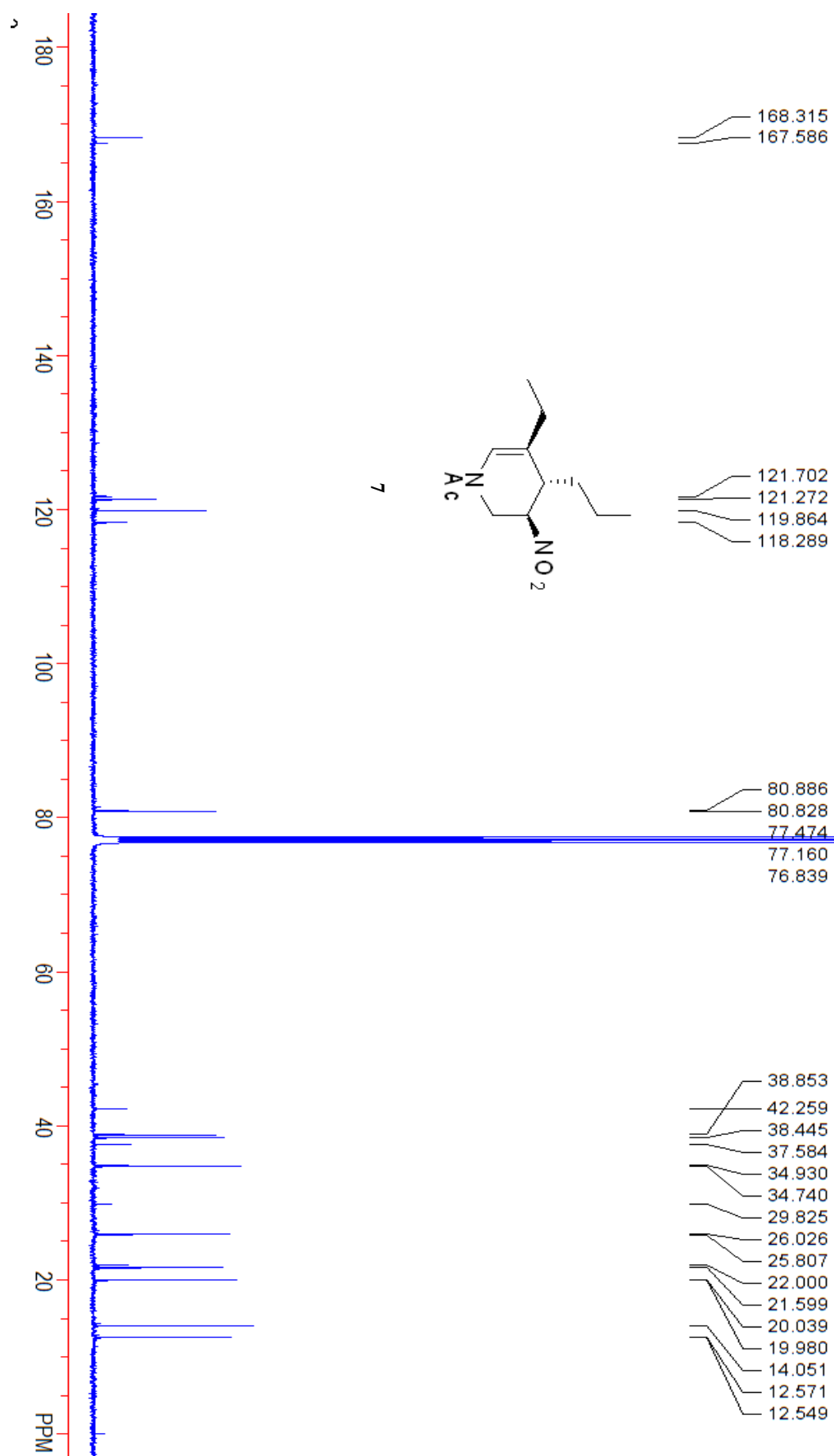
Time : 16:40

Operator:



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		12.527	55.6	1831.4	0.0367
2	2		13.777	817.9	24950.0	0.4996
3	3		19.485	340.9	19602.6	0.3925
4	4		24.027	101902.7	4947892.8	99.0713
Total				103117.1	4994276.8	100.0000





HPLC Report

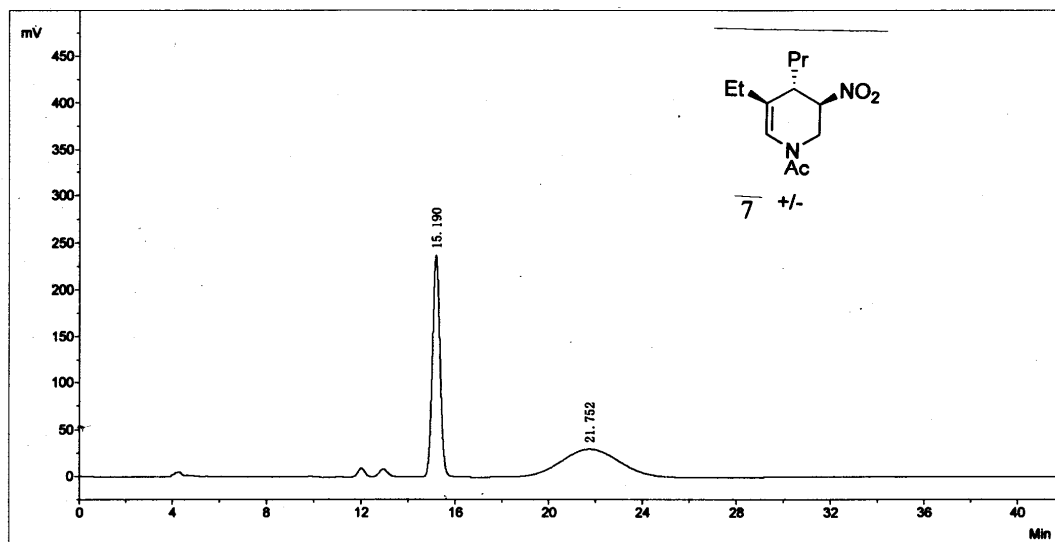
Sample Name:

Data File: ZSL-6-45-0+- PA-2 73 214 0.7. che

Operator:

Date: 2010-08-20

Time: 13:53



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		15.190	237616.4	5339516.5	50.7161
2	2		21.752	30110.3	5188736.6	49.2839
Total				267726.7	10528253.1	100.0000

HPLC Report

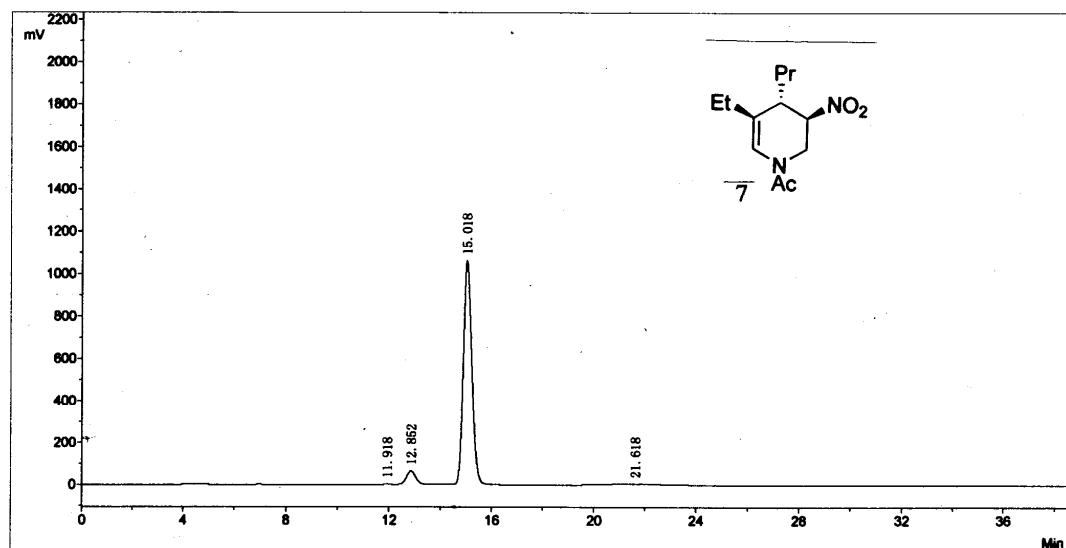
Sample Name:

Data File:ZSL-6-49-0..che

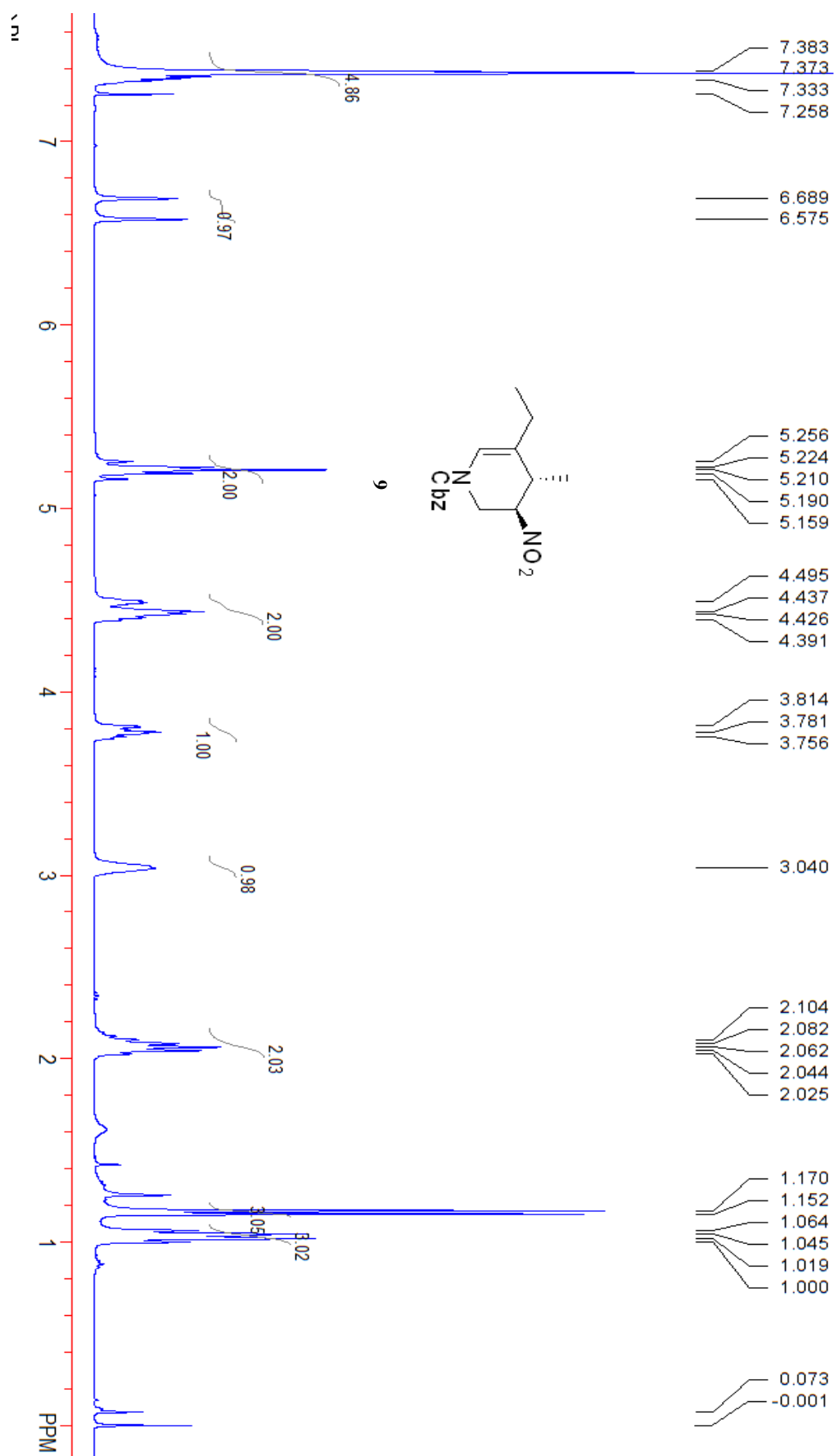
Operator:

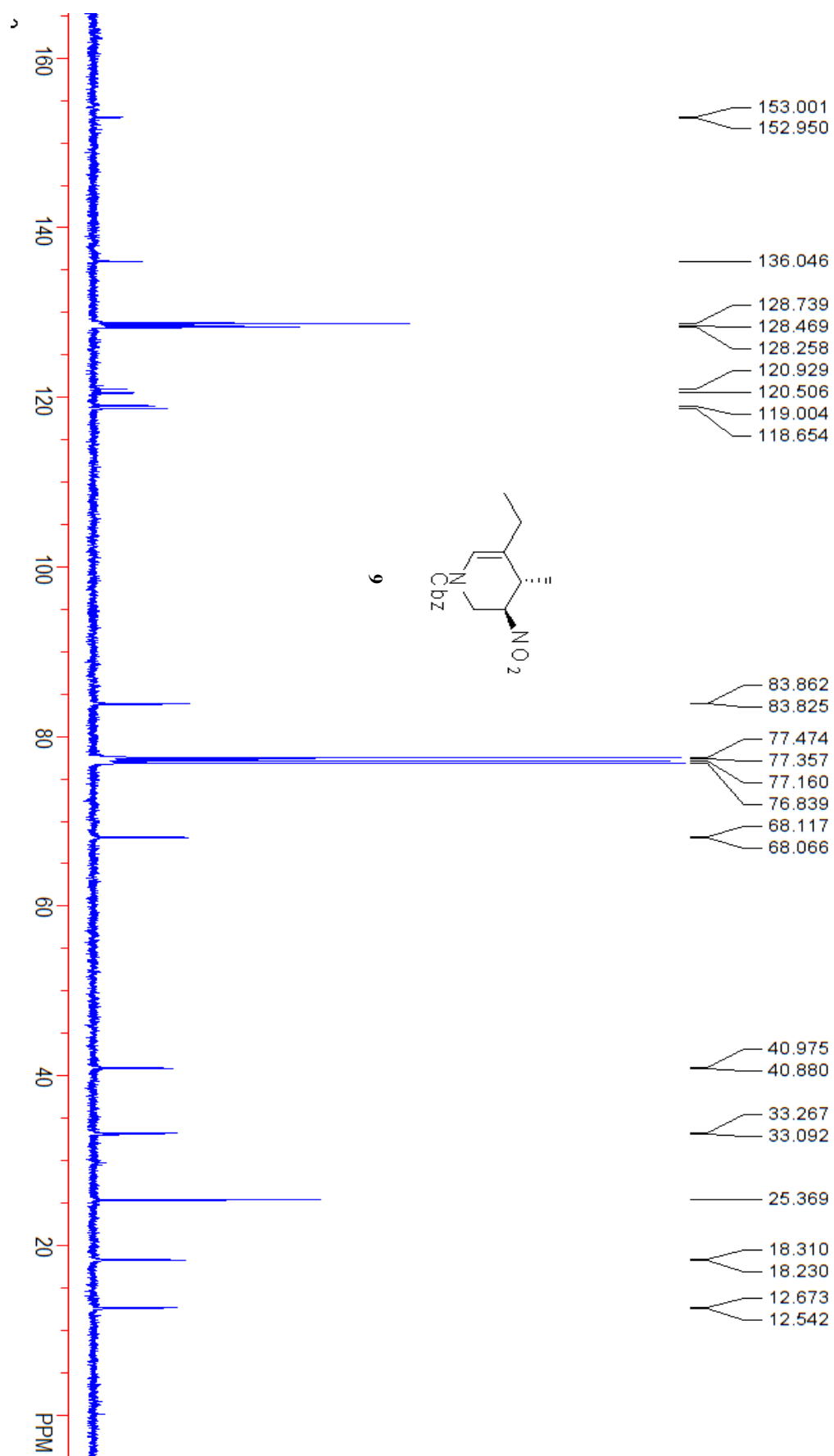
Date:2010-08-20

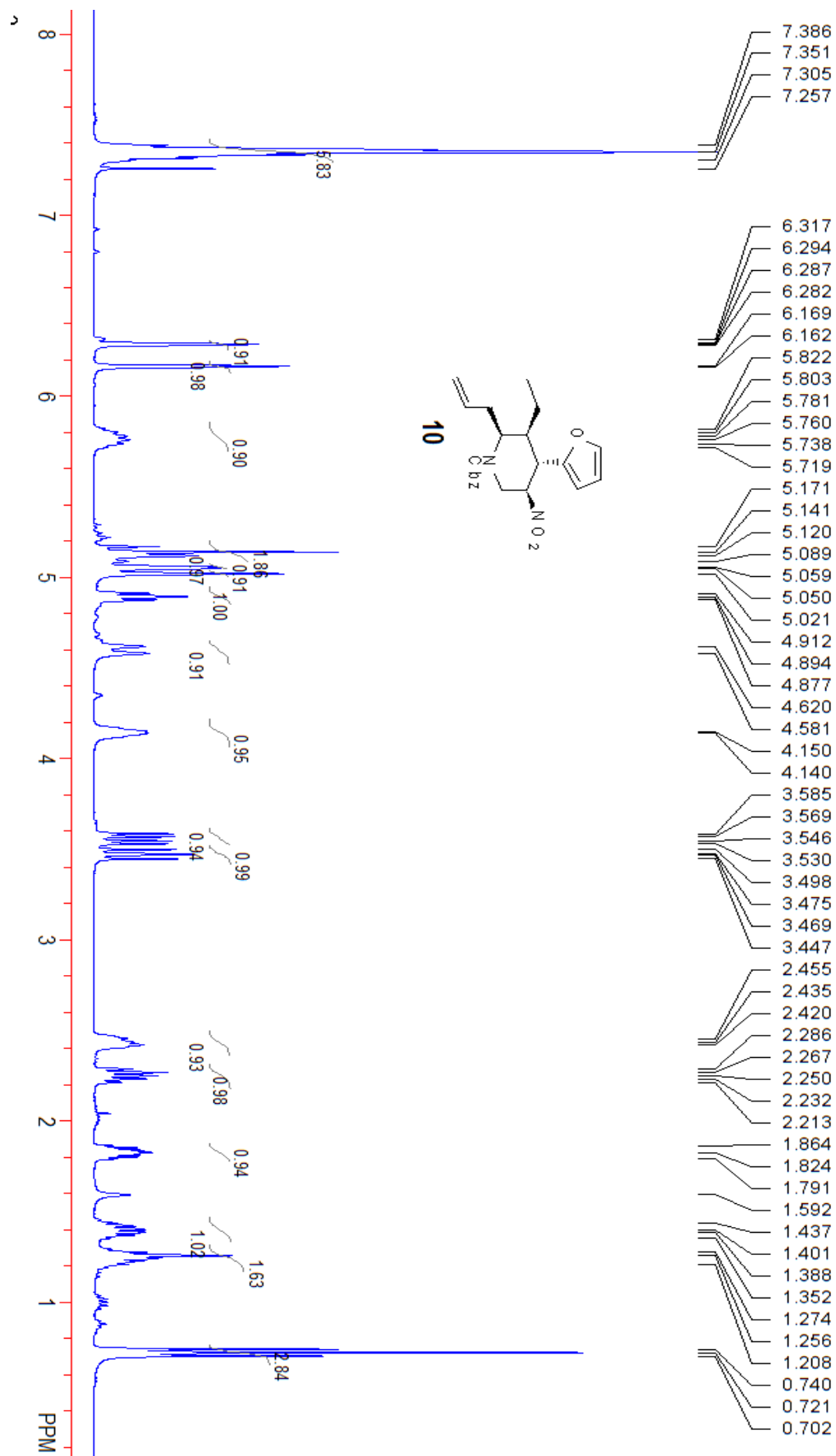
Time:14:37

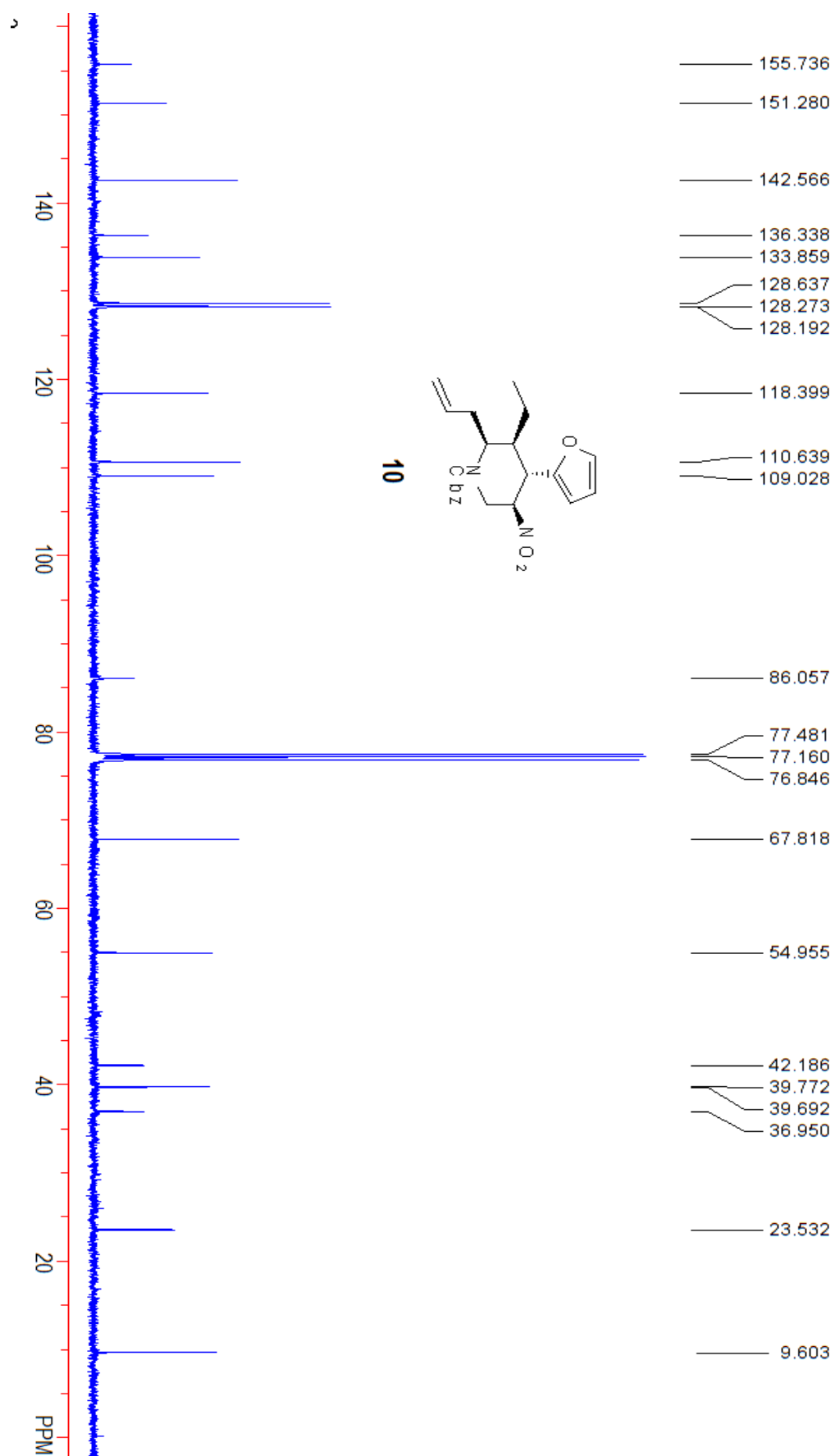


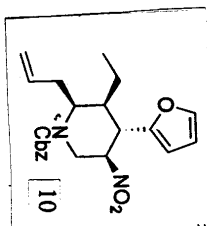
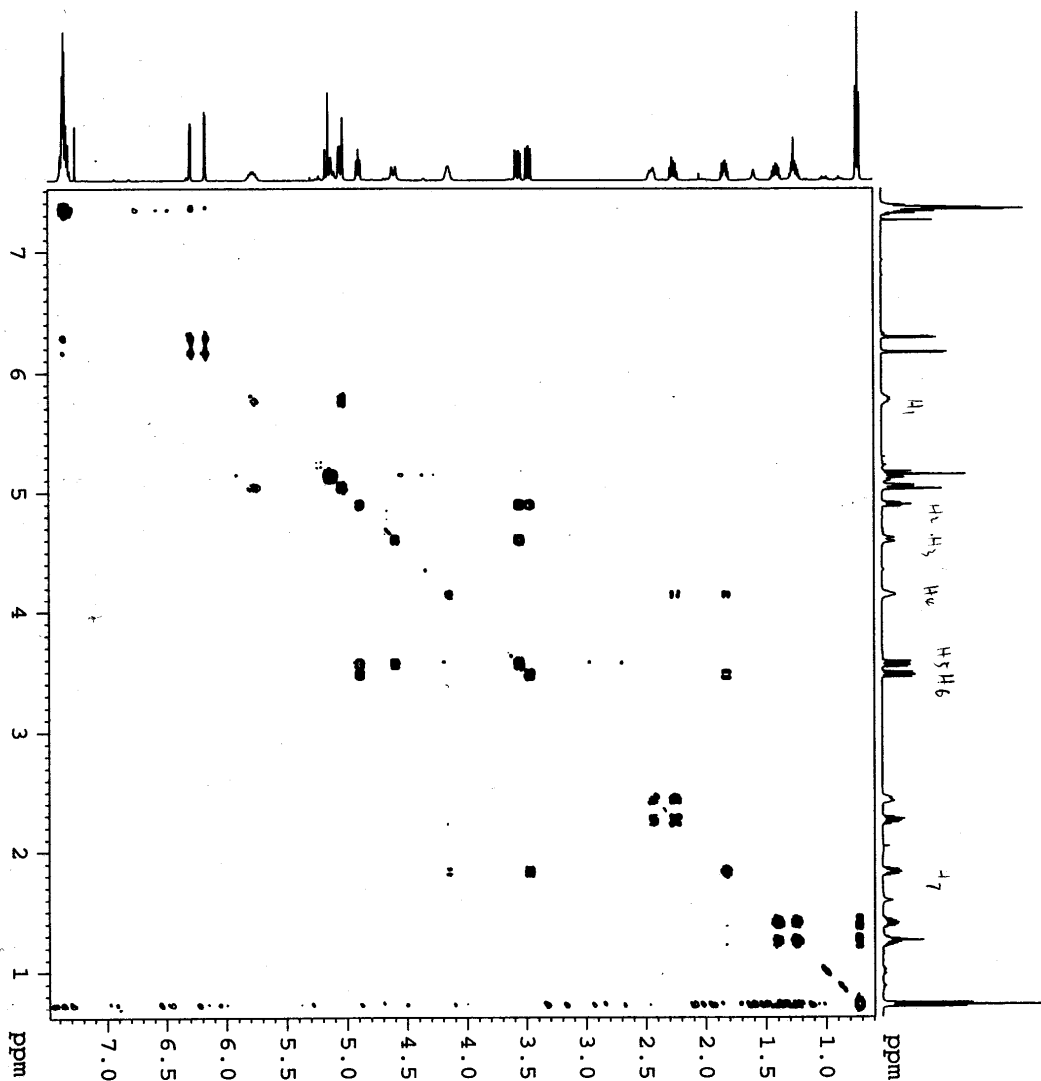
No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		11.918	1116.1	22220.3	0.0819
2	2		12.852	64588.6	1637058.0	6.0356
3	3		15.018	1061127.9	24532317.4	90.4475
4	4		21.618	5505.1	931660.9	3.4349
Total				1132337.8	27123256.6	100.0000



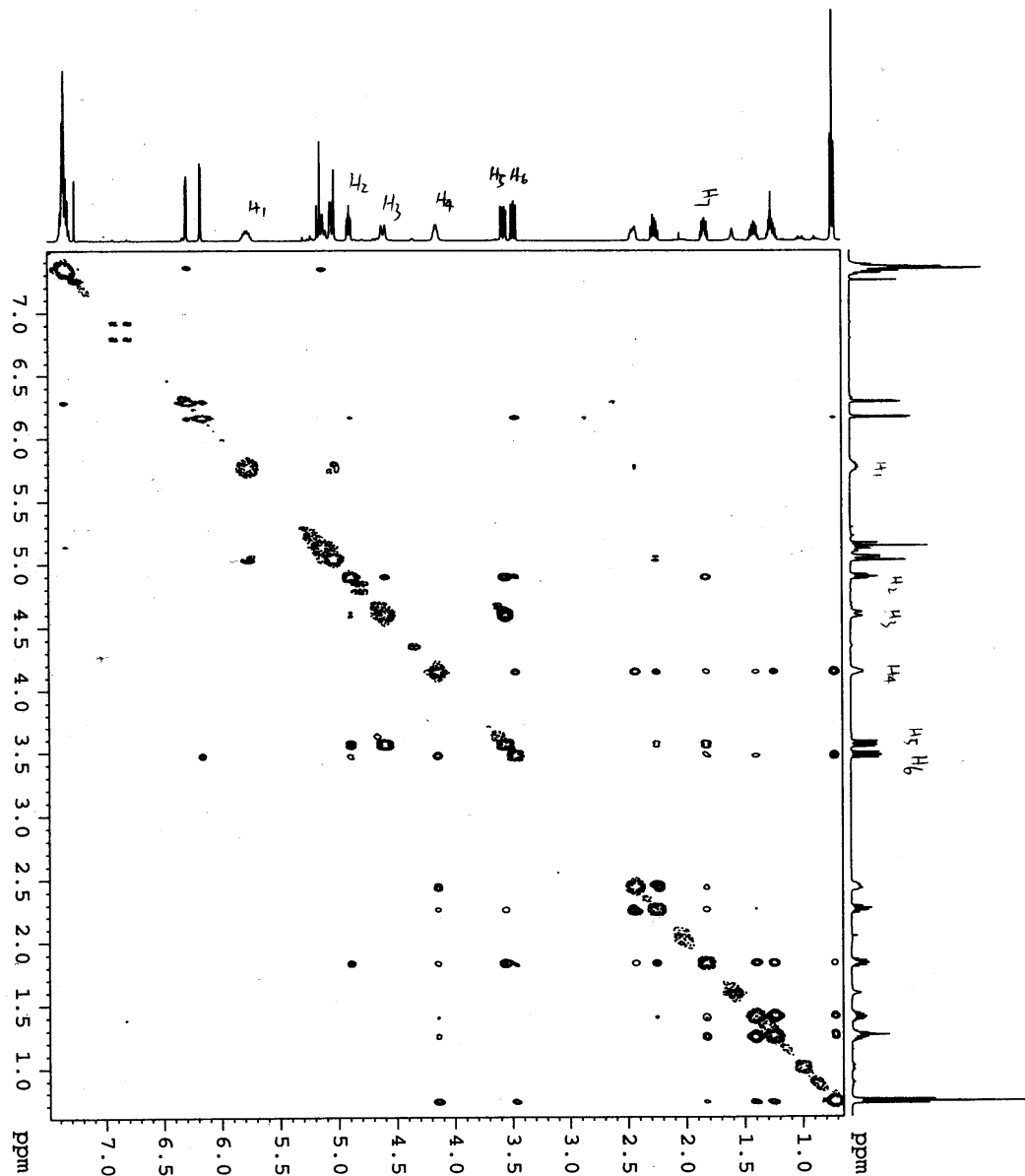








Current Data Parameters
 NAME 2010039com-6-95
 EXPTNO 1
 PROCNO 1
 F2 - Acquisition Parameters
 Date_ 20101123
 Time 13:48
 INSTRUM spect
 PROBD 5 mm QNP 1H/13
 PULPROG cosygmzgpg
 TD 65536
 FIDRES 0.139200 Hz
 AQ 0.0000000 sec
 DE 126.000 usec
 DB 6.50 usec
 TS 300.0 K
 DO 0.00000300 sec
 D1 2.00000000 sec
 D13 0.00000000 sec
 D16 0.00000000 sec
 INO 0.0002500 sec
 ===== CHANNEL f1 =====
 NUC1 1H
 P1 10.50 usec
 PL1 4.00 dB
 SFO1 500.1318950 MHz
 ===== GRADIENT CHANNEL =====
 GPMAX1 SINE.100
 GPMAX2 SINE.100
 GPMAX3 SINE.100
 GPZ1 15.00 %
 GPZ2 15.00 %
 GPZ3 40.00 %
 P16 1000.00 usec
 F1 - Acquisition parameters
 MH0 1
 SFO1 500.1319 MHz
 FIDRES 10.046212 Hz
 SW 7.934 ppm
 FPMODE QF
 F2 - Processing parameters
 SI 32768
 SF 500.1300159 MHz
 WDM SINE
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.40
 F1 - Processing parameters
 SI 1024
 OF 1300159 MHz
 SINE 0.00 Hz
 0



Current Data Parameters
NAME 2010035noe-6-95
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20101123
Time 11:13
INSTRUM spect
PROBHD 5 mm QNP 1H/13
PULPROG zgpg30
SOLVENT noesyspph
NS 2
DS 2
SWH 398.224 Hz
FIDRES 3.87241 Hz
AQ 0.129200 sec
RG 128
DW 126.000 usec
DE 18.30 usec
TE 300.2 K
D0 0.001263 sec
D1 2.00000000 sec
D16 0.00120000 sec
Z 0.001200 sec
TMO 0.0002500 sec
SFO 286
SFOCWT 0.4988001 sec
TNU

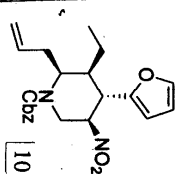
===== CHANNEL f1 =====
NUC1 1H
P1 10.50 usec
P2 21.00 usec
PL 0.00 dB
SFO1 500.131850 MHz

===== GRADIENT CHANNEL =====
GPM1 SINE 100
GPM2 SINE 100
OP1 40.00 %
OP2 -40.00 %
P15 1000.00 usec

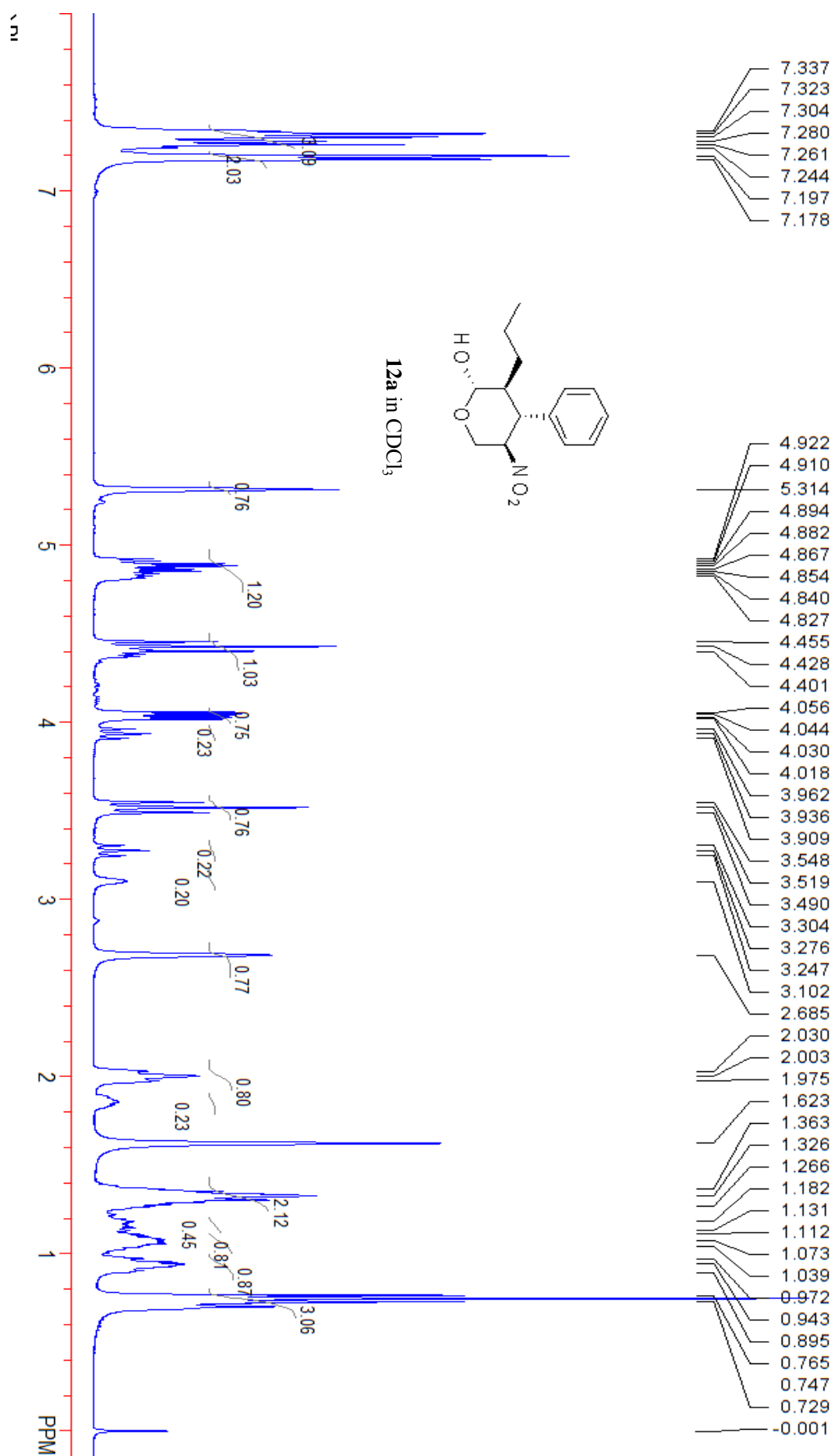
F1 - Acquisition Parameters
MD 1
TD 280
SFO1 500.1319 MHz
SF 500.1319 MHz
FIDRES 14.17216 Hz
SFO2 125.76115 MHz
P15002 States-tppl

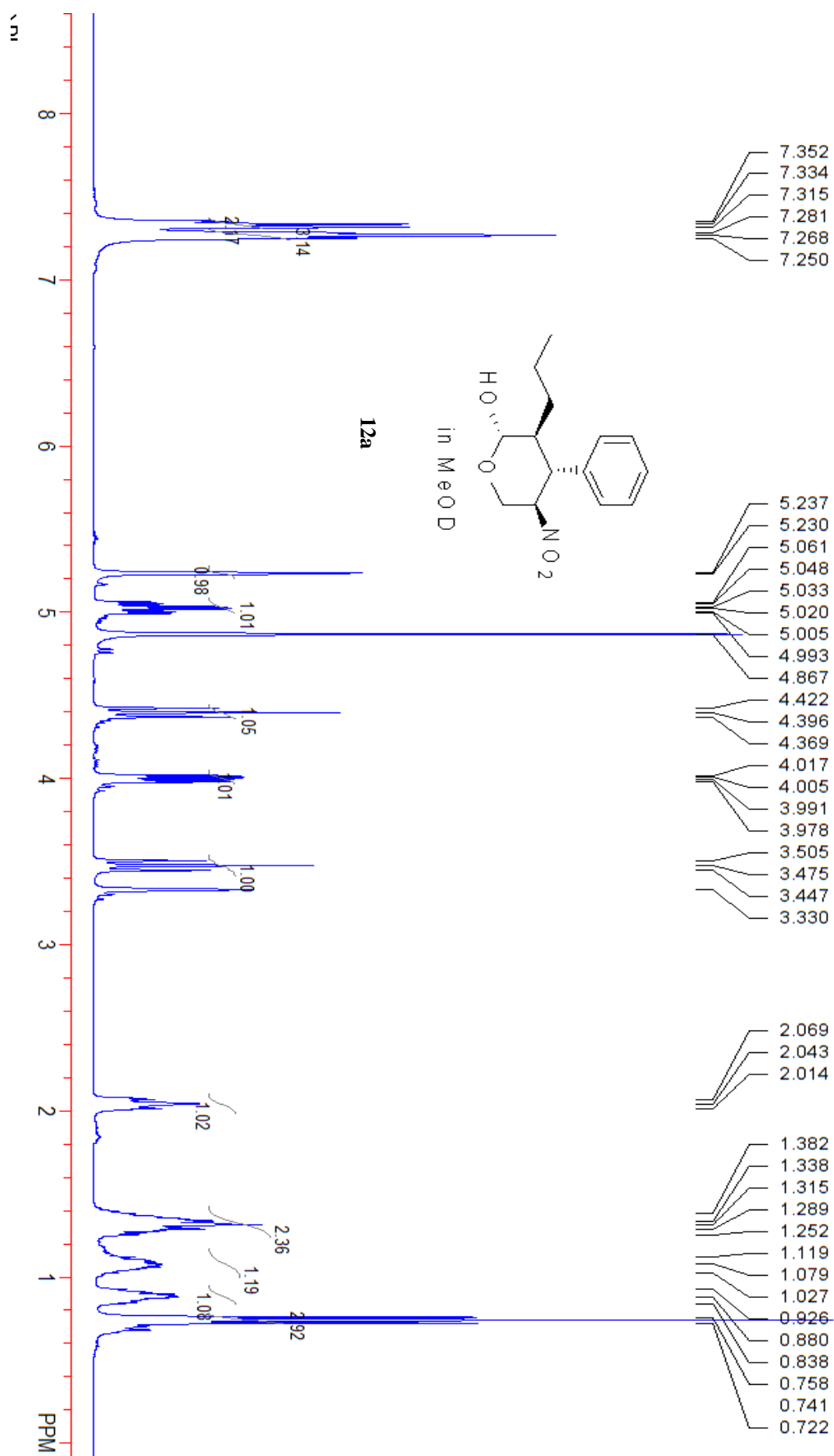
F2 - Processing parameters
SI 32768
SF 500.1300159 MHz
WDW 500.1300159 MHz
SSB 2
LB 0.00 Hz
GB 0.00 Hz
PC 1.40

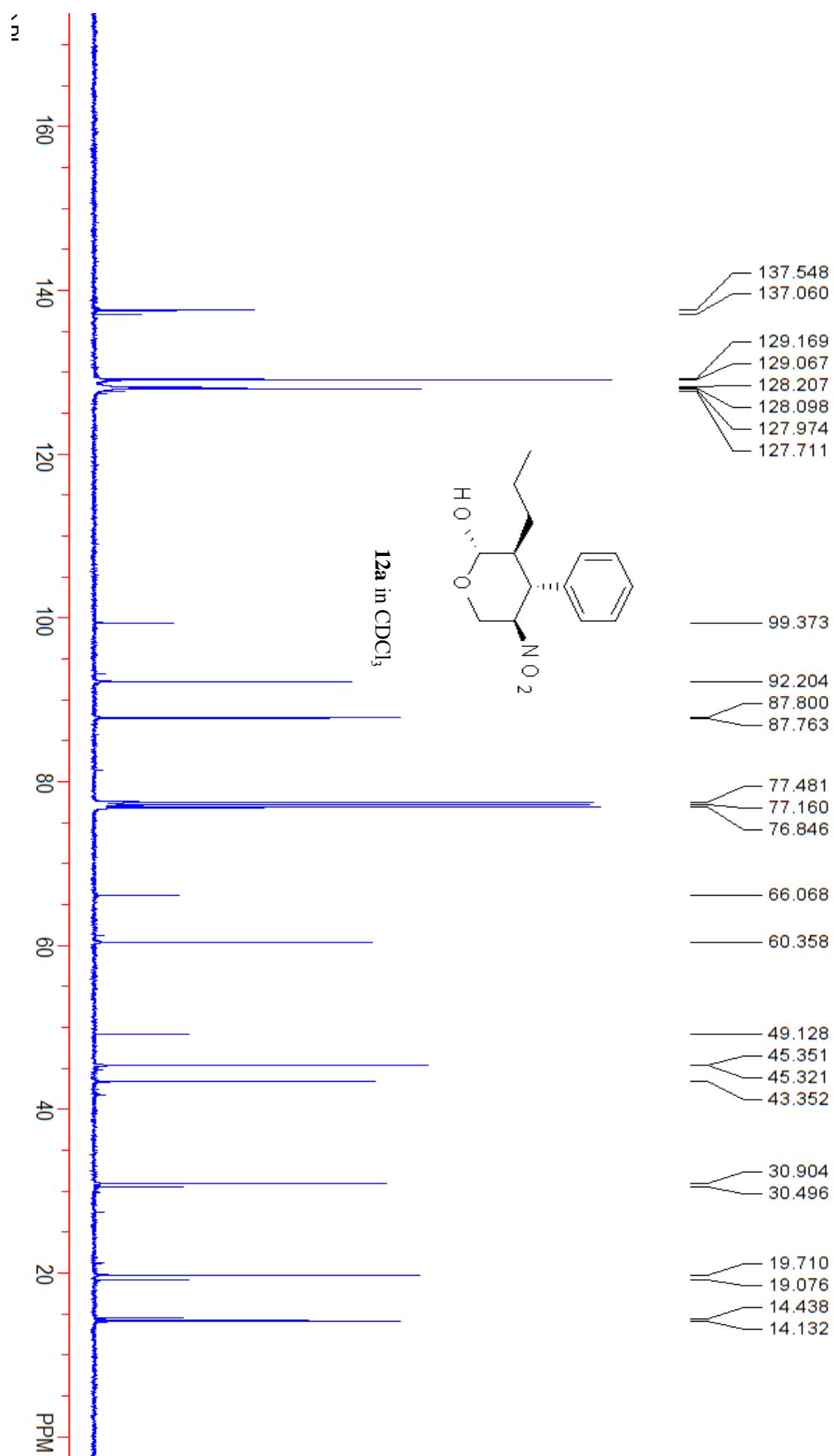
F1 - Processing parameters
SI 1024
SF 500.1300159 MHz
WDW 500.1300159 MHz
SSB 2
LB 0.00 Hz
GB 0.00 Hz
PC 1.40

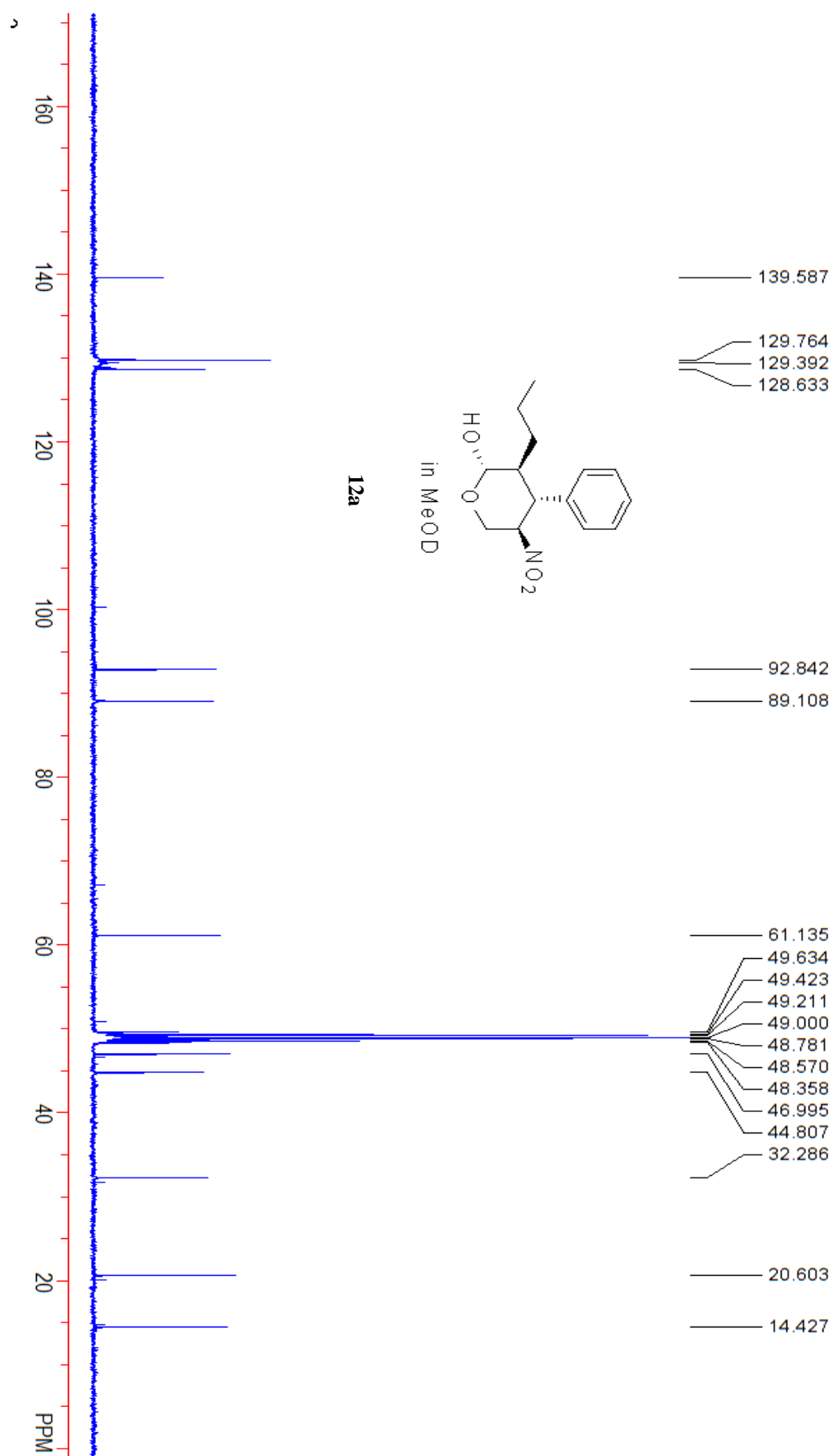


10









HPLC Report

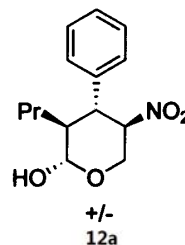
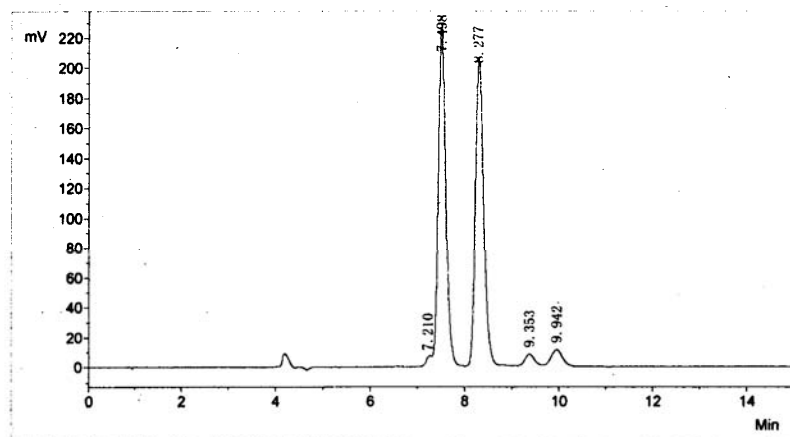
Sample Name:

Data File: ZSL-6-56+- IC 91 214 0.7. che

Operator:

Date: 2010-09-09

Time: 14:49



No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	7.210	89.0	29.2	0.0006
2	2	7.498	217720.6	2319712.6	45.5464
3	3	8.277	198141.8	2522132.3	49.5208
4	4	9.353	7290.2	92011.1	1.8066
5	5	9.942	10316.2	159188.7	3.1256
Total			433557.8	5093073.9	100.0000

HPLC Report

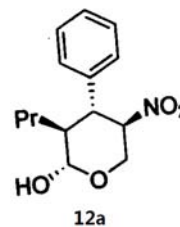
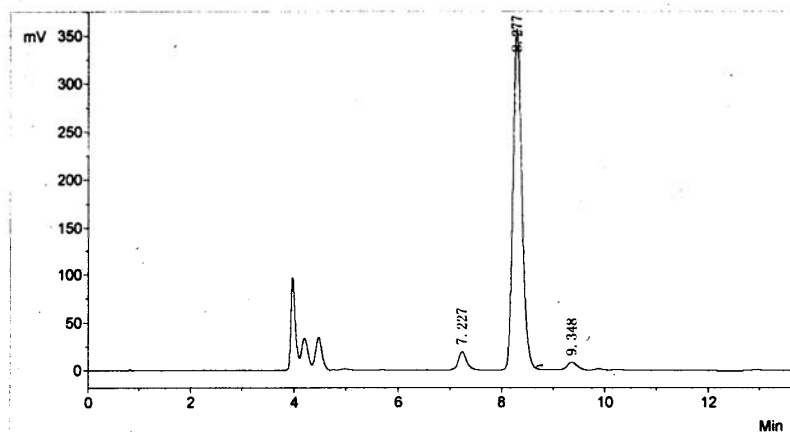
Sample Name:

Data File: ZSL-6-54. che

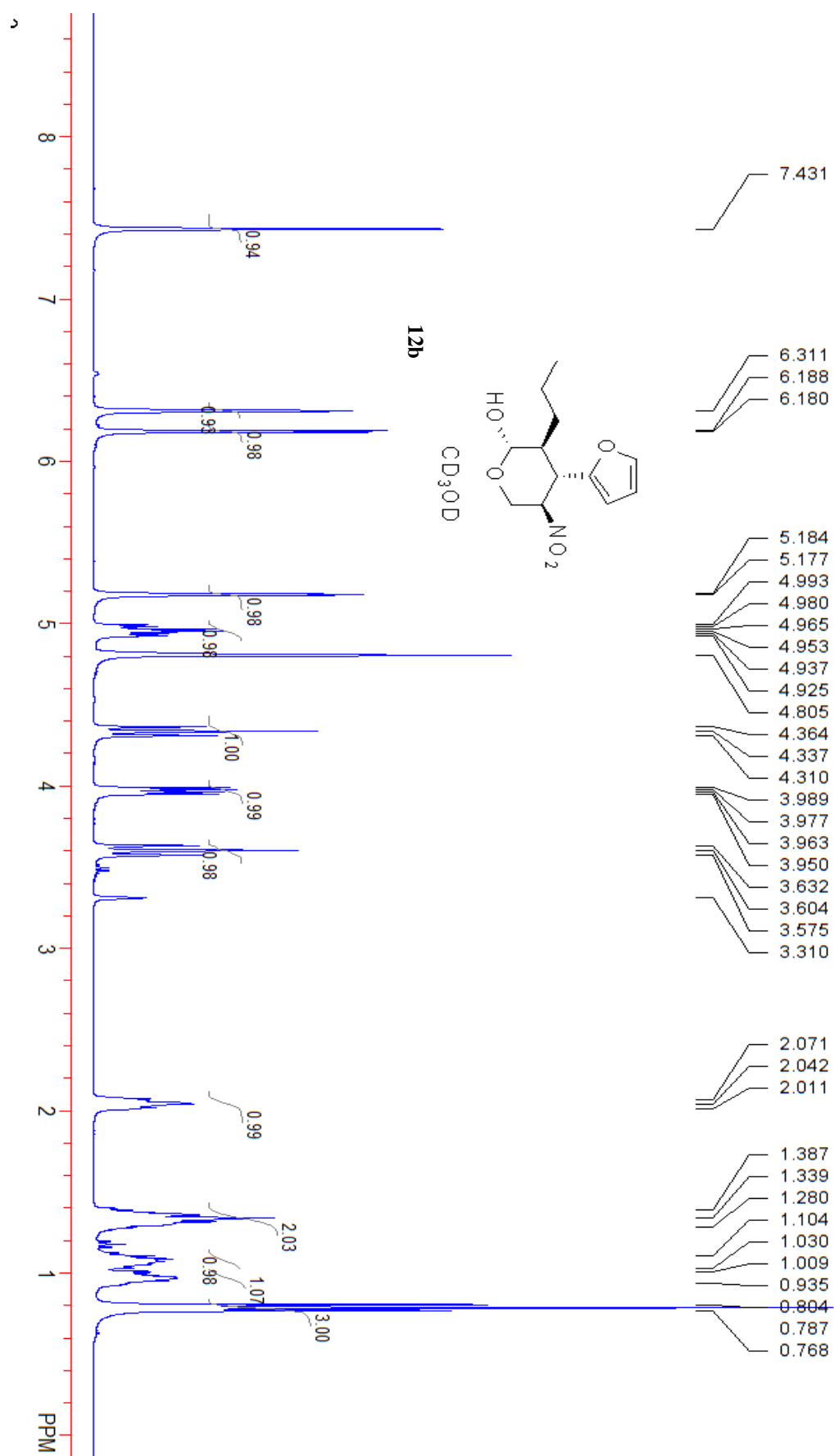
Operator:

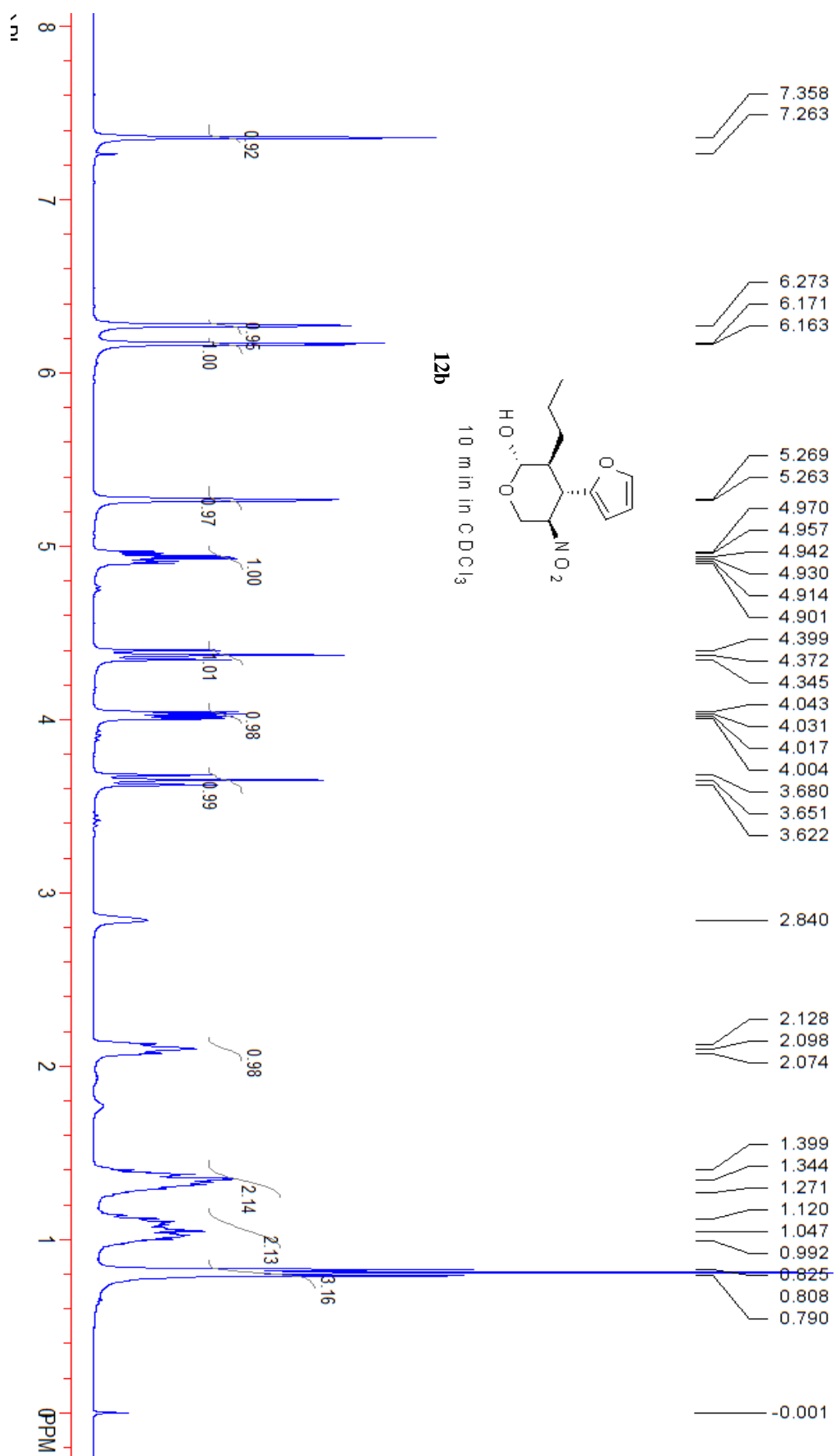
Date: 2010-09-09

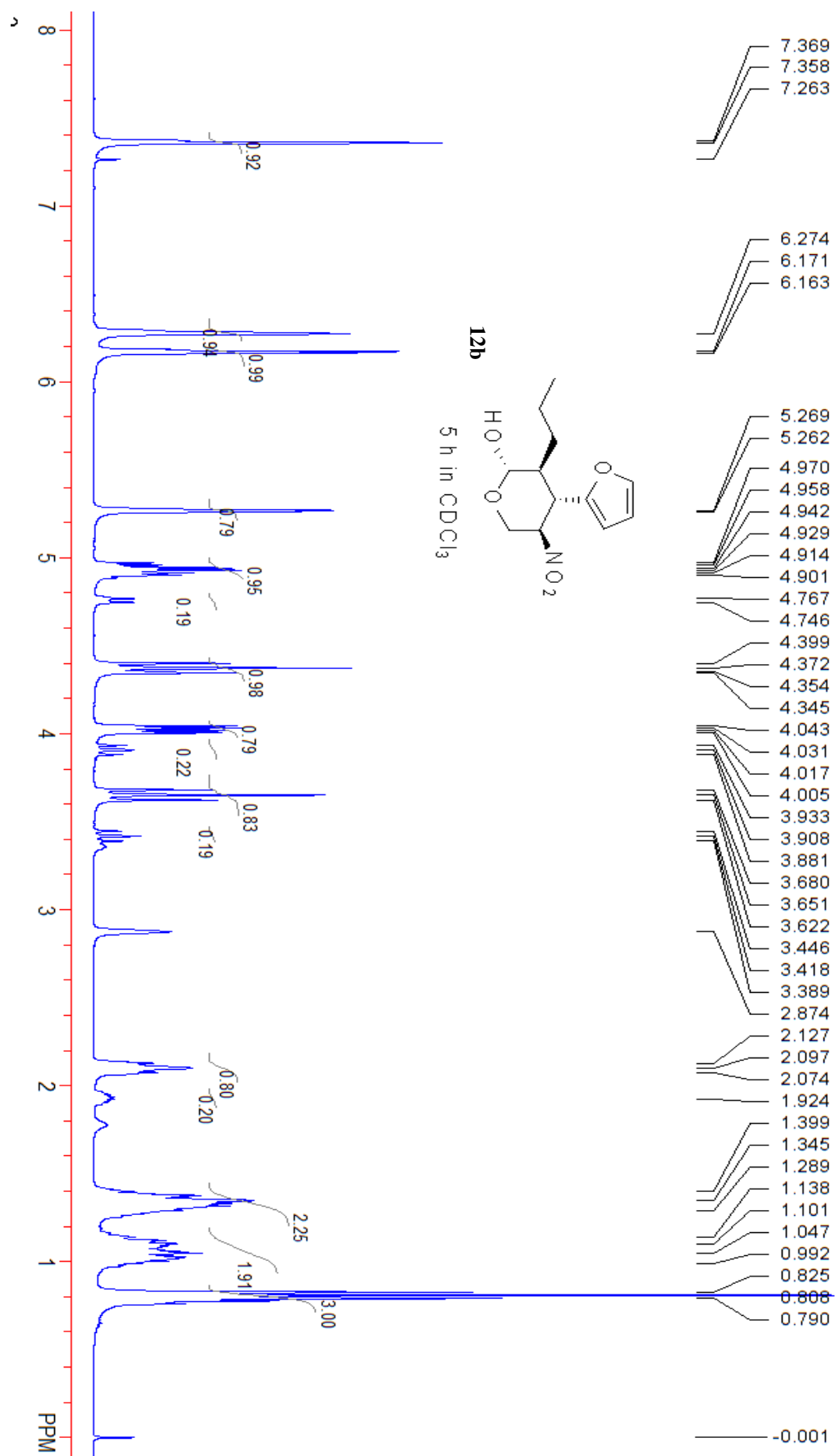
Time: 15:11

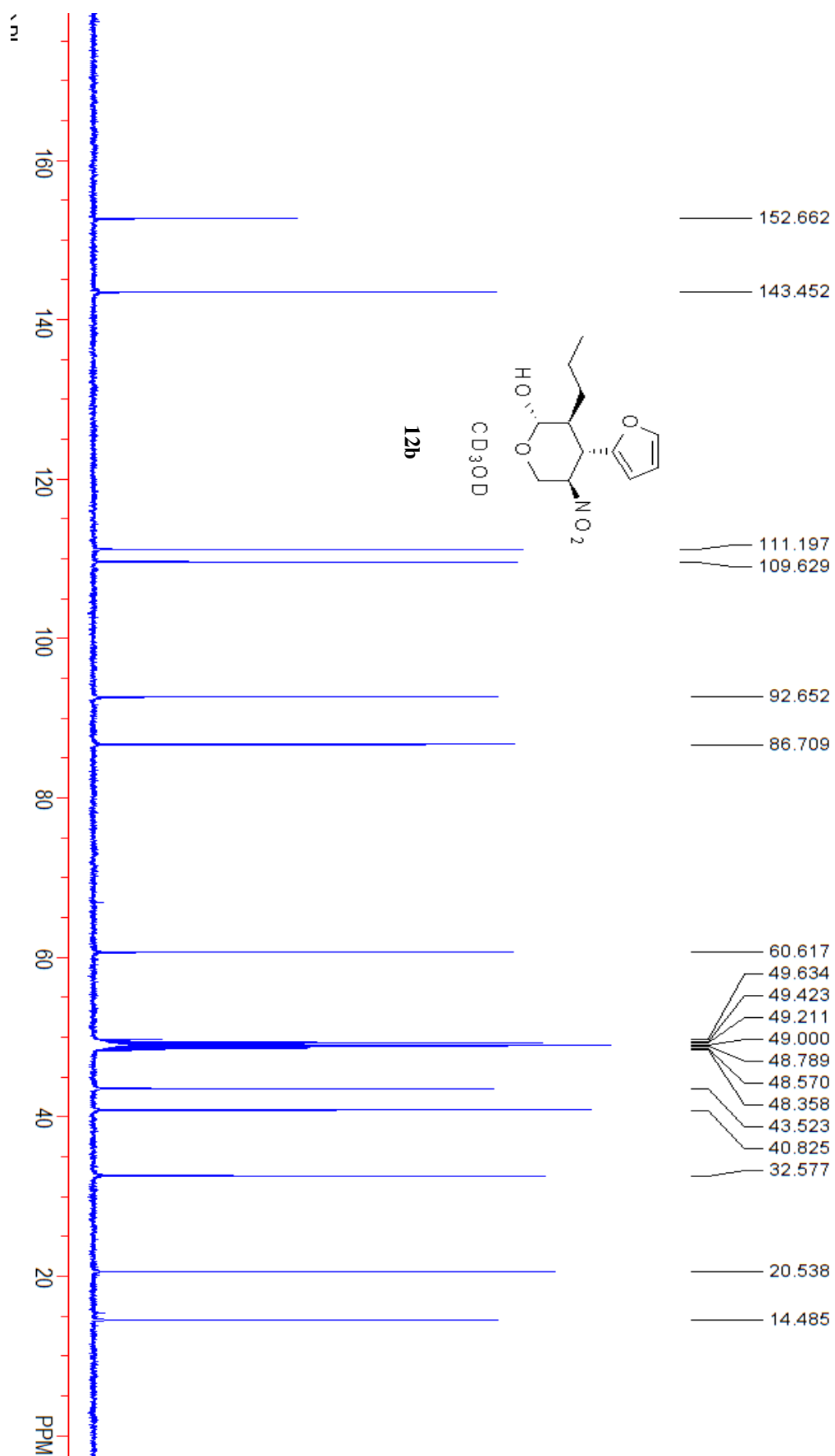


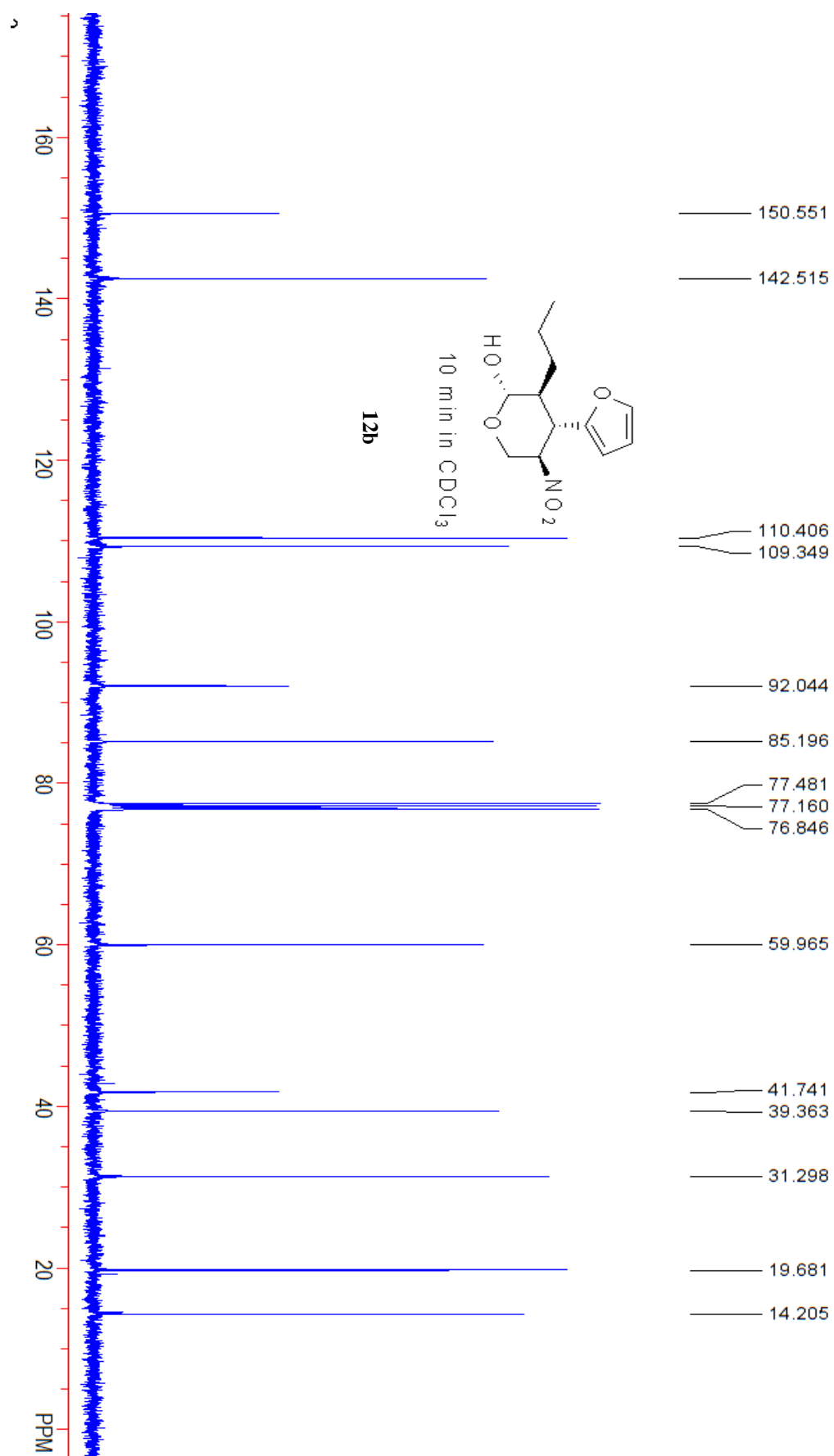
No.	PeakNo	R. Time	PeakHeight	PeakArea	PerCent
1	1	7.227	19627.9	222910.9	4.8456
2	2	8.277	353362.9	4282969.2	93.1032
3	3	9.348	7533.0	94360.6	2.0512
Total			380523.8	4600240.7	100.0000

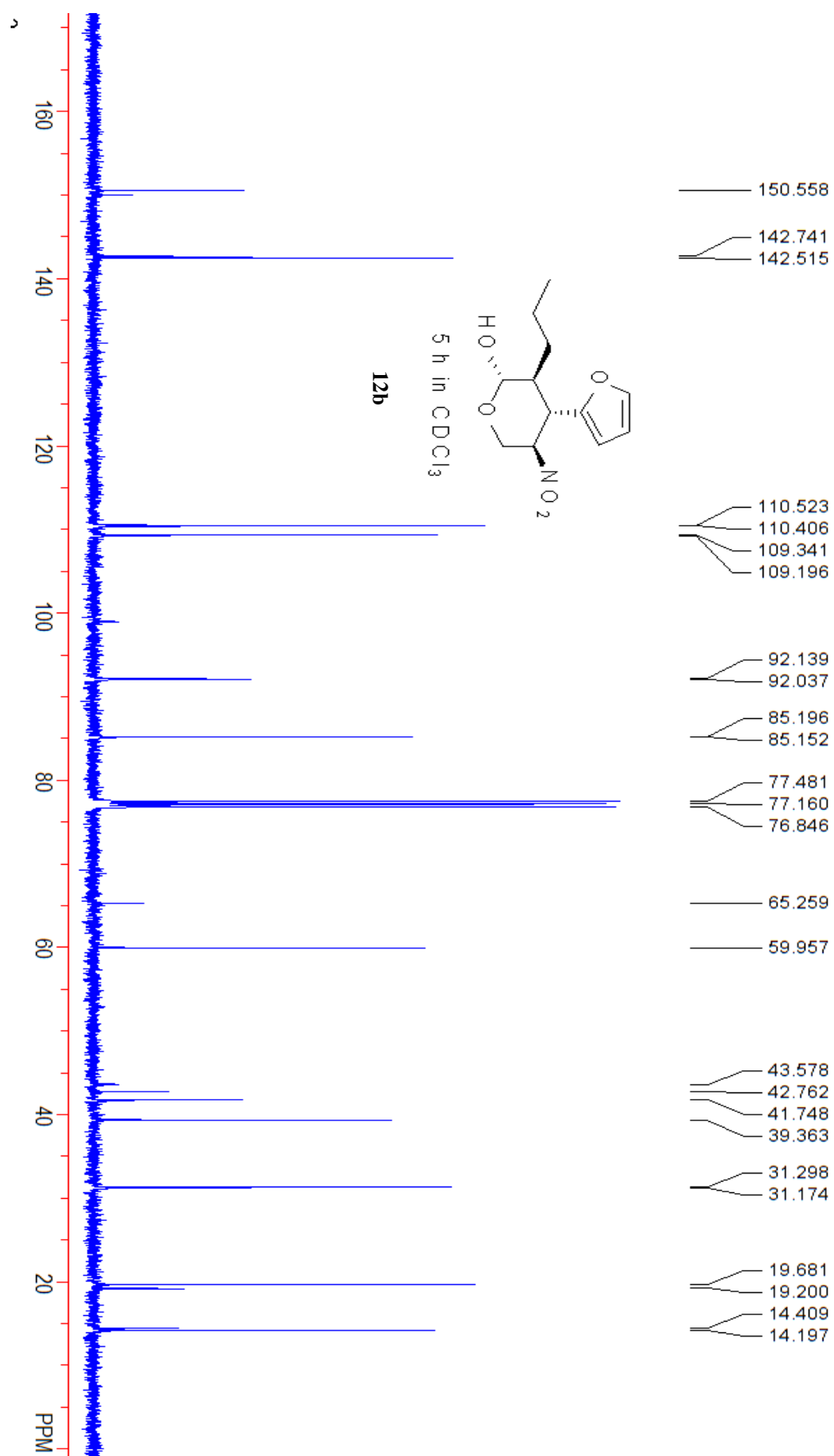






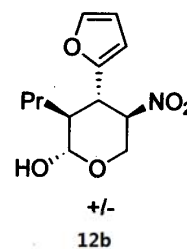
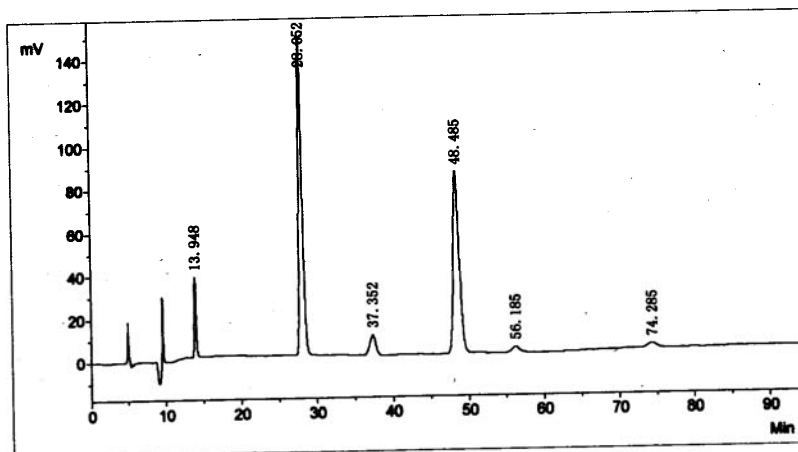






HPLC REPORT

Data File: ZSL-590+- AD-H 95EtOH5 214 0.7. che Sample Name:
 Date: 2009-09-27 Time: 14:54
 Operator:



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		13.948	37192.9	627017.1	5.6261
2	2		28.052	147968.1	4884582.5	43.8285
3	3		37.352	9507.6	515955.4	4.6296
4	4		48.485	84533.0	4735838.7	42.4939
5	5		56.185	2813.2	184939.2	1.6594
6	6		74.285	2300.2	196428.4	1.7625
Total				284315.0	11144761.3	100.0000

HPLC REPORT

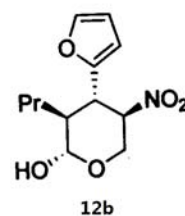
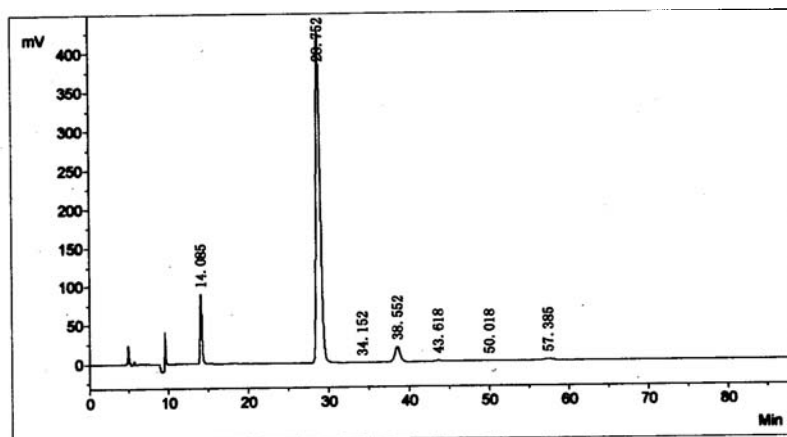
Data File:ZSL-13-85-2.che

Sample Name:

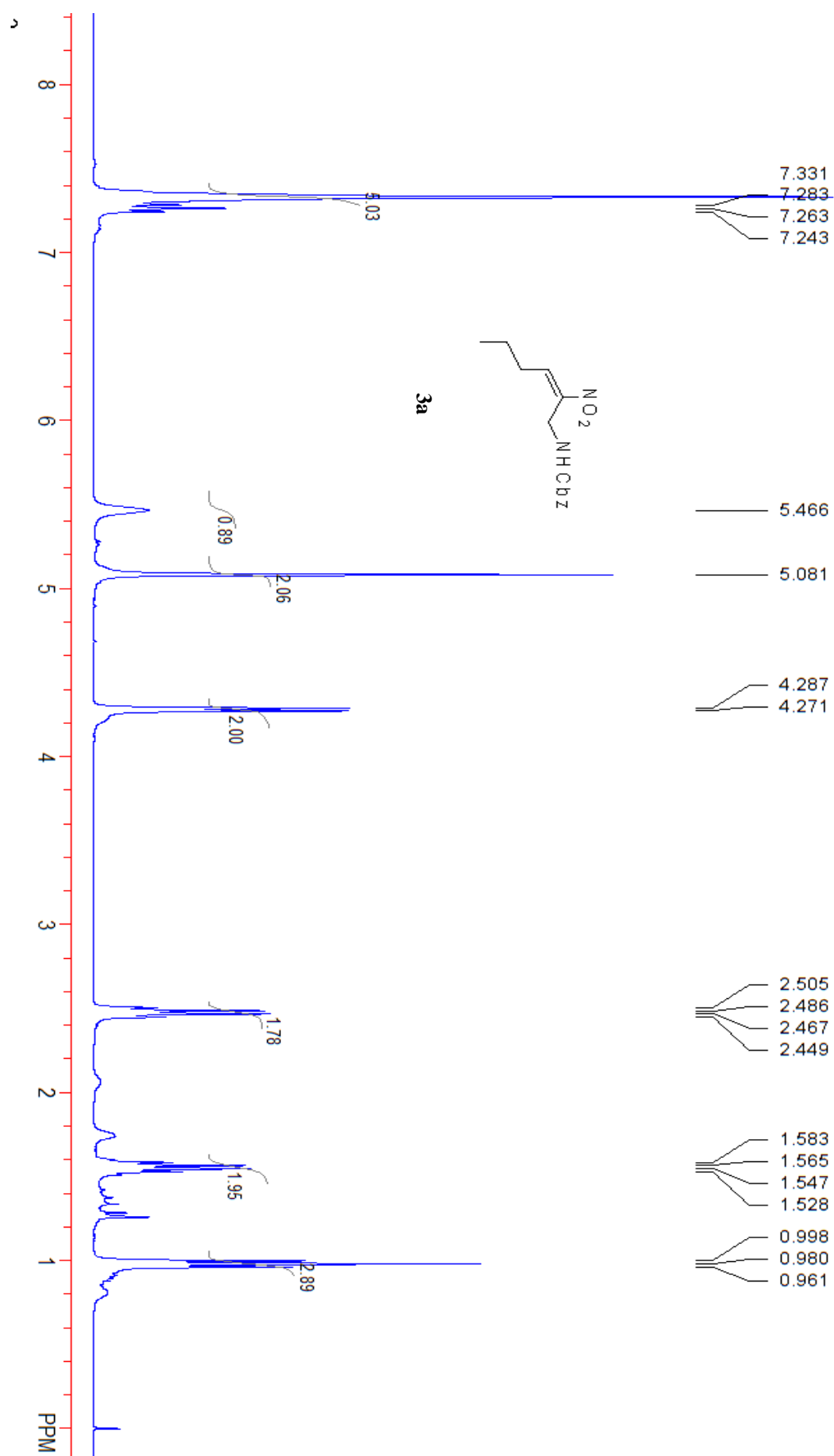
Date:2009-09-27

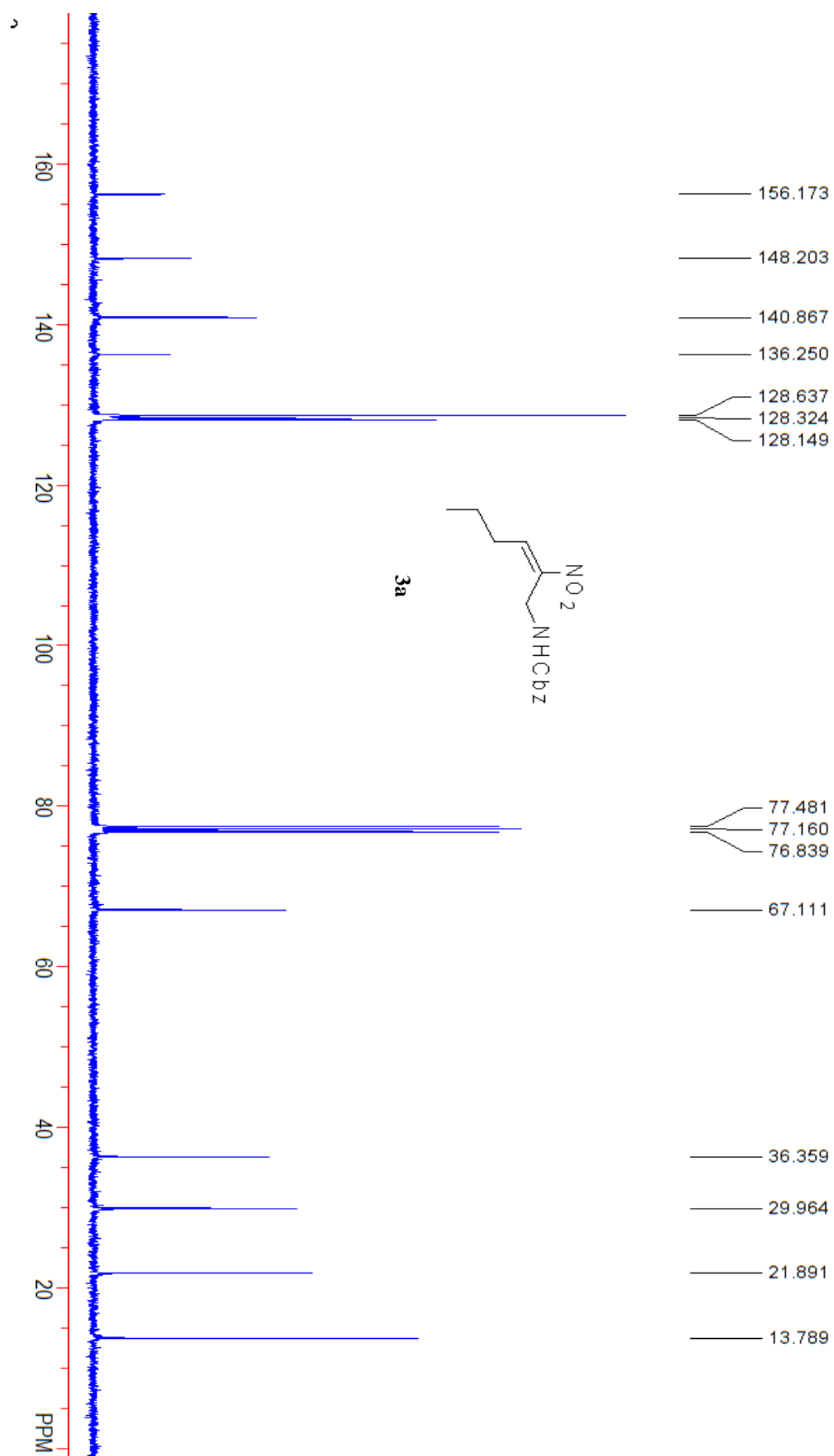
Time:16:39

Operator:



No.	PeakNo	ID. Name	R. Time	PeakHeight	PeakArea	PerCent
1	1		14.085	89367.0	1549891.6	8.5802
2	2		28.752	426646.6	15416639.9	85.3469
3	3		34.152	164.6	5826.5	0.0323
4	4		38.552	19655.5	905280.1	5.0117
5	5		43.618	1106.2	53651.2	0.2970
6	6		50.018	260.5	14094.0	0.0780
7	7		57.385	1781.9	118127.1	0.6540
Total				538982.3	18063510.3	100.0000





Std proton

File: mp

Pulse Sequence: NOESY

Solvent: cdcl3

Temp: 25.0 C / 298.1 K

Operator: vnmr1

VNMR-400 "400MHz"

Relax. delay 2.000 sec

Mixing 1.000 sec

Acq. time 0.242 sec

Width 3811.0 Hz

2D Width 3811.0 Hz

4 repetitions

2 x 128 increments

OBSERVE H1, 399.622490 MHz

DATA PROCESSING

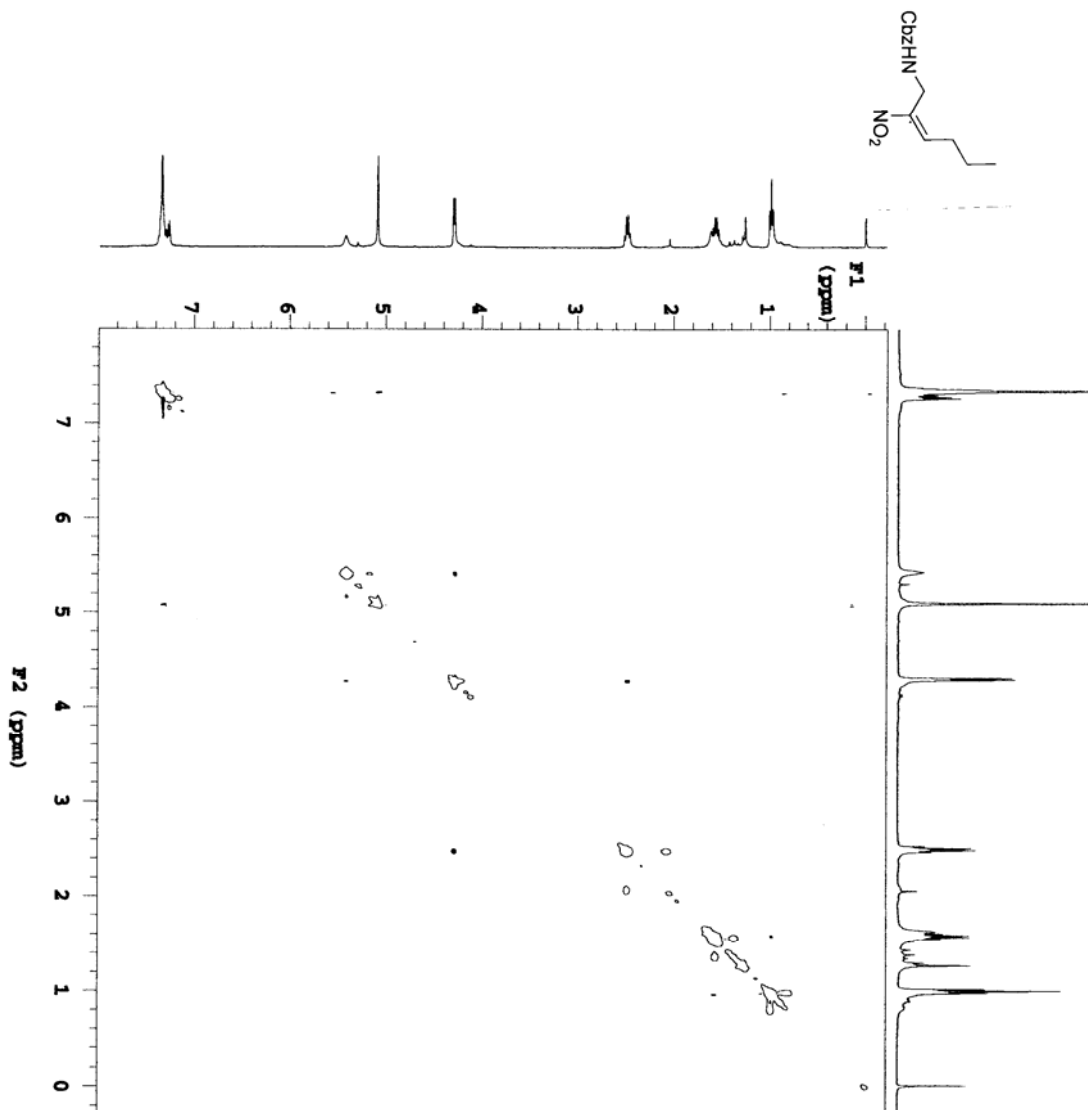
Gauss apodization 0.112 sec

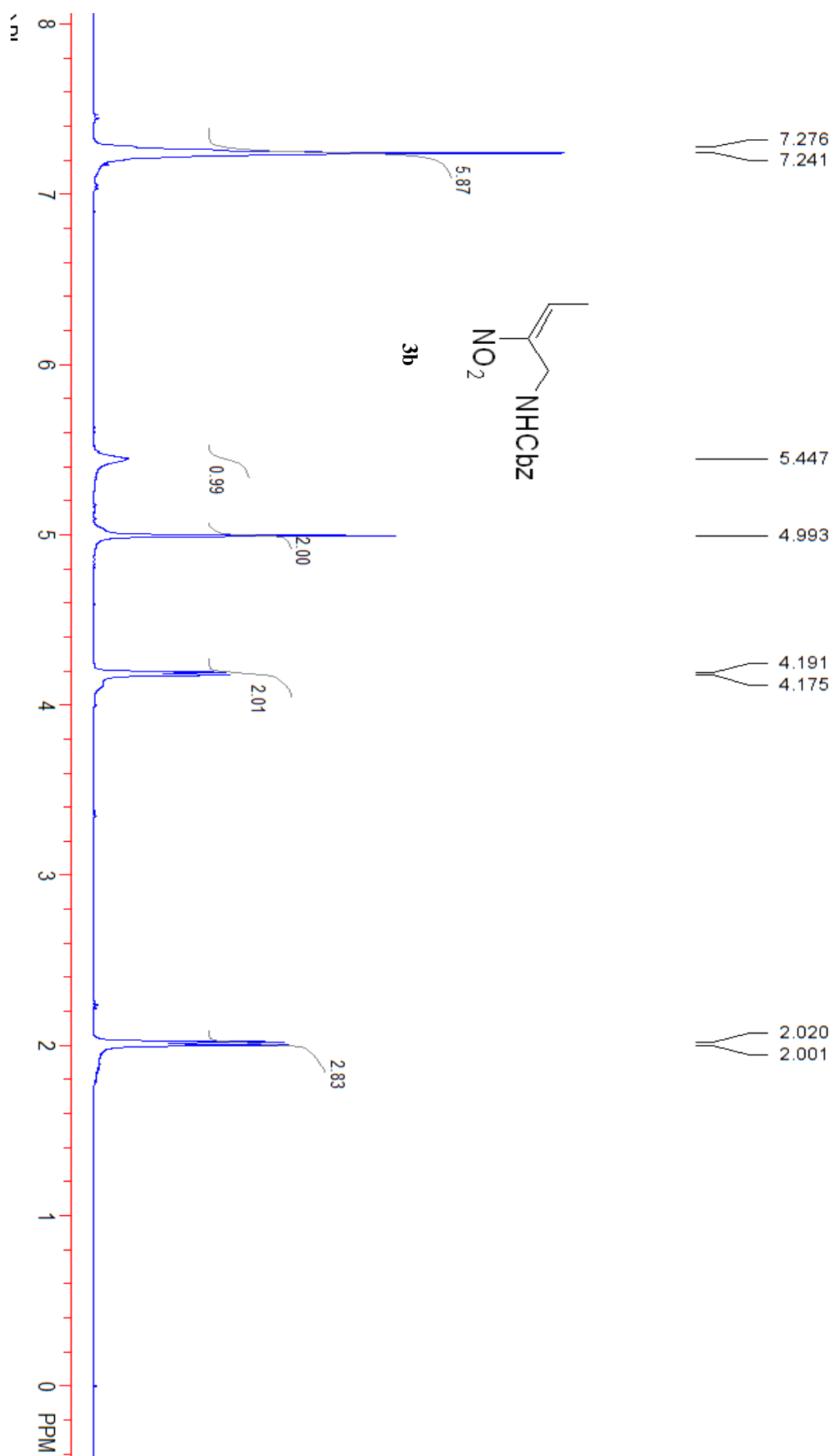
F1 DATA PROCESSING

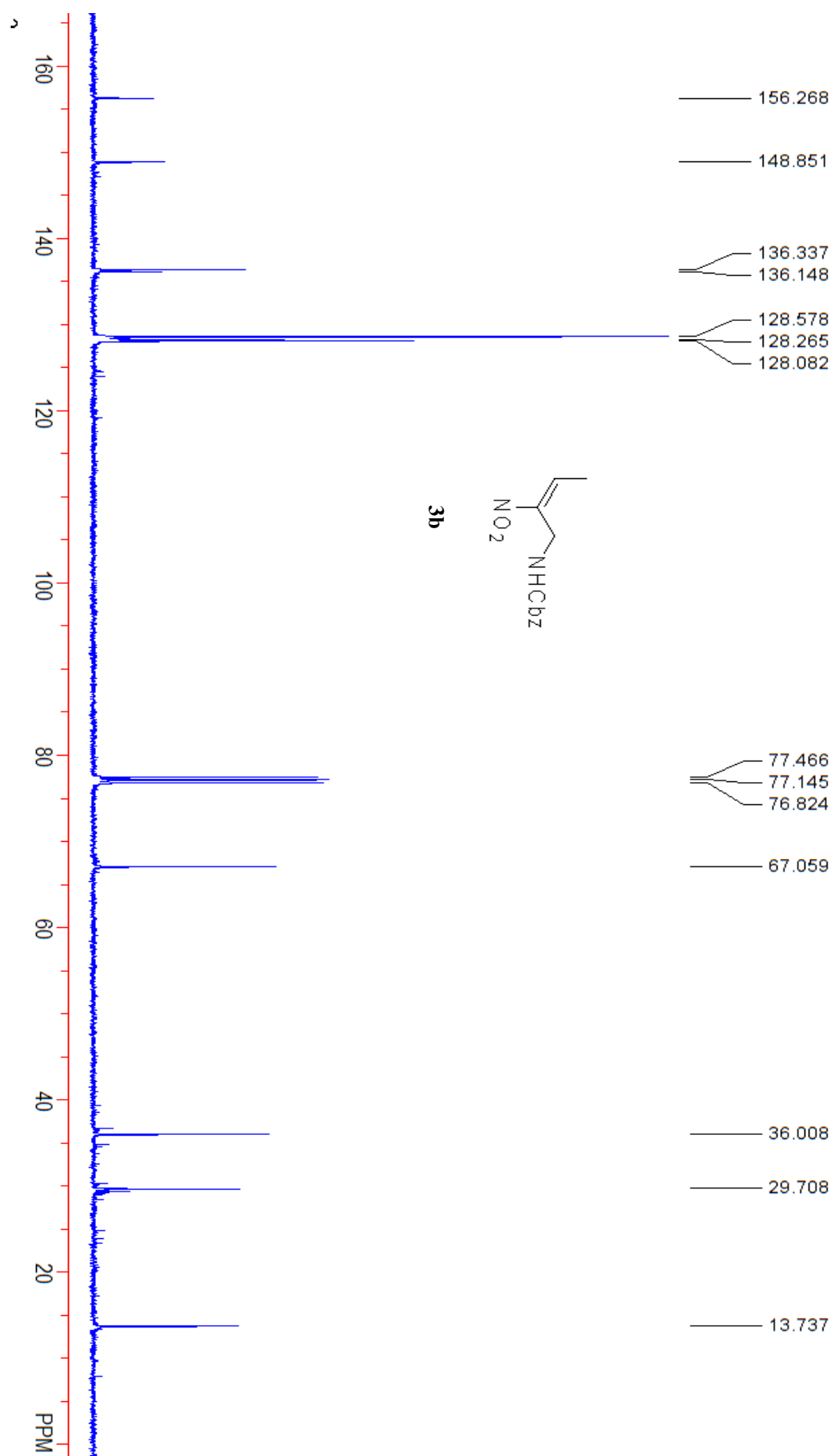
Gauss apodization 0.062 sec

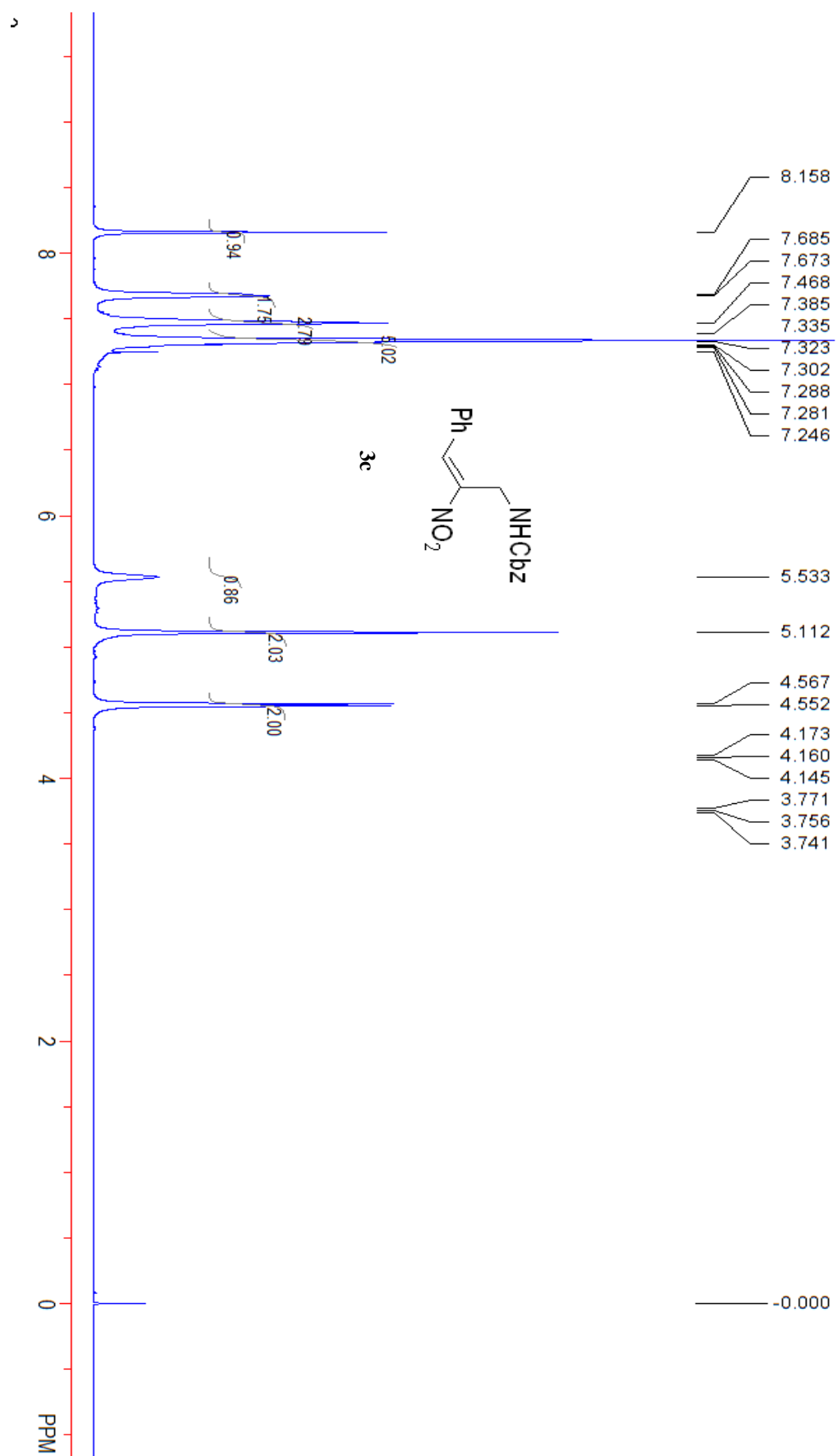
FT size 2048 x 2048

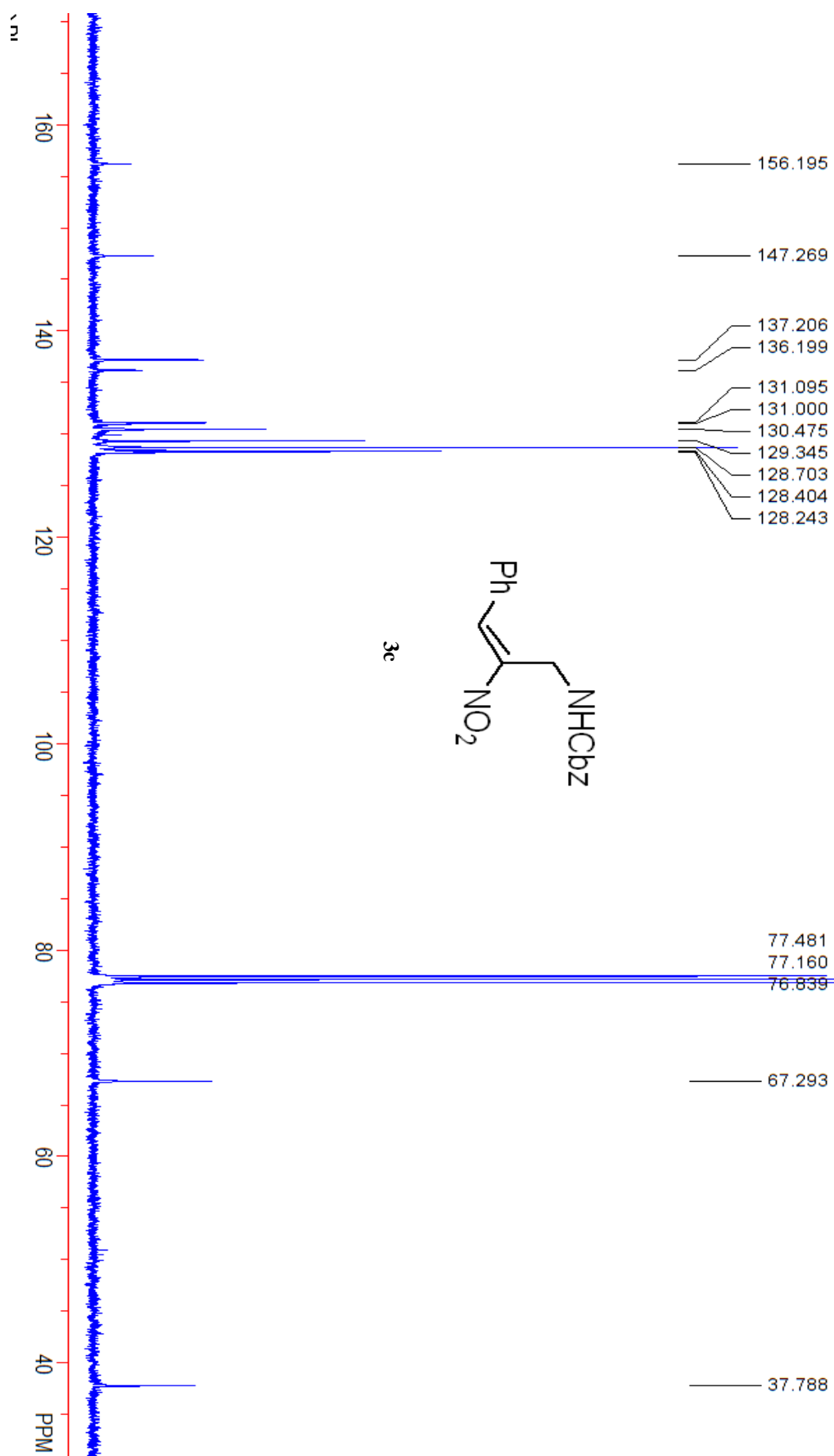
Total time 55 min, 54 sec

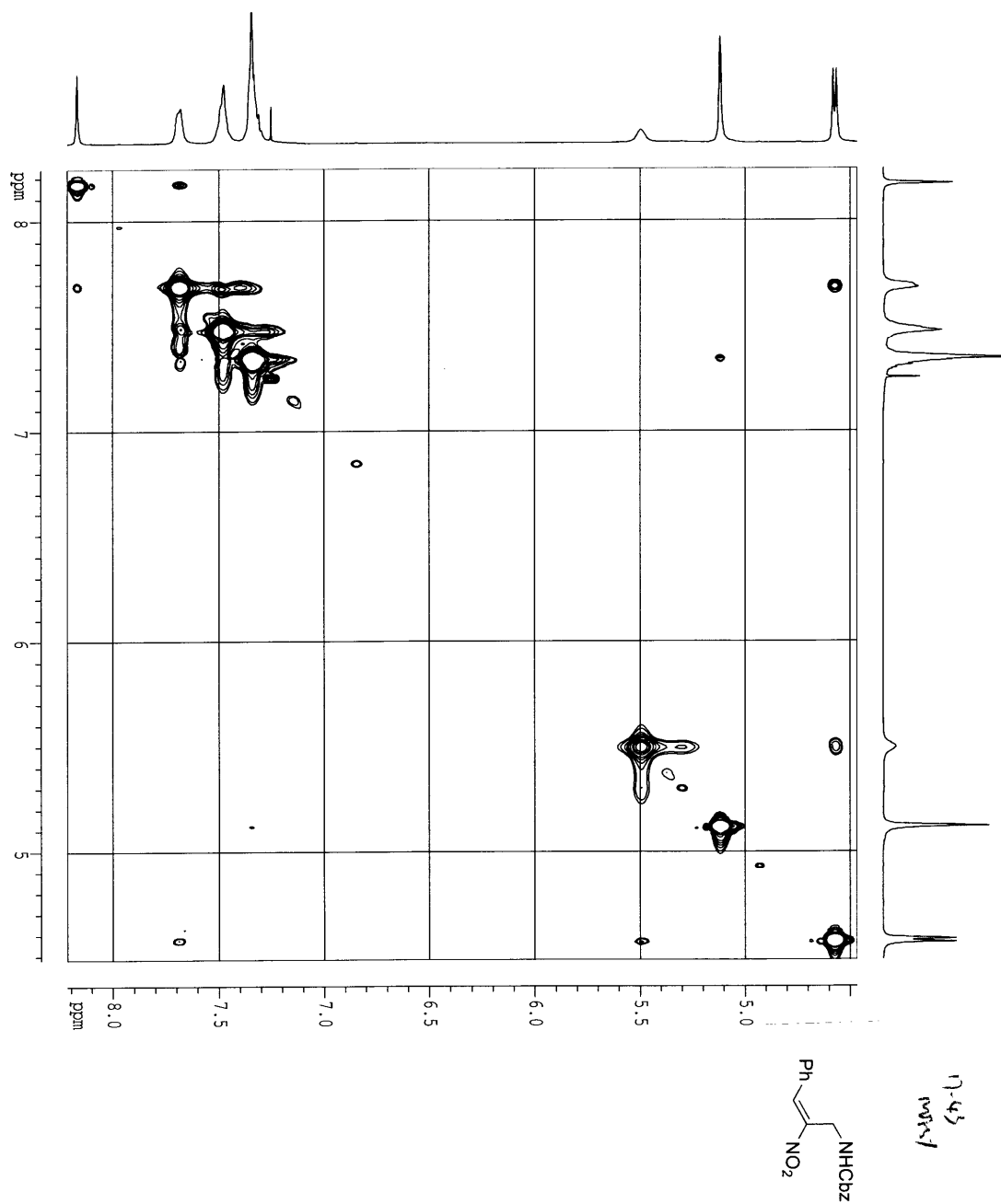


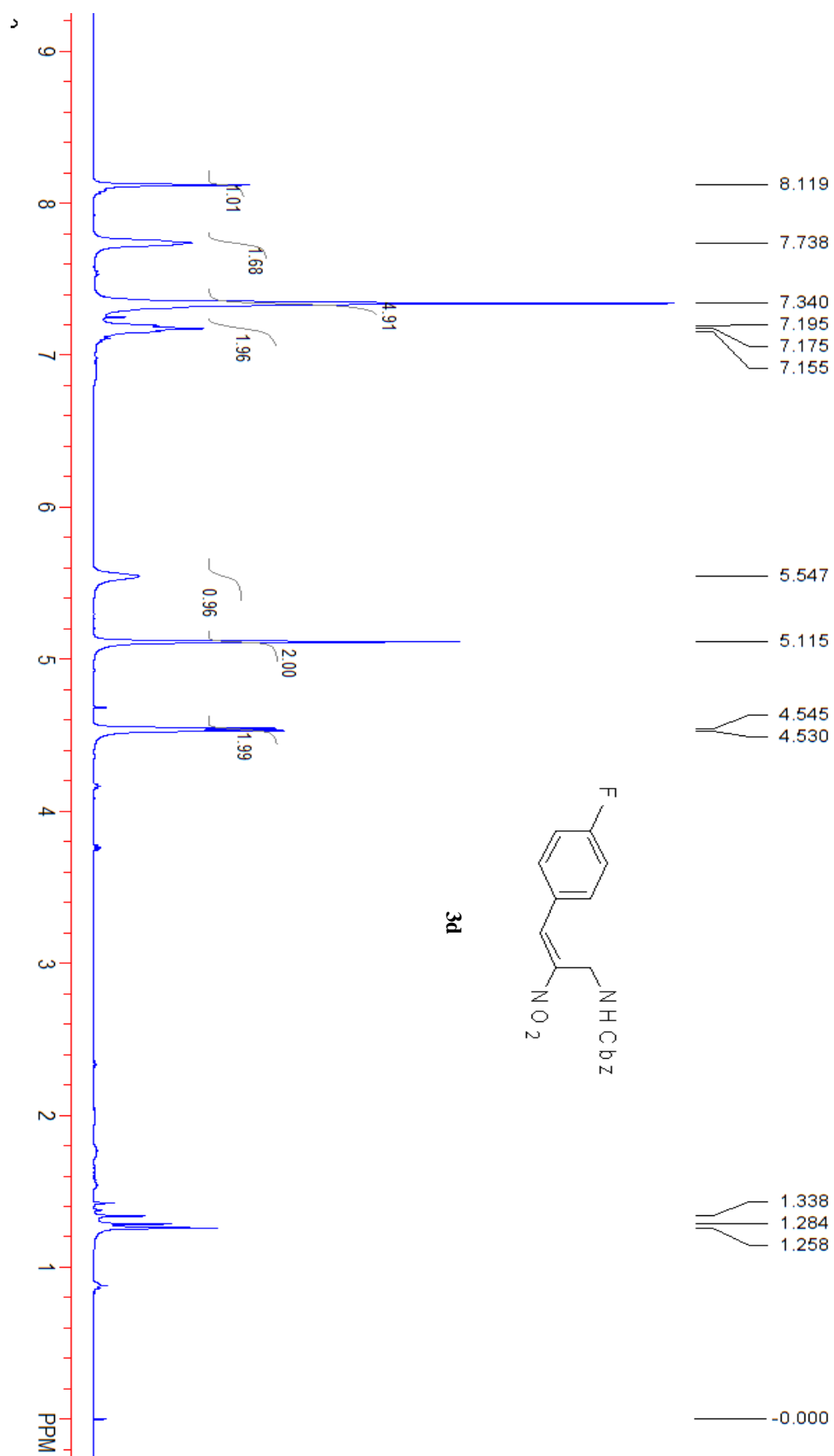


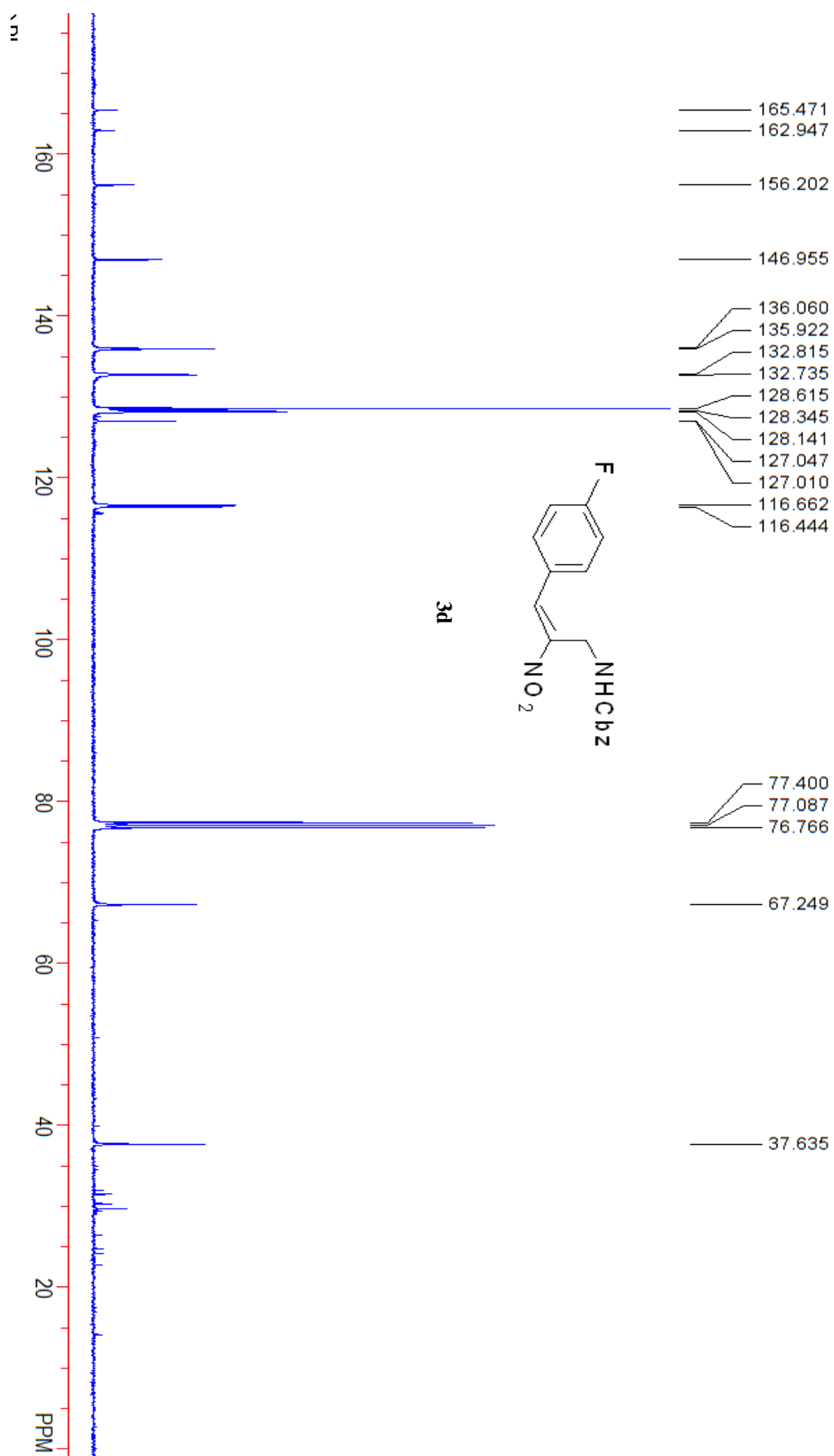


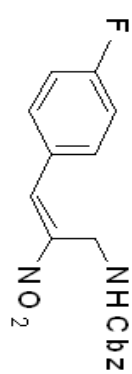




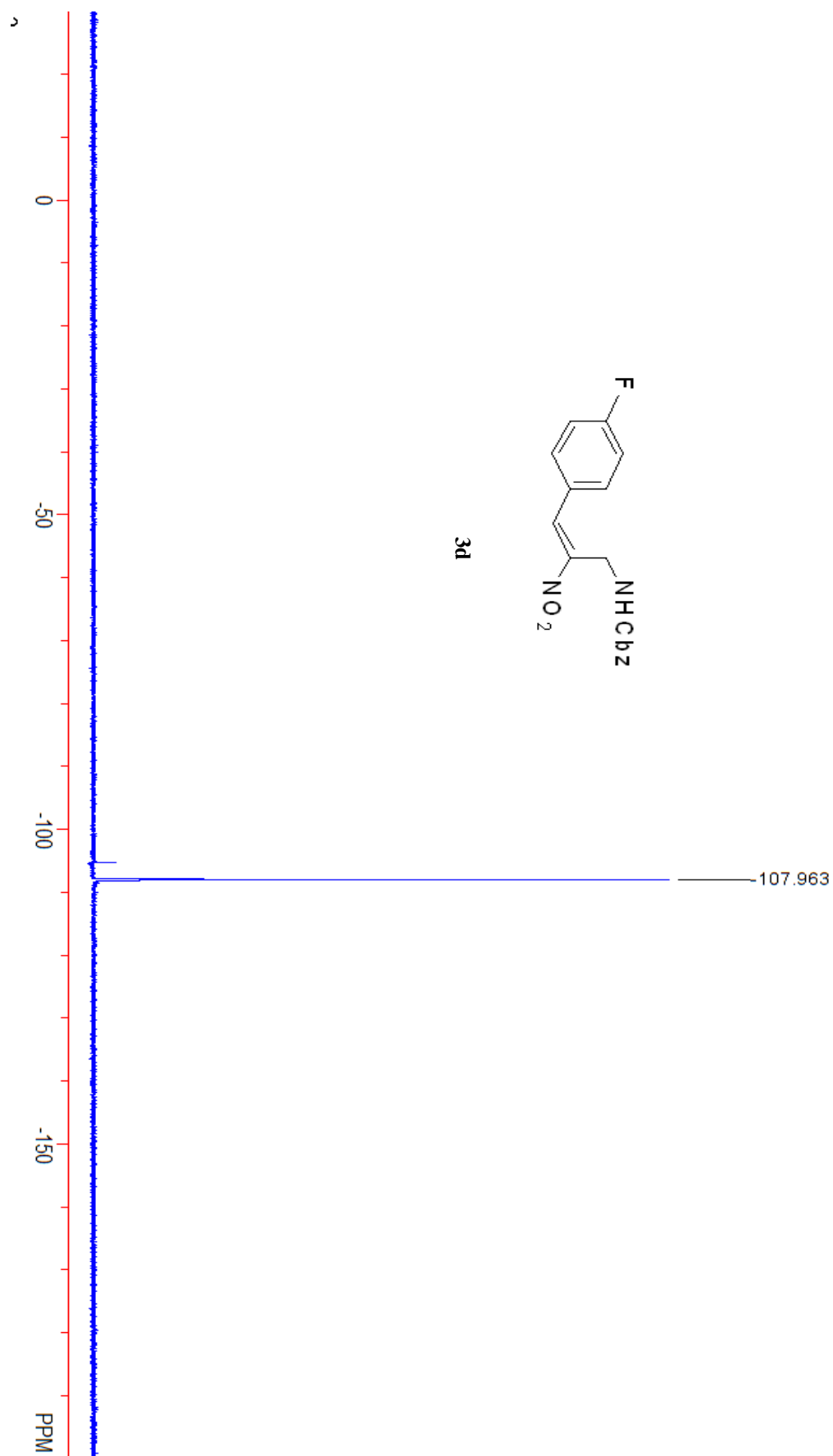


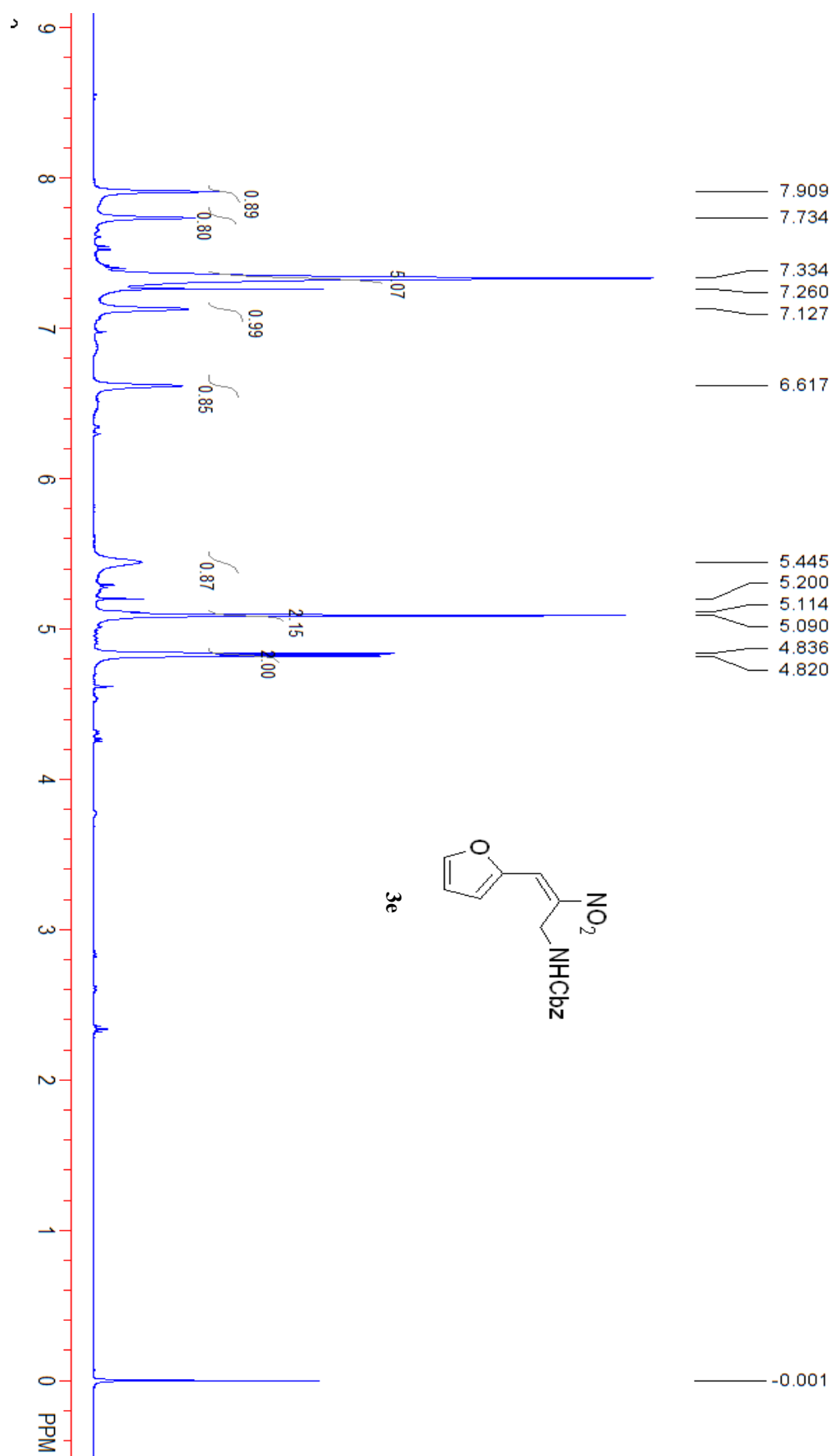


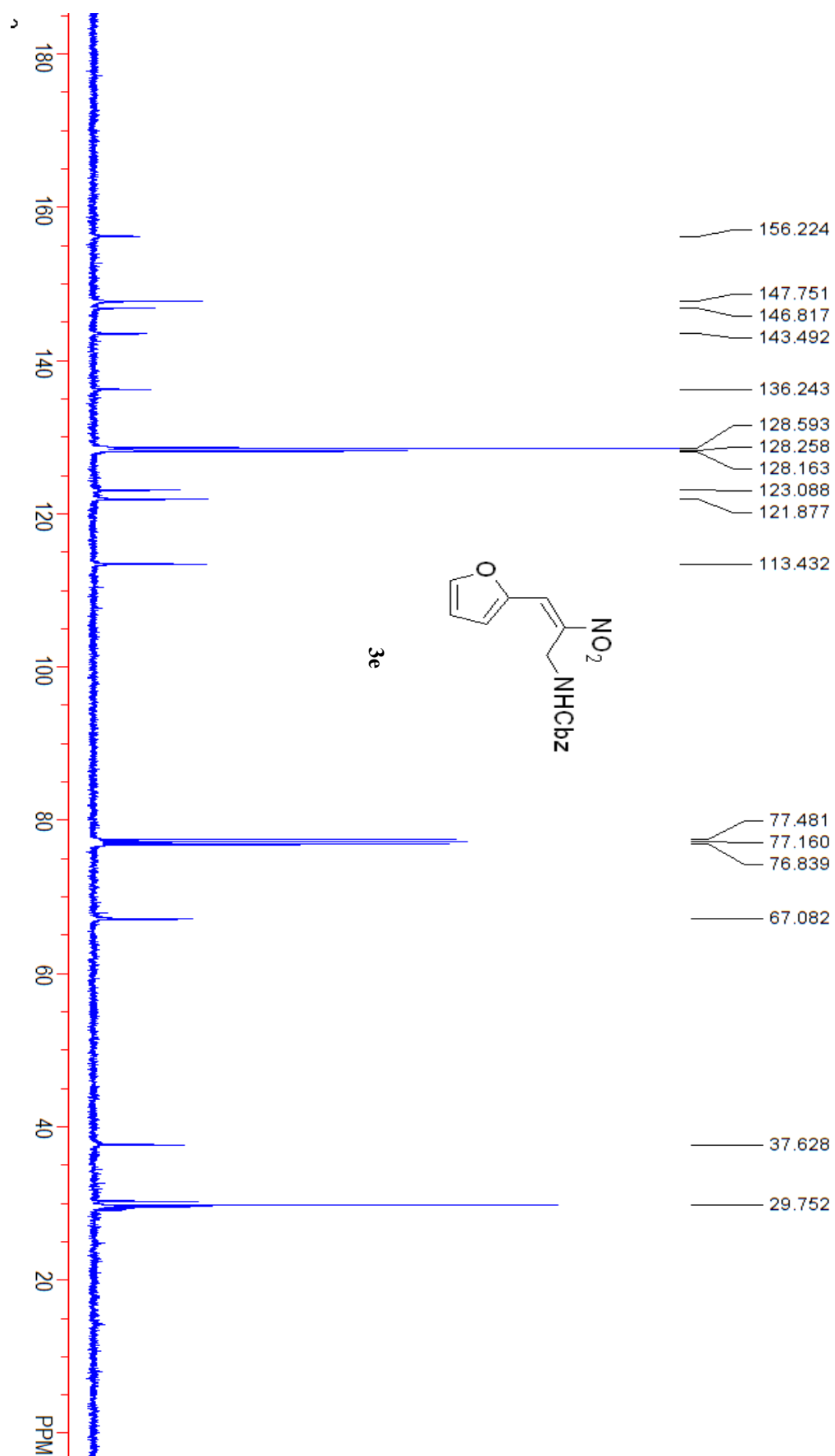


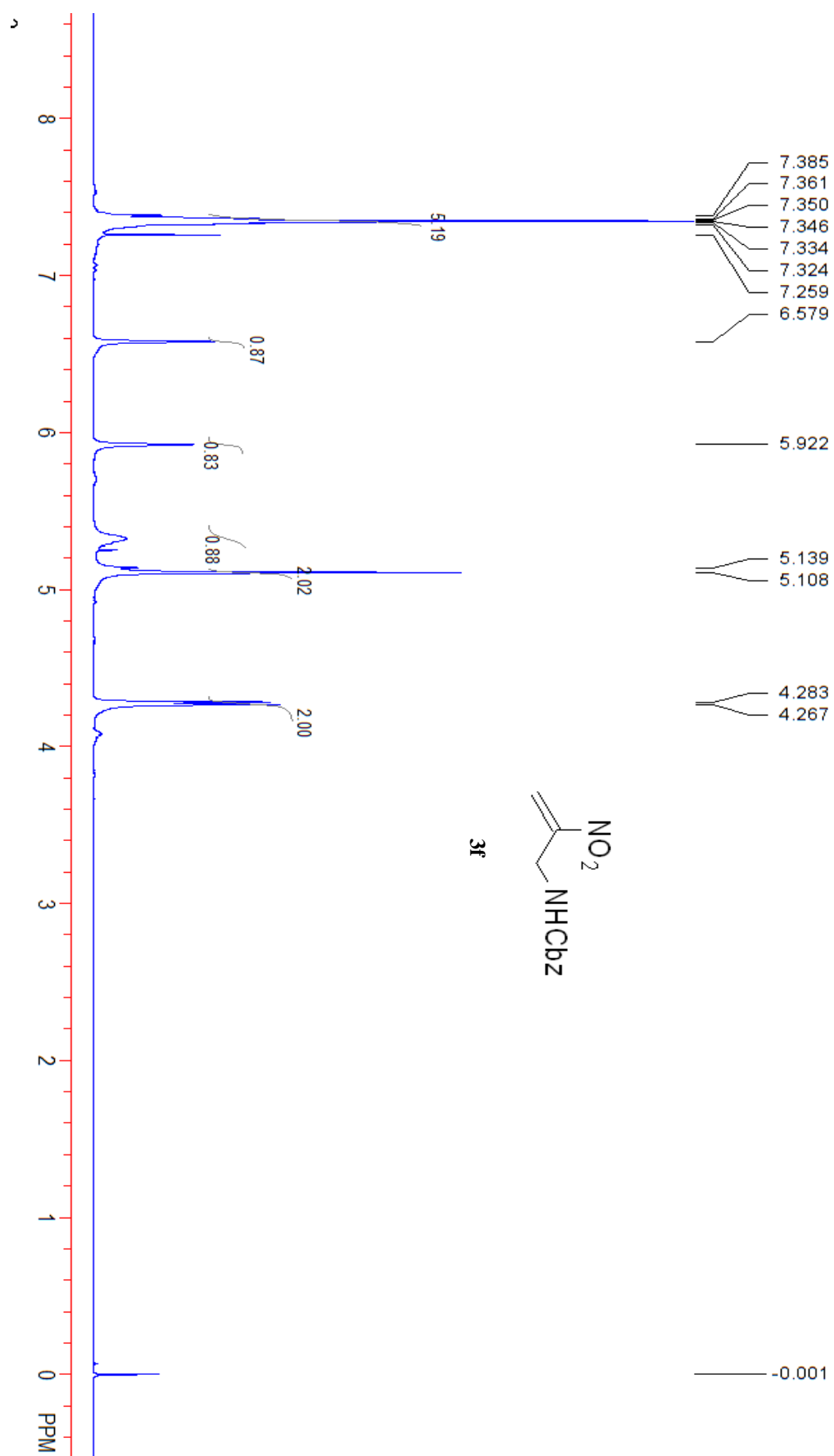


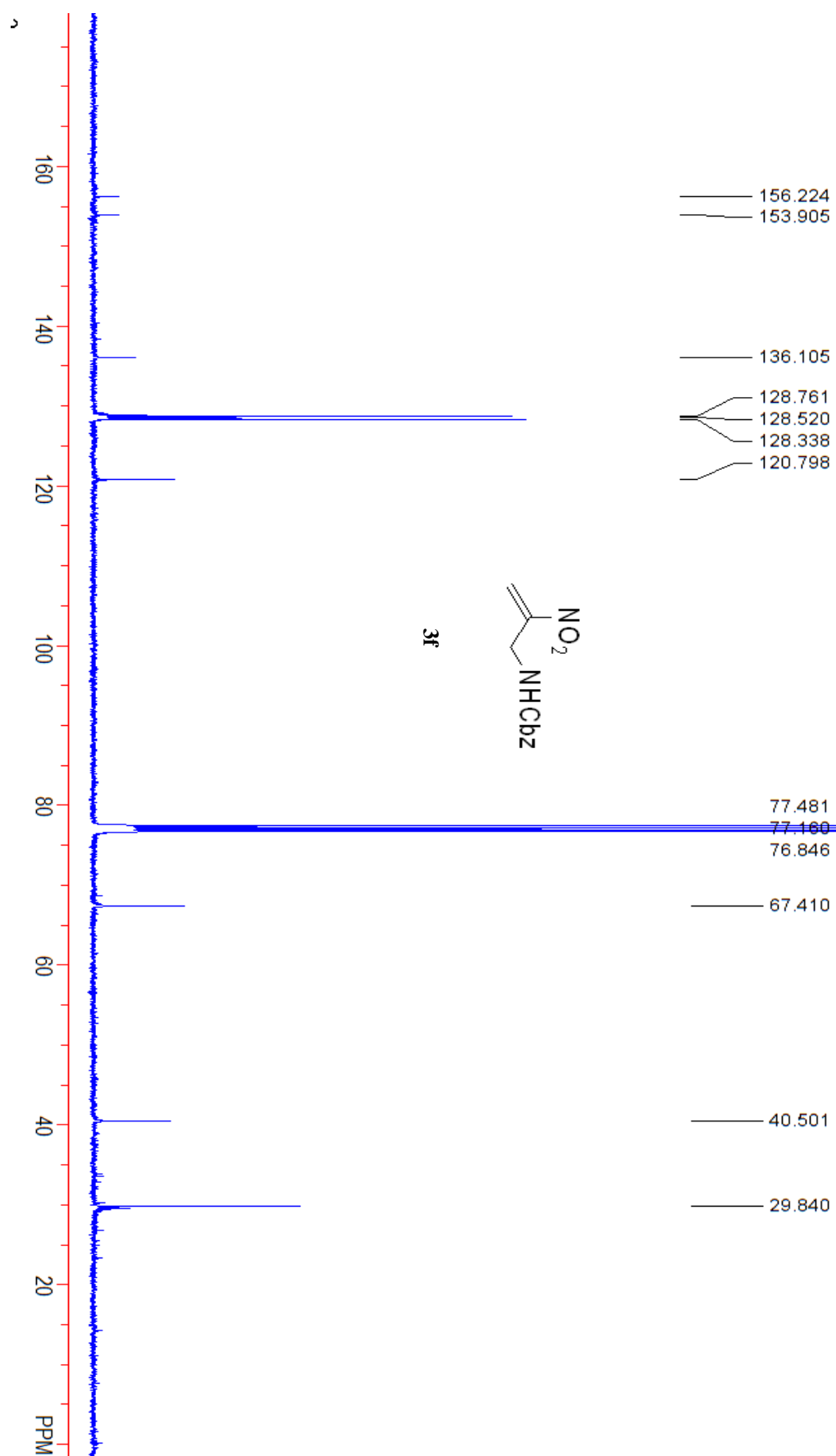
3d

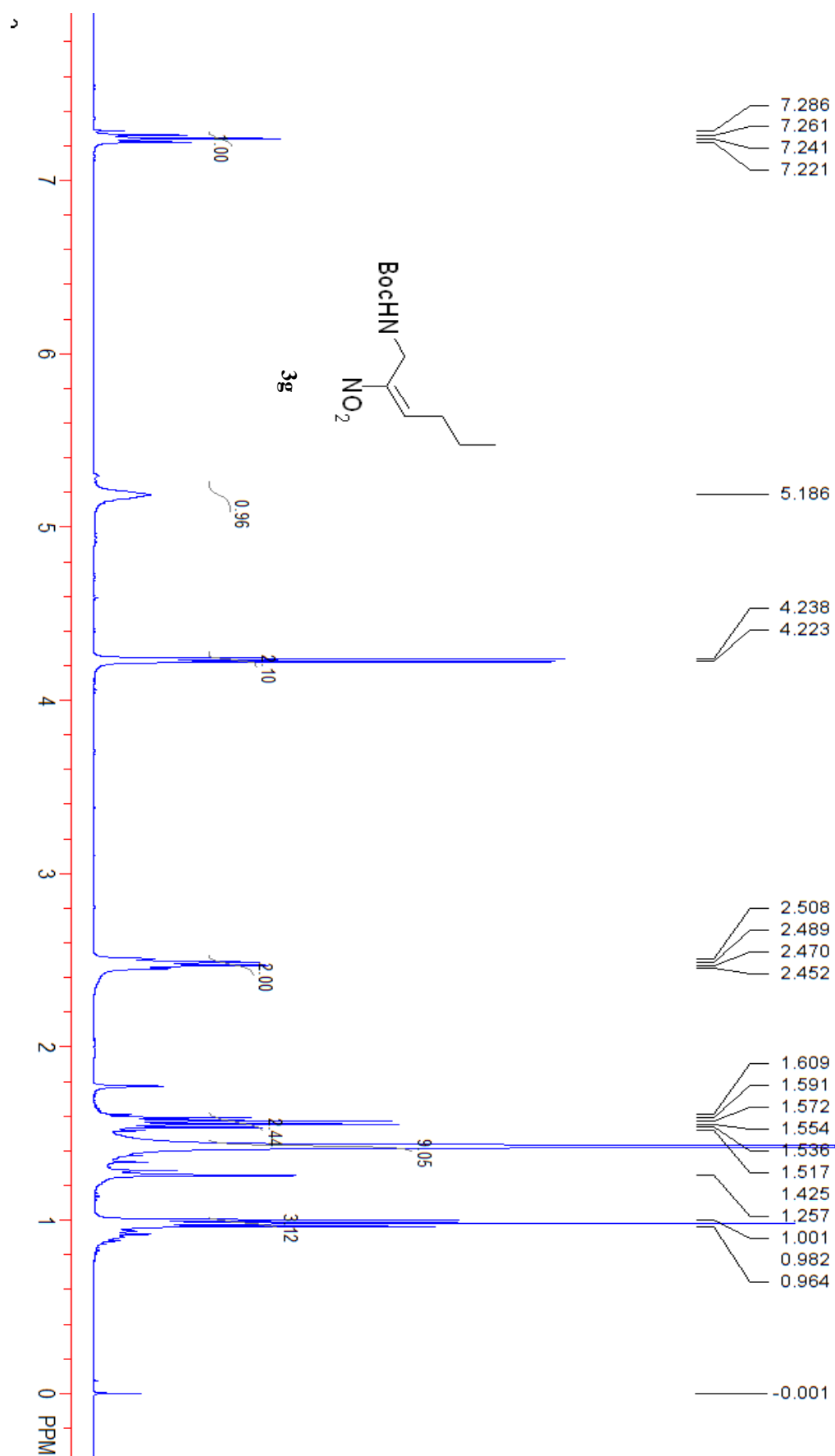


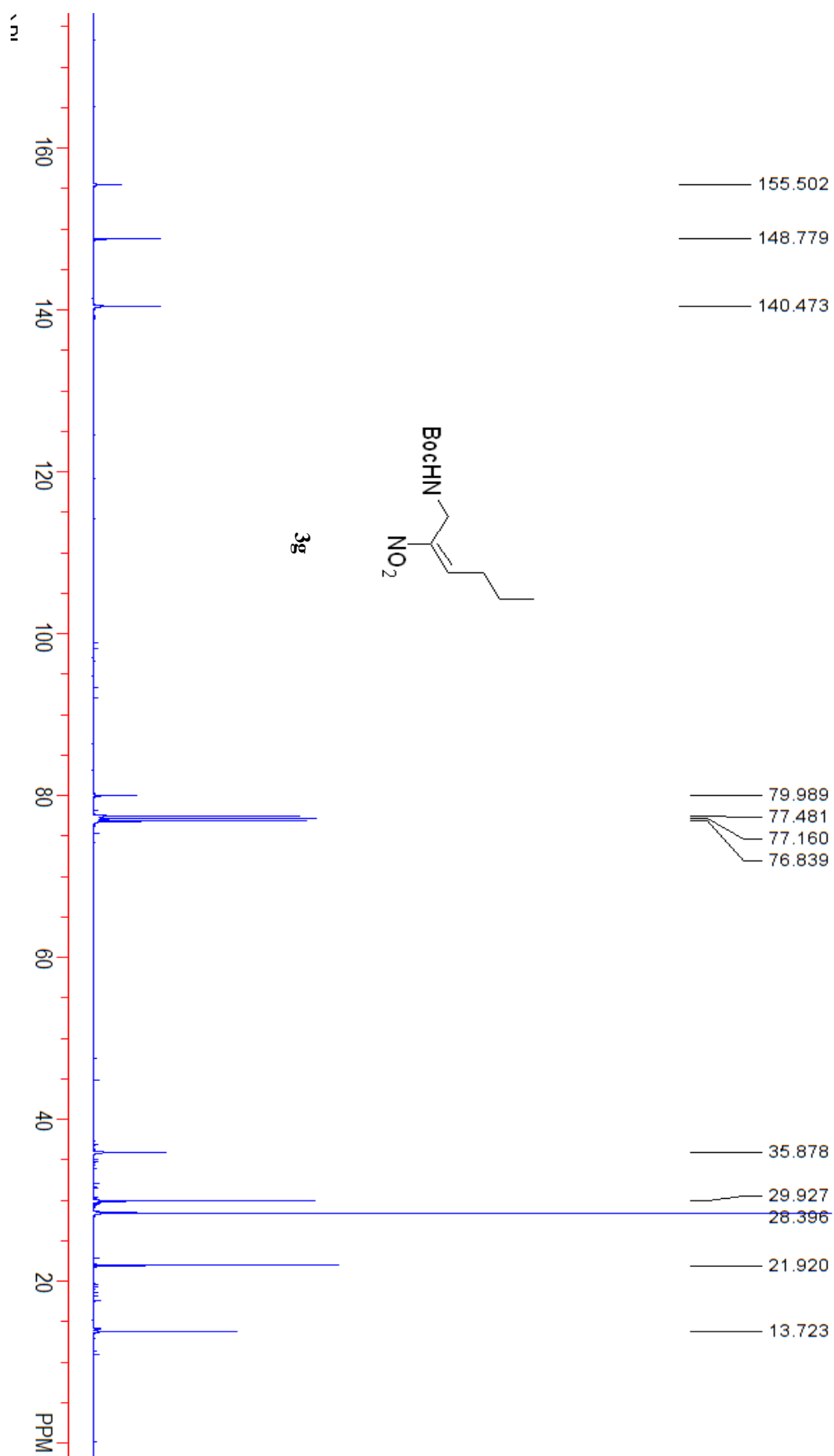


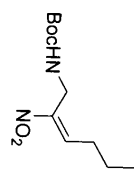
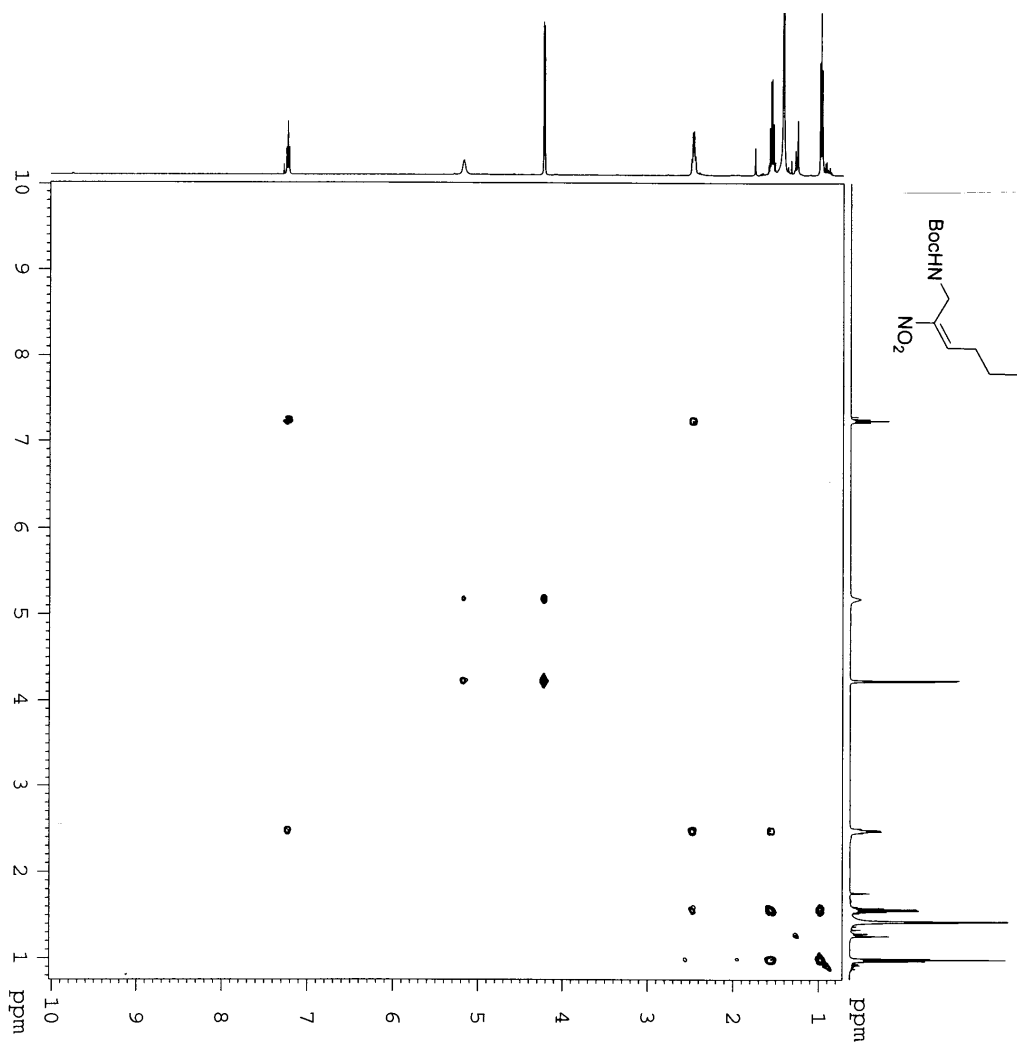












```
Current Data Parameters
NAME      2005183c6v 6.38
PROCNO    1

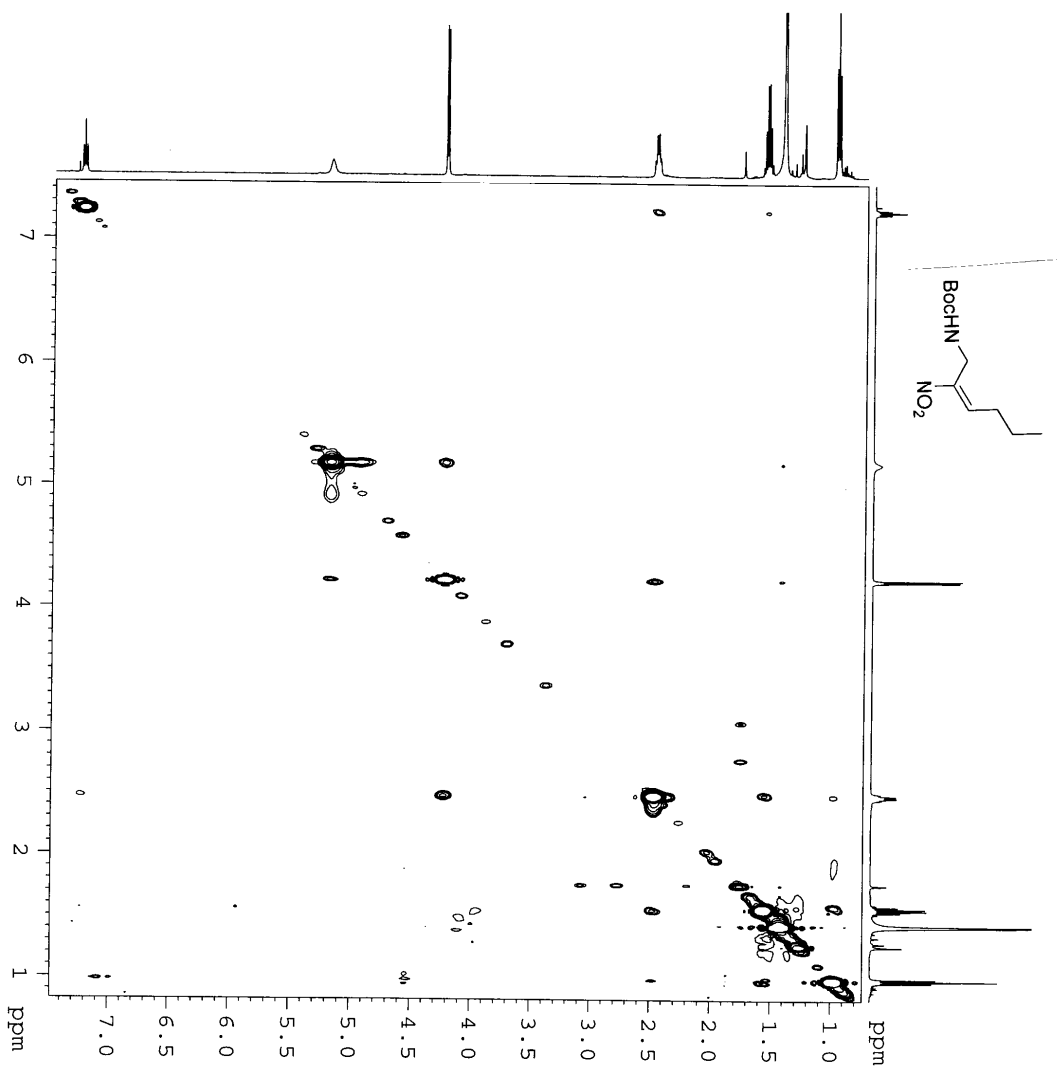
===== Acquisition Parameters =====
Date_     20100824
Time      12.56
INSTRUM   spect
PROBHD    5 mm QNP 1H/13
PULPROG   zgpg30
TD         65536
FIDRES     0.097000 Hz
AQ         0.0002000 sec
RG         91.024
DS         4
SFO        500.136250 MHz
DE         300.0 K
TE         6.50 usec
T1         0.0000300 sec
D1         2.0000000 sec
DELTA      2.0000000 sec
d13        0.0002000 sec
D16        0.00018240 sec

===== CHANNEL f1 =====
NUC1       13C
P1          1H usec
PL1         -1.00 dB
SFO1       500.136250 MHz

===== GRADIENT PARAMETERS =====
GPRAM2     SINE: 100
GPRAM2     SINE: 100
GPRAM2     SINE: 100
GPRAM2     SINE: 100
GP21        16.00 %
GP22        12.00 %
GR21        10.00 usec
P16         1000.00 usec

f1 - Acquisition parameters
SI         500.136250 MHz
SF         500.1360016 MHz
WDW         SINE
SSB         0 Hz
GB          0.00 Hz
PC          1.40

f2 - Processing parameters
SI         1024
SC2        500.1360000 MHz
WDW         SINE
SSB         0
GB          0.00 Hz
```



Current Data Parameters
 EXPNO 200518hpc-6-38
 PROCNO 1
 F2 - Acquisition Parameters
 Date_ 20100824
 Time 10:07
 INSTRUM spect
 PROBRD 5 mm QNP 1H/13
 PULPROG zgpg30
 TD 1024
 SFO1 500.1324200 MHz
 NS 16
 DS 16
 SWH 5482.456 Hz
 FIDRES 5.353961 Hz
 AQ 0.0935300 sec
 RG 362
 DW 91.320 usec
 DE 6.50 usec
 TE 300.0 K
 D1 0.00007783 sec
 D11 2.00000000 sec
 D8 1.00000000 sec
 INO 0.00018240 sec
 STICNT 256
 TAU 0.4988001 sec
 ===== CHANNEL f1 =====
 NUC1 1H
 P1 10.50 usec
 P2 21.00 usec
 PL1 -4.00 dB
 SFO1 500.1324200 MHz
 ===== GRADIENT CHANNEL =====
 GENAM1 SINE.100
 GENAM2 SINE.100
 GP21 40.00 %
 GP22 40.00 %
 PL6 1000.00 usec
 F1 - Acquisition Parameters
 NDO 1
 ND0 256
 SFO1 500.1324200 MHz
 FIDRES 21.41584 Hz
 SW 10.962 ppm
 FMODE States-Tppl
 F2 - Processing parameters
 SI 1024
 MC2 States-Tppl
 MDW 500.130016 MHz
 SSB 2
 LB 0.00 Hz
 GB 0
 FC 1.40
 F1 - Processing parameters
 SI 1024
 MC2 States-Tppl
 MDW 500.130016 MHz
 SSB 2
 LB 0.00 Hz
 GB 0

