

Supporting information

Synthesis, biological evaluation and automated docking of constrained analogs of the opioid peptide H-Dmt-D-Ala-Phe-Gly-NH₂ using the 4- or 5-methyl substituted 4-amino-1,2,4,5-tetrahydro-2-benzazepin-3-one scaffold.

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Content:

S1-2: Synthesis, optical rotation and HPLC retention times of compounds **3** to **6**

S2-3: Figure S1: The amino acid sequence alignments of (boxed) transmembrane helices (TM1-7) and the intra- and extracellular loops (Ils en ELS) of bovine rhodopsin (OPSD_BOVIN), the human OPRM and OPRD.

S3: IFP bit strings for compound **15**

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Enantiopure β -methylphenylalanine isomers.

The diastereomeric *erythro*-(2*S*,3*S* and 2*R*,3*R*)- and *threo*-(2*S*,3*R* and 2*R*,3*S*)- β -MePhe racemates were obtained by fractional crystallization¹ of the isomeric mixture prepared by the method of Kataoka.² After Cbz-protection, resolution was performed by crystallization of the quinine or quinidine salts according to Kataoka.² The enantiomeric ratio (ee) was determined with FDAA derivatisation³ (after small scale Cbz-deprotection) via RP-HPLC and UV detection at $\lambda=340$ nm. The retention times after derivatisation are: 16.3 min (for *erythro*-(2*S*,3*S*)- β -Me-Phe), 17.1 min (for *erythro*-(2*R*,3*R*)- β -MePhe), 16.33 min (for *threo*-(2*S*,3*R*)- β -Me-Phe) and 17.05 min (for *threo*-(2*R*,3*S*)- β -MePhe).

Enantiopure Boc-*erythro*-(4*S*,5*S*) (3), -*erythro*-(4*R*,5*R*) (4), -*threo*-(4*S*,5*R*) (5) and -*threo*-(4*R*,5*S*)-5-Me-Aba-Gly-OH (6) The enantiopure isomers of **3** to **6** were prepared using the previously described procedure for the racemates.⁴ Briefly, the enantiopure β -methylphenylalanine isomers were phthaloyl protected using methyl-2-((succinimidooxy)carbonyl)benzoate, and coupled with methyl glycinate. The esters were subsequently hydrolyzed, followed by formation of the oxazolidinone by reaction with formaldehyde using p-TosOH catalysis. Ring closure was effected using trifluoromethanesulfonic acid (TFMSA) to provide the Phth-protected *erythro* and *threo*-5-Me-Aba-Gly-OH isomers. After phthaloyl deprotection by hydrazinolysis, final Boc protection was followed by crystallization of the *N,N*-dicyclohexylamine (DCHA) salts in Et₂O.⁵ The crystals contained a small amount of di-Boc hydrazide which could not be removed. Before performing the peptide synthesis, the DCHA salts were converted into the free acid⁵ and the Boc-protected enantiomers were purified by an additional acid-base extraction which removed the di-Boc-hydrazide. The optical rotation data and RP-HPLC retention times of the different stereoisomers are collected in Table 3:

Table S1: Optical rotations and HPLC retention times of **3-6**

Compound	Optical rotation	RP-HPLC
	α_D (solvent)	t_{Ret}
Boc- <i>erythro</i> -(4 <i>S</i> ,5 <i>S</i>)-5-Me-Aba-Gly-OH 3	+90.6° (EtOAc)	15.4 min
Boc- <i>erythro</i> -(4 <i>R</i> ,5 <i>R</i>)-5-Me-Aba-Gly-OH 4	-93.0° (EtOAc)	15.3 min
Boc- <i>threo</i> -(4 <i>S</i> ,5 <i>R</i>)-5-Me-Aba-Gly-OH 5	Not determined*	14.6 min
Boc- <i>threo</i> -(4 <i>R</i> ,5 <i>S</i>)-5-Me-Aba-Gly-OH 6	Not determined*	14.6 min

* HPLC analysis showed that the Boc-*threo*-(4*S*,5*R*)-5-Me-Aba-Gly-OH.DCHA had epimerized to the erythro derivative [46% *threo*-(4*S*,5*R*) + 54% *erythro*-(4*S*,5*S*)] and the Boc-*threo*-(4*R*,5*S*)-5-Me-Aba-Gly-OH to the (4*R*,5*R*) erythro analogue [48% *threo*-(4*R*,5*S*) + 52% *erythro*-(4*R*,5*R*)]

Supplementary Figure S1. The amino acid sequence alignments of (boxed) transmembrane helices (TM1-7) and the intra- and extracellular loops (Ils en ELS) of bovine rhodopsin (OPSD_BOVIN), the human OPRM and OPRD. Conserved reference residues according to the Ballesteros-Weinstein numbering scheme⁶ are indicated in red (N1.50 in TM 1, D2.50 in TM2, R3.50 in TM3, W4.50 in TM4, P5.50 in TM5, P6.50 in TM6, and P7.50 in TM7).

	TM1	IL1
OPRM_HUMAN	[ITAITIMALYSIVCVVGLFG N FLVMYVIVR]	YTKMKT
OPRD_HUMAN	[ALAIAITALYSAVCAVGLL G NVLVMFGIVR]	YTKMKT
ADRB2_HUMAN	[VWVVGMGIVMSLIVLAIIVFG N VLVITAIK]	FERLQT
CCR5_HUMAN	[IAARLLPPLYSLVFI F GFVGNMLVILILIN]	CKRLKS
OPSD_BOVIN	[WQFSMLAAYMFLIIMLGFP I NFLTLYVTVQ]	HKKLRT
	TM2	EL1
OPRM_HUMAN	[ATNIYIFNLALADALATSTL P FQSVNYLMG]	-TWPFQ
OPRD_HUMAN	[ATNIYIFNLALADALATSTL P FQSAKYLME]	-TWPFQ
ADRB2_HUMAN	[VTNYFITSLACADL V MGLAVV P F G AAHILM]	KMWTFQ
CCR5_HUMAN	[MTDIYLLNLAI S DLFFLLTV P F W AH Y AAAQ]	--WDFG
OPSD_BOVIN	[PLNYILLNLAVADL F MV F GGFTTTLYTSLH]	GYFVFG
	TM3	IL2
OPRM_HUMAN	[TILCKIVISIDYNNMFTSIFTLCTMSVD R YIAV]	CHPVKALDFRTP
OPRD_HUMAN	[ELLCKAVLSIDYNNMFTSIFTLTMM S VD R YIAV]	CHPVKALDFRTP
ADRB2_HUMAN	[NFWCEFWTSIDVLCVTASIE T LCVIAVD R YFAI]	TSPFKYQSL L TK
CCR5_HUMAN	[NTMCQLLTGLYF I GF F SG I FF I ILLTID R YLAV]	VHAVFALKARTV
OPSD_BOVIN	[PTGCNLEGFFATLGG E IALW S LVVLAIE R YVVV]	CKPMSNFR F GE
	TM4	EL2
OPRM_HUMAN	[RNAKIINVCN W ILSSAIGLPV M F]	MATTKYRQ G --SID C TLTF S HPTW--YW
OPRD_HUMAN	[AKAKLINIC I WVLASGVGP I MV]	MAVTRPRD G --AV V C M LQ F PSP S W--YW
ADRB2_HUMAN	[NKARVILMV W IVSGLT S FL P IQ]	MHWYRATHQ--EAIN C YAN E TCCD F FT-
CCR5_HUMAN	[TFGVVTSVIT W VAVFAS L PG I I]	--F T RSQ K EGLHY T C S SH F PYSQ Y Q F W
OPSD_BOVIN	[NHAIMGVA F T W VMALACA A AP L V]	-GWSRY I PEGM Q CS C GIDY Y TP H E E T N
	TM5	IL3
OPRM_HUMAN	[ENLLKICV F IFAF I MPV L IITVC Y GL]	MILRLK--SV R ML S GS K E K DR N

IFP bit-strings:

1. hydrophobics
2. aromatic face to face
3. aromatic edge to face
4. H-bond (acceptor-donor)
5. H-bond (donor-acceptor)
6. Ionic (negative-positive)
7. Ionic (positive-negative)

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