

*Supporting Information*

*for*

**Substrate/Product-Targeted NMR Monitoring of Pyrimidine  
Catabolism and Its Inhibition by a Clinical Drug**

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## 1. Preparation

**General.** Reagents and solvents were purchased from standard suppliers and used without further purification. NMR spectra were obtained on a JEOL JNM-A500 (500 MHz) NMR or a JEOL JNM-ECX400 (400 MHz) spectrometer. The coupling constants ( $J$  values) are reported in Hertz. The EI mass spectra were recorded on a JEOL JMS SX102A mass spectrometer.

**[<sup>13</sup>C3]-Ethyl *N*-(2-cyano-3-ethoxyacryloyl)carbamate (2):** A solution of ethyl *N*-(2-cyanoacetyl)carbamate (**1**) (1.57 g, 10.1 mmol) and labeled triethyl orthoformate ( $\text{H}^{13}\text{C}(\text{OCH}_2\text{CH}_3)_3$ ) (99%, <sup>13</sup>C) (1.5 g, 10.1 mmol) in acetic anhydride (4 mL) was refluxed for 7 h. After the reaction mixture was allowed to cool to room temperature, the insoluble solid was collected by filtration and washed with petroleum ether and cold diethyl ether to give compound **2** (1.84 g, 86%) as a white solid; <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 500 MHz)  $\delta$ : 10.58 (s, 1H), 8.57, 8.20 (d,  $J$  = 187.0 Hz, 1H), 4.37 (m, 2H), 4.13 (q,  $J$  = 7.0 Hz, 2H), 1.32 (t,  $J$  = 7.0 Hz, 3H), 1.22 (t,  $J$  = 7.5 Hz, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 125 MHz)  $\delta$ : 173.8, 158.8, 149.9, 113.2, 86.9 (d,  $J_{\text{CC}}$  = 85.8 Hz), 74.6, 62.5, 15.2, 14.1; HRMS (EI):  $m/z$  calcd. for  $\text{C}_8^{13}\text{CH}_{12}\text{O}_4\text{N}_2$  [ $\text{M}^+$ ] 213.0831, found 213.0834.

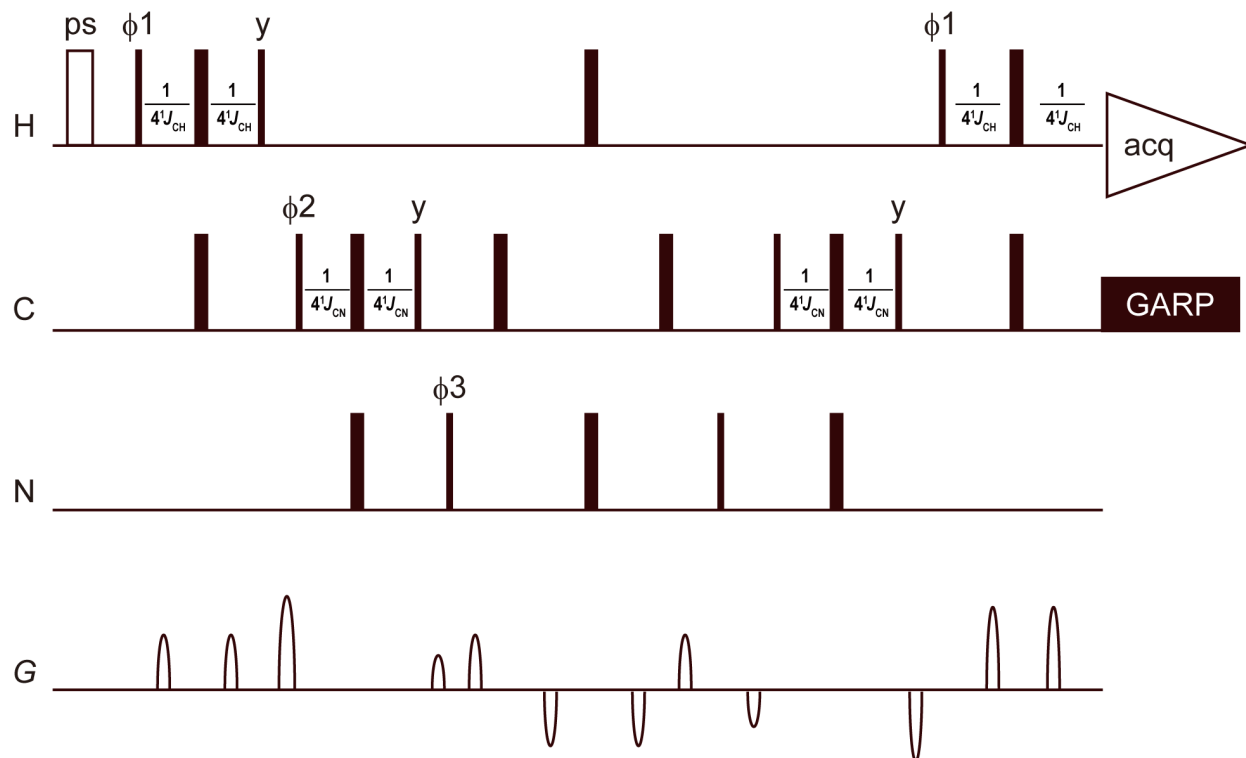
**[<sup>15</sup>N1, <sup>13</sup>C6]-5-Cyanouracil (3):** A solution of compound **2** (1.74 g, 8.18 mmol) and labeled ammonium chloride (<sup>15</sup>NH<sub>4</sub>Cl) (99%, <sup>15</sup>N) (490 mg, 8.99 mmol) in 0.25 M aqueous NaOH (46 mL) was stirred for 4 h at room temperature, and then refluxed for 5 h. The reaction mixture was concentrated in vacuo. The crude residue was recrystallized from water to give compound **3** (1.05 g, 93%) as a white solid; <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 500 MHz)  $\delta$  8.14 (dd,  $J$  = 178.5, 7.5 Hz, 1H); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 100 MHz)  $\delta$  161.1, 152.1 (d,  $J_{\text{CN}}$  = 12.0 Hz), 150.0 (d,  $J_{\text{CN}}$  = 17.7 Hz), 114.6, 87.1 (d,  $J_{\text{CC}}$  = 70.9 Hz); <sup>15</sup>N NMR (DMSO-*d*<sub>6</sub>, 40 MHz) for labeled <sup>15</sup>N  $\delta$  198.6; HRMS (EI):  $m/z$  calcd. for  $\text{C}_4^{13}\text{CH}_3\text{O}_2\text{N}_2^{15}\text{N}$  [ $\text{M}^+$ ] 139.0229, found 139.0229.

**[<sup>15</sup>N1, <sup>13</sup>C6]-Uracil:** A solution of compound **3** (1.38 g, 9.89 mmol) in aqueous 6N HCl (60 mL) was refluxed for 27 h. The reaction mixture was concentrated in vacuo. The crude residue was recrystallized from water to give doubly labeled [<sup>15</sup>N1, <sup>13</sup>C6]-uracil (1.07 g, 95%) as a white solid; <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 400 MHz)  $\delta$  11.0 (s, NH), 10.8 (d,  $J$  = 97.2 Hz, NH), 7.37 (ddd,  $J$  = 182.4, 7.8,

3.6 Hz, 1H), 5.45-5.41 (1H);  $^{13}\text{C}$  NMR (DMSO- $d_6$ , 100 MHz)  $\delta$  164.3, 151.0 (d,  $J_{\text{CN}} = 11.0$  Hz), 142.1 (d,  $J_{\text{CN}} = 10.5$  Hz), 100.2 (d,  $J_{\text{CC}} = 66.1$  Hz);  $^{15}\text{N}$  NMR (DMSO- $d_6$ , 40 MHz) labeled for  $^{15}\text{N}$   $\delta$  127.4 (d,  $J_{\text{CN}} = 10.4$  Hz); HRMS (EI):  $m/z$  calcd. for  $\text{C}_3^{13}\text{CH}_4\text{O}_2\text{N}^{15}\text{N}$   $[\text{M}^+]$  114.0277, found 114.0277.

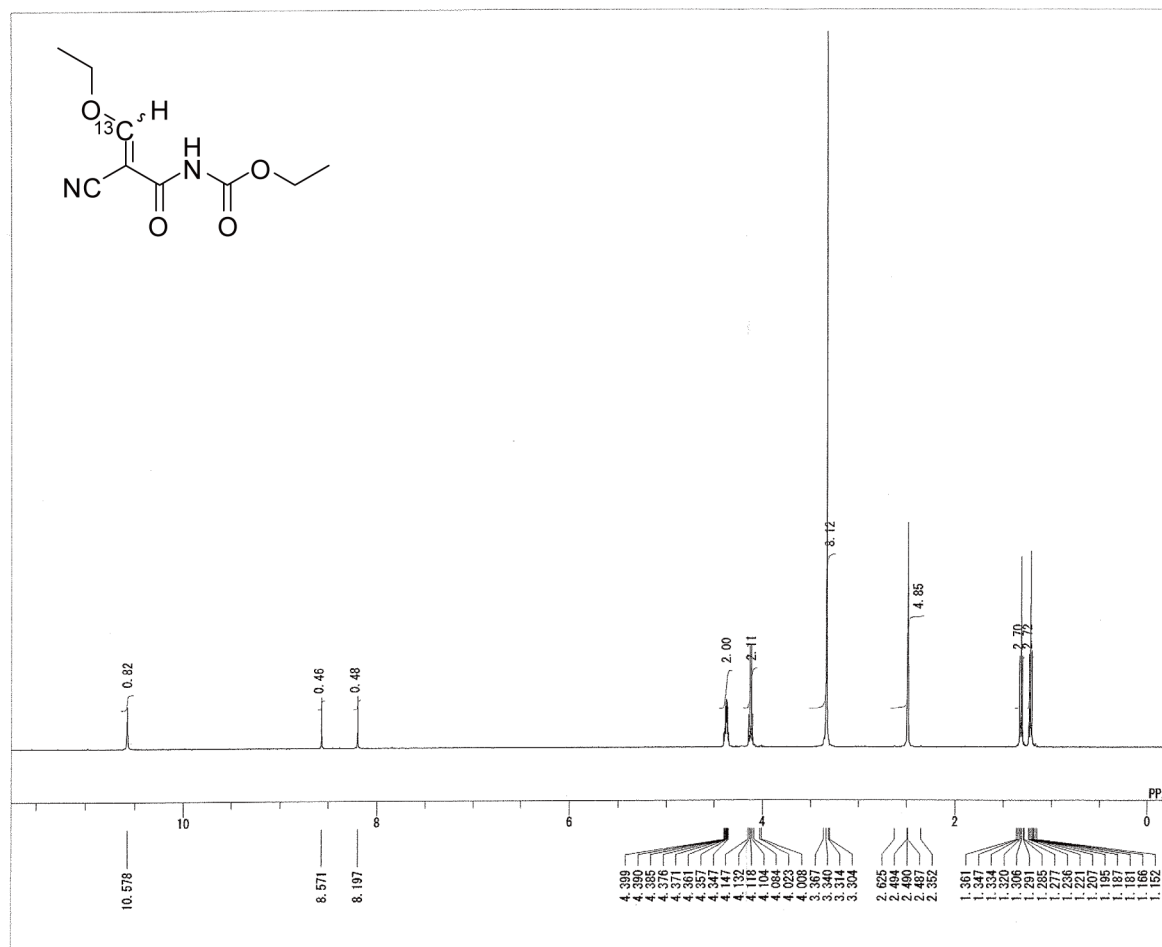
The NMR spectra for compounds **2** and **3** and [ $^{15}\text{N}1$ ,  $^{13}\text{C}6$ ]-uracil are shown below in Figures S2-S9.

## 2. Pulse Scheme for One-dimensional $^1\text{H}\{-^{13}\text{C}\{-^{15}\text{N}\}$ Triple Resonance NMR

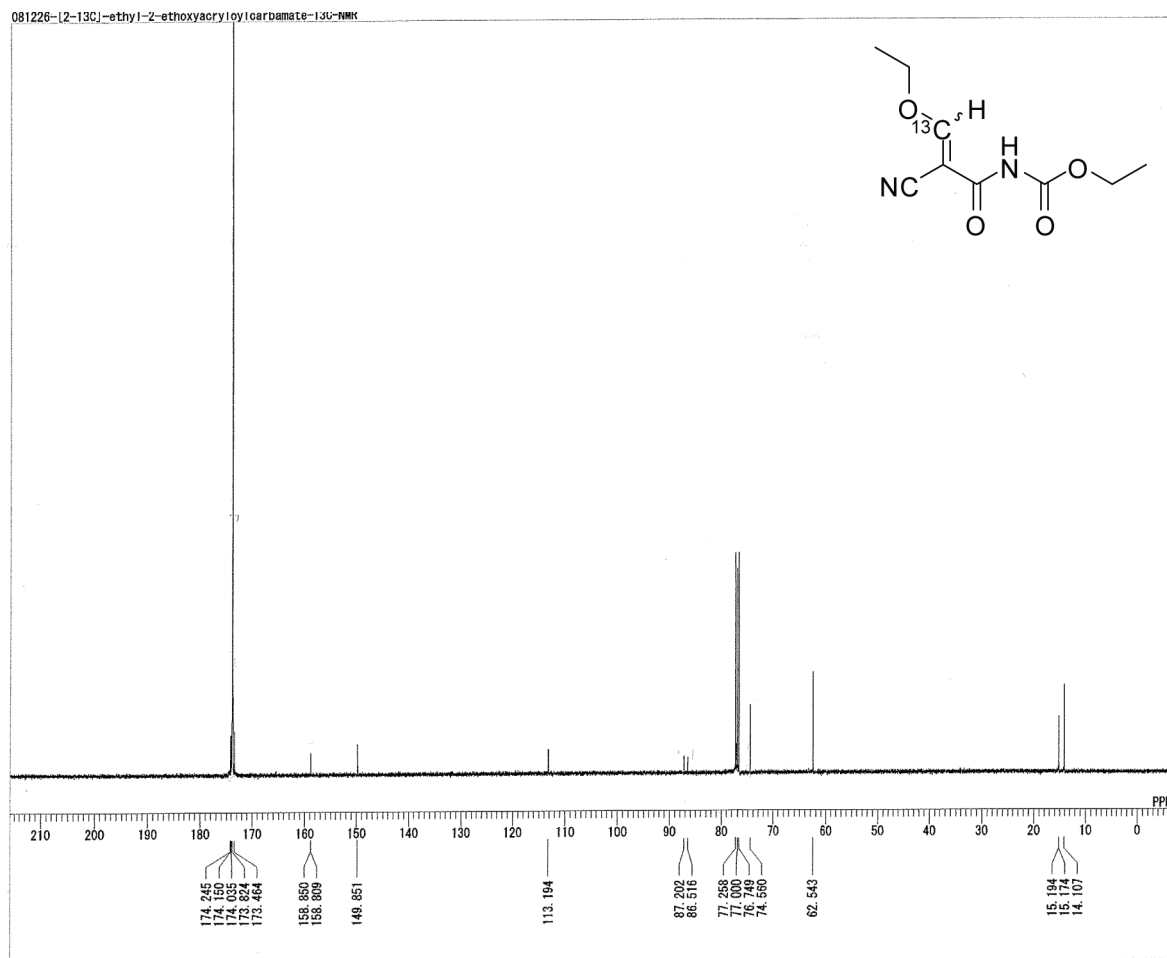


**Figure S1.** Pulse scheme of one-dimensional  $^1\text{H}\{-^{13}\text{C}\{-^{15}\text{N}\}$  triple resonance NMR experiments used in this study. The narrow and broad filled bars represent  $90^\circ$  and  $180^\circ$  pulses, respectively. All pulses have phase =  $x$  unless otherwise indicated. Parameters for detection of  $^1\text{H}\{-^{13}\text{C}\{-^{15}\text{N}\}$  in labeled uracil were transmitter offsets of  $^{13}\text{C} = 143$  ppm and  $^{15}\text{N} = 110$  ppm, and delay intervals of  $1/4^1J_{\text{CH}} = 1.36$  ms and  $1/4^1J_{\text{CN}} = 32$  ms. Those for detection of  $^1\text{H}\{-^{13}\text{C}\{-^{15}\text{N}\}$  in labeled  $\beta$ -alanine were  $^{13}\text{C} = 36.3$  ppm,  $^{15}\text{N} = 30.5$  ppm,  $1/4^1J_{\text{CH}} = 1.72$  ms, and  $1/4^1J_{\text{CN}} = 48$  ms. PS denotes a pre-saturation pulse (1.5 ms) used for water suppression. The phase cycle is  $\phi1 = x, -x$ ;  $\phi2 = 2(x), 2(-x)$ ;  $\phi3 = 4(y), 4(-y)$  and receiver =  $2(y), 4(-y), 2(y)$ . During the detection period,  $^{13}\text{C}$  GARP decoupling is used. All gradients were applied along the  $z$  axis.

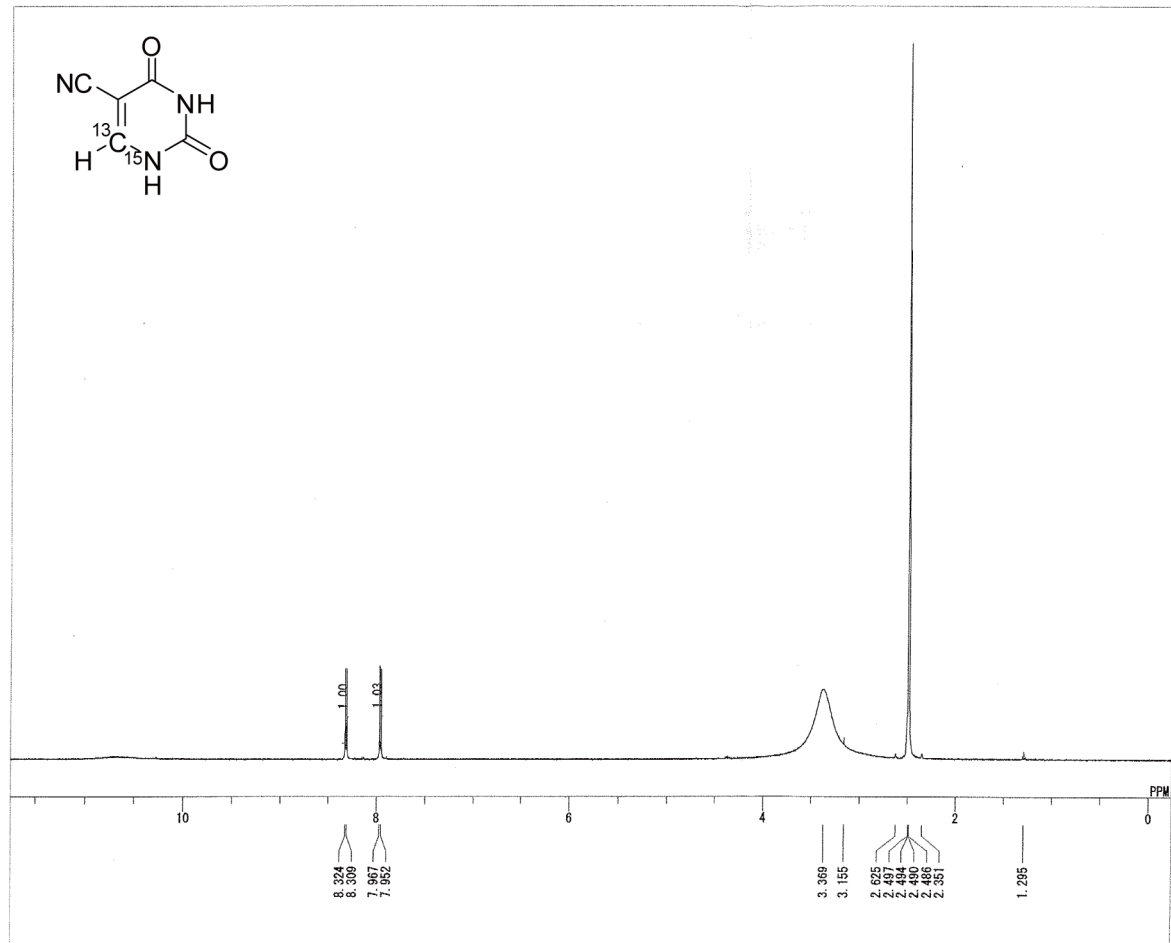
### 3. NMR Spectra of Labeled Compounds



**Figure S2.**  $^1\text{H}$  NMR spectrum of compound **2** in  $\text{DMSO}-d_6$  (500 MHz).



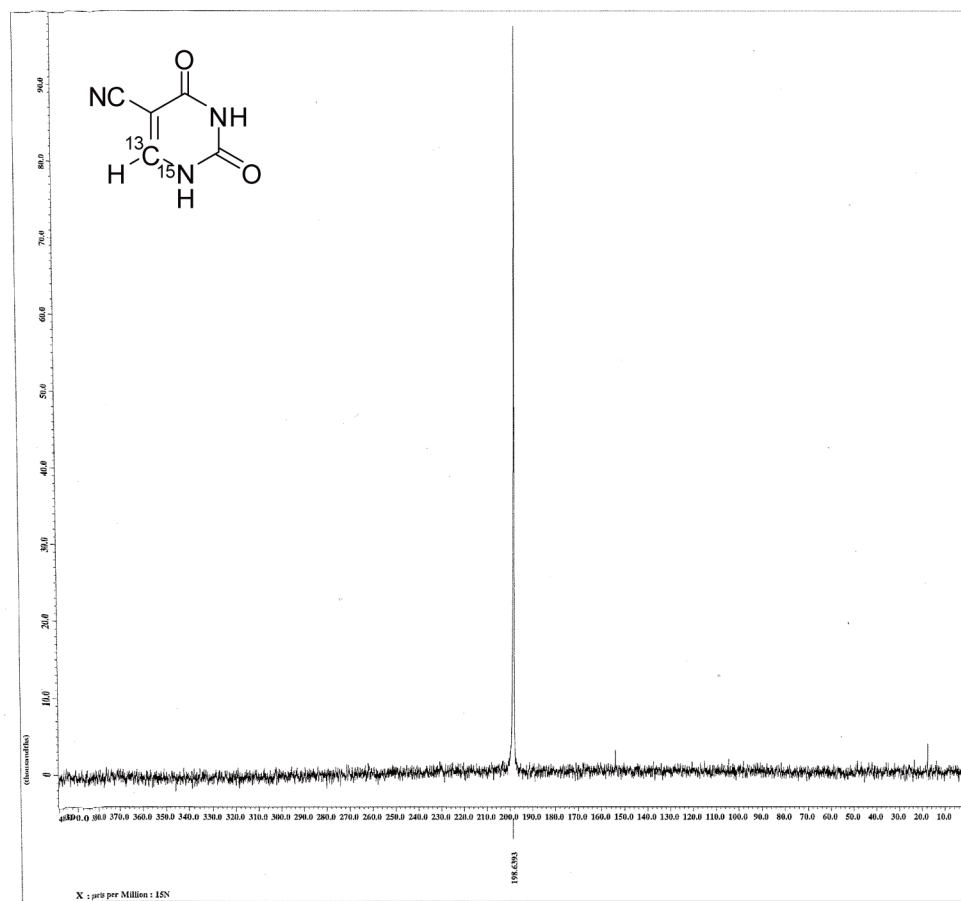
**Figure S3.** <sup>13</sup>C NMR spectrum of compound **2** in DMSO-*d*<sub>6</sub> (125 MHz).



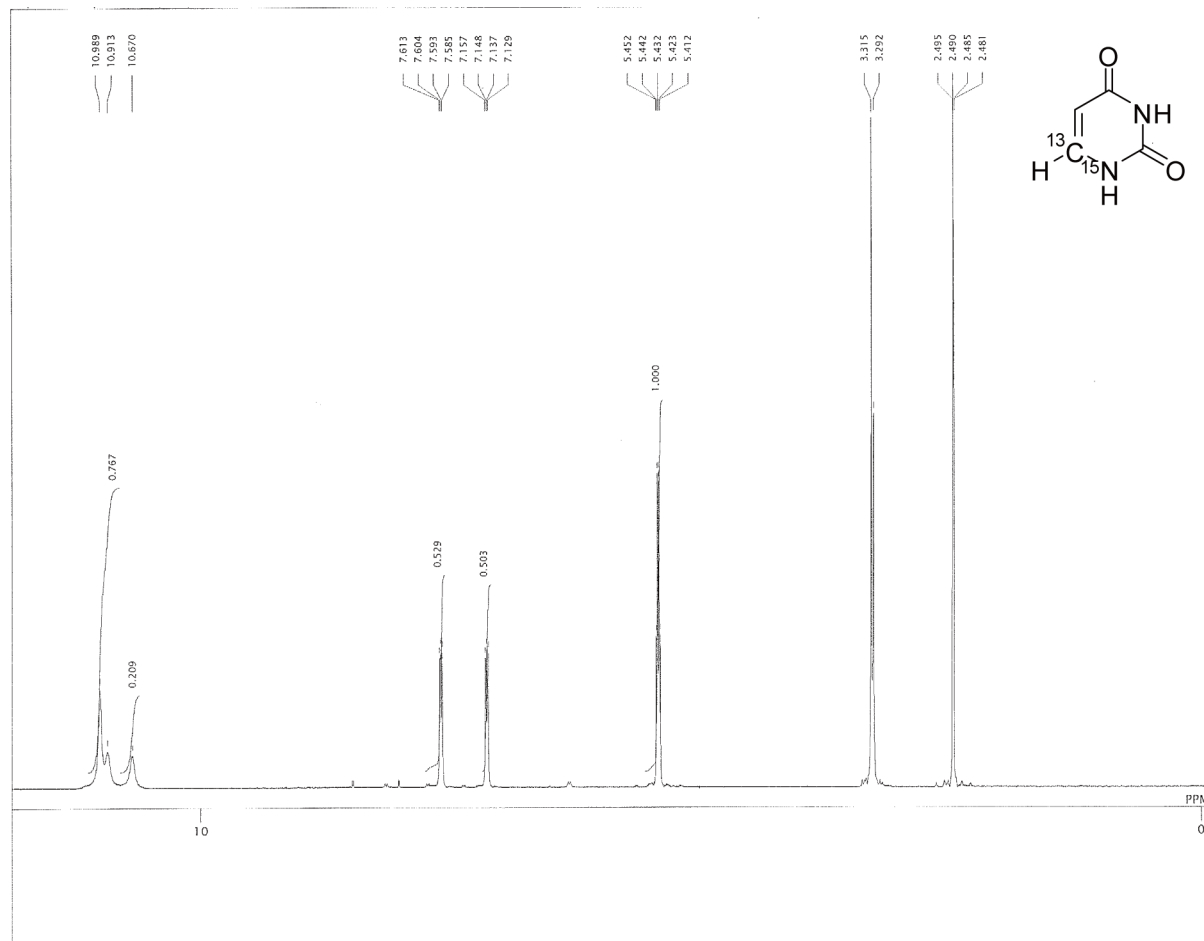
**Figure S4.**  $^1\text{H}$  NMR spectrum of compound **3** in  $\text{DMSO}-d_6$  (500 MHz).



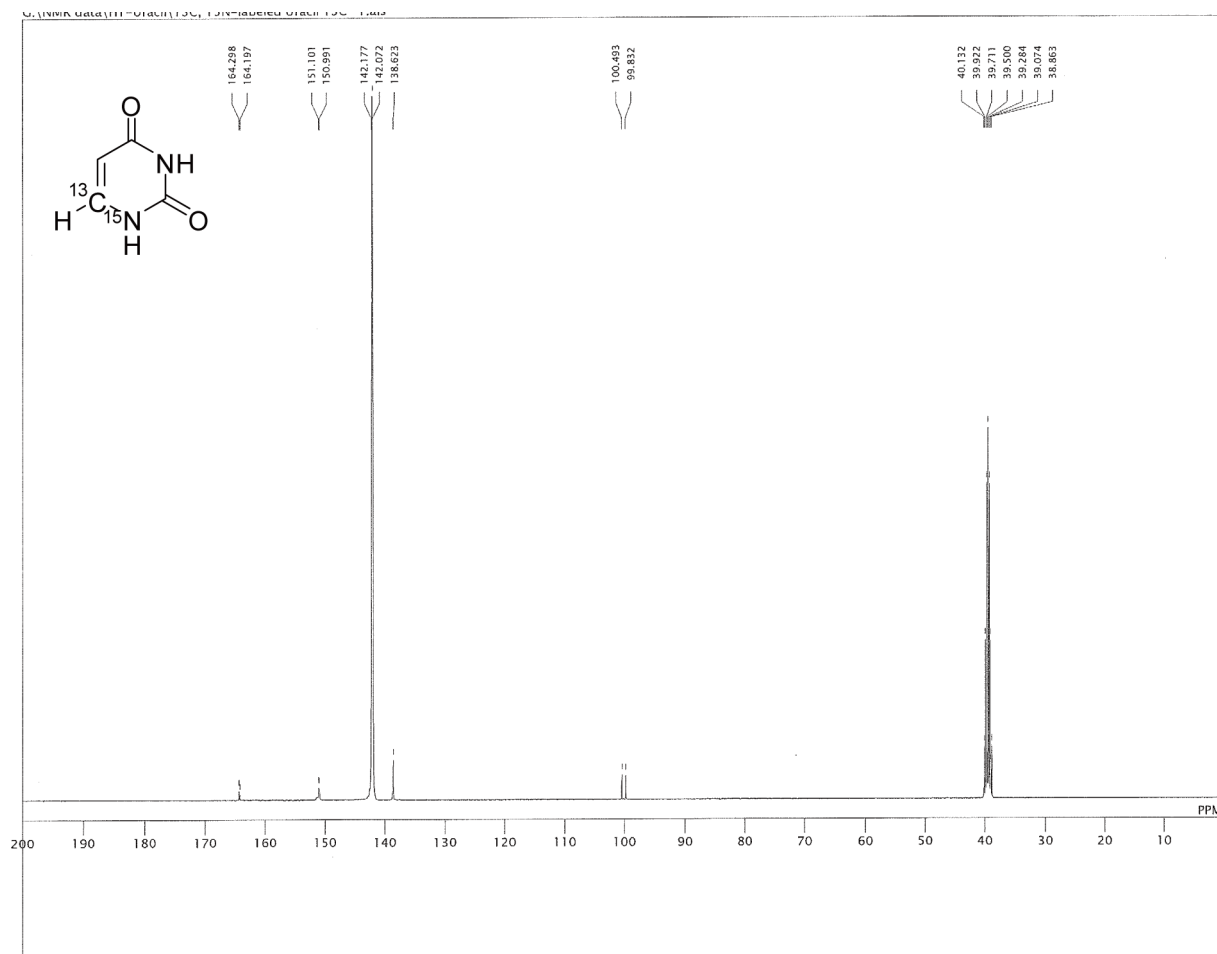




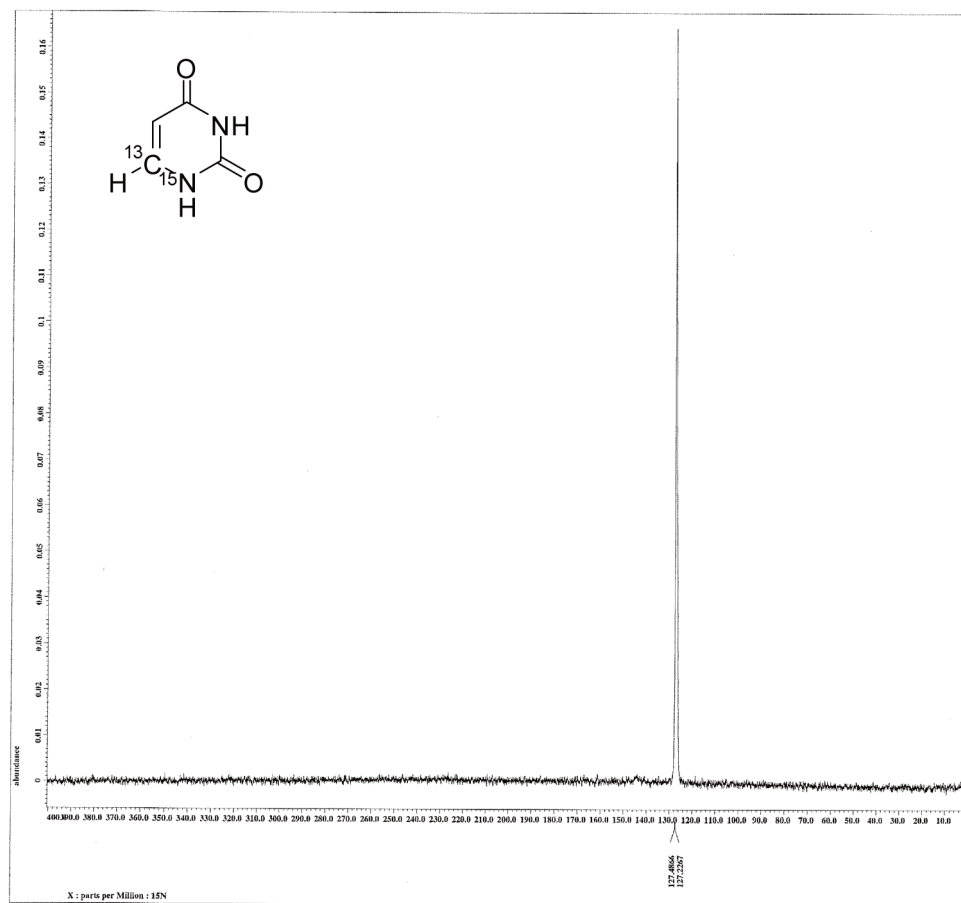
**Figure S6.**  $^{15}\text{N}$  NMR spectrum of compound **3** in  $\text{DMSO}-d_6$  (40 MHz).



**Figure S7.**  $^1\text{H}$  NMR spectrum of  $[\text{}^{15}\text{N}1, \text{}^{13}\text{C}6]\text{-uracil}$  in  $\text{DMSO-}d_6$  (400 MHz).

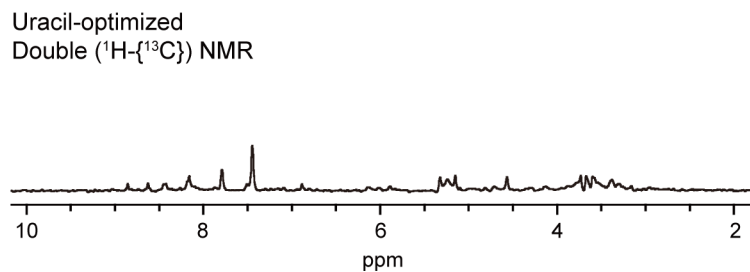


**Figure S8.**  $^{13}\text{C}$  NMR spectrum of  $[\text{}^{15}\text{N}1, \text{}^{13}\text{C}6]\text{-uracil}$  in  $\text{DMSO-}d_6$  (100 MHz).



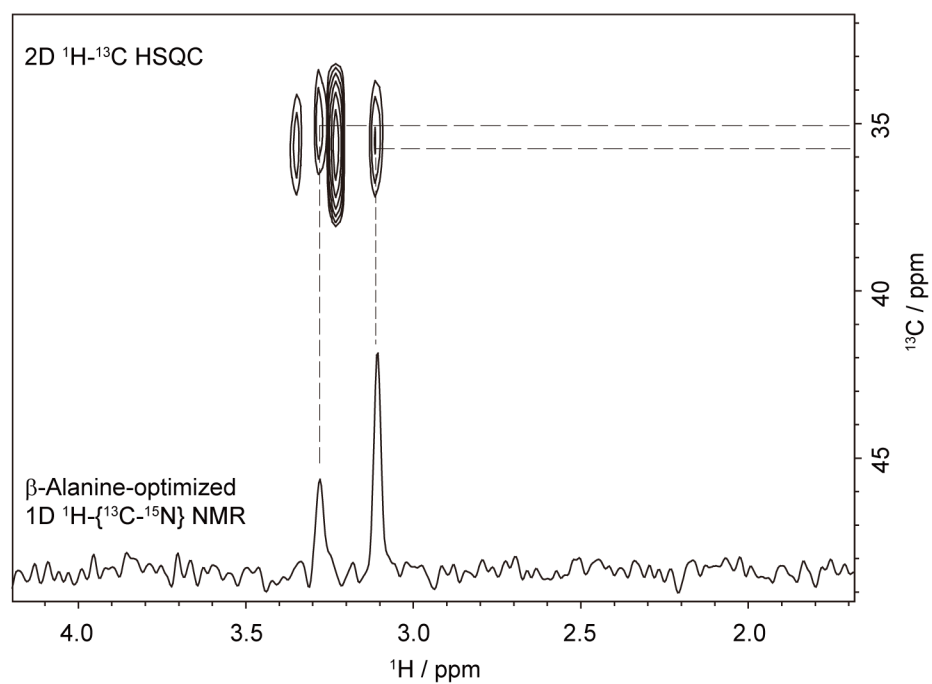
**Figure S9.**  $^{15}\text{N}$  NMR spectrum of  $[\text{}^{15}\text{N}1, \text{}^{13}\text{C}6]\text{-uracil}$  in  $\text{DMSO-}d_6$  (40 MHz).

#### 4. Uracil-Optimized 1D $^1\text{H}\{-^{13}\text{C}\}$ HSQC Spectrum

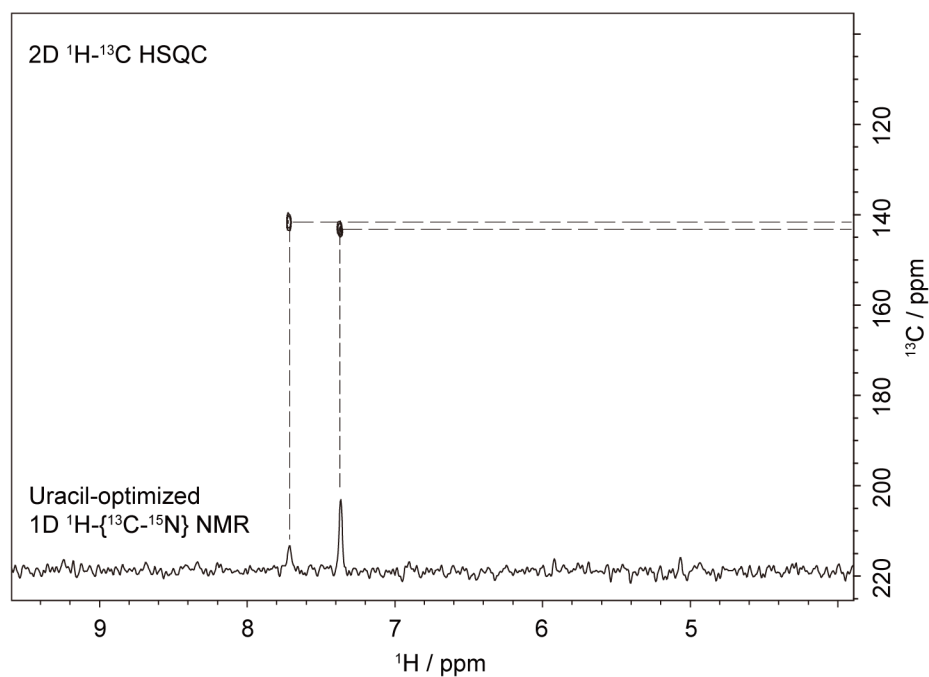


**Figure S10.** Uracil-optimized 1D  $^1\text{H}\{-^{13}\text{C}\}$  double-resonance NMR spectrum of a mouse liver lysate (10% v/v) containing [ $^{15}\text{N}1$ ,  $^{13}\text{C}6$ ]-labeled uracil (0.5 mM) in 10 mM Tris-HCl (pH 8.0), 0.5 mM EDTA, 0.5 mM 2-mercaptoethanol, 2 mM dithiothreitol, 5 mM  $\text{MgCl}_2$ , 2 mM NADPH, incubated at 37 °C for 1 h. Uracil-optimized parameters: chemical-shift ( $^{13}\text{C}$  = 142.5 ppm) and coupling-constant data ( $J_{\text{C-H}}$  = 184 Hz).

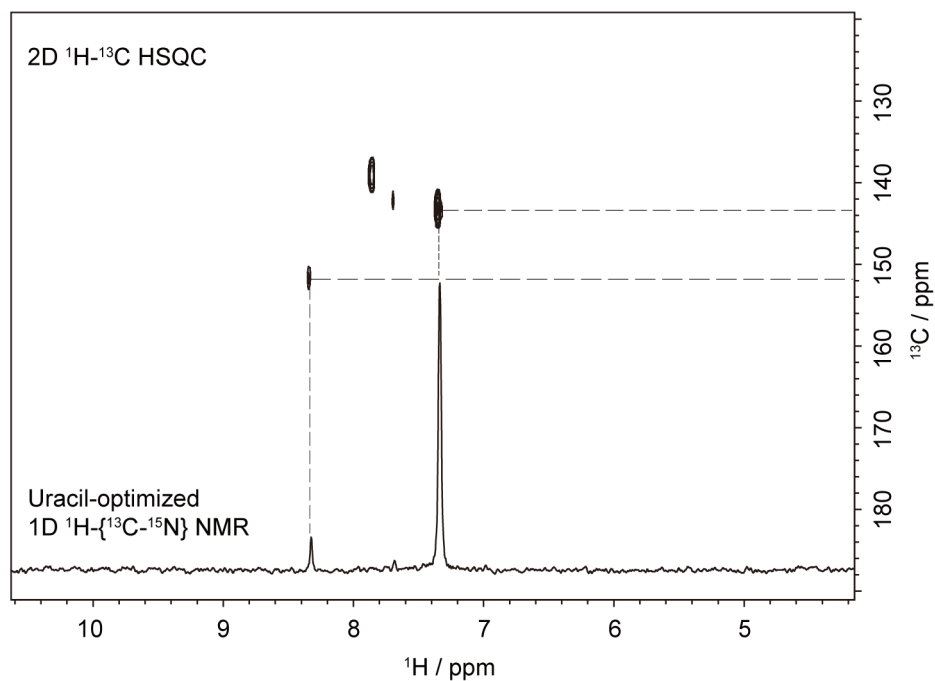
## 5. 2D $^1\text{H}$ - $^{13}\text{C}$ HSQC Spectra



**Figure S11.** 2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC spectrum of the extracts from kidney of a mouse injected intraperitoneally with a solution of [ $^{15}\text{N}1$ ,  $^{13}\text{C}6$ ]-labeled uracil ( $300\ \mu\text{g g}^{-1}$  body weight). The 2D HSQC spectrum is merged with the  $\beta$ -alanine-optimized 1D triple-resonance NMR spectrum of the kidney extracts (Figure 5A).



**Figure S12.** 2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC spectrum of the extracts from liver of a mouse injected intraperitoneally with a solution of [ $^{15}\text{N}1$ ,  $^{13}\text{C}6$ ]-labeled uracil ( $300\ \mu\text{g g}^{-1}$  body weight). The 2D HSQC spectrum is merged with the uracyl-optimized 1D triple-resonance NMR spectrum of the liver extracts (Figure 5B).



**Figure S13.** 2D  $^1\text{H}$ - $^{13}\text{C}$  HSQC spectrum of the extracts from kidney of a mouse injected intraperitoneally with a solution of [ $^{15}\text{N}1$ ,  $^{13}\text{C}6$ ]-labeled uracil ( $300\text{ }\mu\text{g g}^{-1}$  body weight). The 2D HSQC spectrum is merged with the uracil-optimized 1D triple-resonance NMR spectrum of the kidney extracts (Figure 5B).