

Hg(II)-mediated intramolecular cyclization reaction in aqueous media and its application as Hg(II) selective indicator

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Contents

General Experimental	S2
X-ray structural data of 5a	S4
Synthetic Experimental	S7
NMR Spectra	S10

General Experiments

Reagents were purchased at the highest commercial quality and used without further purification, unless otherwise stated. Yields of synthesized compounds were measured after chromatographic purification. UV-vis spectra were recorded on a UV 1800 (Shimadzu) spectrophotometer. Proton and ^{13}C -NMR spectra were measured at 25 °C using Varian Unity Innova 400 and 500 instruments. The electrospray ionization (ESI) source was coupled to a hybrid quadrupole orthogonal time-of-flight (Q-TOF) mass spectrometer (SYNAPT G2, Waters, MS Technologies, Manchester, U.K.) was used to mass spectra acquirement in positive- and negative-ion mode. A capillary and cone voltage of $\pm 3.0\text{kV}$ and 30 V, and capillary temperature of 120 °C were used for both polarities, respectively. The desolvation source conditions were as follows; for the desolvation gas 800 L/h was used and the desolvation temperature was kept at 600 °C. Data acquisition took place over the mass range of m/z 50 to m/z 1200 for MS modes. The sample was introduced into the ESI source at a constant flow rate of 20 $\mu\text{L}/\text{min}$ by using an external syringe pump (HARVARD 11Plus, Holliston, MA, USA).

Stock solutions of all of the compounds studied were made up in DMSO with the final concentrations being between $1.3 \times 10^{-5}\text{ M}$ and $9.5 \times 10^{-6}\text{ M}$. ACS grade solvents were purchased and used without purification. The stock solutions were appropriately diluted with the solvents for the ensuing studies.

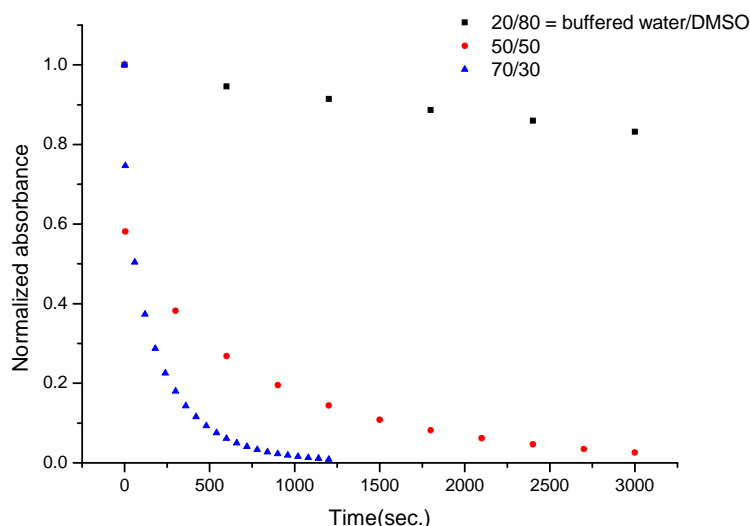


Figure S1. Plots of absorbance change of **6** ($1.1 \times 10^{-5}\text{ M}$) at 608 nm as a function of time with 2 equiv of Hg(II) in various percentages of buffered water and DMSO (v/v) (10 mM PBS buffer, pH = 7.0).

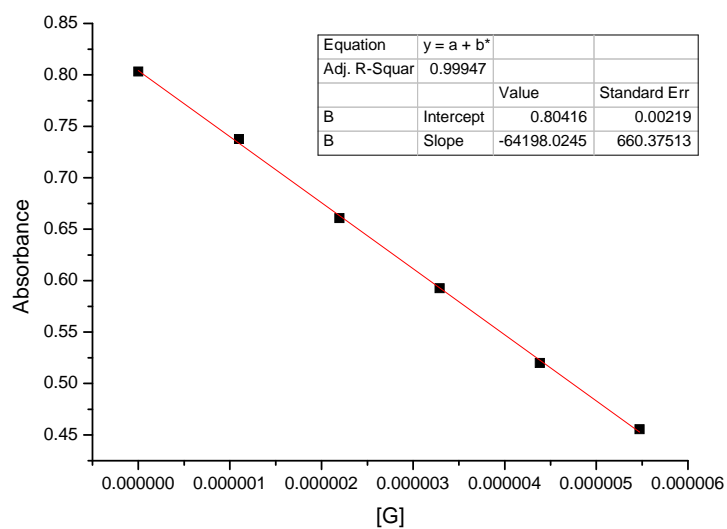


Figure S2. Plot of absorbance change of **6** (9.56×10^{-6} M) at 608 nm as a function of Hg(II) concentration in DMSO/water (3:7, v/v) (10 mM PBS buffer, pH = 7.0). Data were acquired 20 min. after the addition of a given amount of Hg(II) from 0 to 5.5 μ M.

Table 1. Crystal data and structure refinement for **5a**

Identification code	5a	
Empirical formula	C13 H14 Cl Hg N O3	
Formula weight	468.29	
Temperature	296(2) K	
Wavelength	0.71073 Å	
Crystal system	Triclinic	
Space group	P-1	
Unit cell dimensions	a = 6.98880(10) Å	$\alpha = 92.8870(10)^\circ$.
	b = 10.8795(2) Å	$\beta = 91.0330(10)^\circ$.
	c = 19.1331(4) Å	$\gamma = 90.1320(10)^\circ$.
Volume	1452.69(5) Å ³	
Z	4	
Density (calculated)	2.141 g/cm ³	
Absorption coefficient	10.781 mm ⁻¹	
F(000)	880	
Crystal size	0.220 x 0.180 x 0.050 mm ³	
Theta range for data collection	1.07 to 28.29°.	
Index ranges	-9 ≤ h ≤ 9, -14 ≤ k ≤ 14, -25 ≤ l ≤ 25	
Reflections collected	38046	
Independent reflections	7193 [R(int) = 0.0555]	
Completeness to theta = 28.29°	99.3 %	
Absorption correction	'SADABS'	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	7193 / 21 / 293	
Goodness-of-fit on F ²	1.014	
Final R indices [I > 2σ(I)]	R1 = 0.0453, wR2 = 0.1227	
R indices (all data)	R1 = 0.0773, wR2 = 0.1459	
Largest diff. peak and hole	1.793 and -1.211 e.Å ⁻³	

Table 2. Atomic coordinates ($\times 10^4$) and equivalent isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for cho. $U(\text{eq})$ is defined as one third of the trace of the orthogonalized U^{ij} tensor.

	x	y	z	$U(\text{eq})$
Hg(1)	-71(1)	-1524(1)	2987(1)	50(1)
Hg(2)	4903(1)	6156(1)	2978(1)	49(1)
Cl(1)	-19(5)	-3616(2)	2770(2)	71(1)
Cl(2)	4982(5)	8213(2)	2751(2)	70(1)
N(1)	2000(12)	2300(8)	5360(4)	53(2)
N(2)	7008(11)	2765(8)	5347(4)	50(2)
O(1)	4339(10)	2301(6)	2624(3)	53(2)
O(2)	7082(12)	3786(7)	5632(4)	64(2)
O(3)	7458(13)	1808(8)	5648(4)	75(2)
O(4)	-639(10)	2252(6)	2621(4)	52(2)
O(5)	2100(13)	1322(7)	5636(4)	68(2)
O(6)	2503(13)	3299(7)	5651(4)	73(2)
C(1)	-766(15)	1005(10)	2486(5)	53(2)
C(2)	-135(13)	342(8)	3024(5)	44(2)
C(3)	360(12)	1234(8)	3583(5)	42(2)
C(4)	20(12)	2379(8)	3302(5)	42(2)
C(5)	351(13)	3516(9)	3666(6)	50(2)
C(6)	1013(12)	3470(8)	4341(5)	45(2)
C(7)	1273(12)	2315(8)	4628(5)	41(2)
C(8)	977(12)	1198(8)	4267(5)	41(2)
C(9)	-1548(18)	638(12)	1779(6)	70(3)
C(10)	-140(30)	770(20)	1197(13)	170(10)
C(11)	1380(30)	-230(20)	1159(13)	150(8)
C(12)	3020(60)	-460(50)	670(30)	400(30)
C(13)	3430(60)	-1810(40)	750(20)	290(20)
C(14)	4245(14)	3561(9)	2461(5)	47(2)
C(15)	4821(14)	4303(8)	3025(5)	45(2)
C(16)	5313(12)	3499(8)	3578(5)	41(2)
C(17)	5972(12)	3664(8)	4263(5)	42(2)
C(18)	6320(12)	2590(8)	4618(5)	42(2)
C(19)	6025(12)	1429(8)	4334(5)	47(2)
C(20)	5375(14)	1260(8)	3655(5)	49(2)

C(21)	5022(13)	2290(8)	3300(5)	45(2)
C(22)	3455(17)	3788(11)	1766(6)	69(3)
C(23)	4780(20)	3520(18)	1181(9)	113(6)
C(24)	6530(30)	4270(20)	1193(14)	162(9)
C(25)	6470(40)	5470(30)	930(20)	115(11)
C(26)	8420(50)	5930(40)	800(20)	290(20)
C(25')	7850(60)	4660(30)	660(20)	154(16)

Experimental Section

General procedure for synthesis of benzofuran derivative by Hg(II)

To a well-stirred solution of 2-alkynylhydroxybenzene (0.086 mmol) in DMSO (4.1 mL) was added HgCl₂ (1.03 mL, 0.17 M in H₂O). The reaction mixture was stirred at room temperature. The reaction mixture was poured into H₂O, and the product was extracted with Et₂O/EtOAc. The combined organic layers were dried over anhydrous Na₂SO₄ and then filtered; the filtrate was concentrated under reduced pressure to obtain the residue. The residue was purified over silica gel to afford the benzofuran derivative.

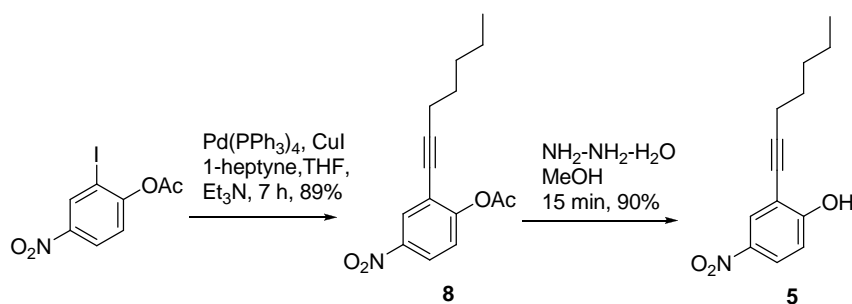
(5-nitro-2-phenylbenzofuran-3-yl)mercury(II) chloride (3a)

By following the general procedure, the reaction of **3** (20.6 mg, 0.086 mmol) yielded **3a** (37.9 mg, 93% yield).

¹H NMR (400 MHz, DMSO) δ 9.17 (d, *J* = 2.5 Hz, 1H), 8.19 (dd, *J* = 9.0, 2.5 Hz, 1H), 8.08 (m, 2H), 7.83 (d, *J* = 9.0 Hz, 1H), 7.54 (m, 3H). HRMS–ESI: *m/z* [M + Cl]⁺ calcd for C₁₄H₈HgNO₃: 509.9587; found: 509.9564.

2-((3-nitrophenyl)ethynyl)phenol (**4**)

1-Iodo-3-nitrobenzene (0.21 g, 0.85 mmol) was added to the solution of 2-ethynylphenol (0.10 g, 0.85 mmol) in anhydrous THF (7 mL) and triethylamine (0.7 mL), and the resulting mixture was flushed with argon and then Pd(PPh₃)₂Cl₂ (0.018 g, 0.025 mmol) and CuI (0.003 g, 0.013 mmol) were added to the solution. The resulting mixture was heated at 60 °C for 3 h. After cooling to room temperature, the reaction mixture was poured into an aqueous saturated solution of NaCl and the product was extracted with EtOAc. The combined organic layers were dried over anhydrous Na₂SO₄ and then filtered; the filtrate was concentrated under reduced pressure to obtain the residue. The residue was purified over silica gel to afford **4** (0.142 g, 70%). ¹H-NMR (400 MHz, CDCl₃) [ppm] ¹H NMR (400 MHz, CDCl₃) δ 8.39 (dd, *J* = 1.8, 1.8 Hz, 1H), 8.21 (ddd, *J* = 8.3, 2.3, 1.0 Hz, 1H), 7.83 (m, 1H), 7.56 (dd, *J* = 7.9, 7.9 Hz, 1H), 7.45 (dd, *J* = 7.7, 1.5 Hz, 1H), 7.32 (ddd, *J* = 8.3, 7.4, 1.7 Hz, 1H), 7.00 (dd, *J* = 8.3, 0.7 Hz, 1H), 6.94 (ddd, *J* = 7.6, 7.6, 1.1 Hz, 1H), 5.74 (s, 1H). ¹³C-NMR (100 MHz, DMSO-*d*₆) [ppm] ¹³C NMR (101 MHz, CDCl₃) δ 156.93, 148.40, 137.35, 132.29, 131.52, 129.77, 126.55, 124.53, 123.56, 120.89, 115.35, 108.78, 93.70, 86.15. HRMS–ESI: *m/z* [M - H]⁺ calcd for C₁₄H₈NO₃: 238.0503; found: 238.0503.



Scheme S1. Synthetic scheme of **5**.

2-(hept-1-ynyl)-4-nitrophenyl acetate (**8**)

To a well stirred solution of 2-iodo-4-nitrophenyl acetate (0.20 g, 0.65 mmol) in THF (6 mL) and Et₃N (0.6 mL) were added Pd(PPh₃)₄ (0.012 g, 0.01 mmol), 1-heptyne (0.13 mL, 0.98 mmol). After 5-10 min. CuI (0.004 g, 0.02 mmol) was added and the resulting solution was stirred at room temperature for 7 h. Then the reaction mixture was extracted with EtOAc. The combined organic layers were dried over anhydrous Na₂SO₄ then filtered, and the filtrate was concentrated under reduced pressure to afford a residue. The residue was purified over silica gel to afford **8** (0.19 g, 89%). ¹H-NMR (400 MHz, CDCl₃) [ppm] 8.31 (d, *J* = 2.8 Hz, 1H), 8.14 (dd, *J* = 8.9, 2.8 Hz, 1H), 7.25 (d, *J* = 8.9 Hz, 1H), 2.44 (t, *J* = 7.1 Hz, 2H), 2.37 (s, 3H), 1.61 (m, 2H), 1.40 (m, 4H), 0.93 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) [ppm] 168.1, 156.3, 145.5, 128.7, 123.8, 123.3, 120.0, 98.9, 74.1, 31.2, 28.3, 22.4, 21.0, 19.7, 14.1. HRMS–ESI: *m/z* [M - CH₃CO]⁺ calcd for C₁₃H₁₄NO₃: 232.0974; found: 232.0976.

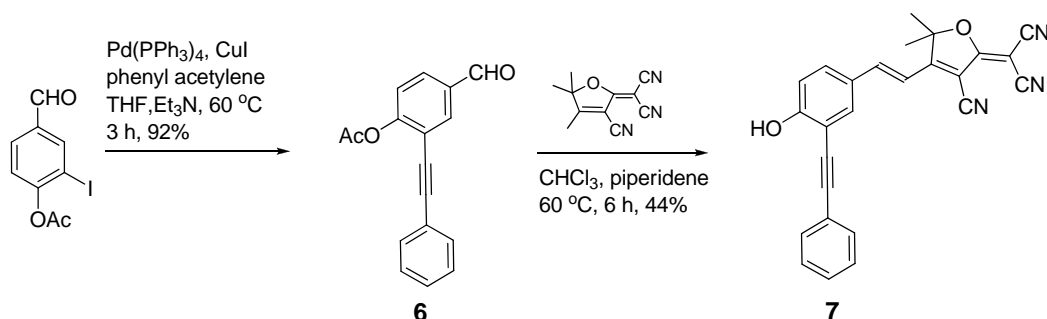
2-(hept-1-ynyl)-4-nitrophenol (**5**)

Compound **8** (0.065 g, 0.23 mmol) was dissolved in MeOH (7 mL). After the addition of hydrazine monohydrate (0.016 mL, 0.34 mmol), the reaction mixture was stirred at room temperature for 15 min. After completion of reaction dil HCl was added and the solvent was removed under reduced pressure to afford a residue. The residue was purified over silica gel to afford **5** (0.05 g, 90%).

¹H-NMR (400 MHz, CDCl₃) [ppm] 8.22 (d, *J* = 2.8 Hz, 1H), 8.10 (dd, *J* = 9.1, 2.8 Hz, 1H), 7.01 (d, *J* = 9.1 Hz, 1H), 6.48 (bs, 1H), 2.50 (t, *J* = 7.2 Hz, 2H), 1.66 (m, 2H), 1.39 (m, 4H), 0.93 (t, *J* = 7.2 Hz, 3H), ¹³C-NMR (100 MHz, CDCl₃) [ppm] 161.6, 141.3, 127.8, 125.7, 114.9, 111.5, 100.9, 72.8, 31.3, 28.3, 22.4, 19.7, 14.1. HRMS–ESI: *m/z* [M - H]⁺ calcd for C₁₃H₁₄NO₃: 232.0974; found: 232.0975.

(5-nitro-2-pentylbenzofuran-3-yl)mercury(II) chloride (**5a**)

By following the general procedure, the reaction of **5** (20.0 mg, 0.086 mmol) yielded **5a** (38.2 mg, 95% yield). ¹H-NMR (400 MHz, DMSO) [ppm] 8.87 (d, *J* = 2.2 Hz, 1H), 8.09 (dd, *J* = 9.0, 2.5 Hz, 1H), 7.67 (dd, *J* = 9.0, 0.4 Hz, 1H), 2.85 (t, *J* = 7.5 Hz, 2H), 1.70 (dd, *J* = 14.6, 7.5 Hz, 2H), 1.30 (m, 4H), 0.87 (t, *J* = 7.2 Hz, 3H), HRMS–ESI: *m/z* [M + Cl]⁺ calcd for C₁₃H₁₄Cl₂HgNO₃: 504.0057; found: 504.0060.



Scheme S2. Synthetic scheme of **7**.

4-formyl-2-(phenylethynyl)phenyl acetate (**6**)

To a well-stirred solution of iodoaldehyde acetate (0.205 g, 0.71 mmol) in THF (8 mL) and Et₃N (0.8 mL) were added phenyl acetylene (0.16 mL, 1.06 mmol), Pd(PPh₃)₄ (0.25 g, 0.021 mmol), and CuI (0.002 g, 0.011 mmol). The resulting mixture was heated at 60 °C for 3 h. After cooling to room temperature, the reaction mixture was poured into an aqueous saturated solution of NaCl and the product was extracted with EtOAc. The combined organic layers were dried over anhydrous Na₂SO₄ and then filtered; the filtrate was concentrated under reduced pressure to obtain the residue. The residue was purified over silica gel to afford **6** (0.174 g, 92%).

¹H NMR (400 MHz, CDCl₃) [ppm] 9.99 (d, *J* = 0.4 Hz, 1H), 8.10 (dd, *J* = 7.9, 0.4 Hz, 1H), 7.88 (dd, *J* = 8.3, 2.0 Hz, 1H), 7.55 – 7.47 (m, 2H), 7.40 – 7.35 (m, 3H), 7.32 (ddd, *J* = 8.3, 0.4, 0.4 Hz, 1H), 2.41 (s, 3H), ¹³C NMR (100 MHz, CDCl₃) [ppm] 190.5, 168.4, 156.1, 134.9, 134.3, 131.9, 130.4, 129.3, 128.7, 123.5, 122.5, 119.0, 95.9, 83.2, 21.1. HRMS–ESI: *m/z* [M – CH₃CO]⁺ calcd for C₁₅H₉O₂: 221.0608; found: 221.0610.

(E)-2-(3-cyano-4-(4-hydroxy-3-(phenylethynyl)styryl)-5,5-dimethylfuran-2(5H)-ylidene)malononitrile (**7**)

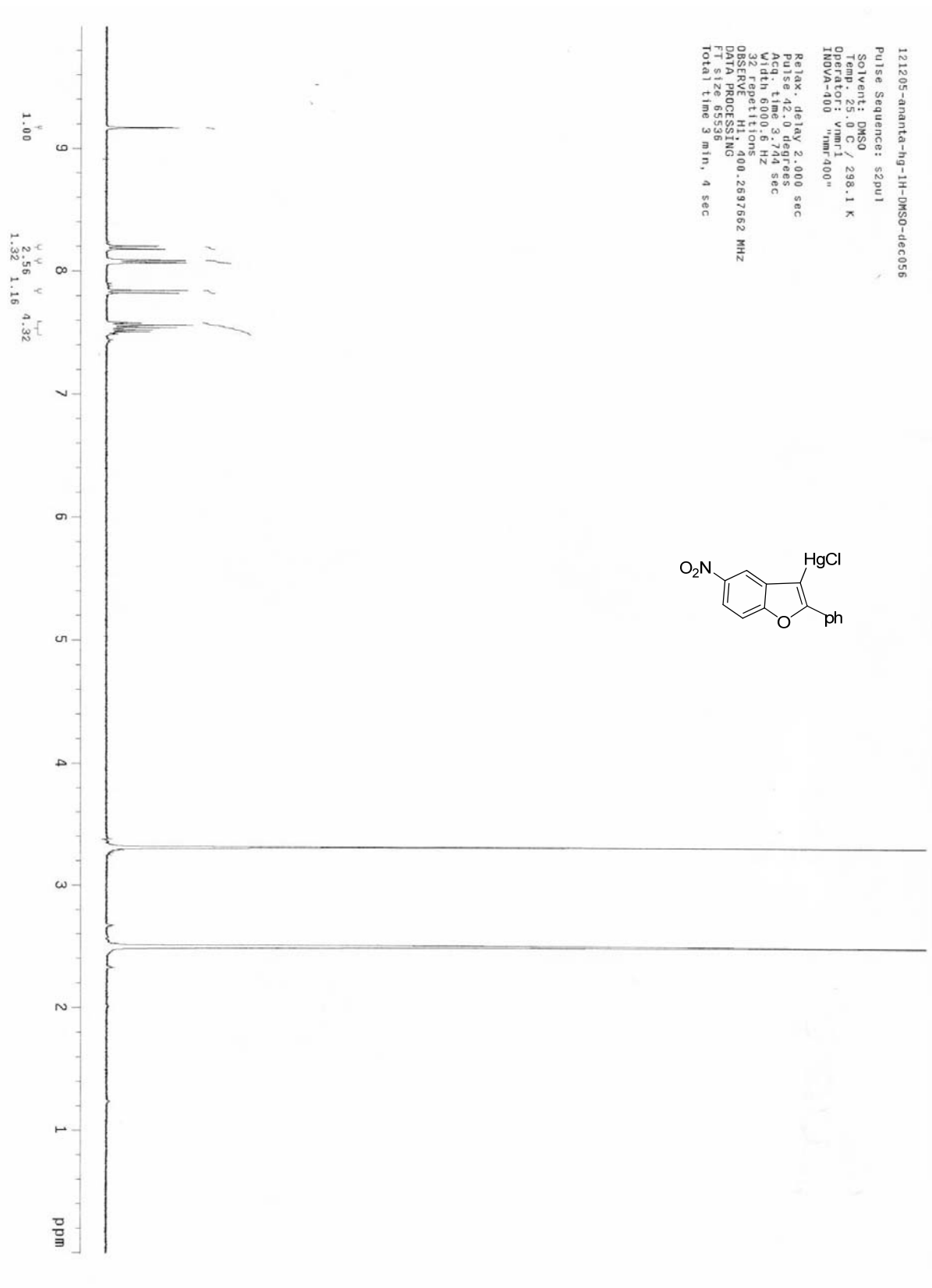
In a round-bottom flask fitted with a reflux condenser, compound **6** (0.1 g, 0.38 mmol) and 2-dicyanomethylene-3-cyano-4,5,5-trimethyl-2,5-dihydrofuran (TCF) (0.68 g, 0.34 mmol) were dissolved in CHCl₃ (4 mL). Piperidine (cat.) was added to the mixture, which was then stirred at reflux for 6 h. After cooling to room temperature, the mixture was concentrated under reduced pressure to afford a residue. The residue was purified over silica gel to afford **7** (0.067 g, 44%). ¹H NMR (400 MHz, DMSO) δ 11.20 (s, 1H), 8.08 (d, *J* = 2.0 Hz, 1H), 7.88 (d, *J* = 16.3 Hz, 1H), 7.83 (dd, *J* = 8.7, 2.0 Hz, 1H), 7.55 (m, 2H), 7.44 (m, 3H), 7.10 (d, *J* = 16.3 Hz, 1H), 7.05 (d, *J* = 8.6 Hz, 1H), 1.79 (s, 6H), ¹³C NMR (100 MHz, DMSO) δ 177.2, 175.6, 162.3, 147.1, 135.3, 131.9, 131.2, 128.8, 125.9, 122.5, 116.5, 112.83, 112.80, 112.0, 111.1, 111.0, 99.2, 97.4, 93.3, 85.4, 53.6, 25.2. HRMS–ESI: *m/z* [M + Na]⁺ calcd for C₂₆H₁₇N₃NaO₂: 426.1218; found: 426.1210.

(E)-(5-(2-(4-cyano-5-(dicyanomethylene)-2,2-dimethyl-2,5-dihydrofuran-3-yl)vinyl)-2-phenylbenzofuran-3-yl)mercury(II) chloride (**7a**)

By following the general procedure, the reaction of **7** (34.7 mg, 0.086 mmol) yielded **7a** (43.9 mg, 80% yield). ¹H NMR (400 MHz, DMSO) δ 8.49 (s, 1H), 8.04 (m, 2H), 7.87 (d, *J* = 8.7 Hz, 1H), 7.72 (d, *J* = 8.4 Hz, 1H),

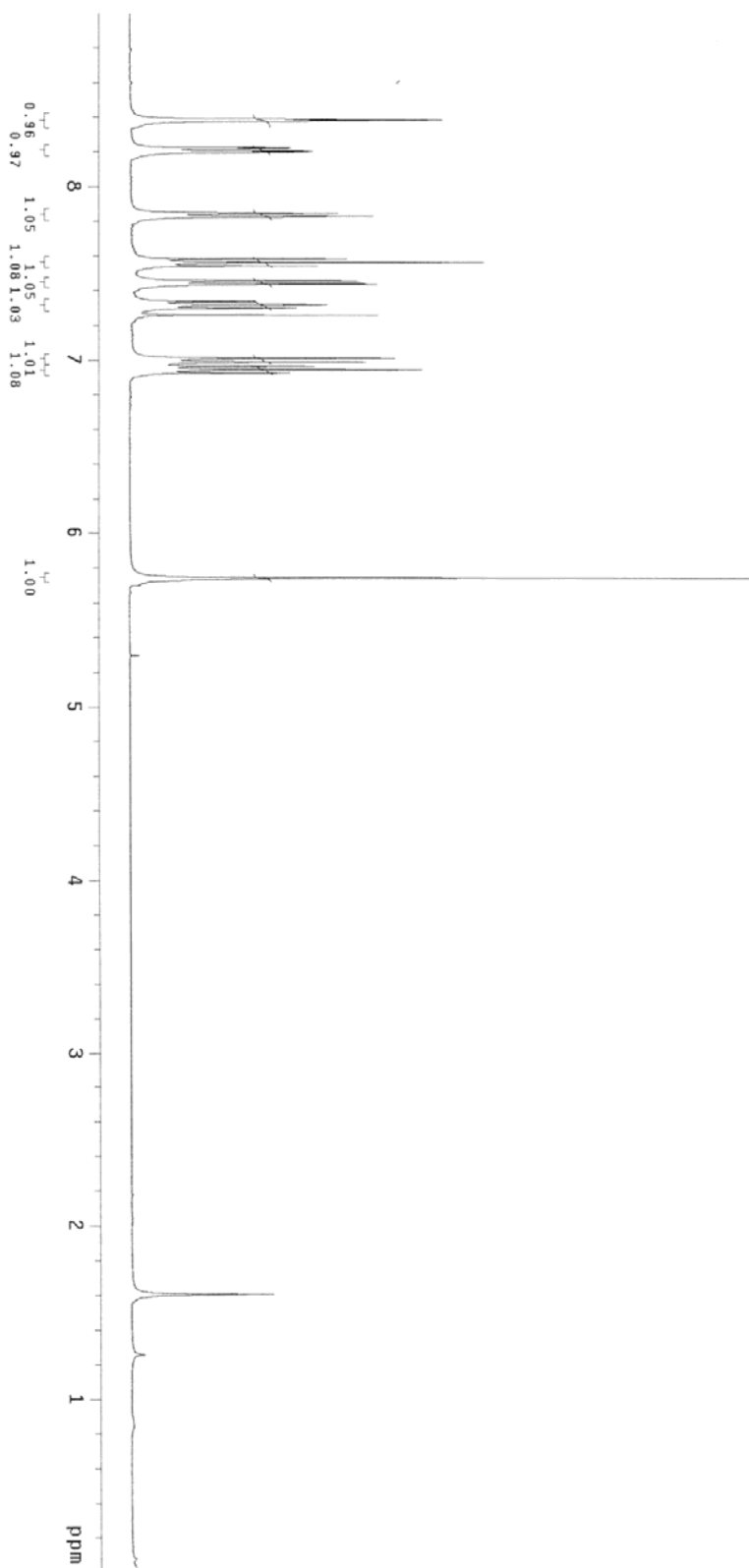
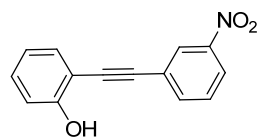
7.51 (m, 4H), 7.18 (d, $J = 16.3$ Hz, 1H), 1.79 (s, 6H), HRMS–ESI: m/z $[M + Cl]^-$ calcd for $C_{26}H_{16}Cl_2HgN_3O_2$: 674.0326; found: 674.0316.

NMR Spectra

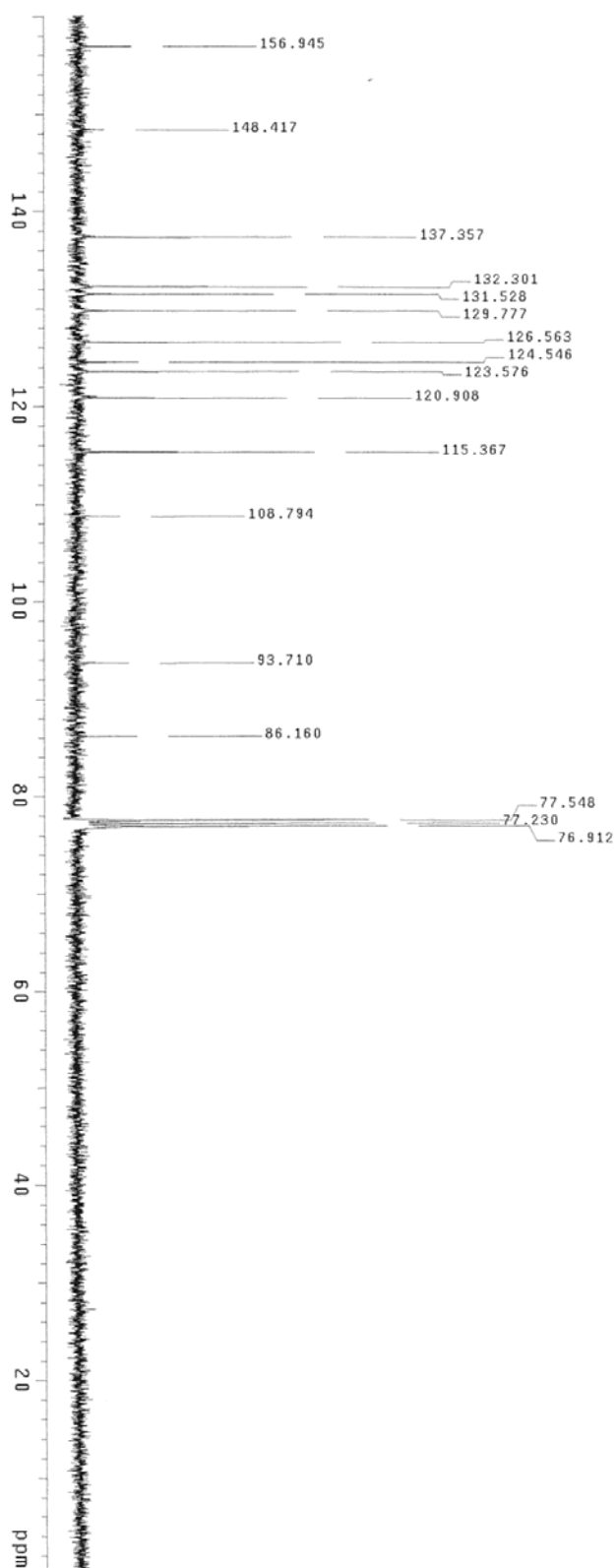


¹H NMR spectrum of **3a** recorded in CDCl₃

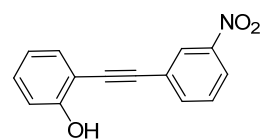
120919-ananta-9p-74-1H-CDCl3-sep161
Pulse Sequence: s2pu1
Solvent: CDCl3
Temp: 25.0 C / 298.1 K
Operator: vnmr1
INOVA-400 "nmr400"
Relax. delay 2.000 sec
Pulse 42.0 degrees
Acq. time 3.744 sec
Width 6000.6 Hz
32 repetitions
OBSERVE H1, 400.2678727 MHz
DATA PROCESSING
FT size 65536
Total time 3 min, 4 sec



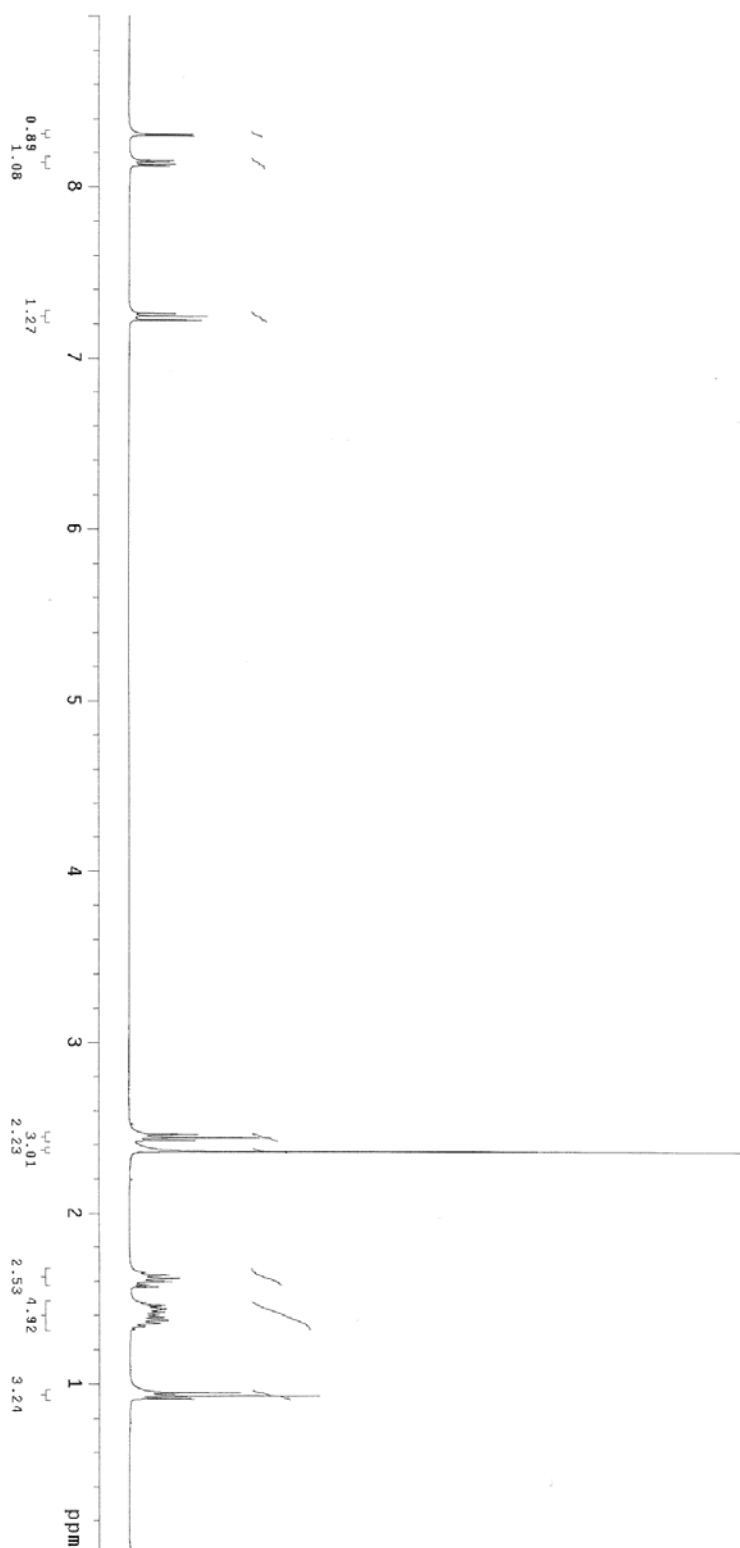
^1H NMR spectrum of **4** recorded in CDCl_3

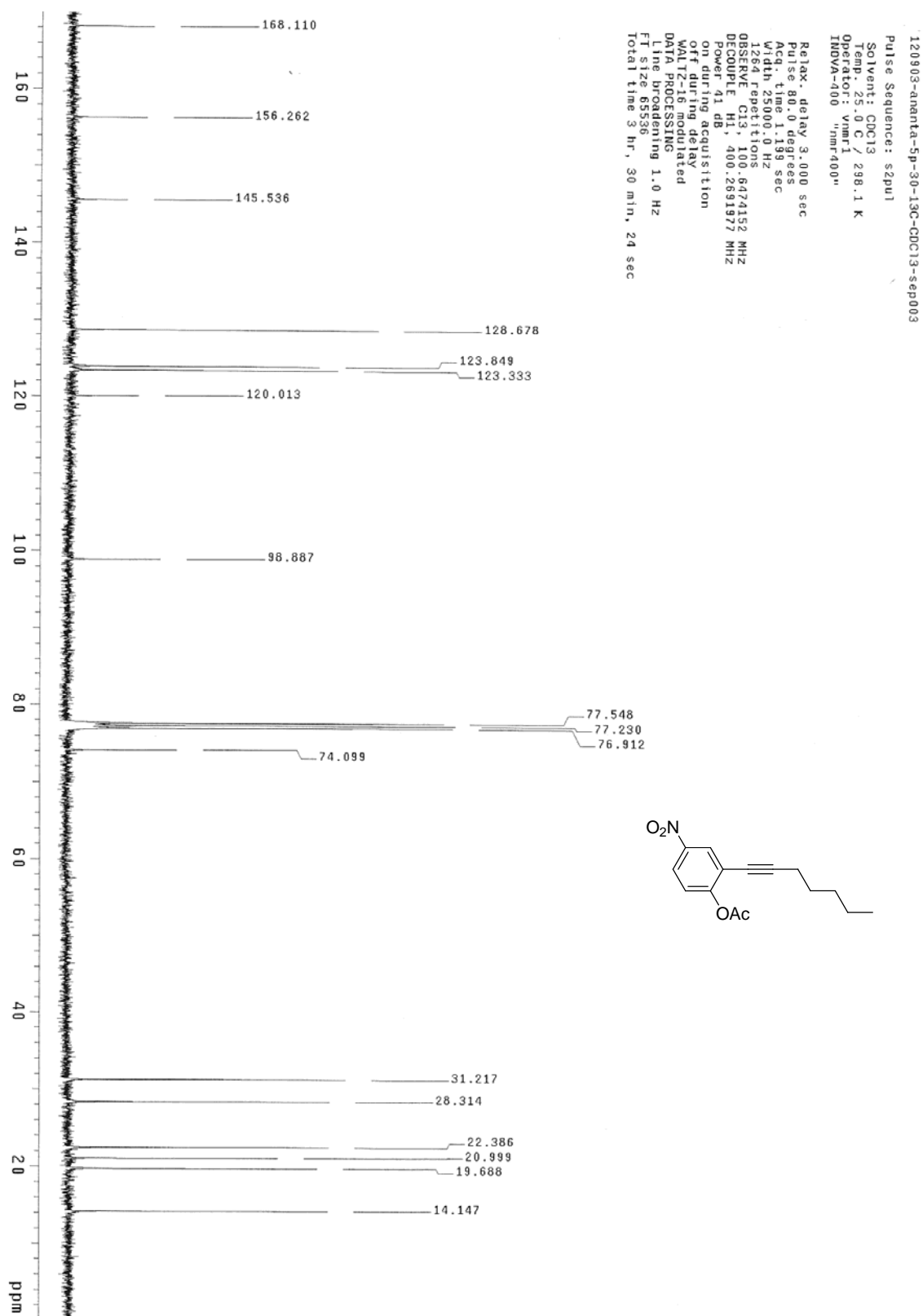


120913-ananta-9p-74-13C-CDCl3-sep162
Pulse Sequence: s2pul
Solvent: CDCl3
Temp: 25.0 C
Observed: 101.327 MHz
INNOVA-400 "nmr400"
Relax. delay 3.000 sec
Pulse 80.0 degrees
Acq. time 0.139 sec
Width 2500.0 Hz
440 repetitions
OBSERVE C13, 100.627414 MHz
DECOUPLE H1, 400.2631977 MHz
Power 41 dB
on during acquisition
off during delay
WALTZ-16 modulated
DATA PROCESSING
Line broadening 1.0 Hz
FT size 65536
Total time 2 hr, 20 min, 16 sec



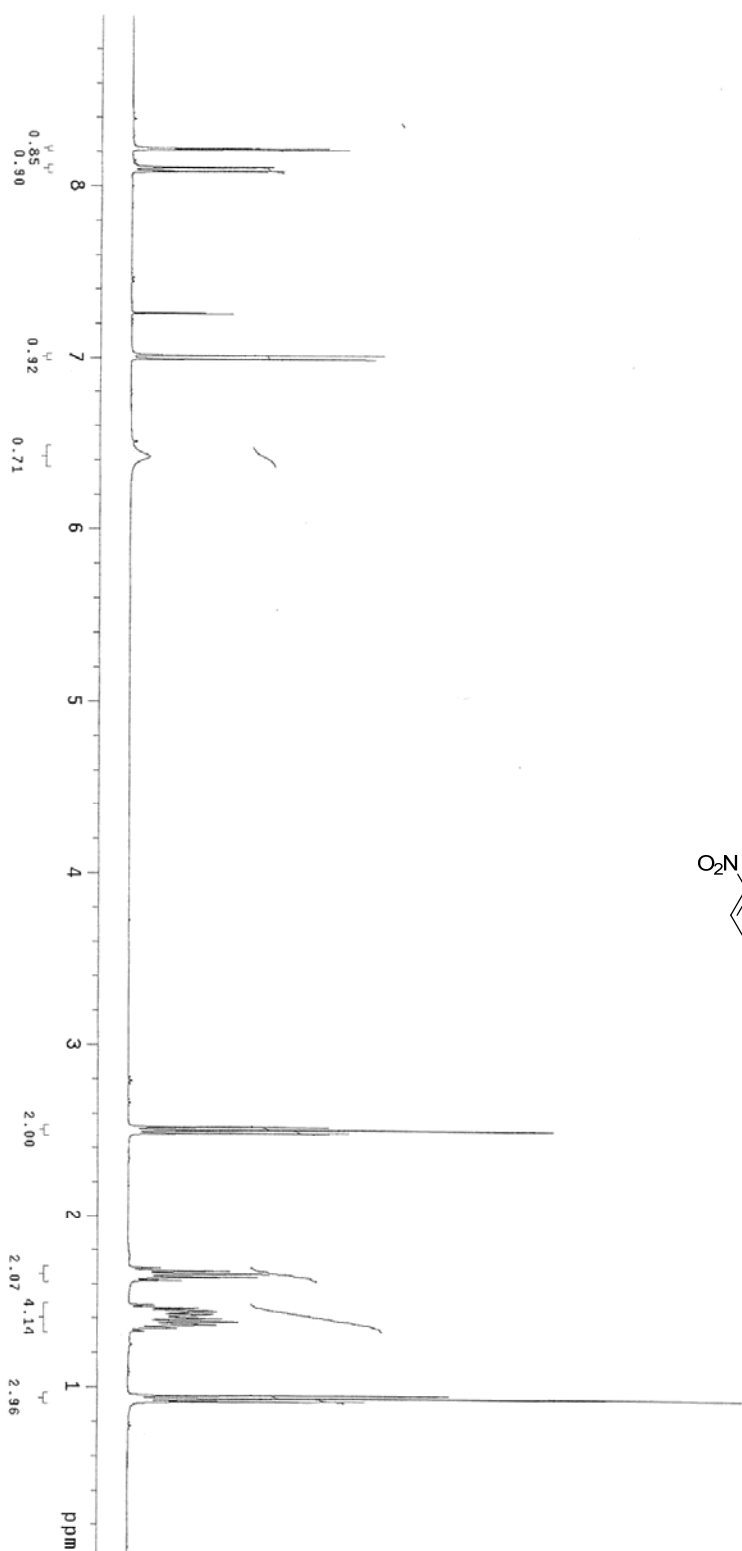
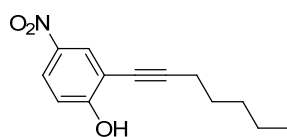
^{13}C NMR spectrum of **4** recorded in DMSO-d_6

CC(=O)Oc1ccccc1C#CCCC¹H NMR spectrum of **8** recorded in CDCl₃



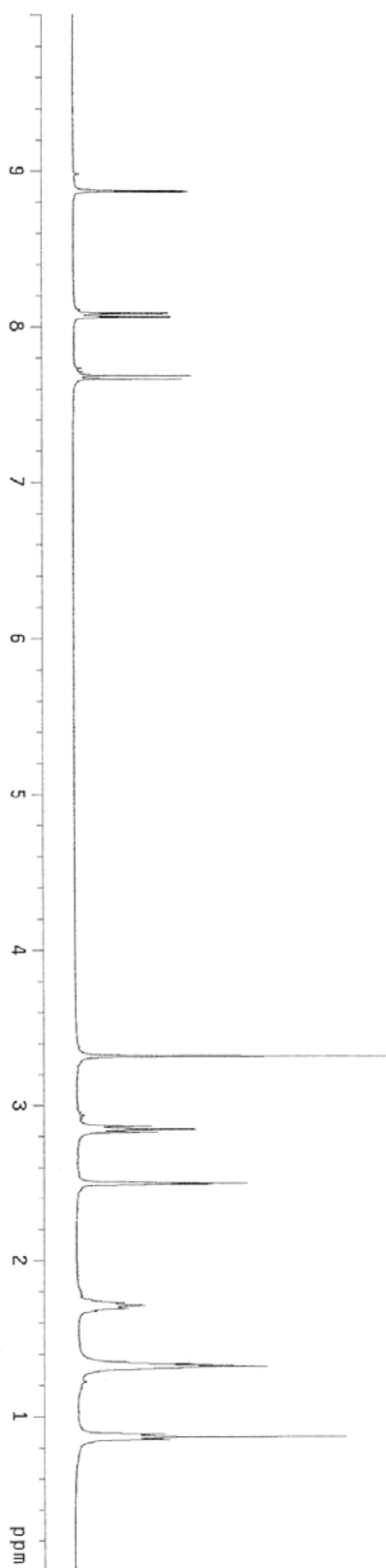
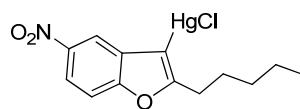
^{13}C NMR spectrum of **8** recorded in CDCl_3

120926-amanita-NO2-OH-1H-CDCl3-sep224
Pulse Sequence: szpu1
Solvent: CDCl3
Temp: 25.0 C / 298.1 K
Operator: vnmr1
INOVA-100 "nmr400"
Relax. delay 2.000 sec
Pulse 42.0 degrees
Acq. time 3.744 sec
Width 100.000 Hz
32 repetitions
OBSERVE H1 400.2678729 MHz
DATA PROCESSING
FT size 65536
Total time 3 min, 4 sec

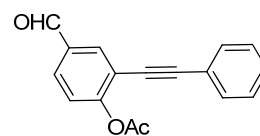
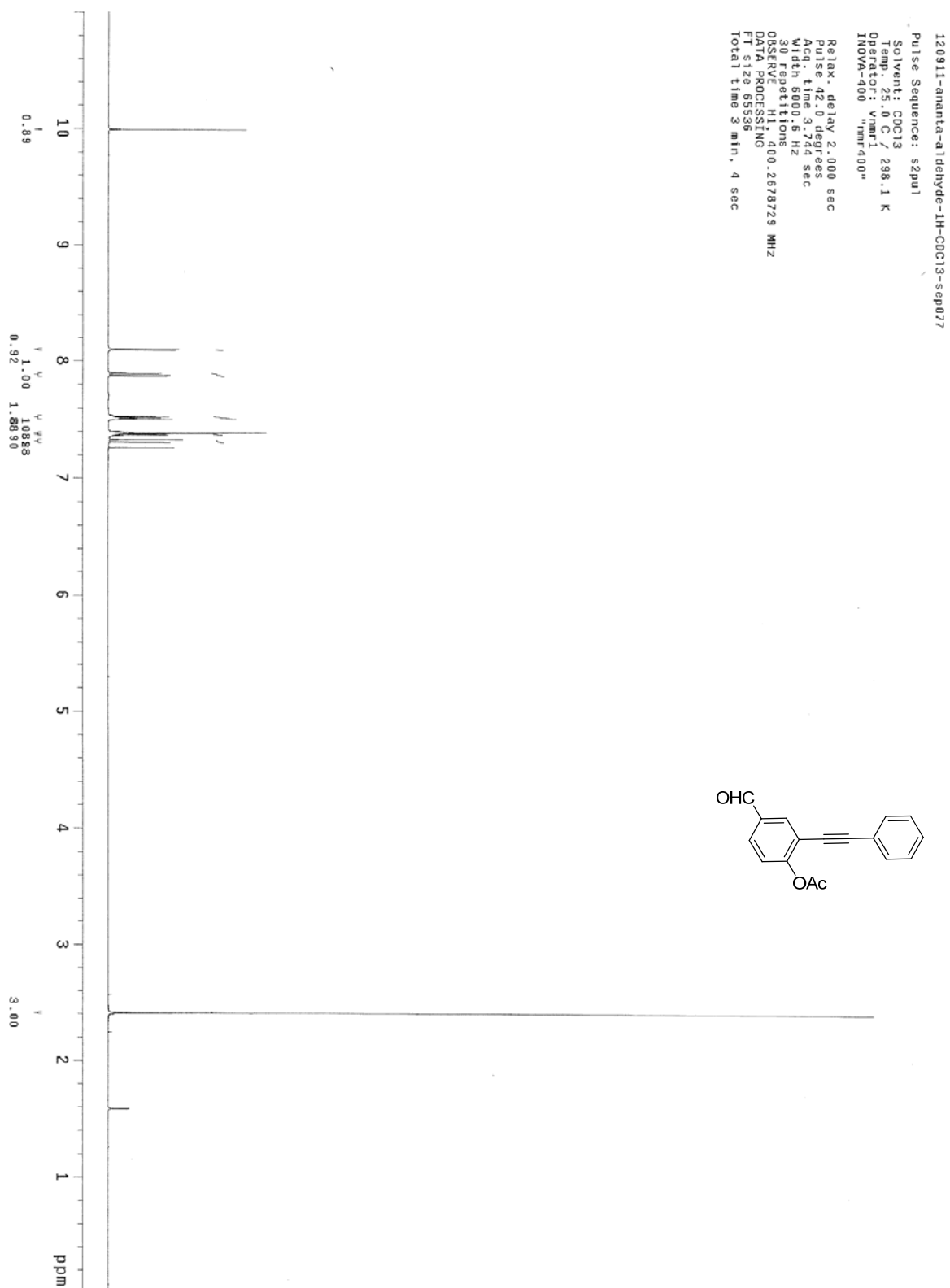


^1H NMR spectrum of **5** recorded in CDCl_3

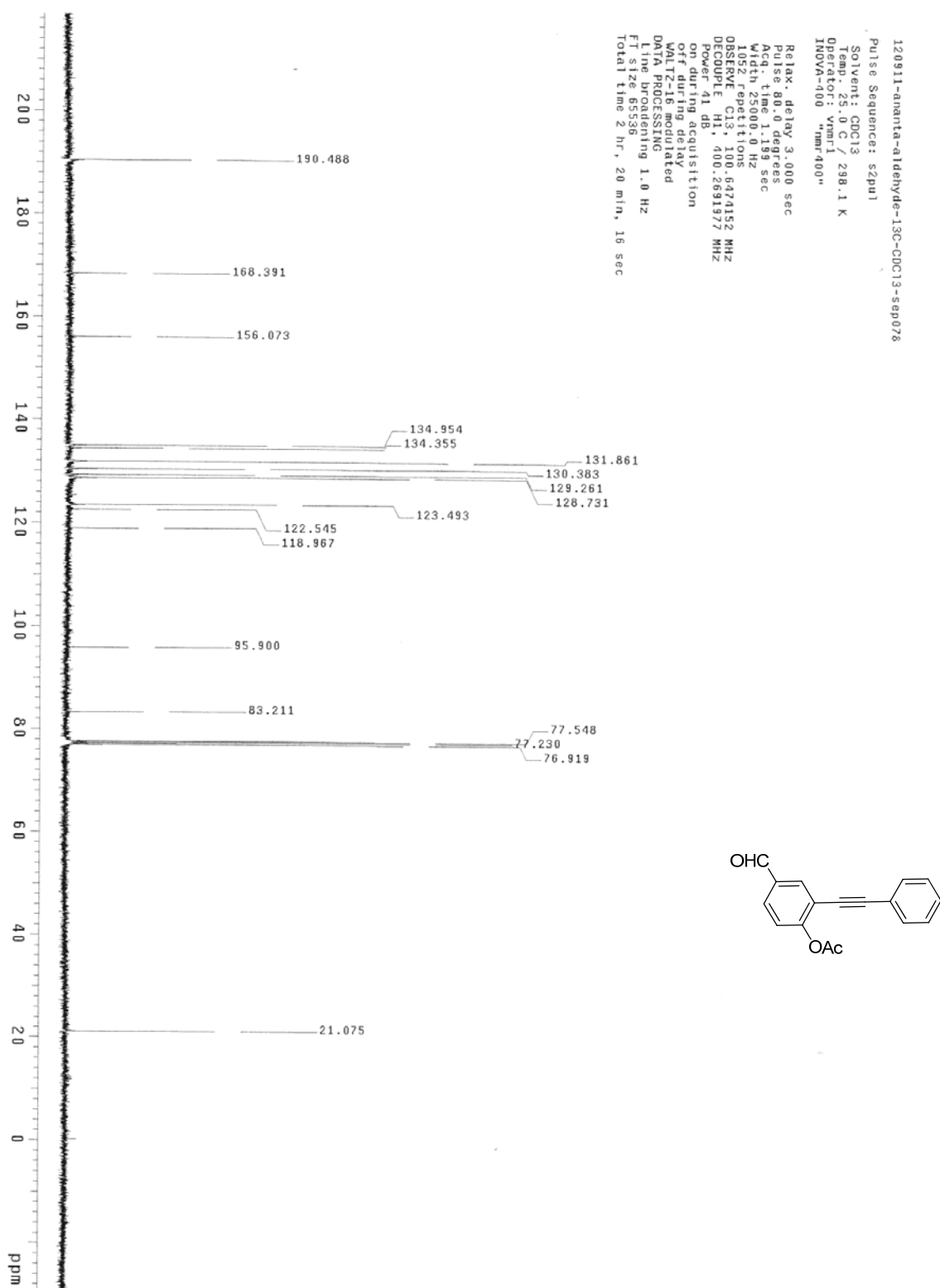
121113-ananta-9-p-140-1H-DMSO-nov158
Pulse Sequence: s2pul
Solvent: DMSO
Temp: 25.0 C / 298.1 K
Operator: vmmr1
INOVA-400 "nmr400"
Relax. delay 2.000 sec
Pulse 42.0 degrees
Acq. time 3.744 sec
Width 6000.6 Hz
10 repetitions
OBSERVE H1, 400.2697669 MHz
DATA PROCESSING
FT size 65536
Total time 3 min, 4 sec



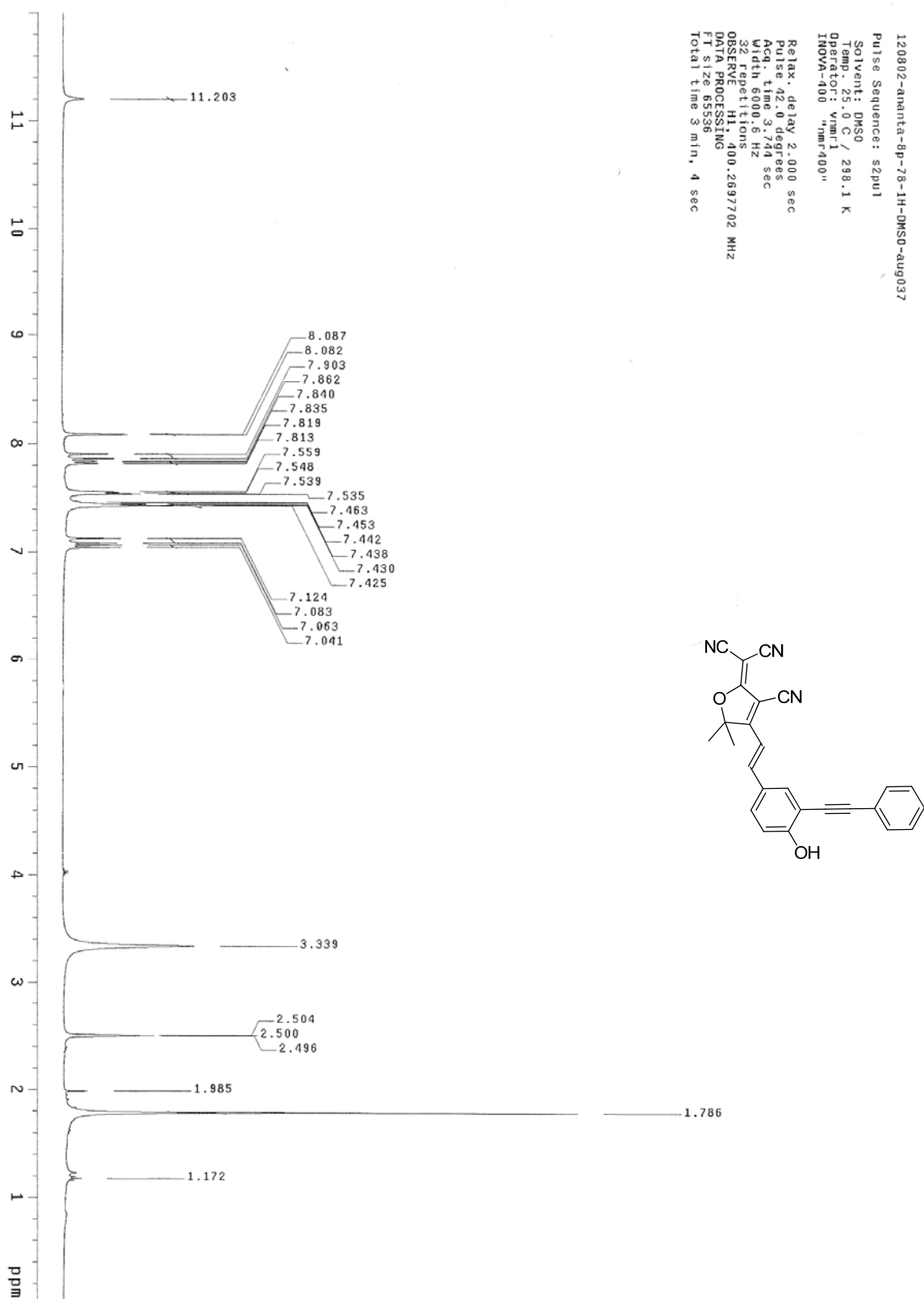
^1H NMR spectrum of **5a** recorded in DMSO- d_6



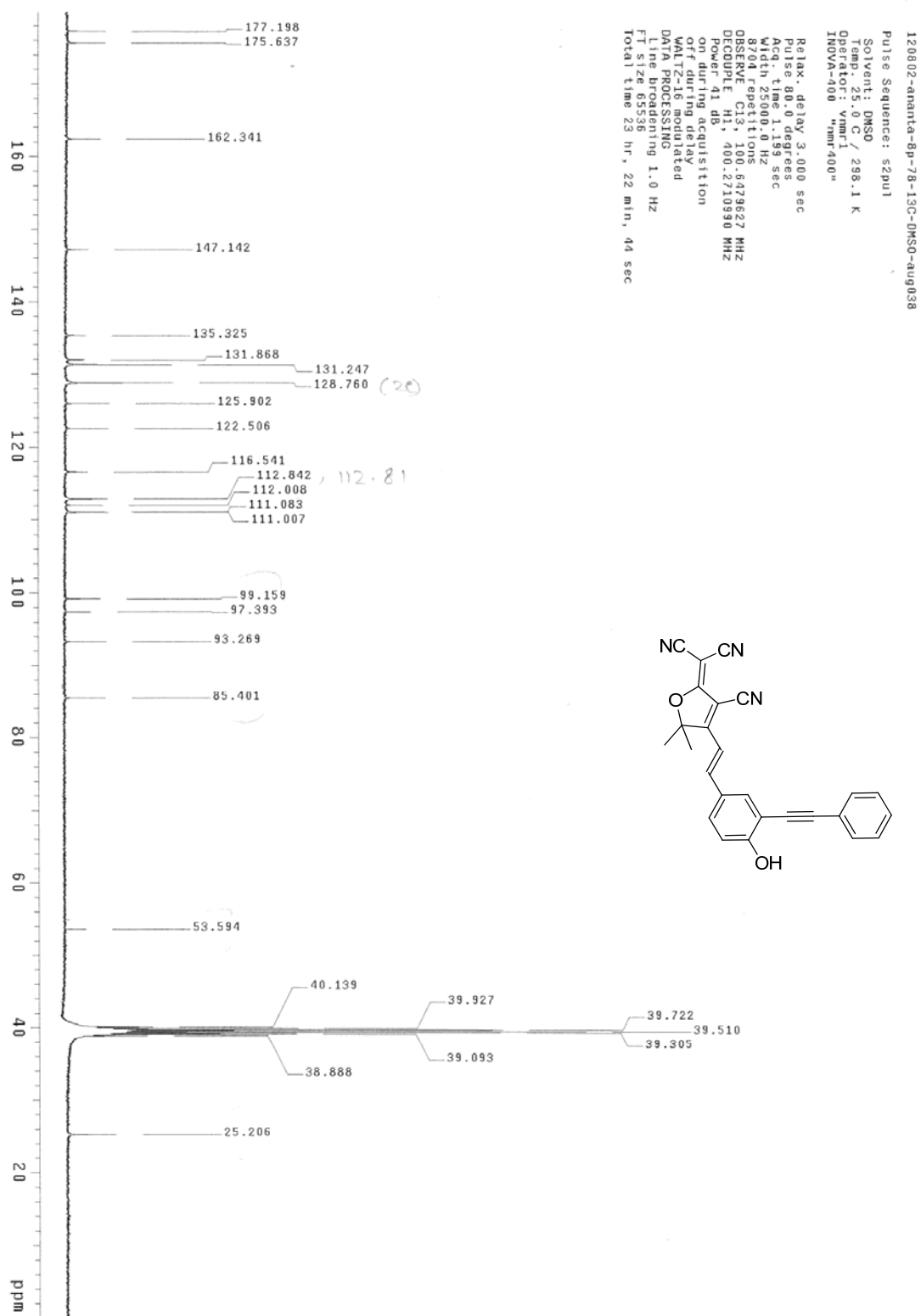
^1H NMR spectrum of **6** recorded in CDCl_3



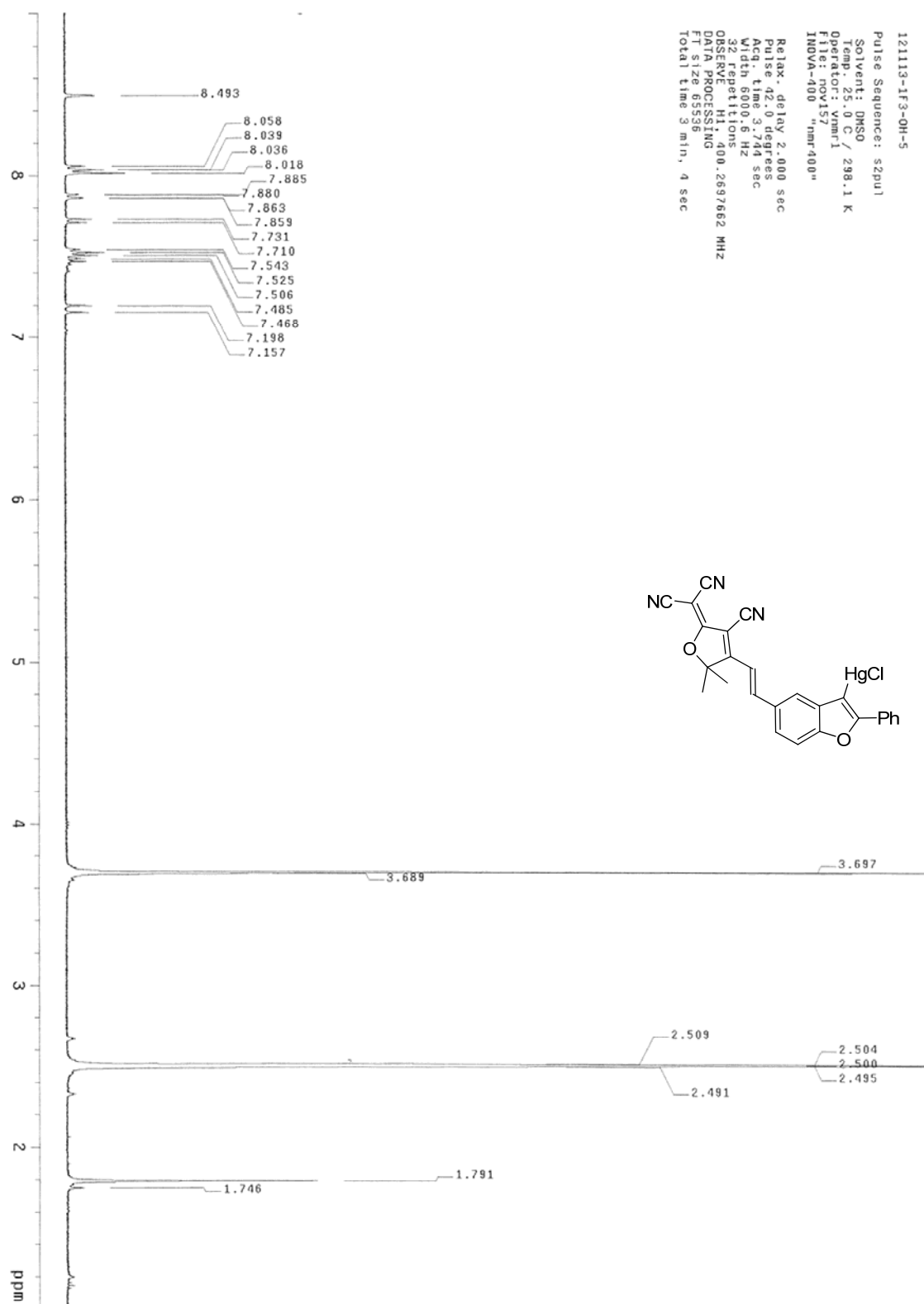
^{13}C NMR spectrum of **6** recorded in CDCl_3



^1H NMR spectrum of **7** recorded in CDCl_3



^{13}C NMR spectrum of **7** recorded in CDCl_3



^1H NMR spectrum of **7a** recorded in DMSO