

Supporting Information

A PARP inhibitor with selectivity toward ADP-ribosyltransferase ARTD3/PARP3

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Supplementary Tables

Supplementary Table 1. STO1131 stabilizes ARTD3/PARP3 and other ADP-ribose transferases in thermal shift assays.^a

Name	Alt. Name	T _m shift (°C)
ARTD1	PARP1	2.0±0.1
ARTD2	PARP2	1.4±0.3
ARTD3	PARP3	8.5±0.2
ARTD4	PARP4	0.3±0.4
ARTD5	TNKS1	0.0±0.04
ARTD6	TNKS2	0.2±0.1
ARTD7	PARP15	0.2±0.5
ARTD8	PARP14	-0.3±0.5
ARTD9	PARP9	-0.1±0.6
ARTD10	PARP10	0.5±0.5
ARTD11	PARP11	n.d.
ARTD12	PARP12	0.1±0.1
ARTD13	PARP13	0.2±0.1
ARTD14	PARP7	n.d.
ARTD15	PARP16	0.5±0.2
ARTD16	PARP8	n.d.
ARTD17	PARP6	n.d.

^a Determined by differential scanning fluorimetry. Values represent means ± S.D. of two to four independent experiments. n.d., not determined.

Supplementary Table 2. Crystallographic data collection and refinement statistics.^a

Data collection	ARTD3•ME0355	ARTD3•ME0354	ARTD3•ME0328	ARTD1•ME0328
Synchrotron	ESRF	Diamond	Diamond	Bessy
Beam line	ID14-1	I04	I04	BL14.1
Wavelength (Å)	0.93340	0.99190	0.99190	0.91841
Space group	P2 ₁	P2 ₁	P2 ₁	P2 ₁
Unit cell dimensions				
a, b, c (Å)	54.8, 56.8, 56.9	55.2, 56.8, 56.6	54.7, 56.7, 57.0	87.1, 49.4, 183.5
α, β, γ (°)	90, 112.77, 90	90, 112.66, 90	90, 112.74, 90	90, 101.82, 90
Resolution (Å)	52.5 – 1.86 (1.96 – 1.86)	30.0 – 1.80 (1.85 – 1.80)	30.0 – 1.80 (1.85 – 1.80)	30.0 - 2.89 (2.90 - 2.89)
Unique reflections	25790 (2775)	29876 (2165)	29828 (2148)	34835 (376)
R-merge (%) ^b	4.8 (19.8)	9.1 (48.0)	8.0 (44.1)	10.0 (56.4)
Completeness (%)	99.0 (94.5)	99.2 (98.2)	99.4 (97.6)	99.9 (99.7)
Redundancy	7.4 (6.4)	7.5 (7.2)	7.4 (7.1)	6.1 (6.1)
<I/σI>	30.4 (9.1)	14.5 (3.4)	21.5 (3.9)	15.5 (3.1)
Refinement				
Resolution (Å)	52.5 - 1.90 (2.00 - 1.90)	28.4 - 1.80 (1.85 - 1.80)	28.4 - 1.80 (1.85 - 1.80)	27.2 - 2.89 (2.98 - 2.89)
R-all (%) ^c	17.2 (27.1)	16.3 (22.0)	16.4 (21.5)	21.5 (25.6)
R-free (%) ^c	21.9 (37.7)	19.4 (24.8)	20.3 (25.8)	28.5 (37.4)
r.m.s.d. bond length (Å)	0.019	0.018	0.018	0.010
r.m.s.d. bond angle (°)	1.7	1.6	1.5	1.2
B-factors (Å ²)				
Protein	18.3	15.9	15.1	61.9
Ligand	15.0	18.2	14.2	35.2
Water	23.0	23.0	21.8	-
Ramachandran plot ^d				
Most favoured (%)	98.9	99.1	98.6	94.0
Allowed (%)	100	100	99.7	99.6

^a Values in parentheses are for the outermost resolution shell.

^b $R_{\text{merge}} = \sum |I - \langle I \rangle| / \sum I$, where I is the intensity measurement for a given reflection and $\langle I \rangle$ is the average intensity for multiple measurements of this reflection.

^c $R = \sum ||F_{\text{obs}}| - |F_{\text{calc}}|| / \sum |F_{\text{obs}}|$, where R_{free} is calculated for a randomly chosen 5-10% of reflections, which were not used for structure refinement, and R_{all} is calculated for all reflections.

^d The Ramachandran plot was calculated using the Molprobit server.¹

Supplementary Table 3. ME0328 and ME0355 specifically inhibit the enzymatic activity of ARTD3/PARP3 *in vitro*.^a

		ME0328			ME0355	
Name	Alt. Name	K _m (μM)	T _m shift (°C)	IC ₅₀ (μM)	T _m shift (°C)	IC ₅₀ (μM)
ARTD1	PARP1	10±1.9	1.8±0.1	6.3±0.62	2.0±0.2	9.1±2.6
ARTD2	PARP2	159±5	1.2±0.1	10.8±4.0	0.7±0.6	
ARTD3	PARP3	230±47	8.5±0.6	0.89±0.28	9.7±0.4	1.3±0.2
ARTD4	PARP4	n.d.		n.d. ^b		n.d. ^b
ARTD5	TNKS1	41±11	0.1±0.1	47.3±29.0	0.1±0.1	n.d.
ARTD6	TNKS2	174±43		34.3±17.0		n.d.
ARTD7	PARP15	6.3±0.6		>100		n.d.
ARTD8	PARP14	164±17	-0.2±0.1	>100	0.3±0.3	n.d.
ARTD9	PARP9	n.d. ^c		n.d. ^c		n.d. ^c
ARTD10	PARP10	95±34		71.3±22.0		>50
ARTD11	PARP11	n.d. ^d		n.d. ^d		n.d. ^d
ARTD12	PARP12	n.d.		>100		>50
ARTD13	PARP13	n.d. ^c		n.d. ^c		n.d. ^c
ARTD14	PARP7	n.d. ^d		n.d. ^d		n.d. ^d
ARTD15	PARP16	n.d.		n.d. ^b		n.d. ^b
ARTD16	PARP8	n.d. ^d		n.d. ^d		n.d. ^d
ARTD17	PARP6	n.d. ^d		n.d. ^d		n.d. ^d

^a Values represent means ± S.D. of three independent experiments (T_m shift data) or means ± S.E. of the fitted parameter (IC₅₀ data; based on triplicates).

^b Not determined – no transferase activity in presence of DMSO.

^c Not determined – protein reportedly lacks ADP-ribose transferase activity.²

^d Not determined – transferase domain could not be produced.

Supplementary Table 4. Physicochemical and metabolic stability profiling of ME0328.

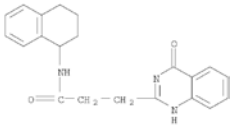
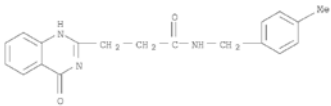
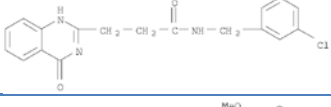
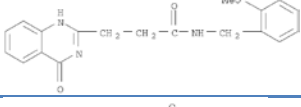
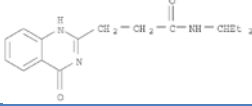
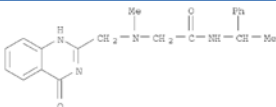
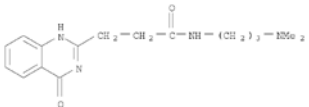
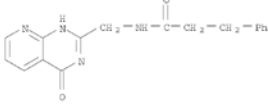
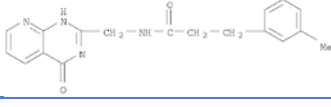
Property/mol. descriptor	ME0328	Range ^a	Comment
Formula molecular weight	321.27	Not available	
QPlogPo/w ^b	2.5	-2.0 – 6.5	Predicted octanol/water partition coefficient
QPPCaco ^b	559	<25 poor, >500 great	Predicted apparent Caco-2 cell permeability in nm/s
QPPMDCK ^b	371	<25 poor, >500 great	Predicted apparent MDCK cell permeability in nm/s
QPlogBB ^b	-1.0	-3.0 – 1.2	Predicted brain/blood partition coefficient
QPlogKhsa ^b	-0.1	-1.5 – 1.5	Prediction of binding to human serum albumin
Mean solubility (μM) ^c	25.4		
HLM CL _{int} (μl/min/mg) ^c	16.28		Internal clearance in human liver microsomes
HW rat hep CL _{int} (μl/min/1E6 cells) ^c	36.98		
HuCaco2 pH 6.5 A-to-B P _{app} (1E6 x cm/s) ^c	43.54		Apparent permeability coefficient in Caco-2 cells

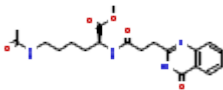
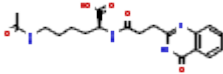
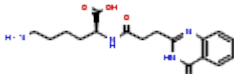
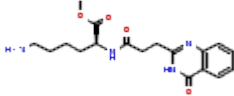
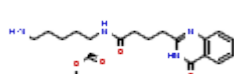
^a For 95% of known drugs.

^b Calculated using Qikprop 3.3 (Schrödinger Inc.).

^c Determined by AstraZeneca R&D (Mölndal, Sweden).³

Supplementary Table 5. Database search results for similarity of ME0328/ME0355 among known bioactive compounds.

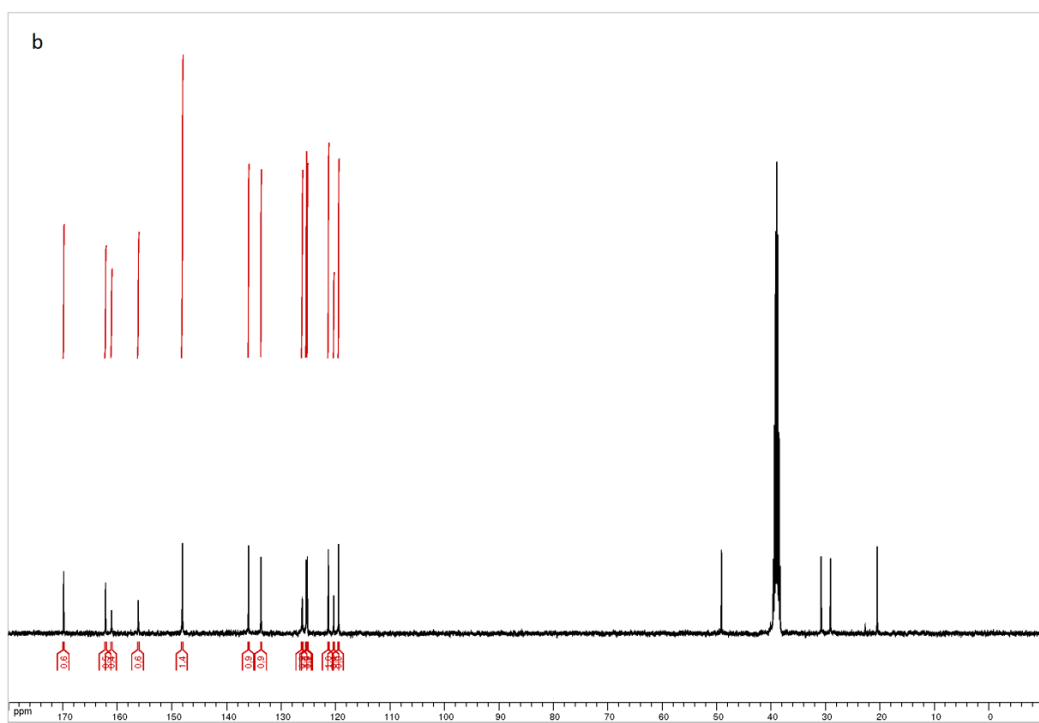
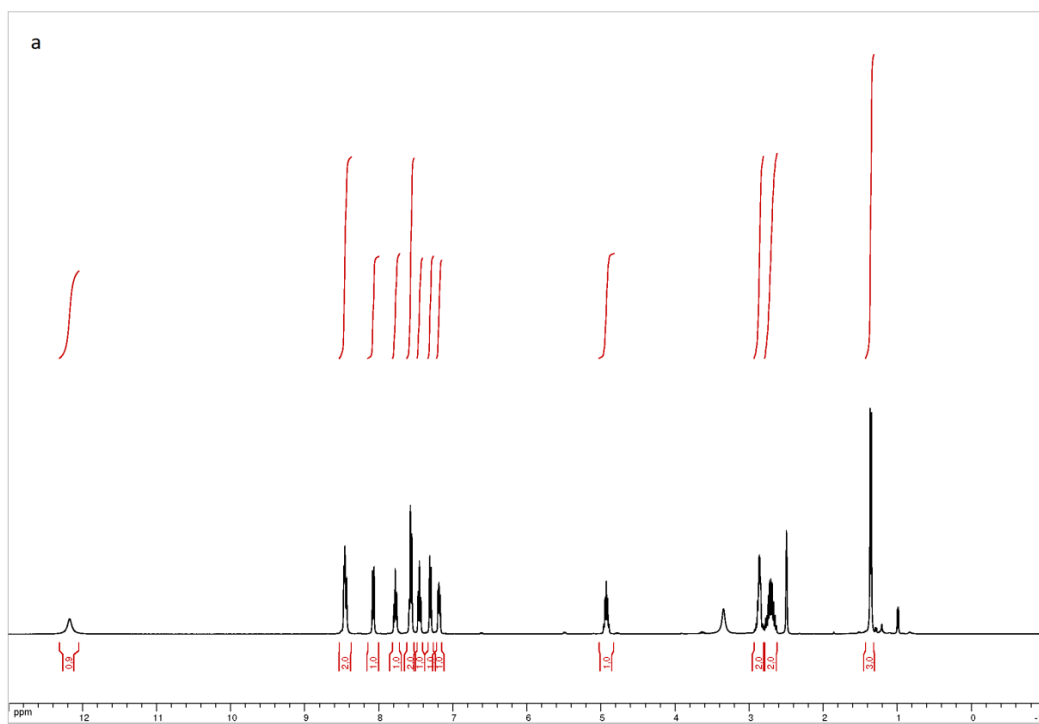
Substructure search using ME0328/ME0355 ^a		
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870768-51-7		Variety of antiprion compounds discovered through an in silico screen based on cellular-form prion protein structure: correlation between antiprion activity and binding affinity. <i>Antimicrobial Agents and Chemotherapy</i> (2009), 53(2), 765-771.
Similarity search of compounds with ≥ 80 % similarity to ME0328/ME0355 ^a		
CAS	Structure	Reference
743451-47-0		Method using lifespan-altering compounds for altering the lifespan of eukaryotic organisms, and screening for such compounds. U.S. Pat. Appl. Publ. (2009), US 20090163545 A1 20090625.
851105-00-5		Same as 743451-47-0
785702-46-7		Same as 743451-47-0
731817-17-7		Same as 743451-47-0
794562-86-0		Same as 743451-47-0
749901-14-2		1. Identification of Non-Peptide Malignant Brain Tumor (MBT) Repeat Antagonists by Virtual Screening of Commercially Available Compounds. <i>Journal of Medicinal Chemistry</i> (2010), 53(21), 7625-7631 2. Same as 743451-47-0
958356-12-2		1. Pyrido pyrimidinones as selective agonists of the high affinity niacin receptor GPR109A: Optimization of in vitro activity. <i>Bioorganic & Medicinal Chemistry Letters</i> (2010), 20(18), 5426-5430. 2. Preparation of pyrido[2,3-d]pyrimidin-4-one derivatives as HM74A agonists. PCT Int. Appl. (2007), WO 2007134986 A1 20071129
958361-10-9		Preparation of pyrido[2,3-d]pyrimidin-4-one derivatives as HM74A agonists. PCT Int. Appl. (2007), WO 2007134986 A1 20071129

CHEMBL1783023	<p>Antibacterial activity against <i>Xanthomonas oryzae</i>. Antibacterial activity against <i>Xanthomonas campestris</i> pvs. Antibacterial activity against <i>Escherichia coli</i>. Antibacterial activity against <i>Bacillus subtilis</i>. Antibacterial activity against <i>Pseudomonas fluorescens</i>.</p>
	Same as CHEMBL1783023
CHEMBL1783025	Same as CHEMBL1783023
	Same as CHEMBL1783023
CHEMBL1789526	Same as CHEMBL1783023
	Same as CHEMBL1783023
CHEMBL1789619	Same as CHEMBL1783023
	Same as CHEMBL1783023
CHEMBL1789671	Same as CHEMBL1783023
	

^a Using SciFinder (<https://www.cas.org/products/scifinder>).

^b Using ChEMBL (<https://www.ebi.ac.uk/chembl>).

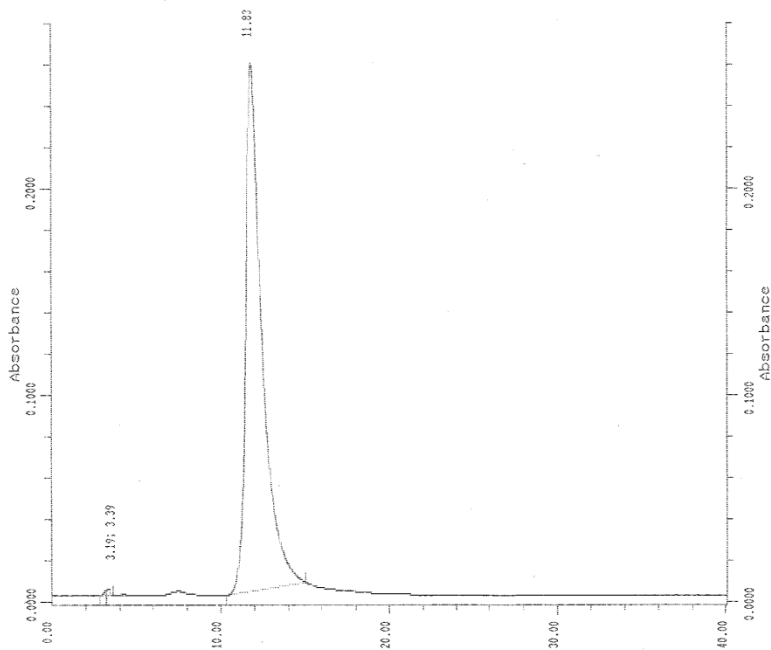
Supplementary Figures



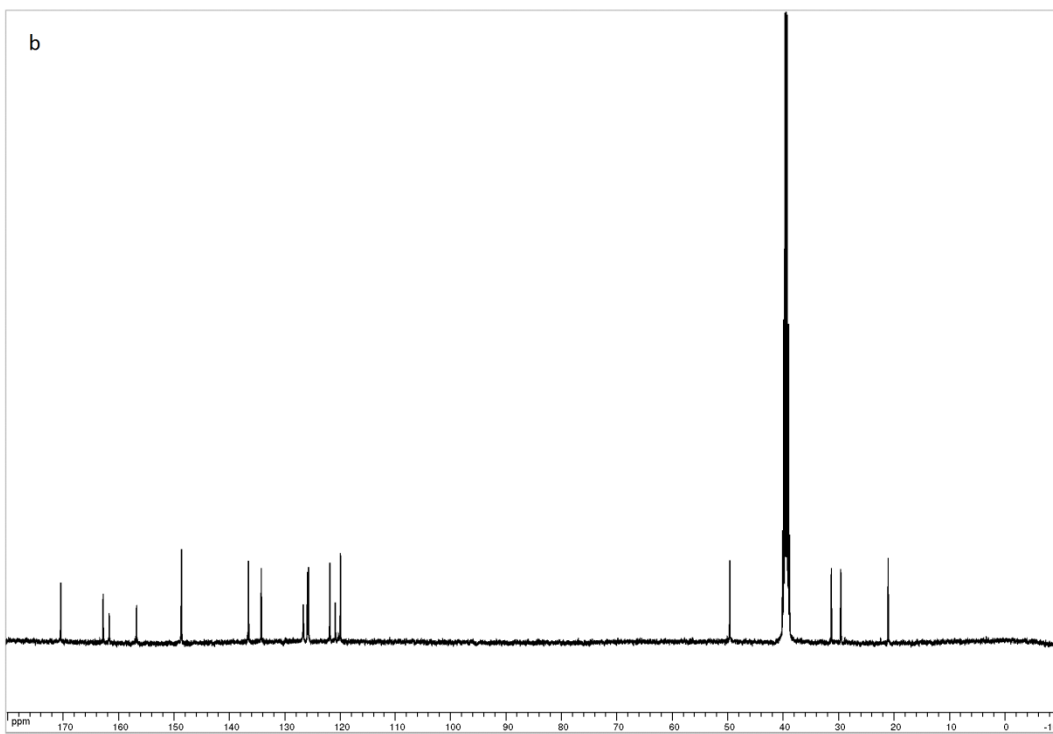
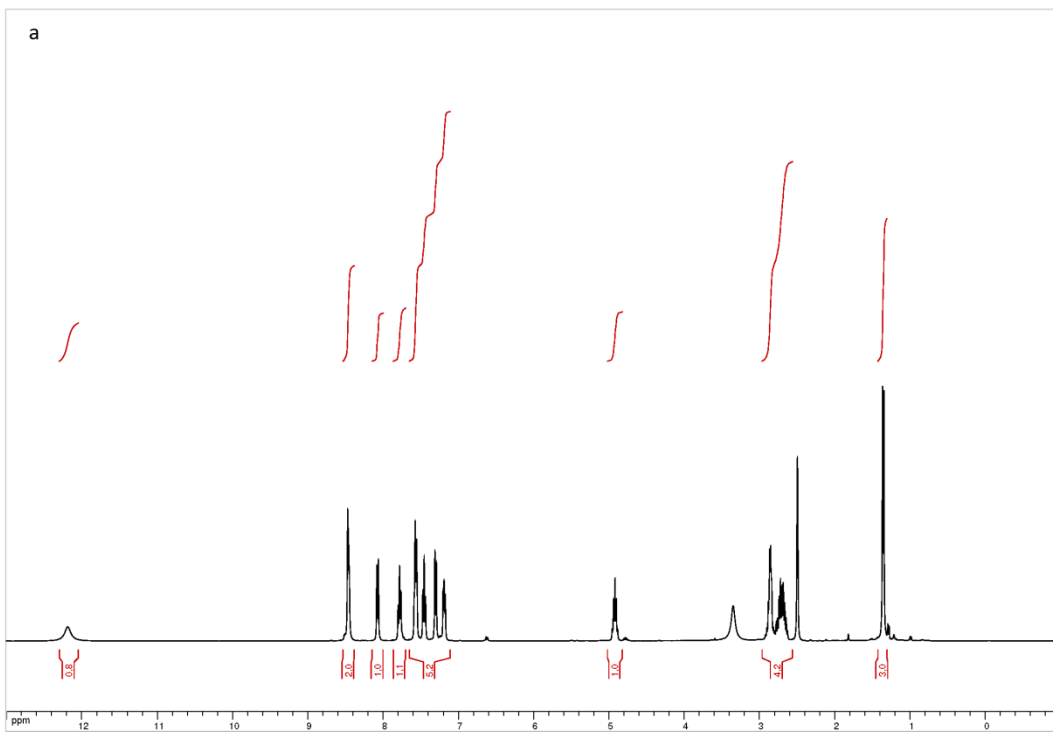
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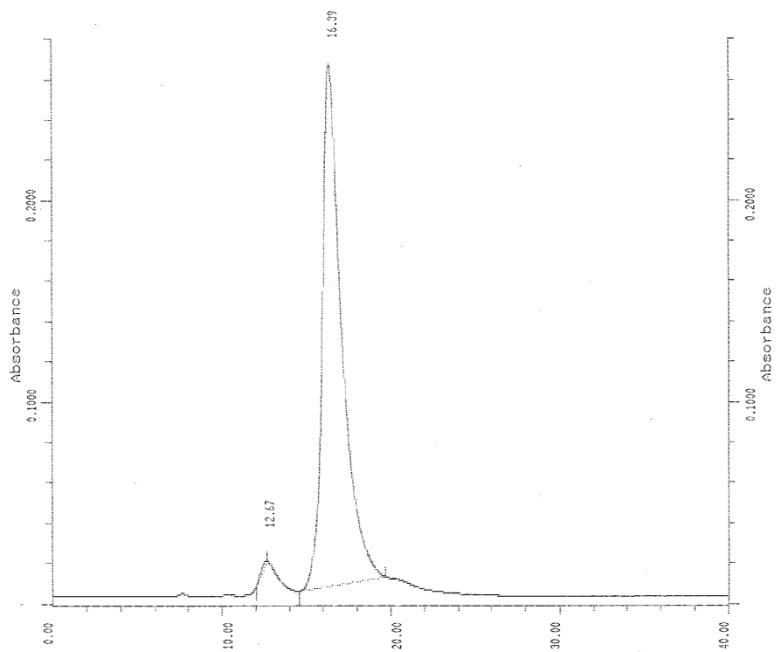
Supplementary Figure 1. Characterization of ME0354. (a) ^1H -NMR spectrum. (b) ^{13}C -NMR spectrum. (c) Chiral HPLC UV trace.



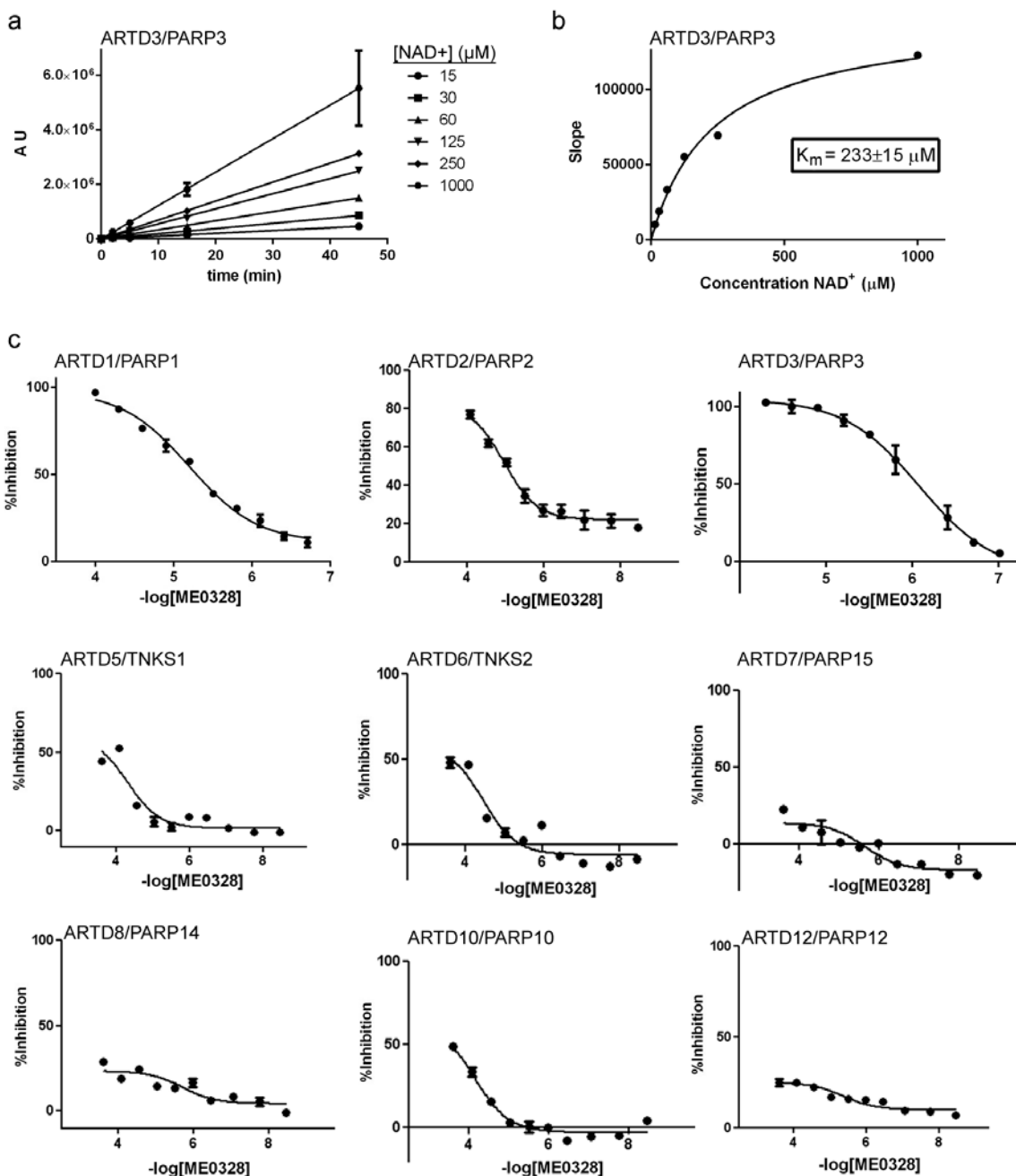
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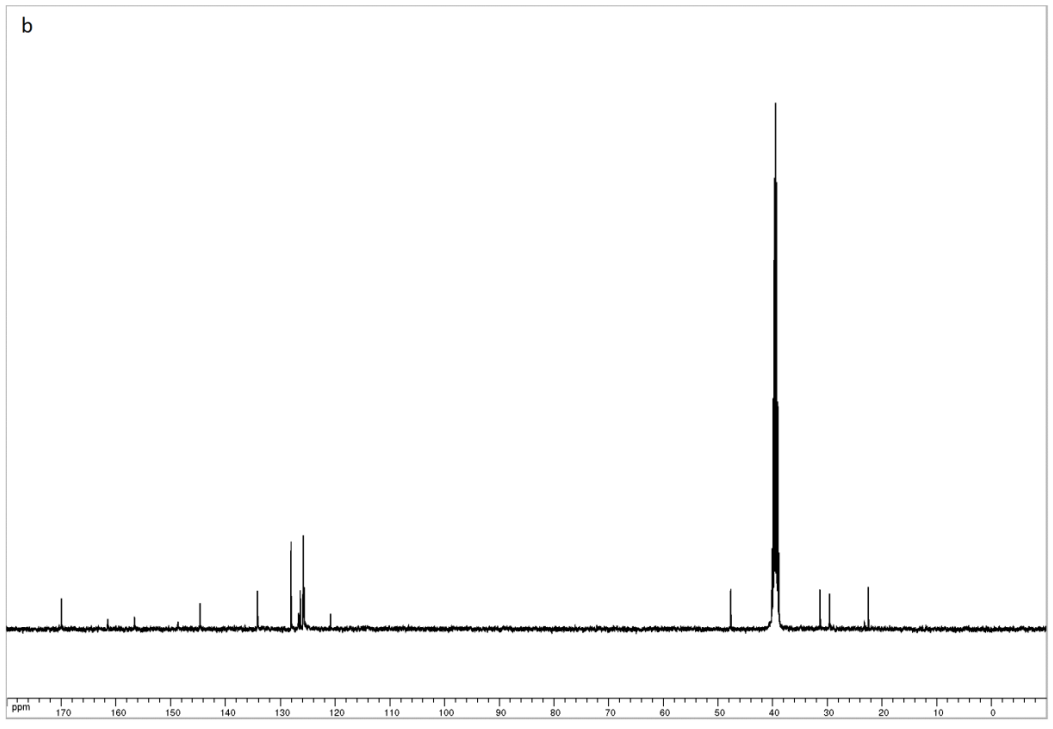
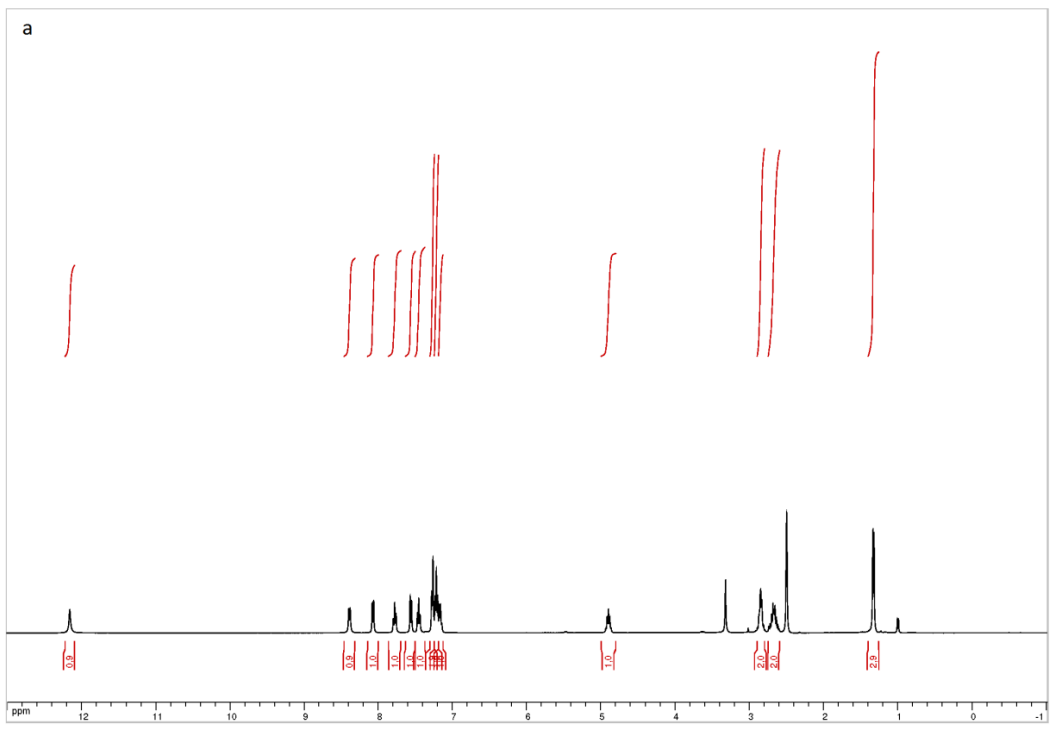
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Supplementary Figure 2. Characterization of ME0355. (a) ^1H -NMR spectrum. (b) ^{13}C -NMR spectrum. (c) Chiral HPLC UV trace.



Supplementary Figure 3. Examples of enzymatic assay data. (a) Representative raw data of a K_m determination for ARTD3. All measurements were performed within the linear range of the assay. (b) K_m determination for ARTD3. Values listed in Supplementary Table 3 were determined accordingly. (c) IC_{50} determinations for ME0328 and 9 ARTD family members. Representative curves are shown. Values listed in Supplementary Table 3 represent means \pm S.E. of the fitted parameter based on triplicates of such measurements.

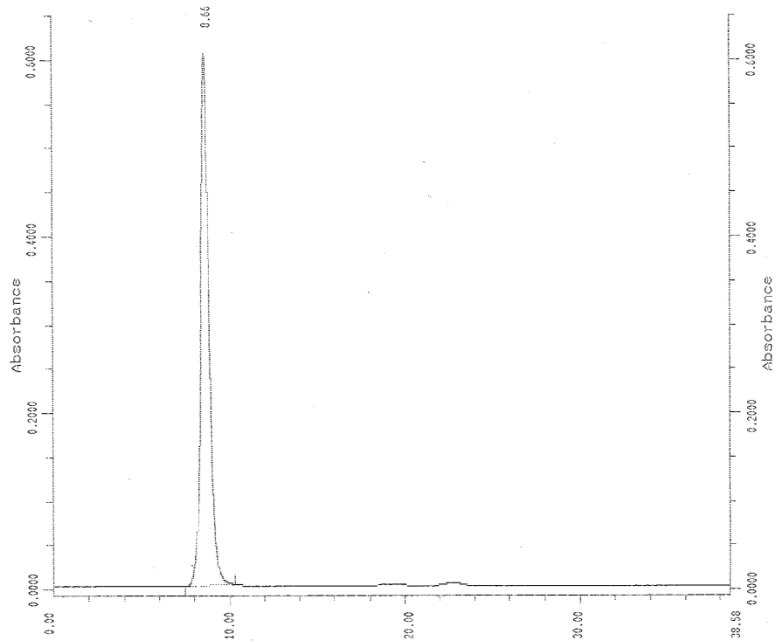


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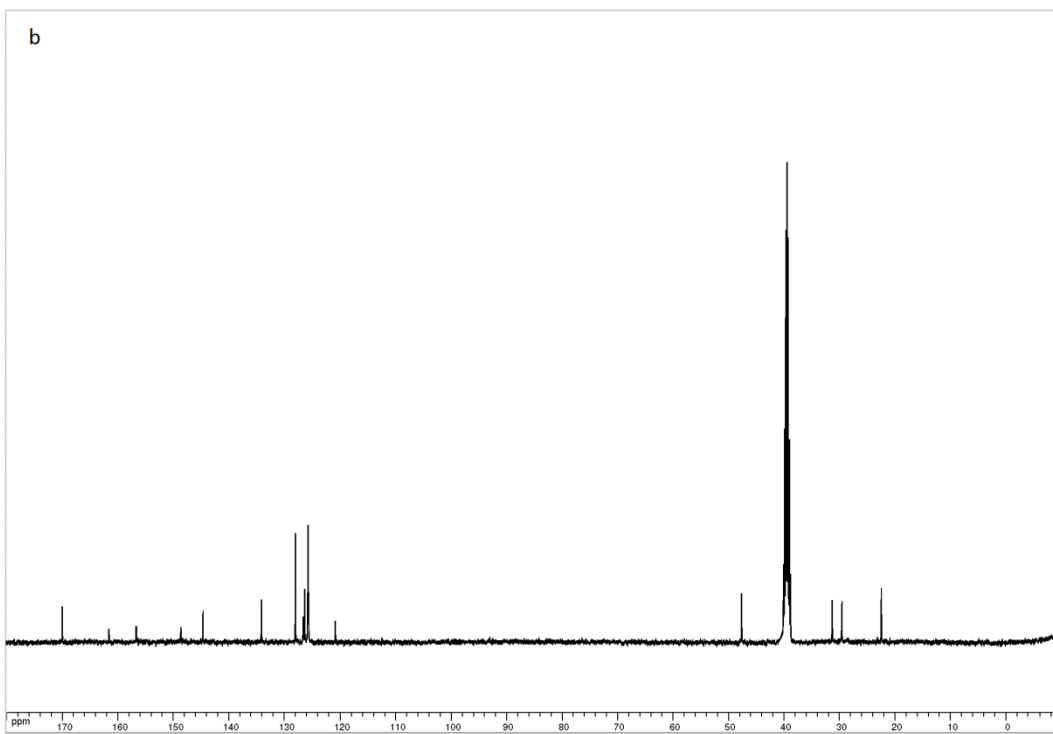
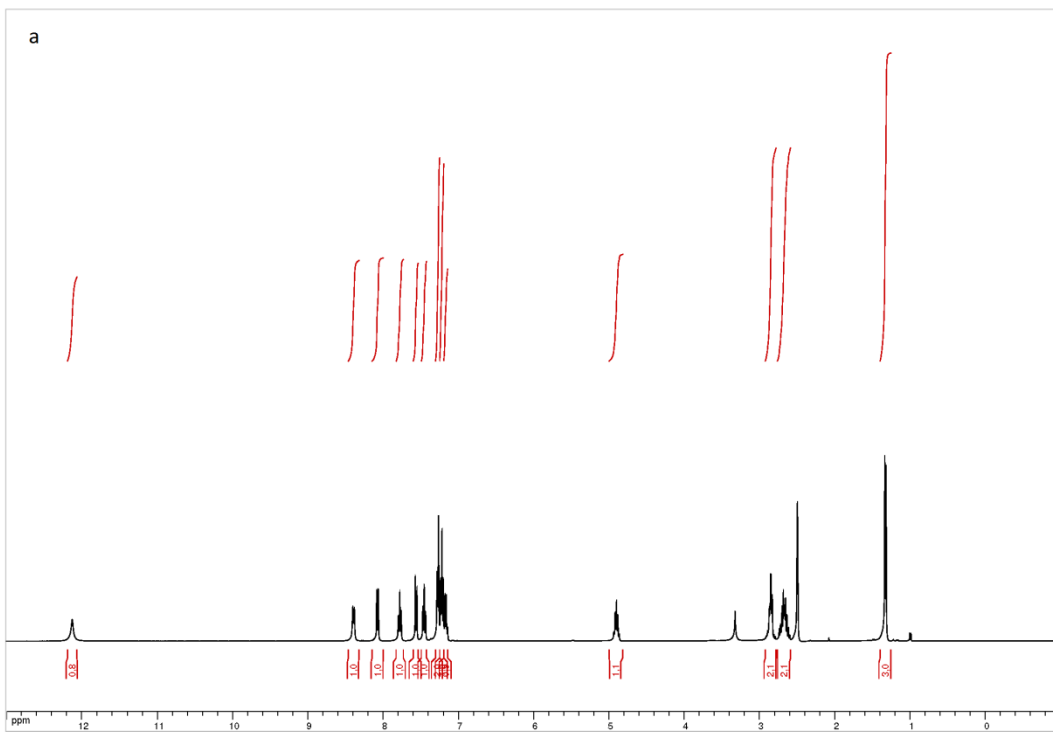
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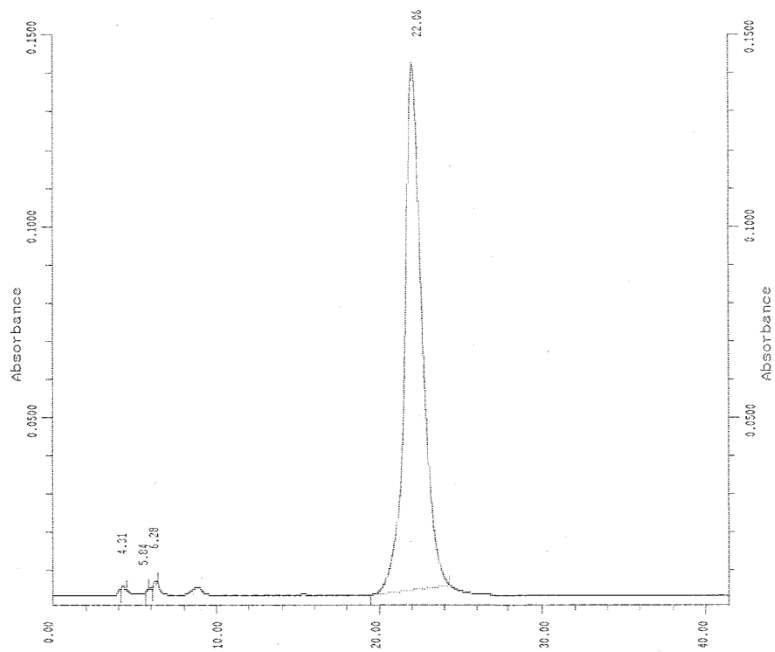
Supplementary Figure 4. Characterization of ME0327. (a) ^1H -NMR spectrum. (b) ^{13}C -NMR spectrum. (c) Chiral HPLC UV trace.



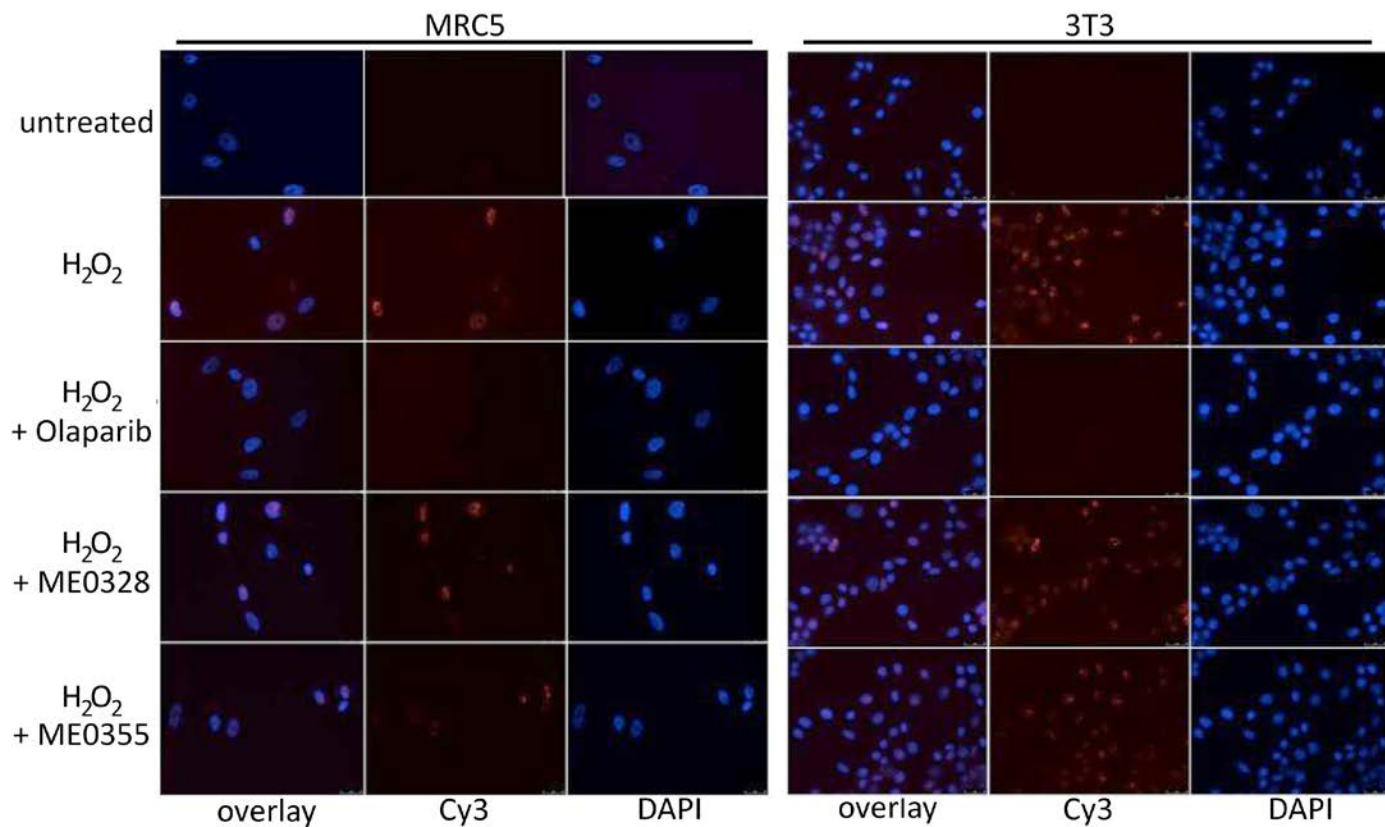
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Supplementary Figure 5. Characterization of ME0328. (a) ^1H -NMR spectrum. (b) ^{13}C -NMR spectrum. (c) Chiral HPLC UV trace.



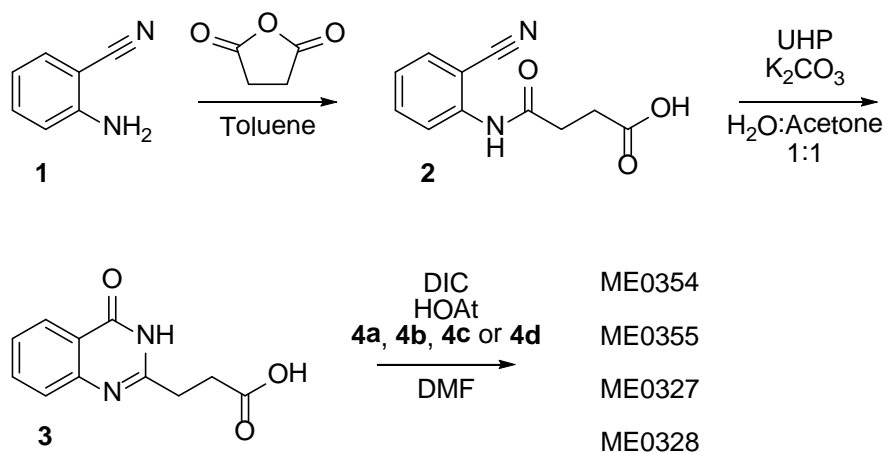
Supplementary Figure 6. ME0328 and ME0355 have no effect on ARTD1/PARP1 activity in cells after H₂O₂-induced DNA damage. MRC5 or 3T3 cells were pretreated for 1 hour with either Olaparib or ME0328 or ME0355 (20 μ M) prior to H₂O₂ treatment (1 mM for 10 minutes). Cells were fixed (methanol/acetic acid) and stained with anti-PAR 10H antibody followed by Cy3-coupled anti-mouse antibody. Cells were co-stained with DAPI to visualize nuclei.

Supplementary Methods

General Chemical Procedures. LCMS analysis was carried out using a Waters LC system equipped with an Xterra MS C18 18.5 μm 4.6*50 mm column and an eluent system consisting of MeCN in water, both of which contained 0.2% formic acid. Detection was performed at 214 and 254 nm. Mass spectra were obtained using a Waters Micromass ZG 2000 using both positive and negative electrospray ionization. ^1H NMR and ^{13}C NMR spectra were recorded using a Bruker DRX-400 spectrometer in CDCl_3 solution (residual CHCl_3 (δ_{H} 7.26 ppm, δ_{C} 77.16 ppm) as internal standard) or in $(\text{CD}_3)_2\text{SO}$ solution (residual $(\text{CH}_3)_2\text{SO}$ (δ_{H} 2.50 ppm, δ_{C} 39.52 ppm) as internal standard). Optical rotations were measured on a Perkin Elmer Polarimeter 343 at 20 $^\circ\text{C}$. All target compounds were $\geq 95\%$ pure according to LCMS UV-traces. Chiral HPLC was performed using a Beckman system equipped with a Pirkle Covalent (S,S) Whelk-O 1 10/100 Krom FEC 25 cm x 4.6 mm column and an eluent system consisting of isopropanol and hexane in a one to one ratio. Detection was performed at 254 nm using a System Gold 166 Detector.

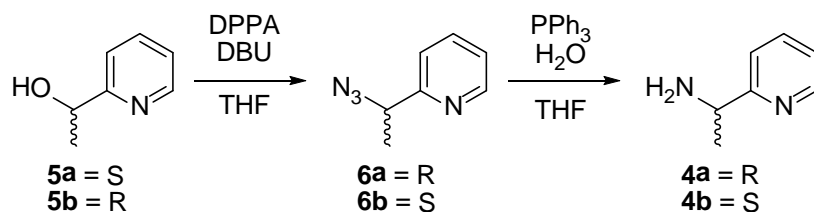
Synthetic strategy. The synthetic route to compounds ME0327, ME0328, ME0354 and ME0355 is presented in scheme 1. Reacting 2-aminobenzonitrile (**1**) with succinic anhydride in toluene gave the resulting anilide (**2**) in 96% yield. Compound **2** was then ring closed to the quinazolinone main fragment (**3**) in 68% yield using UHP and K_2CO_3 in a H_2O :acetone 1:1 mixture.⁴ Finally, ME0327, ME0328, ME0354 and ME0355 were obtained from **3** in 50-78% yield using standard amide coupling reagents DIC and HOAt together with the appropriate amine in DMF.

Scheme 1. Synthesis of compounds ME0327, ME0328, ME0354 and ME0355.

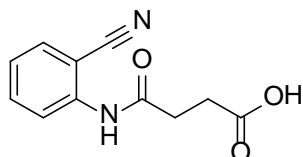


The two amines (**4a** and **4b**) corresponding to ME0354 and ME0355 were not commercially available and thus had to be synthesized from their respective alcohols (scheme 2). This was done by reacting **5a** or **5b** with DPPA and DBU in THF to produce the azides, **6a** and **6b**, in 53-64% yield with inversion of absolute configuration. The azides were then reduced to the desired amines in 79-97% yield using PPh_3 and H_2O in THF.

Scheme 2. Synthesis and absolute configuration of amine building blocks **4a** and **4b**.

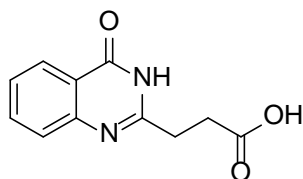


4-(2-cyanophenylamino)-4-oxobutanoic acid (**2**)



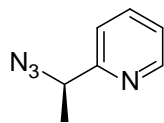
2-aminobenzonitrile (**1**) (1500 mg, 12.7 mmol) and succinic anhydride (1525 mg, 15.24 mmol) in toluene (15 ml) were heated to 90 °C for 3 hours. The solid material was filtered off, washed with Et₂O and dried *in vacuo* to give 4-(2-cyanophenylamino)-4-oxobutanoic acid (**2**) (2670 mg, 96%). ¹H NMR (400 MHz, (CD₃)₂SO) δ 12.16 (s, 1H), 10.18 (s, 1H), 7.79 (dd, J = 7.8, 1.6 Hz, 1H), 7.67 (ddd, J = 8.6, 7.4, 1.2 Hz, 1H), 7.59 (d, 7.7 Hz, 1H), 7.33 (t, J = 7.6 Hz, 1H), 2.63 (t, J = 6.8 Hz, 2H), 2.53 (t, 6.8 Hz, 2H); ¹³C NMR (100 MHz, (CD₃)₂SO) δ 173.6, 170.6, 140.2, 133.7, 133.2, 125.4, 125.2, 116.8, 106.9, 30.5, 28.8; ESIMS *m/z* calcd [M+H]⁺, 219.08; found 219.13.

3-(4-oxo-3H-quinazolin-2-yl)propanoic acid (**3**)



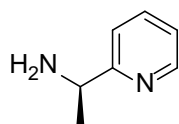
4-(2-cyanophenylamino)-4-oxobutanoic acid (**2**) (2100 mg, 9.62 mmol) was dissolved in Acetone:H₂O 1:1 (120 ml) and K₂CO₃ (2660 mg, 19.24 mmol) followed by UHP (2715 mg, 28.8 mmol) were added. The reaction mixture was heated to 82 °C for 40 hours. K₂CO₃ (2660 mg, 19.24 mmol) and UHP (2715 mg, 28.8 mmol) were added and the mixture was heated to 82 °C for another 48 hours. The pH of the water phase was adjusted to 4 using 6 M HCl. The solid material was filtered off and washed with MeOH and DCM to give 3-(4-oxo-3H-quinazolin-2-yl)propanoic acid (**3**) (1422 mg, 58%). ¹H NMR (400 MHz, (CD₃)₂SO) δ 12.27-12.15 (m, 2H), 8.07 (d, J = 7.7 Hz, 1H), 7.76 (ddd, J = 8.2, 7.2, 1.0 Hz, 1H), 7.57 (d, J = 8.3 Hz, 1H), 7.45 (t, J = 7.5 Hz, 1H), 2.85 (t, 6.9 Hz, 2H), 2.75 (t, 6.8 Hz, 2H); ¹³C NMR (100 MHz, (CD₃)₂SO) δ 173.5, 161.6, 156.3, 148.6, 134.3, 126.7, 126.0, 125.7, 120.9, 29.8, 29.1; ESIMS *m/z* calcd [M+H]⁺, 219.08; found 219.26.

(R)-2-(1-azidoethyl)pyridine (**6a**)



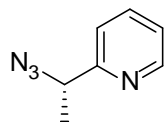
DBU (0.18 ml, 1.22 mmol) followed by DPPA (0.26 ml, 1.22 mmol) were added dropwise to a stirred solution of (S)- α -methyl-2-pyridinemethanol (**5a**) (100 mg, 0.1 mmol) in THF (3.5 ml). The reaction mixture was stirred at room temperature for 20 hours, concentrated and purified using column chromatography on silica gel (EtOAc:Heptane 25:75) to give (R)-2-(1-azidoethyl)pyridine (**6a**) (64 mg, 53%). ¹H NMR (400 MHz, CDCl₃) δ 8.57 (dd, J = 4.9, 1.8 Hz, 1H), 7.69 (td, J = 7.7, 1.9 Hz, 1H), 7.33 (d, J = 7.8 Hz, 1H), 7.20 (dd, J = 7.6, 4.9 Hz, 1H), 4.65 (q, 6.9 Hz, 1H), 1.59 (d, J = 6.9 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 160.1, 149.5, 137.1, 122.9, 120.6, 61.7, 20.2; ESIMS *m/z* calcd [M-N₂+H]⁺, 121.08; found 121.23.

(R)-1-(pyridin-2-yl)ethanamine (**4a**)



(R)-2-(1-azidoethyl)pyridine (**6a**) (64 mg (0.43 mmol) and triphenylphosphine (135 mg, 0.51 mmol) were dissolved in THF (2.5 ml) and H₂O (0.12 ml, 6.4 mmol) was added. The reaction mixture was stirred at room temperature for 26 hours, diluted with DCM and extracted with 1 M HCl. The pH of the water phase was adjusted to 12 and the water phase was extracted five times with DCM. The combined organic phases were dried (Na₂SO₄), filtered and concentrated to give (R)-1-(pyridin-2-yl)ethanamine (**4a**) (51 mg, 97%). ¹H NMR (400 MHz, CDCl₃) δ 8.53 (d, 4.8 Hz, 1H), 7.62 (td, J = 7.7, 1.8 Hz, 1H), 7.28 (d, J = 7.8 Hz, 1H), 7.12 (dd, 7.5, 4.9 Hz, 1H), 4.13 (q, J = 6.7 Hz, 1H), 1.90 (s, 2H), 1.41 (d, 6.7 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 165.8, 149.2, 136.6, 121.9, 120.1, 52.5, 24.5; ESIMS *m/z* calcd [M+H]⁺, 123.09; found 123.27.

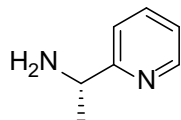
(S)-2-(1-azidoethyl)pyridine (**6b**)



DBU (0.18 ml, 1.22 mmol) followed by DPPA (0.26 ml, 1.22 mmol) were added dropwise to a stirred solution of (R)- α -methyl-2-pyridinemethanol (**5b**) (100 mg, 0.1 mmol) in THF (3.5 ml). The reaction mixture was stirred at room temperature for 20 hours, concentrated and purified using column chromatography on silica gel (EtOAc:Heptane 25:75) to give (S)-2-(1-azidoethyl)pyridine (**6b**) (77 mg, 64%). ¹H NMR (400 MHz, CDCl₃) δ 8.57 (dd, J = 4.9, 1.8 Hz,

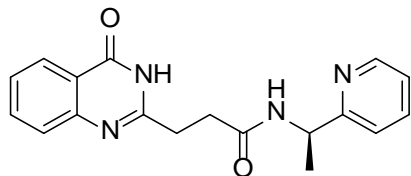
1H), 7.69 (td, $J = 7.7, 1.9$ Hz, 1H), 7.33 (d, $J = 7.8$ Hz, 1H), 7.20 (dd, $J = 7.6, 4.9$ Hz, 1H), 4.65 (q, 6.9 Hz, 1H), 1.59 (d, $J = 6.9$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 160.1, 149.5, 137.1, 122.9, 120.6, 61.7, 20.2.

(S)-1-(pyridin-2-yl)ethanamine (4b)



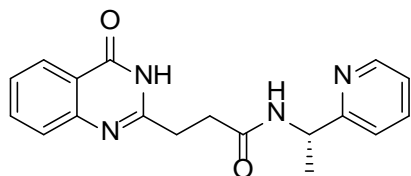
(S)-2-(1-azidoethyl)pyridine (**6b**) (60 mg (0.40 mmol) and triphenylphosphine (127 mg, 0.48 mmol) were dissolved in THF (2.5 ml) and H_2O (0.11 ml, 6.0 mmol) was added. The reaction mixture was stirred at room temperature for 26 hours, diluted with DCM and extracted with 1 M HCl. The pH of the water phase was adjusted to 12 and the water phase was extracted five times with DCM. The combined organic phases were dried (Na_2SO_4), filtered and concentrated to give (S)-1-(pyridin-2-yl)ethanamine (**4b**) (39 mg, 79%). ^1H NMR (400 MHz, CDCl_3) δ 8.53 (d, 4.8 Hz, 1H), 7.62 (td, $J = 7.7, 1.8$ Hz, 1H), 7.28 (d, $J = 7.8$ Hz, 1H), 7.12 (dd, 7.5, 4.9 Hz, 1H), 4.13 (q, $J = 6.7$ Hz, 1H), 2.39 (s, 2H), 1.41 (d, 6.7 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 165.8, 149.2, 136.6, 121.9, 120.1, 52.5, 24.5.

(R)-3-(4-oxo-3H-quinazolin-2-yl)-N-(1-(pyridin-2-yl)ethyl)propanamide (ME0354)



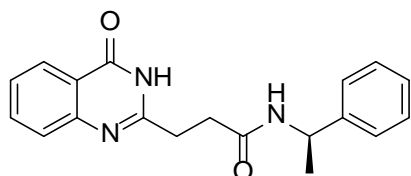
3-(4-oxo-3H-quinazolin-2-yl)propanoic acid (**3**) (40 mg, 0.183 mmol), (R)-1-(pyridin-2-yl)ethanamine (**4a**) (26.9 mg, 0.22 mmol) and HOAt (29.9 mg, 0.22 mmol) were dissolved in DMF (1.5 ml) and DIC (34.3 μl , 0.22 mmol) was added. The reaction mixture was stirred at room temperature for 23 hours, diluted with EtOAc and washed twice with saturated aqueous NaHCO_3 . The organic phase was dried (Na_2SO_4), filtered and concentrated. The residue was purified using column chromatography on silica gel (DCM:MeOH 95:5) to give (R)-3-(4-oxo-3H-quinazolin-2-yl)-N-(1-(pyridin-2-yl)ethyl)propanamide (ME0354) (41 mg, 69%). $[\alpha]_{\text{D}}^{27}$ (c 0.57, DMSO); 99% enantiomeric excess; ^1H NMR (400 MHz, $(\text{CD}_3)_2\text{SO}$) δ 12.17 (s, 1H), 8.49-8.42 (m, 2H), 8.08 (dd, $J = 7.9, 1.6$ Hz, 1H), 7.78 (ddd, $J = 8.2, 7.2, 1.3$ Hz, 1H), 7.60-7.53 (m, 2H), 7.45 (t, $J = 7.4$ Hz, 1H), 7.31 (d, $J = 8.0$ Hz, 1H), 7.19 (dd, $J = 7.5, 4.8$ Hz, 1H), 4.93 (quin, $J = 7.3$ Hz, 1H), 2.94-2.81 (m, 2H), 2.80-2.63 (m, 2H), 1.36 (d, $J = 7.1$ Hz, 3H); ^{13}C NMR (100 MHz, $(\text{CD}_3)_2\text{SO}$) δ 169.9, 162.2, 161.1, 156.2, 148.1 (2C), 136.0, 133.7, 126.1, 125.4, 125.2, 121.3, 120.4, 119.5, 49.2, 30.8, 29.1, 20.6; HRMS (ES+) calcd $[\text{M}+\text{Na}]^+$, 345.1322; found 345.1336.

(S)-3-(4-oxo-3H-quinazolin-2-yl)-N-(1-(pyridin-2-yl)ethyl)propanamide (ME0355)



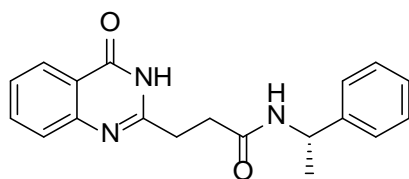
3-(4-oxo-3H-quinazolin-2-yl)propanoic acid (**3**) (40 mg, 0.183 mmol), (S)-1-(pyridin-2-yl)ethanamine (**4b**) (26.9 mg, 0.22 mmol) and HOAt (29.9 mg, 0.22 mmol) were dissolved in DMF (1.5 ml) and DIC (34.3 μ l, 0.22 mmol) was added. The reaction mixture was stirred at room temperature for 23 hours, diluted with EtOAc and washed twice with saturated aqueous NaHCO₃. The organic phase was dried (Na₂SO₄), filtered and concentrated. The residue was purified using column chromatography on silica gel (DCM:MeOH 95:5) to give (R)-3-(4-oxo-3H-quinazolin-2-yl)-N-(1-(pyridin-2-yl)ethyl)propanamide (ME0355) (31 mg, 53%). [α]_D -24° (*c* 0.65, DMSO); 90% enantiomeric excess; ¹H NMR (400 MHz, (CD₃)₂SO) δ 12.17 (s, 1H), 8.49-8.42 (m, 2H), 8.08 (dd, *J* = 7.9, 1.6 Hz, 1H), 7.78 (ddd, *J* = 8.2, 7.2, 1.3 Hz, 1H), 7.60-7.53 (m, 2H), 7.45 (t, *J* = 7.4 Hz, 1H), 7.31 (d, *J* = 8.0 Hz, 1H), 7.19 (dd, *J* = 7.5, 4.8 Hz, 1H), 4.93 (quin, *J* = 7.3 Hz, 1H), 2.94-2.81 (m, 2H), 2.80-2.63 (m, 2H), 1.36 (d, *J* = 7.1 Hz, 3H); ¹³C NMR (100 MHz, (CD₃)₂SO) δ 169.9, 162.2, 161.1, 156.2, 148.1 (2C), 136.0, 133.7, 126.1, 125.4, 125.2, 121.3, 120.4, 119.5, 49.2, 30.8, 29.1, 20.6; HRMS (ES⁺) calcd [M+Na⁺]⁺, 345.1322; found 345.1321.

(R)-3-(4-oxo-3H-quinazolin-2-yl)-N-(1-phenylethyl)propanamide (ME0327)



3-(4-oxo-3H-quinazolin-2-yl)propanoic acid (**3**) (40 mg, 0.183 mmol), (R)-(+)-1-phenylethylamine (**4c**) (28.1 μ l, 0.22 mmol) and HOAt (29.9 mg, 0.22 mmol) were dissolved in DMF (1.5 ml) and DIC (34.3 μ l, 0.22 mmol) was added. The reaction mixture was stirred at room temperature for 19 hours, diluted with EtOAc and washed twice with saturated aqueous NaHCO₃. The organic phase was dried (Na₂SO₄), filtered and concentrated. The residue was purified using column chromatography on silica gel (DCM:MeOH 96:4) to give (S)-3-(4-oxo-3H-quinazolin-2-yl)-N-(1-phenylethyl)propanamide (ME0327) (46 mg, 78%). [α]_D 21° (*c* 0.63, DMSO); 98% enantiomeric excess; ¹H NMR (400MHz, (CD₃)₂SO) δ 12.13 (s, 1H), 8.39 (d, *J* = 8.0 Hz, 1H), 8.08 (dd, *J* = 7.9, 1.6 Hz, 1H), 7.78 (ddd, *J* = 9.0, 7.1, 1.8 Hz, 1H), 7.56 (d, 8.3 Hz, 1H), 7.46 (t, *J* = 7.7 Hz, 1H), 7.30-7.25 (m, 2H), 7.25-7.20 (m, 2H), 7.20-7.15 (m, 1H), 4.90 (quin, *J* = 7.3 Hz, 1H), 2.93-2.79 (m, 2H), 2.76-2.60 (m, 2H), 1.33 (d, *J* = 7.0 Hz, 1H); ¹³C NMR (100 MHz, (CD₃)₂SO) δ 170.1, 161.7, 156.8, 148.7, 144.7, 134.2, 128.1 (2C), 126.7, 126.4, 125.8 (2C), 125.8, 125.7, 120.9, 47.7, 31.4, 29.7, 22.5; HRMS (ES⁺) calcd [M+Na⁺]⁺, 344.1369; found 344.1370.

(S)-3-(4-oxo-3H-quinazolin-2-yl)-N-(1-phenylethyl)propanamide (ME0328)



3-(4-oxo-3H-quinazolin-2-yl)propanoic acid (**3**) (40 mg, 0.183 mmol), (S)-(-)-1-phenylethylamine (**4d**) (28.1 μ l, 0.22 mmol) and HOAt (29.9 mg, 0.22 mmol) were dissolved in DMF (1.5 ml) and DIC (34.3 μ l, 0.22 mmol) was added. The reaction mixture was stirred at room temperature for 19 hours, diluted with EtOAc and washed twice with saturated NaHCO₃. The organic phase was dried (Na₂SO₄), filtered and concentrated. The residue was purified using column chromatography on silica gel (DCM:MeOH 96:4) to give (S)-3-(4-oxo-3H-quinazolin-2-yl)-N-(1-phenylethyl)propanamide (ME0328) (46 mg, 78%). [α]_D -22° (c 0.65, DMSO); 98% enantiomeric excess; ¹H NMR (400 MHz, (CD₃)₂SO) δ 12.13 (s, 1H), 8.39 (d, J = 8.0 Hz, 1H), 8.08 (dd, J = 7.9, 1.6 Hz, 1H), 7.78 (ddd, J = 9.0, 7.1, 1.8 Hz, 1H), 7.56 (d, 8.3 Hz, 1H), 7.46 (t, J = 7.7 Hz, 1H), 7.30-7.25 (m, 2H), 7.25-7.20 (m, 2H), 7.20-7.15 (m, 1H), 4.90 (quin, J = 7.3 Hz, 1H), 2.93-2.79 (m, 2H), 2.76-2.60 (m, 2H), 1.33 (d, J = 7.0 Hz, 1H); ¹³C NMR (100 MHz, (CD₃)₂SO) δ 170.1, 161.7, 156.8, 148.7, 144.7, 134.2, 128.1 (2C), 126.7, 126.4, 125.8 (2C), 125.8, 125.7, 120.9, 47.7, 31.4, 29.7, 22.5; HRMS (ES⁺) calcd [M+Na⁺]⁺, 344.1369; found 344.1374.

Abbreviations

Ac	Acetate
DBU	1,8-Diazabicyclo[5.4.0]undec-7-ene
DCM	Dichloromethane
DIC	<i>N,N'</i> -Diisopropylcarbodiimide
DMF	<i>N,N</i> -Dimethylformamide
DMSO	Dimethylsulfoxide
DPPA	Diphenyl phosphoryl azide
Et	Ethyl
FCS	Fetal calf serum
HOAt	1-Hydroxybenzotriazole
HRMS	High-resolution Mass Spectrometry
LCMS	Liquid Chromatography Mass Spectrometry
MDCK	Madin-Darby Canine Kidney
Me	Methyl
NMR	Nuclear Magnetic Resonance
Ph	Phenyl
THF	Tetrahydrofuran
UHP	Urea Hydrogen Peroxide
UV	Ultra Violet

Supplementary References

- (1) Chen, V. B., Arendall, W. B., 3rd, Headd, J. J., Keedy, D. A., Immormino, R. M., Kapral, G. J., Murray, L. W., Richardson, J. S., Richardson, D. C. (2010) MolProbity: all-atom structure validation for macromolecular crystallography. *Acta crystallogr. D Biol. crystallogr.* 66, 12-21.
- (2) Aguiar, R. C., Takeyama, K., He, C., Kreinbrink, K., Shipp, M. A. (2005) B-aggressive lymphoma family proteins have unique domains that modulate transcription and exhibit poly(ADP-ribose) polymerase activity. *J. Biol. Chem.* 280, 33756-33765.
- (3) McGinnity, D. F., Parker, A. J., Soars, M., and Riley, R. J. (2000) Automated definition of the emzymology of drug oxidation by the major human drug metabolizing cytochtome P450s. *Drug Metabol. Dispos.* 28, 1327-1334.
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