

1 Supporting Information

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4 **Dechlorination of commercial PCBs and other multiple halogenated compounds by**
5 **a sediment-free culture containing *Dehalococcoides* and *Dehalobacter***

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8 Shanquan Wang, and Jianzhong He*

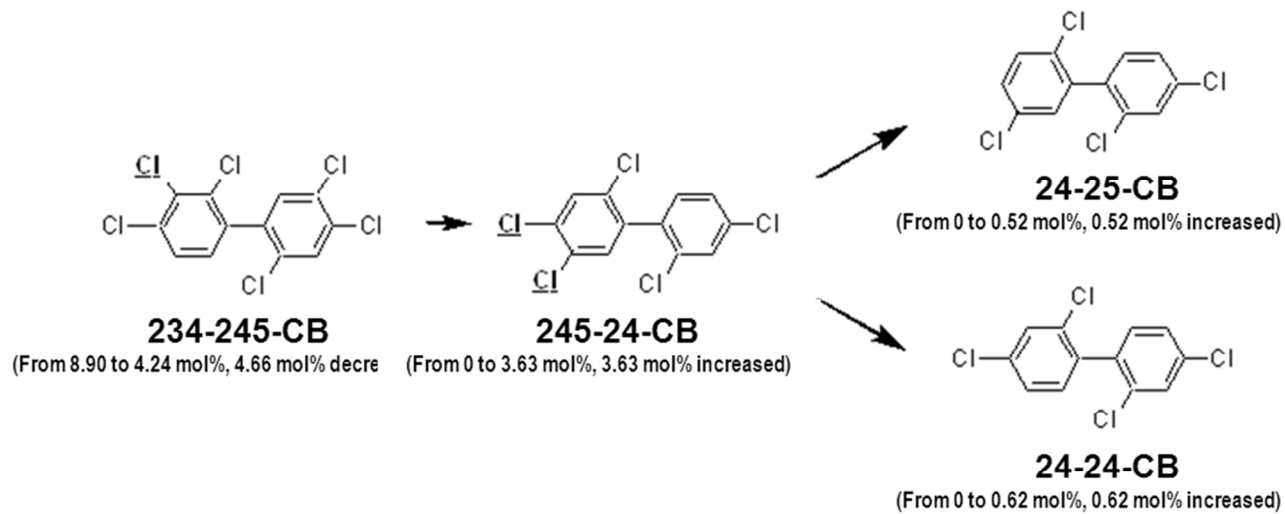
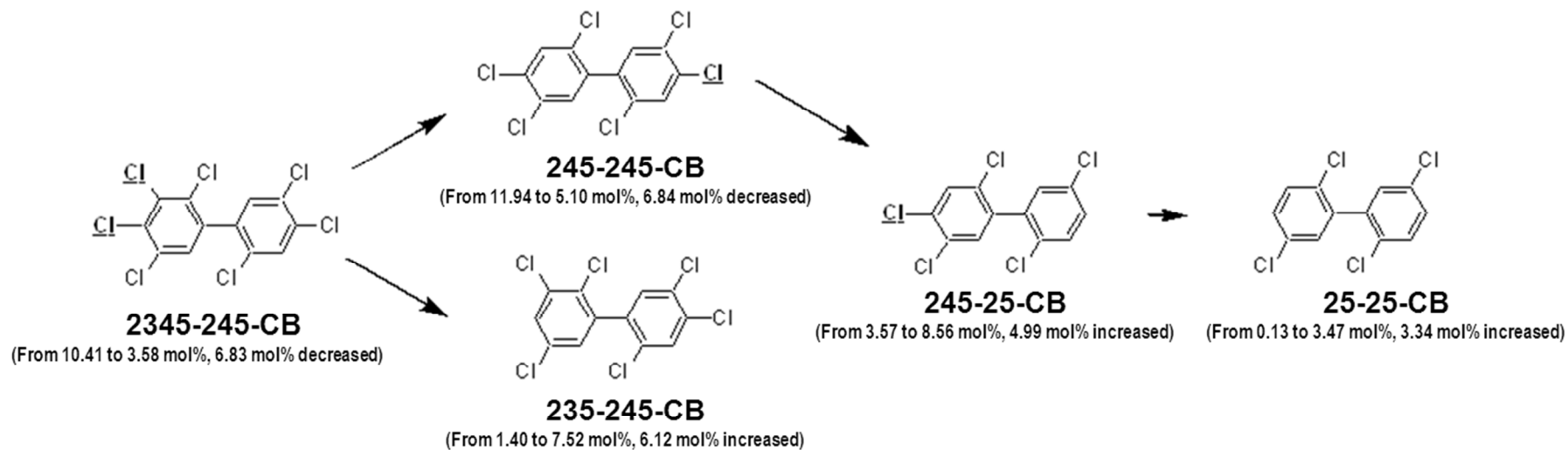
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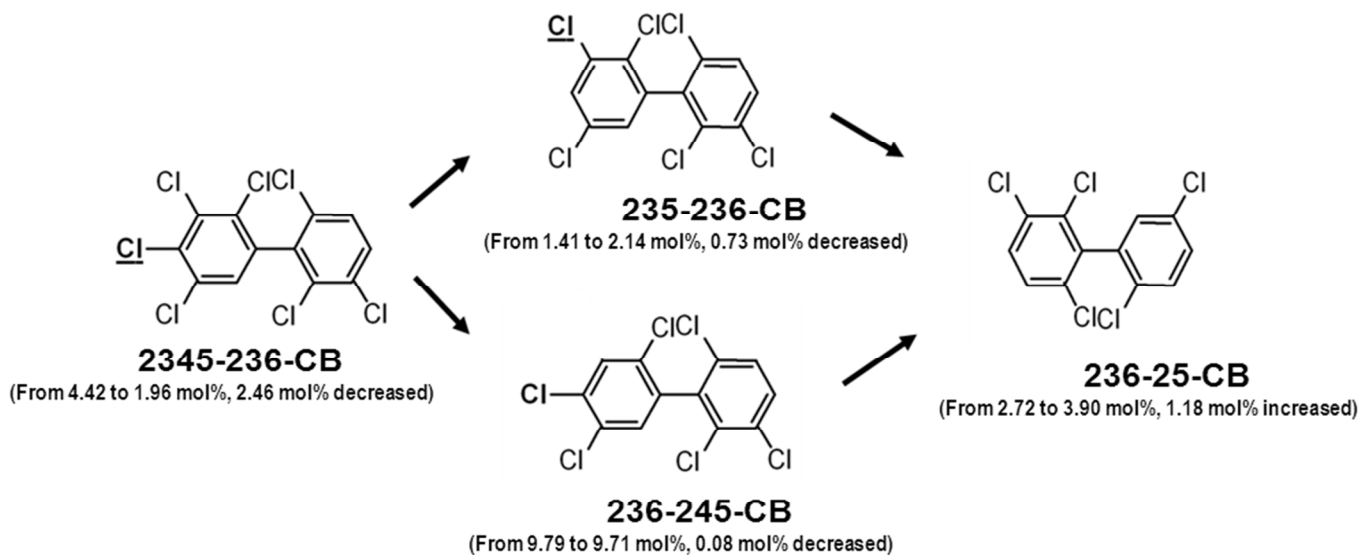
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13 The 13 pages supplementary materials include 8 figures and 3 tables.

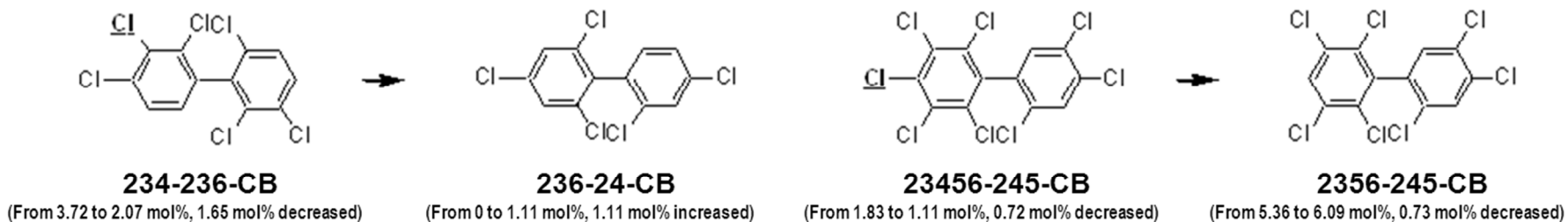


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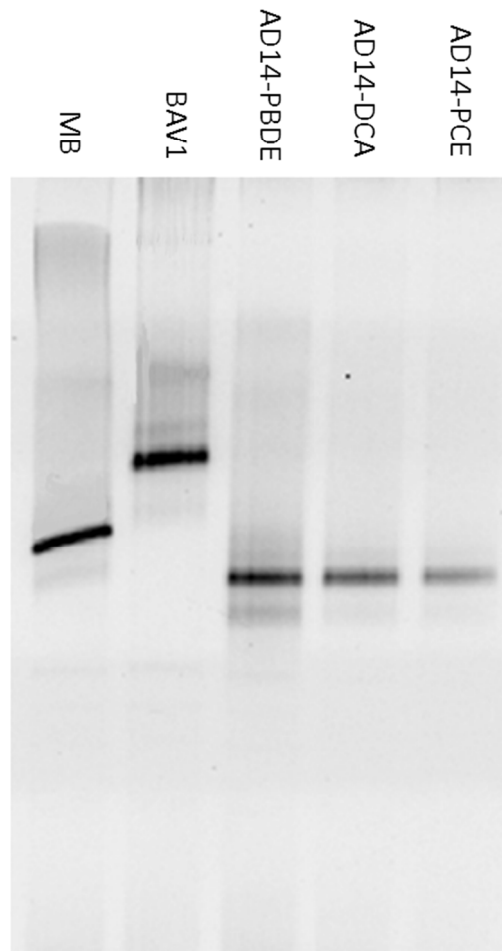


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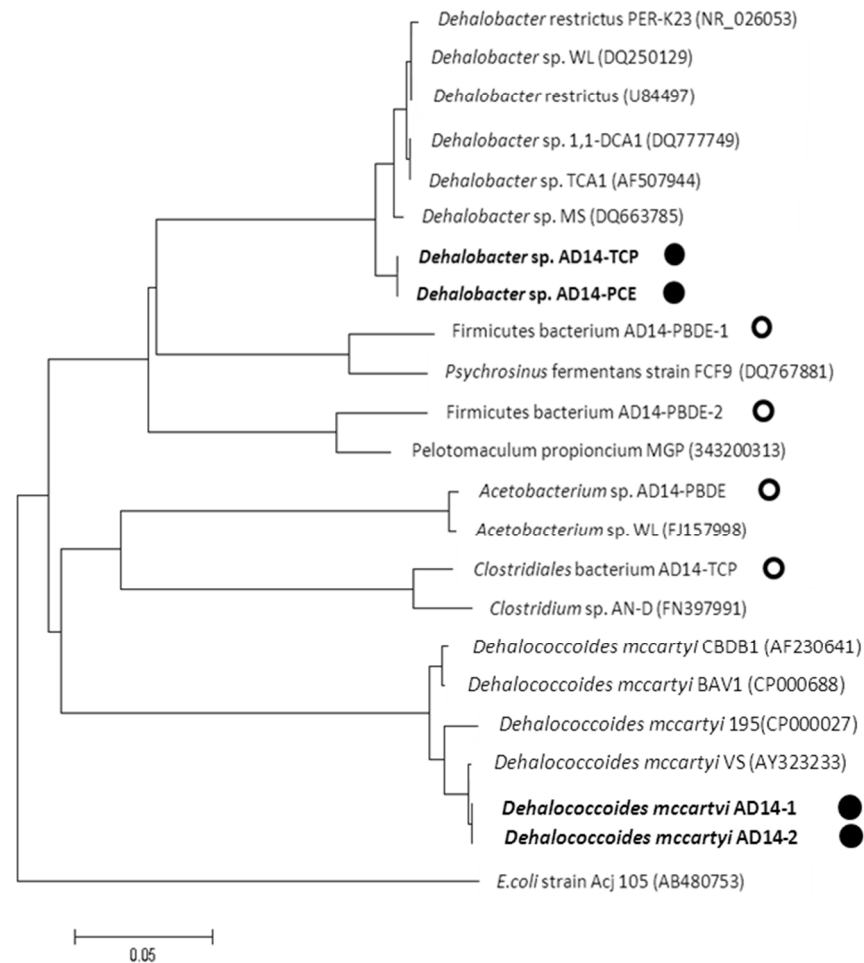
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18 **Figure S1.** Inferred dechlorination pathways and products for most congeners dechlorinated by AD14 culture after four months of
 19 incubation. The dechlorination pathways for 2345-245-CB and 234-245-CB were confirmed by studying the dechlorination of
 20 individual congeners in AD14 cultures.



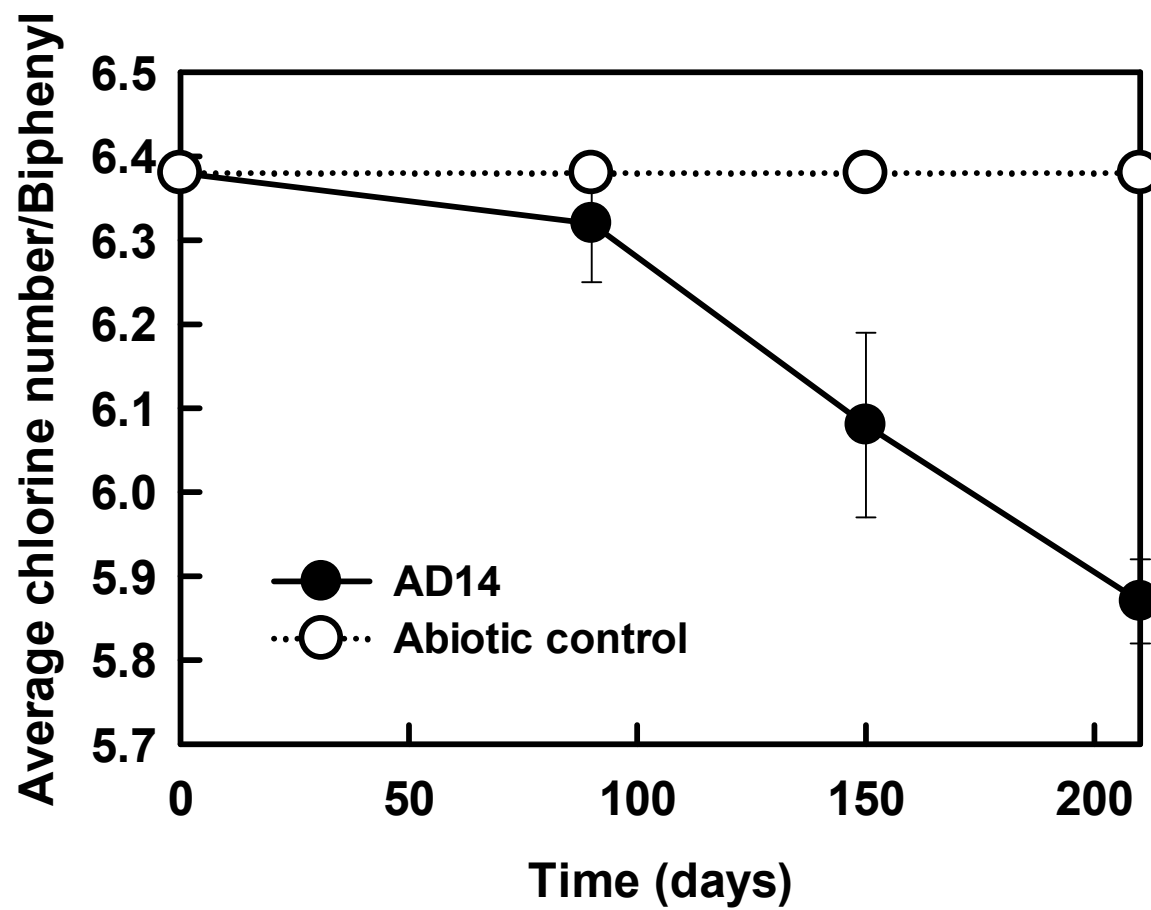
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22 **Figure S2.** DGGE analysis of amplified 16S rRNA gene sequences from highly-enriched subcultures with *Dehalococcoides*-specific
23 1FGC and 259R primer set (Duhamel et al., 2004, AEM).

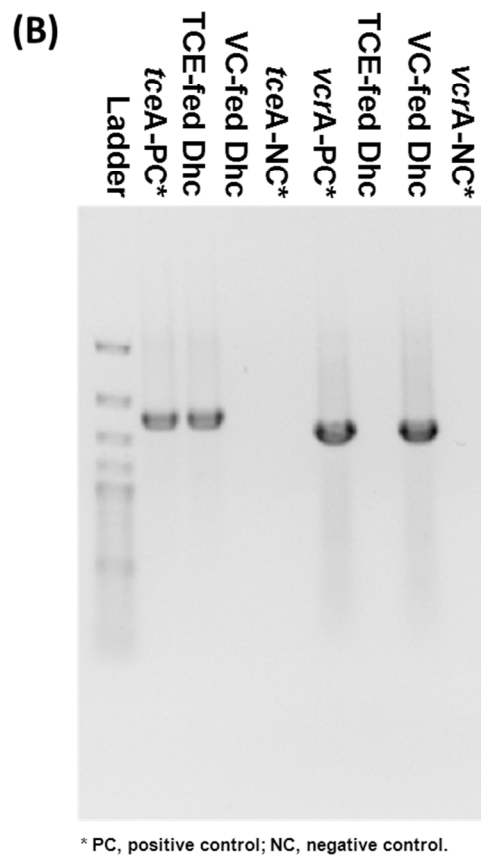
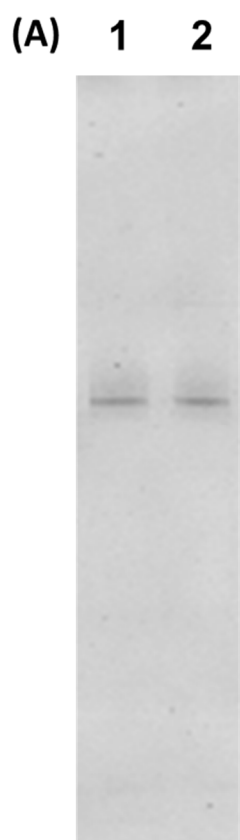


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25 **Figure S3.** Phylogenetic tree of bacterial 16S rRNA genes from enriched subcultures (Dechlorinators and other non-dechlorinating
 26 bacteria were labeled with closed and open circles, respectively) and their closest relatives. Phylogenetic analyses was conducted via
 27 neighbor-joining with MEGA4.²⁷



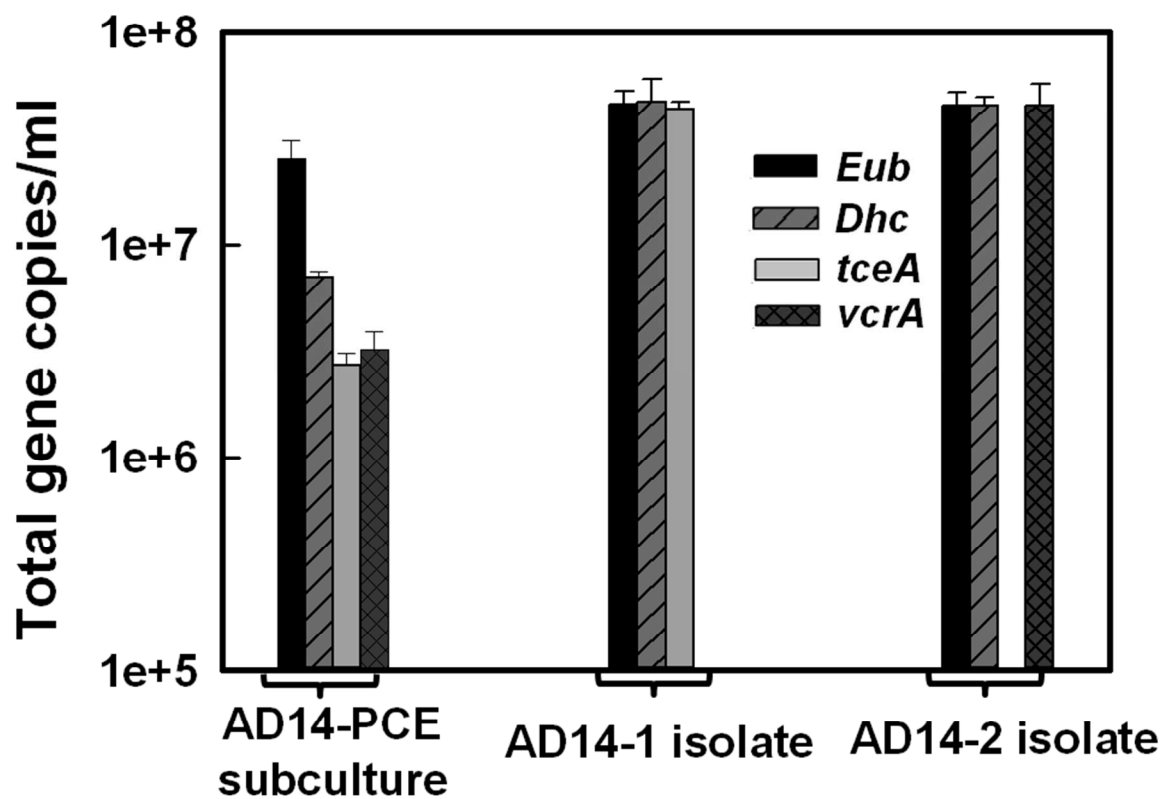
30 **Figure S4.** Decrease in average chlorine number per biphenyl in AD14-PCE subculture and its abiotic control.



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32 **Figure S5.** (A) DGGE analysis (16S rRNA gene sequences were amplified with universal bacterial primers 8FGC and 518R) confirm
 33 the culture purity of *Dehalococcoides mccartyi* strain AD14-1 (TCE-fed, lane 1) and strain AD14-2 (VC-fed, lane 2); (B) PCR
 34 amplification with *tceA*-/*vcrA*-gene specific primers suggest that TCE- and VC-fed *Dehalococcoides* (Dhc) exclusively possess *tceA*
 35 and *vcrA* gene, respectively.

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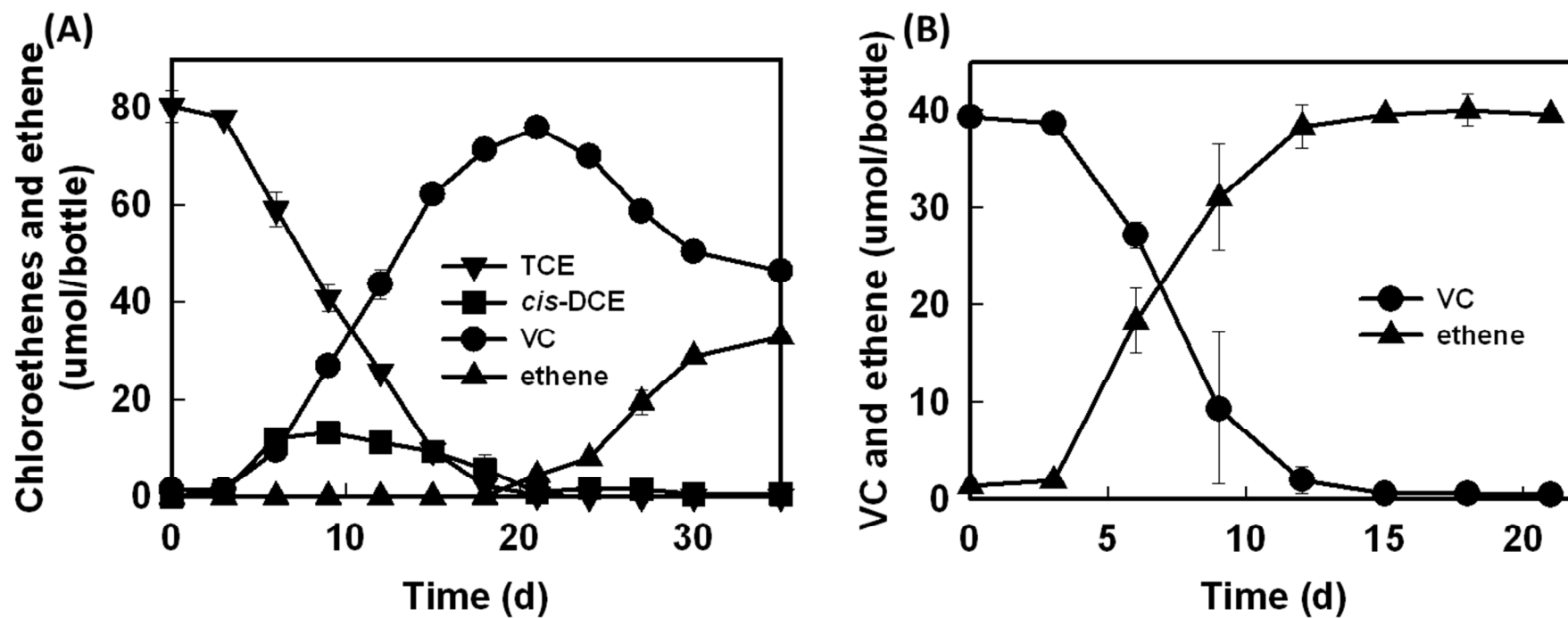
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46 **Figure S7.** qPCR analysis of total bacteria and *Dehalococcoides* during isolating strains AD14-1 and AD14-2 from subculture AD14-
47 PCE with 16S rRNA gene- and functional gene-targeted primers. *Eub*, *Eubacteria*; *Dhc*, *Dehalococcoides*.

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52 **Figure S8.** Dechlorination of (A) TCE by strain AD14-1 and (B) VC by strain AD14-2.

53 **Table S1.** Primers and probes used in this chapter

| Primer | Target | Orientation | Sequence (5' to 3') | Reference or source |
|-----------------------|--------------------------------------|-------------|---|---------------------|
| 8F | <i>Bacteria</i> 16S rRNA genes | Forward | AGAGTTTGATCCTGGCTCAG | 1 |
| 1392R | <i>Bacteria</i> 16S rRNA genes | Reverse | ACGGGCGGTGTGTAC | 2 |
| BacF1 | <i>Bacteria</i> 16S rRNA genes | Forward | TCCTACGGGAGGCAGCAG | 3 |
| BacR1 | <i>Bacteria</i> 16S rRNA genes | Reverse | GGACTACCAGGGTATCTAATCCTG TT | 3 |
| BacR2 | <i>Bacteria</i> 16S rRNA genes | Reverse | GGACTACCAGAGTATCTAATTCTGT T | 3 |
| BacProbe | <i>Bacteria</i> 16S rRNA genes | Probe | FAM- CGTATTACCGCGGCTGCTGGCAC- TAMRA | 3 |
| DhcF | <i>Dehalococcoides</i> 16S rRNA gene | Forward | GGTAATACGTAGGAAGCAAGCG | 3 |
| DhcR | <i>Dehalococcoides</i> 16S rRNA gene | Reverse | CCGGTTAAGCCGGGAAATT | 3 |
| DhcProbe | <i>Dehalococcoides</i> 16S rRNA gene | Probe | VIC- ACATCCAACCTTGAAAGACCACCTA CGCTCACT-TAMRA | 3 |
| tceAF | <i>tceA</i> gene | Forward | ATCCAGATTATGACCCTGGTGAA | 4 |
| tceAR | <i>tceA</i> gene | Reverse | GCGGCATATATTAGGGCATCTT | 4 |
| tceAProbe | <i>tceA</i> gene | Probe | FAM-TGGGCTATGGCGACCGCAGG- TAMRA | 4 |
| vcrAF | <i>vcrA</i> gene | Forward | CTCGGCTACCGAACGGATT | 3 |
| vcrAR | <i>vcrA</i> gene | Forward | GGGCAGGAGGATTGACACAT | 3 |
| vcrAProbe | <i>vcrA</i> gene | Probe | FAM- CGCACTGGTTATGGCAACCACTC- TAMRA | 3 |
| DhcadkF | <i>adk</i> gene | Forward | GATGAGATTACCATTAGTGT | This study |
| DhcadkR | <i>adk</i> gene | Reverse | TTACCGCACTTTATAGCTGA | This study |
| DhcrpoBF | <i>rpoB</i> gene | Forward | CTGTTTGAGGTTATTGATAATGACC | This study |
| DhcrpoBR | <i>rpoB</i> gene | Reverse | GCCAGAGAACCGATAAGACCA | This study |
| DhcatpDF | <i>atpD</i> gene | Forward | TGCCTTTCCTTTACTCCTACCGAT | This study |
| DhcatpDR | <i>atpD</i> gene | Reverse | GCRTCATCAATAGTGCCTACCAT | This study |
| Deb433F | <i>Dehalobacter</i> 16S rRNA gene | Forward | ATACTGTTGTTAGGGAAGAACGGC | This study |
| Dre645R | <i>Dehalobacter</i> 16S rRNA gene | Reverse | CCTCTCCTGTCCTCAAGCCATA | 5 |
| GC-clamp ^a | - | Forward | CGCCCCCGCGCGCGGGCGGGG GCGGGGACGCGGGG | 6 |

54 ^a The GC-clamp was attached to the 5'-end of the 8F and 519F to get primers 8FGC and 519FGC, respectively.

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59 **Table S2.** Effect of AD14 culture on PCB homolog distribution after 120 days of
60 incubation

| PCB homolog ^a | Mole Percent of Total PCBs | | | % Decrease |
|--------------------------|----------------------------|---|------|------------|
| | Aroclor1260 ^b | Dechlorinated Aroclor 1260 ^c | SD | |
| Tri-CB | 0.07 | 0.10 | 0.03 | |
| Tetra-CB | 0.21 | 4.72 | 0.26 | |
| Penta-CB | 9.63 | 30.35 | 2.47 | |
| Hexa-CB | 47.75 | 38.23 | 1.34 | 19.94 |
| Hepta-CB | 36.26 | 21.86 | 1.63 | 39.71 |
| Octa-CB | 5.51 | 4.18 | 0.09 | 24.14 |
| Nona-CB | 0.57 | 0.56 | 0.01 | |

61 ^a No mono-, or dichlorobiphenyls were detected.

62 ^b Data are the means for two unincubated controls.

63 ^c Data are the means for the three AD14 cultures that showed extensive dechlorination of Aroclor 1260.

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69 **Table S3.** Summary of halogenated compound utility profile by *Dehalococcoides*
70 *mccartyi* strain AD14-1 and AD14-2

| Isolates | Halogenated compounds reduced | End-products |
|--|-------------------------------|--------------------------|
| <i>Dehalococcoides mccartyi</i> AD14-1 | TCE | VC and ethene |
| | 1,2-DCA | Ethene |
| | 2,4,6-TCP | No activity ^a |
| | octa-BDE mixture | hexa-BDEs |
| | Aroclor 1260 | No activity ^b |
| <i>Dehalococcoides mccartyi</i> AD14-2 | PCE/TCE | No activity ^a |
| | <i>cis</i> -DCE | Ethene |
| | 1,2-DCA | Ethene |
| | 2,4,6-TCP | 2,4-DCP |
| | octa-BDE mixture | penta- and tetra-BDEs |
| Aroclor 1260 | No activity ^b | |

71 ^a the activities were assessed after 6 months of incubation.

72 ^b PCB dechlorination activity was assessed after 12 months of incubation. Aroclor 1260 dechlorination activity was
73 conducted in defined medium inoculated with AD14-control culture which was obtained by transferring culture AD14
74 for three times (12 months) in defined media without amending Aroclor 1260.

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77 **References**

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79 hyperthermophilic pink filament community in Octopus Spring, Yellowstone National Park. *Appl.*
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