

Supporting information

Total synthesis of callipeltin B and M, peptidyl marine natural products

Mari Kikuchi and Hiroyuki Konno*

Department of Biochemical Engineering, Graduate School of Science and Technology,

Yamagata University, Yonezawa, Yamagata 992-8510, Japan

e-mail: konno@yz.yamagata-u.ac.jp

Contents

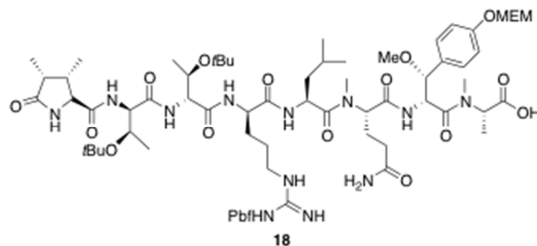
General information.....	S2
Experimental procedures.....	S3-S11
¹ H, ¹³ C NMR and MS spectral data.....	S12-S28

General information

All solvents were reagent grade. CH_2Cl_2 was distilled from CaH_2 . All commercial reagents were of the highest purity available. ^1H (500 or 800 MHz) and ^{13}C NMR (125 or 200 MHz) spectra were recorded on either a JNM-ECX500 or JNM-ECA800. Chemical shifts are expressed in ppm relative to CH_3OH (3.31 ppm for ^1H and 49.0 ppm for ^{13}C). Analytical HPLC was carried out with a COSMOSIL 5C₁₈-AR-II column (4.6ID×150 mm) with a linear gradient of MeCN (0.1% TFA) in H_2O (0.1% TFA) at a run time of 30 min (flow rate of 1 mL/min), on a SHIMADZU SPD-10A as a UV-Vis detector, HITACHI L-6000 Pump and HITACHI L-6200 Intelligent Pump. Preparative HPLC was performed with a COSMOSIL 5C₁₈-AR-II column (10ID×250 mm) with a linear gradient of MeCN (0.1% TFA) in H_2O (0.1% TFA) at a run time of 30 min (flow rate of 2 mL/min), on a SHIMADZU SPD-10Ai as a UV-Vis detector and HITACHI L-6200 Intelligent Pump. UV measurement was recorded at a wavelength of 220 nm. High-resolution mass spectra (HRMS) were obtained using a JEOL AccuTOF JMS-T100LC (ESIMS).

Experimental procedures

pDME-D-*α*Thr(*t*Bu)-D-*α*Thr(*t*Bu)-D-Arg(Pbf)-Leu-MeGln-βMeOTyr(MEM)-MeAla-OH (18)



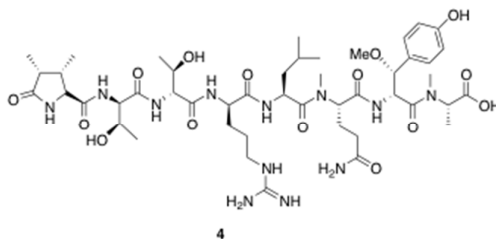
2-Chlorotrityl chloride resin (161 mg, 100-200 mesh, 1% DVB, 1.60 mmol/g) was swelled in DMF (2.5 mL) for 40 min. Fmoc-MeAla-OH (167 mg, 514 μ mol) and *i*-Pr₂NEt (133 μ L, 771 μ mol) were dissolved in DMF (2.5 mL) and added to the preactivated resin, through the reaction for 2 h. The solvent was removed by filtration and the resin washed sequentially with DMF. The resin was treated with a 20% piperidine/DMF solution (2.5 mL) for 30 min. The solvent was removed by filtration and the resin was washed with DMF. Fmoc- β MeOTyr(MEM)-OH (268 mg, 514 μ mol), PyBOP (401 mg, 771 μ mol), HOBT·H₂O (118 mg, 771 μ mol), *i*-Pr₂NEt (177 μ L, 1.03 mmol) dissolved in DMF (2.5 mL) were added to the resin and agitated for 4 h. The solvent was removed by filtration, and the resin was washed with DMF. This procedure was repeated (2 times). The resin was treated with a 20% piperidine/DMF solution (2.5 mL) for 30 min. Fmoc-MeGln-OH (197 mg, 514 μ mol), PyBOP (401 mg, 771 μ mol), HOBT·H₂O (118 mg, 771 μ mol), *i*-Pr₂NEt (177 μ L, 1.03 mmol) dissolved in DMF (2.5 mL) were added to the resin and agitated for 4 h. The solvent was removed by filtration, and the resin was washed with DMF. The resin was treated with a 20% piperidine/DMF solution (2.5 mL) for 30 min. Fmoc-Leu-OH (454 mg, 1.29 mmol), COMU (440 mg, 1.03 mmol), HOAt (140 mg, 1.03 mmol), *i*-Pr₂NEt (266 μ L, 1.54 mmol), dissolved in

DMF (2.5 mL) were added to the resin and agitated for 5 h. The solvent was removed by filtration, and the resin was washed with DMF. The resin was treated with a 20% piperidine/DMF solution (2.5 mL) for 30 min. Fmoc-D-Arg(Pbf)-OH (365 mg, 514 μ mol), PyBOP (401 mg, 771 μ mol), HOBt·H₂O (118 mg, 771 μ mol), *i*-Pr₂NEt (177 μ L, 1.03 mmol) dissolved in DMF (2.5 mL) were added to the resin and agitated for 3 h. The solvent was removed by filtration, and the resin was washed with DMF. The resin was treated with a 20% piperidine/DMF solution (2.5 mL) for 30 min. Fmoc-D-*α*Thr(*t*Bu)-OH (204 mg, 514 μ mol), PyBOP (401 mg, 771 μ mol), HOBt·H₂O (118 mg, 771 μ mol), *i*-Pr₂NEt (177 μ L, 1.03 mmol) dissolved in DMF (2.5 mL) were added to the resin and agitated for 2 h. The solvent was removed by filtration, and the resin was washed with DMF. The resin was treated with a 20% piperidine/DMF solution (2.5 mL) for 30 min.

To the resin (128 μ mol) in DMF (2 mL) was added Fmoc-D-*α*Thr(*t*Bu)-OH (102 mg, 256 μ mol) using PyBOP (200 mg, 384 μ mol), HOBt·H₂O (58.8 mg, 384 μ mol), *i*-Pr₂NEt (88 μ L, 512 μ mol) and agitated for 4 h. The solvent was removed by filtration, and the resin was washed with DMF. The resin was treated with a 20% piperidine/DMF solution (2 mL) for 30 min. pDME (30.2 mg, 192 μ mol), PyBOP (200 mg, 384 μ mol), HOBt·H₂O (58.8 mg, 384 μ mol), *i*-Pr₂NEt (88 μ L, 512 μ mol) dissolved in DMF (2 mL) were added to the resin and agitated for 4 h. The solvent was removed by filtration, and the resin was washed with DMF and Et₂O. The product was cleaved from the resin with a 20% HFIP/CH₂Cl₂ (3.5 mL) for 2.5 h. The mixture was filtrated and evaporated under reduced pressure. To the residue in Et₂O was centrifuged and decanted. The crude product was purified with RP-HPLC (linear gradient from 40% to 80% MeCN) and lyophilized to afford protected callipeltin M (**18**) (7.9 mg, 5.26 μ mol, 4%) as a white powder. **18**: ¹H NMR (500 MHz, CD₃OD) δ 7.30 (d, *J* = 8.5 Hz, 2H),

7.04 (d, $J = 9.0$ Hz, 2H), 5.25, (m, 2H), 5.20 (d, $J = 9.0$ Hz, 1H), 5.12 (q, $J = 7.5$ Hz, 1H), 4.88 (overlapped, 1H), 4.73 (m, 1H), 4.43-4.32 (m, 4H), 4.12 (m, 1H), 4.05 (t, $J = 6.5$ Hz, 1H), 3.86 (d, $J = 3.5$ Hz, 1H), 3.79 (t, $J = 5.0$ Hz, 2H), 3.55 (t, $J = 5.0$ Hz, 2H), 3.33 (s, 3H), 3.18 (m, 2H), 3.14 (s, 3H), 3.09 (s, 2H), 3.01 (s, 3H), 2.83 (s, 3H), 2.67-2.62 (m, 2H), 2.57 (s, 3H), 2.51 (s, 3H), 2.09 (s, 3H), 1.96-1.79 (m, 5H), 1.67-1.59 (m, 6H), 1.46 (s, 6H), 1.40 (d, $J = 7.5$ Hz, 3H), 1.19 (s, 9H), 1.18 (overlapped, 3H), 1.17 (s, 9H), 1.11 (d, $J = 7.5$ Hz, 3H), 1.05 (d, $J = 7.5$ Hz, 3H), 0.95 (d, $J = 6.5$ Hz, 3H), 0.92 (d, $J = 7.0$ Hz, 3H); ^{13}C NMR (125 MHz, CD_3OD) δ 183.3, 175.1, 174.3, 172.4, 170.9, 159.0, 132.1, 130.4, 117.3, 94.7, 85.2, 75.9, 75.9, 72.8, 68.8, 68.2, 63.6, 60.5, 59.1, 57.2, 54.1, 43.9, 40.3, 39.9, 32.7, 32.6, 31.3, 28.7, 26.0, 25.1, 23.8, 21.7, 19.9, 19.6, 18.4, 14.7, 14.7, 12.5, 10.5; HRMS (ESI) m/z : calcd. for $\text{C}_{72}\text{H}_{117}\text{N}_{12}\text{O}_{20}\text{S}$ $[\text{M}+\text{H}]^+$ 1501.8228, found 1501.8207; RP-HPLC: $t_{\text{R}} = 22.2$ min (linear gradient from 10% to 90% MeCN).

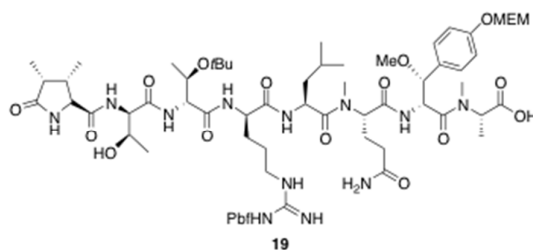
pDME-D-*a*Thr-D-*a*Thr-D-Arg-Leu-MeGln- β MeOTyr-MeAla-OH; callipeltin M (4)



Protected callipeltin M (**18**) (7.9 mg, 5.26 μmol) was added 50% TFA/ CH_2Cl_2 (1 mL) and stirred for 2 h. To the mixture in Et_2O was centrifuged and decanted. The crude material was purified with RP-HPLC (linear gradient from 20% to 90% MeCN) and lyophilized to afford **4** (1.6 mg, 1.53 μmol , 29%) as a white powder. ^1H NMR (500 MHz, CD_3OD) δ 7.19 (d, $J = 8.5$ Hz, 2H), 6.78 (d, $J = 8.5$ Hz, 2H), 5.21 (t, $J = 9.0$ Hz, 1H), 5.13 (q, $J = 7.5$ Hz, 1H), 4.88 (overlapped, 1H), 4.73 (m, 1H),

4.36-4.32 (m, 3H), 4.22 (d, $J = 6.5$ Hz, 1H), 4.09-4.01 (m, 2H), 3.81 (d, $J = 4.0$ Hz, 1H), 3.16 (m, 2H), 3.13 (s, 3H), 3.10 (s, 3H), 2.83 (s, 3H), 2.66 (m, 1H), 2.57 (m, 1H), 1.99-1.88 (m, 3H), 1.72-1.61 (m, 7H), 1.47 (m, 1H), 1.40 (d, $J = 7.5$ Hz, 3H), 1.28 (m, 6H), 1.12 (d, $J = 7.0$ Hz, 3H), 1.07 (d, $J = 7.5$ Hz, 3H), 0.95 (d, $J = 6.0$ Hz, 3H), 0.92 (d, $J = 6.5$ Hz, 3H); ^{13}C NMR (125 MHz, CD_3OD) δ 180.6, 177.6, 174.6, 174.0, 173.6, 172.8, 172.8, 171.0, 169.8, 163.2, 162.9, 130.5, 116.2, 85.3, 68.6, 68.5, 63.9, 61.3, 61.2, 57.4, 57.1, 54.2, 53.8, 48.2, 41.8, 40.6, 40.3, 39.9, 32.6, 32.5, 30.7, 29.4, 26.2, 25.9, 24.9, 23.7, 21.6, 20.7, 20.4, 14.9, 14.7, 10.5; HRMS (ESI) m/z : calcd. for $\text{C}_{47}\text{H}_{77}\text{N}_{12}\text{O}_{15}$ $[\text{M}+\text{H}]^+$ 1049.5631, found 1049.5672; RP-HPLC: $t_{\text{R}} = 10.6$ min (linear gradient from 10% to 90% MeCN).

pDME-D- α Thr-D- α Thr(*t*Bu)-D-Arg(Pbf)-Leu-MeGln- β MeOTyr(MEM)-MeAla-OH
(19)

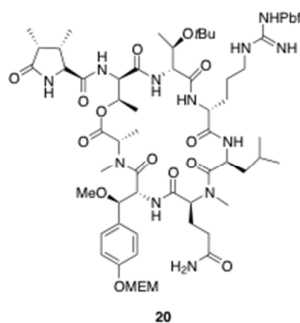


2-Chlorotrityl chloride resin (111 mg, 100-200 mesh, 1% DVB, 1.60 mmol/g) was swelled in DMF (2.5 mL) for 40 min. Fmoc-MeAla-OH (115 mg, 354 μmol) and *i*-Pr₂NEt (91 μL , 531 μmol) were dissolved in DMF (2 mL) and added to the preactivated resin, through the reaction for 2 h. The solvent was removed by filtration and the resin washed sequentially with DMF. The resin was treated with a 20% piperidine/DMF solution (2 mL) for 30 min. The solvent was removed by filtration and the resin washed with DMF. Fmoc- β MeOTyr(MEM)-OH (185 mg, 354 μmol), HATU

(202 mg, 531 μ mol), HOAt (72.3 mg, 531 μ mol), *i*-Pr₂NEt (122 μ L, 708 μ mol) dissolved in DMF (2 mL) were added to the resin and agitated for 4 h. The solvent was removed by filtration, and the resin was washed with DMF. The resin was treated with a 20% piperidine/DMF solution (2 mL) for 30 min. Fmoc-MeGln-OH (135 mg, 354 μ mol), PyBOP (276 mg, 531 μ mol), HOBt·H₂O (81.3 mg, 531 μ mol), *i*-Pr₂NEt (122 μ L, 708 μ mol) dissolved in DMF (2 mL) were added to the resin and agitated for 4 h. The solvent was removed by filtration, and the resin was washed with DMF. The resin was treated with a 20% piperidine/DMF solution (2 mL) for 30 min. Fmoc-Leu-OH (313 mg, 885 μ mol), PyBOP (368 mg, 708 μ mol), HOBt·H₂O (108 mg, 708 μ mol), *i*-Pr₂NEt (183 μ L, 1.06 mmol), dissolved in DMF (2 mL) were added to the resin and agitated for 5 h. The solvent was removed by filtration, and the resin was washed with DMF. The resin was treated with a 20% piperidine/DMF solution (2 mL) for 30 min. Fmoc-D-Arg(Pbf)-OH (251 mg, 354 μ mol), PyBOP (276 mg, 531 μ mol), HOBt·H₂O (81.3 mg, 531 μ mol), *i*-Pr₂NEt (122 μ L, 708 μ mol) dissolved in DMF (2 mL) were added to the resin and agitated for 3 h. The solvent was removed by filtration, and the resin was washed with DMF. The resin was treated with a 20% piperidine/DMF solution (2 mL) for 30 min. Fmoc-D- α Thr(*t*Bu)-OH (141 mg, 354 μ mol), PyBOP (276 mg, 531 μ mol), HOBt·H₂O (81.3 mg, 531 μ mol), *i*-Pr₂NEt (122 μ L, 708 μ mol) dissolved in DMF (2 mL) were added to the resin and agitated for 2 h. The solvent was removed by filtration, and the resin was washed with DMF. The resin was treated with a 20% piperidine/DMF solution (2 mL) for 30 min. Fmoc-D- α Thr-OH (121 mg, 354 μ mol), PyBOP (276 mg, 531 μ mol), HOBt·H₂O (81.3 mg, 531 μ mol), *i*-Pr₂NEt (122 μ L, 708 μ mol) dissolved in DMF (2 mL) were added to the resin and agitated for 4 h. The solvent was removed by filtration, and the resin was washed with DMF. The resin was treated with a 20% piperidine/DMF solution (2 mL) for 30 min. pDME (30.6 mg, 195

μmol), PyBOP (276 mg, 531 μmol), HOBt $\cdot\text{H}_2\text{O}$ (81.3 mg, 531 μmol), *i*-Pr₂NEt (122 μL , 708 μmol) dissolved in DMF (2 mL) were added to the resin and agitated for 2 h. The solvent was removed by filtration, and the resin was washed with DMF and Et₂O. The product was cleaved from the resin with a 20% HFIP/CH₂Cl₂ (4 mL) for 2.5 h. The mixture was filtrated and evaporated under reduced pressure. To the residue in Et₂O was centrifuged and decanted. The crude product was added to 1 M NaOH and stirred for 20 min. The mixture was purified with RP-HPLC (linear gradient from 40% to 75% MeCN) and lyophilized to afford protected callipeltin M (**19**) (17.6 mg, 12.2 μmol , 7%) as a white powder. **19**: ¹H NMR (500 MHz, CD₃OD) δ 8.13 (d, *J* = 6.0 Hz, 1H), 7.87 (d, *J* = 7.5 Hz, 1H), 7.69 (d, *J* = 7.5 Hz, 1H), 7.30 (d, *J* = 8.5 Hz, 2H), 7.03 (d, *J* = 9.0 Hz, 2H), 5.25 (s, 2H), 5.21 (d, *J* = 9.0 Hz, 1H), 5.14 (q, *J* = 7.5 Hz, 1H), 4.87 (overlapped, 1H), 4.76 (m, 1H), 4.44-4.38 (m, 1H), 4.28 (m, 2H), 4.18 (m, 1H), 4.09 (m, 1H), 3.80-3.78 (m, 3H), 3.56-3.54 (m, 2H), 3.33 (s, 3H), 3.17-3.15 (m, 2H), 3.13 (s, 3H), 3.11 (s, 2H), 3.00 (s, 3H), 2.84 (s, 3H), 2.68-2.59 (m, 2H), 2.56 (s, 3H), 2.50 (s, 3H), 2.08 (s, 3H), 1.97-1.58 (m, 11H), 1.45 (s, 6H), 1.40 (d, *J* = 7.5 Hz, 3H), 1.30 (d, *J* = 6.0 Hz, 3H), 1.21 (s, 9H), 1.18-1.09 (m, 6H), 1.07-1.03 (m, 3H), 0.94 (d, *J* = 6.0 Hz, 3H), 0.91 (d, *J* = 6.5 Hz, 3H); ¹³C NMR (125 MHz, CD₃OD) δ 175.3, 174.5, 174.4, 172.8, 172.4, 170.9, 159.0, 132.2, 130.4, 117.3, 94.7, 85.1, 75.9, 72.8, 68.8, 67.4, 63.8, 61.9, 60.3, 59.1, 57.3, 57.2, 54.3, 54.1, 43.9, 40.9, 40.2, 40.1, 32.5, 31.3, 28.7, 25.9, 25.1, 23.8, 21.7, 21.1, 19.6, 19.2, 18.4, 14.9, 14.7, 12.5, 10.5; HRMS (ESI) *m/z*: calcd. for C₆₈H₁₀₉N₁₂O₂₀S [M+H]⁺ 1445.7602, found 1445.7624; RP-HPLC: *t*_R = 20.3 min (linear gradient from 10% to 90% MeCN).

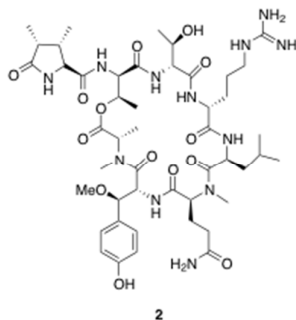
Protected callipeltin B (20)



To the solution of protected callipeltin M (**19**) (0.9 mg, 622 nmol) in DMF (83 μ L) was added the DIPCDI (0.5 μ L, 3.11 μ mol) and DMAP (1.5 mg, 12.4 μ mol). After stirring for 18 h at 45 $^{\circ}$ C, reaction mixture was purified with RP-HPLC (linear gradient from 40% to 80% MeCN) and lyophilized to afford **20** (0.4 mg, 280 nmol, 44%) as a white powder. ^1H NMR (500 MHz, CD_3OD) δ 8.60 (d, J = 10.0 Hz, 1H), 8.38 (d, J = 10.0 Hz, 1H), 8.21 (d, J = 9.0 Hz, 1H), 7.72 (d, J = 7.0 Hz, 1H), 7.66 (d, J = 7.5 Hz, 1H), 7.36 (d, J = 8.5 Hz, 2H), 7.01 (d, J = 8.5 Hz, 2H), 5.42-5.33 (m, 2H), 5.27 (d, J = 7.0 Hz, 1H), 5.22 (d, J = 7.5 Hz, 1H), 4.99-4.92 (m, 2H), 4.73 (m, 1H), 4.54 (d, J = 10.0 Hz, 1H), 4.26 (m, 1H), 4.14 (m, 1H), 4.04 (m, 1H), 3.91 (d, J = 3.0 Hz, 1H), 3.80 (t, J = 5.0 Hz, 2H), 3.57 (t, J = 5.0 Hz, 2H), 3.17 (overlapped, 2H), 3.15 (s, 3H), 3.05 (s, 3H), 3.00 (s, 3H), 2.76 (m, 1H), 2.56 (s, 3H), 2.50 (s, 3H), 2.07 (s, 3H), 1.94-1.49 (m, 11H), 1.45 (s, 6H), 1.33-1.28 (m, 18H), 1.22 (d, J = 7.0 Hz, 3H), 1.19 (d, J = 6.0 Hz, 3H), 0.94 (d, J = 7.0 Hz, 3H), 0.91 (d, J = 6.5 Hz, 3H); ^{13}C NMR (125 MHz, CD_3OD) δ 183.8, 176.8, 175.4, 174.7, 171.7, 171.4, 170.6, 158.9, 156.4, 131.9, 131.4, 130.8, 118.5, 116.9, 94.7, 87.7, 83.9, 76.3, 72.9, 68.8, 67.6, 63.3, 61.4, 59.1, 56.9, 56.6, 53.7, 52.3, 49.9, 44.1, 40.6, 40.3, 40.1, 36.5, 33.1, 32.8, 31.5, 30.8, 30.8, 30.6, 30.5, 30.3, 30.0, 28.7, 28.1, 26.9, 26.1, 25.9, 23.8, 23.7, 21.4, 20.3, 19.6, 18.4, 15.2, 14.5, 13.4, 12.5, 10.5; HRMS (ESI) m/z : calcd. for $\text{C}_{68}\text{H}_{107}\text{N}_{12}\text{O}_{19}\text{S}$ $[\text{M}+\text{H}]^+$ 1427.7496, found

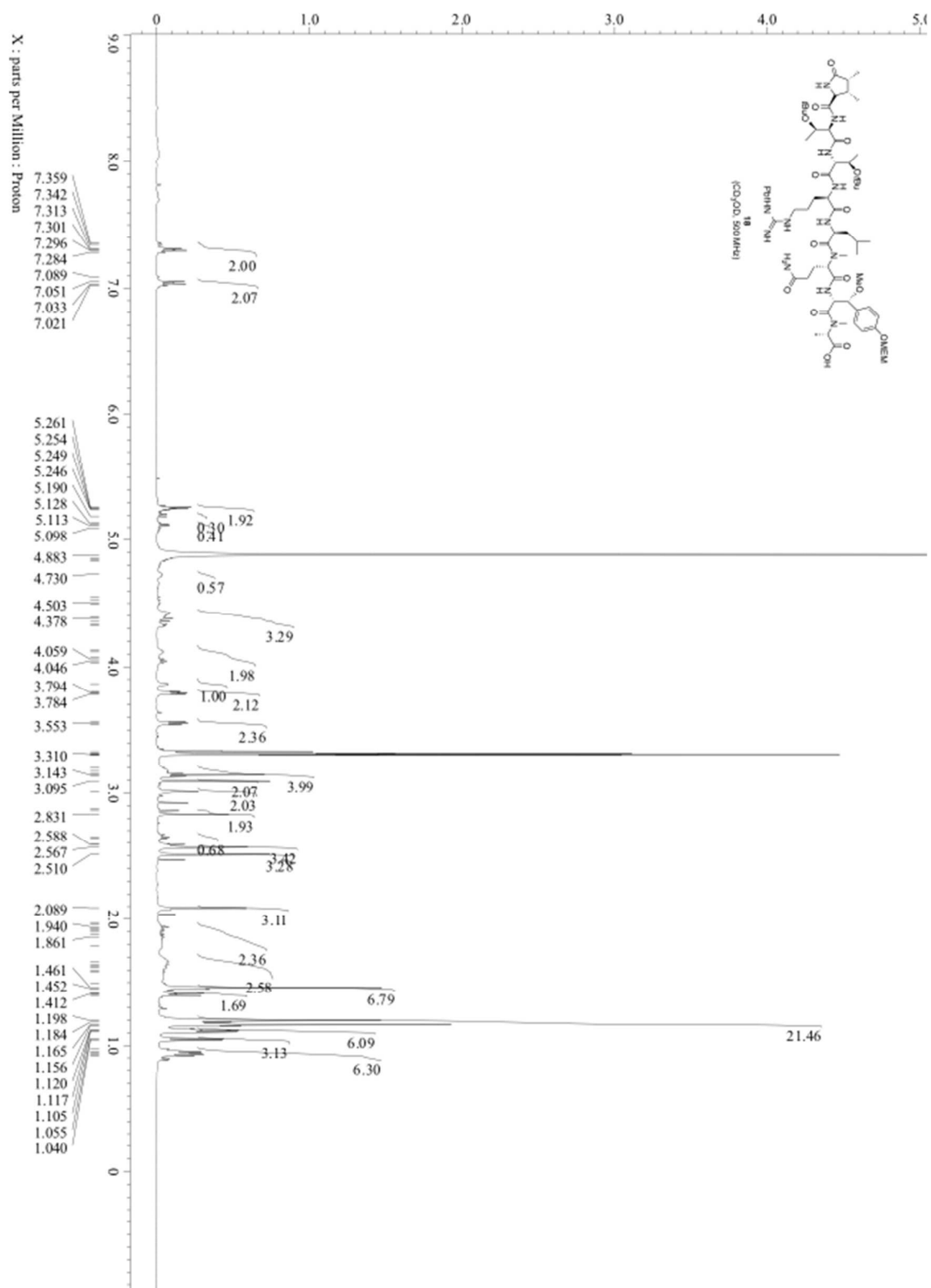
1427.7498; RP-HPLC: t_R = 22.4 min (linear gradient from 10% to 90% MeCN).

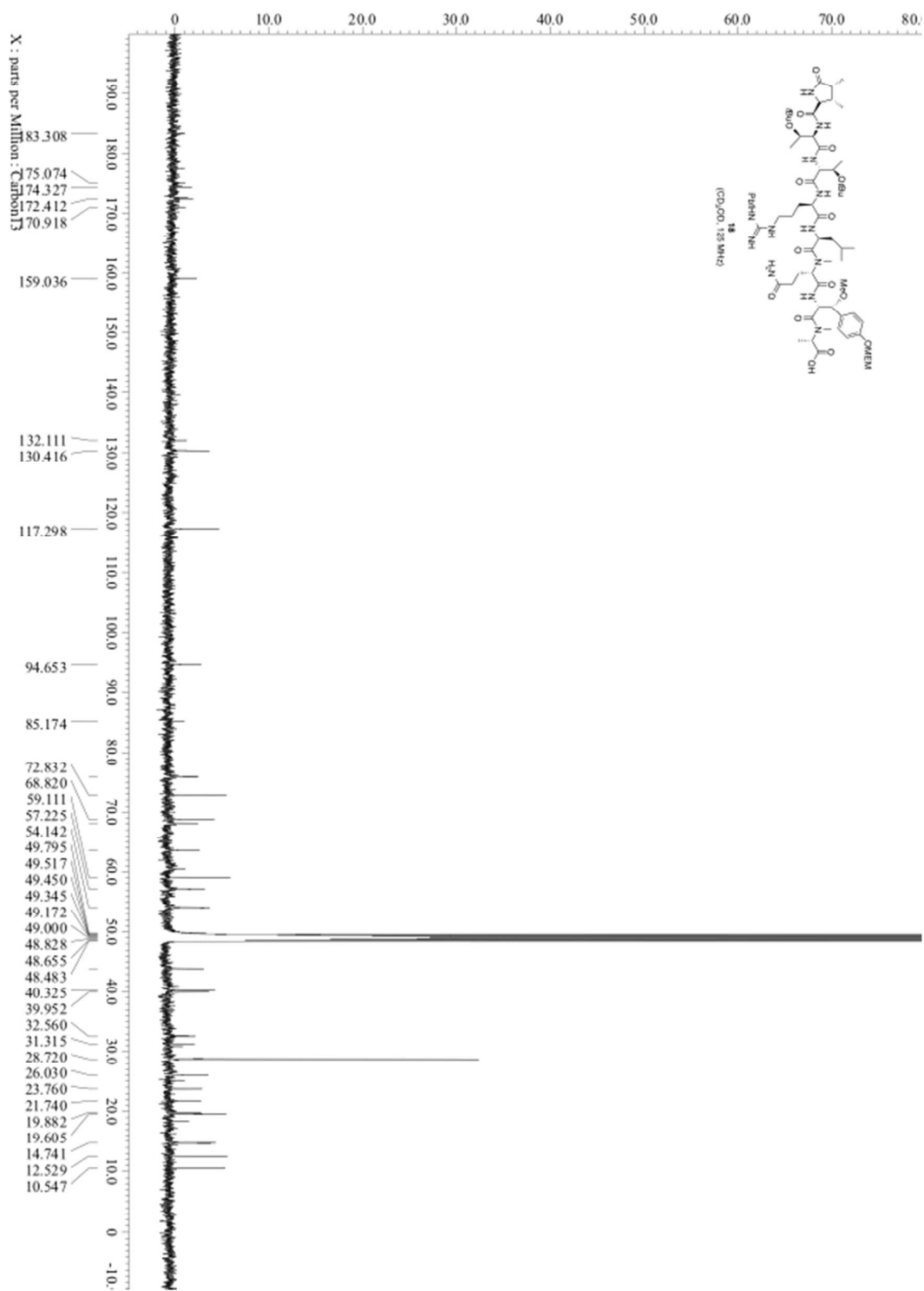
Callipeltin B (2)

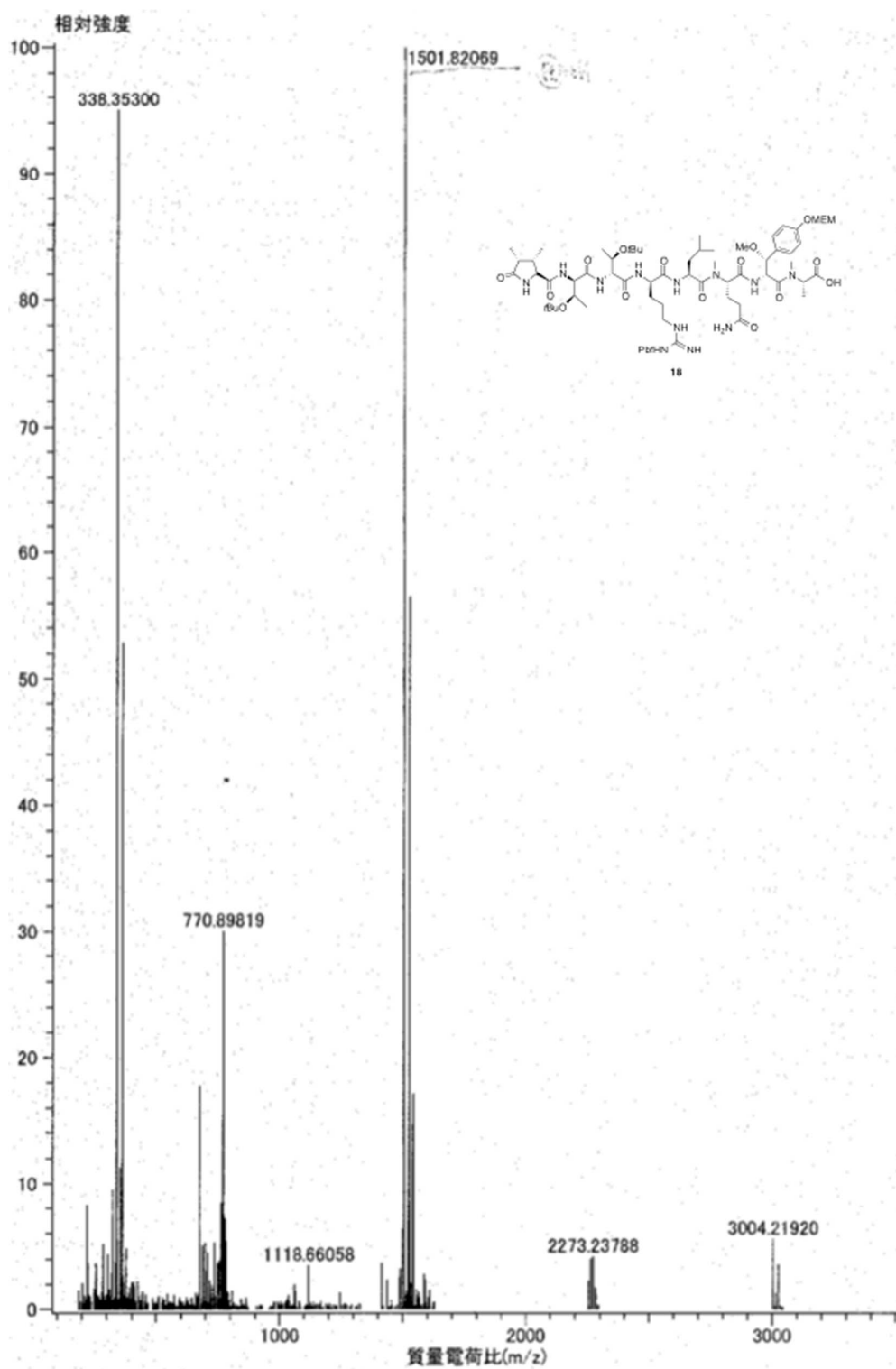


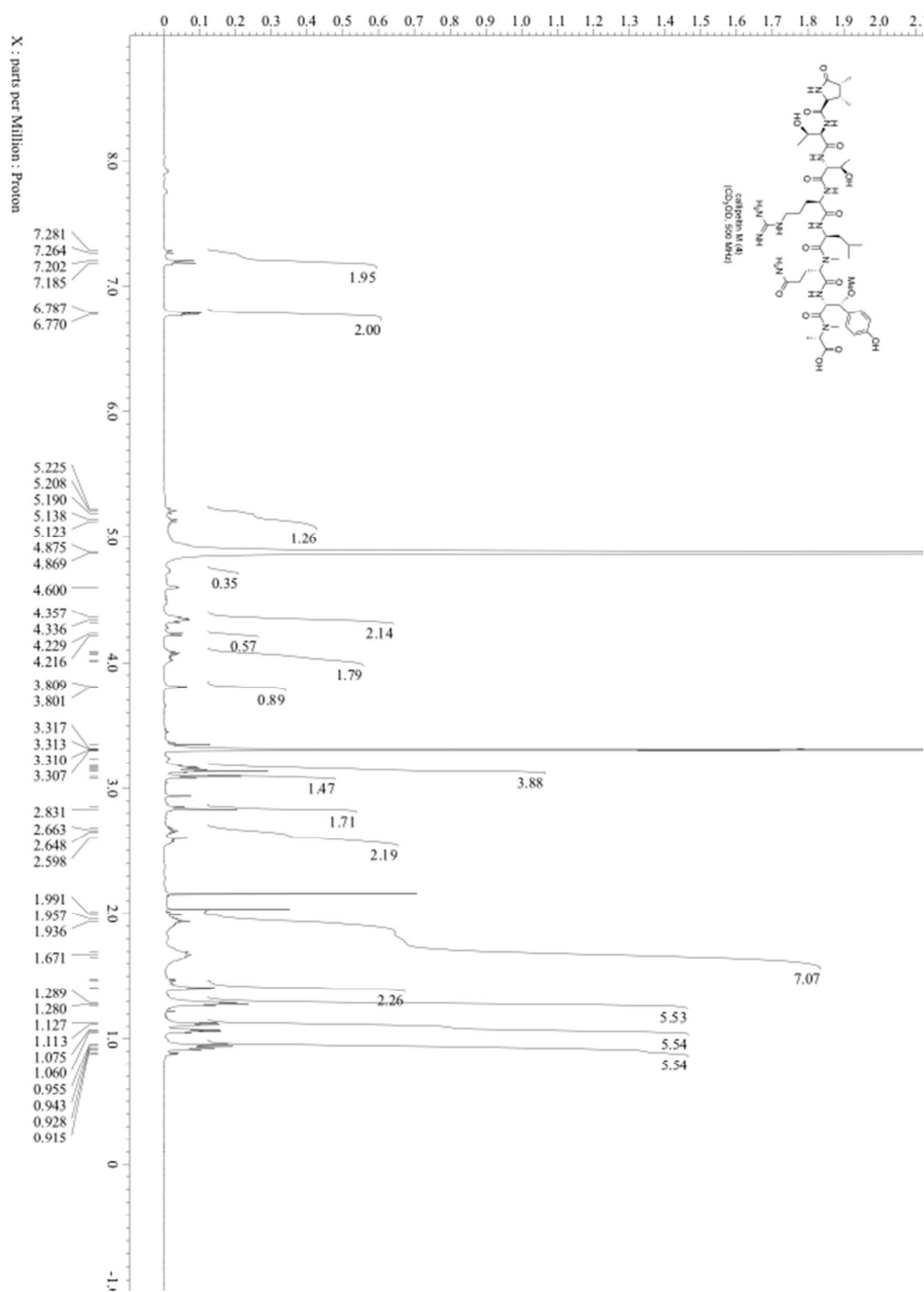
20 (0.5 mg, 350 nmol) was dissolved in 10% TIPS/TFA (50 μ L) and stirred for 1.5 h. To the mixture was added Et₂O and centrifuged to remove the supernatant. The crude product was purified with RP-HPLC (linear gradient from 20% to 60% MeCN) and lyophilized to afford callipeltin B (**2**) (0.3 mg, 290 nmol, 83%) as a white powder. ¹H NMR (800 MHz, CD₃OD) δ 8.61 (d, J = 8.0 Hz, 1H), 8.37 (d, J = 8.8 Hz, 1H), 7.40 (d, J = 7.2 Hz, 1H), 7.23 (d, J = 7.2 Hz, 2H), 6.75 (d, J = 7.2 Hz, 2H), 5.45 (m, 2H), 5.31 (q, J = 7.2 Hz, 1H), 4.92 (overlapped, 2H), 4.73 (m, 1H), 4.49 (d, J = 8.8 Hz, 1H), 4.15 (m, 1H), 4.09 (m, 1H), 4.07 (brs, 1H), 3.93 (brs, 1H), 3.17 (overlapped, 3H), 3.15 (s, 3H), 2.99 (s, 3H), 2.76 (m, 1H), 2.71 (s, 3H), 1.96-1.88 (m, 2H), 1.78-1.62 (m, 4H), 1.58-1.44 (m, 4H), 1.37 (m, 1H), 1.29 (d, J = 7.2 Hz, 3H), 1.26 (brs, 6H), 1.21 (d, J = 6.4 Hz, 3H), 1.18 (d, J = 7.2 Hz, 3H), 0.94 (d, J = 6.4 Hz, 3H), 0.92 (d, J = 5.6 Hz, 3H); ¹³C NMR (200 MHz, CD₃OD) δ 183.9, 183.3, 177.2, 176.0, 174.9, 173.3, 172.0, 171.9, 171.8, 170.9, 158.6, 131.3, 129.5, 115.9, 84.2, 72.7, 67.2, 63.5, 62.0, 56.9, 56.5, 54.2, 54.1, 53.9, 53.8, 52.5, 49.9, 42.1, 40.8, 40.1, 32.5, 31.0, 30.1, 27.8, 26.4, 26.0, 25.7, 23.6, 21.6, 20.3, 15.2, 14.5, 13.6, 10.5; HRMS (ESI) m/z : calcd. for C₄₇H₇₅N₁₂O₁₄ [M+H]⁺ 1031.5526, found 1031.5575; RP-HPLC: t_R = 13.0 min (linear gradient from

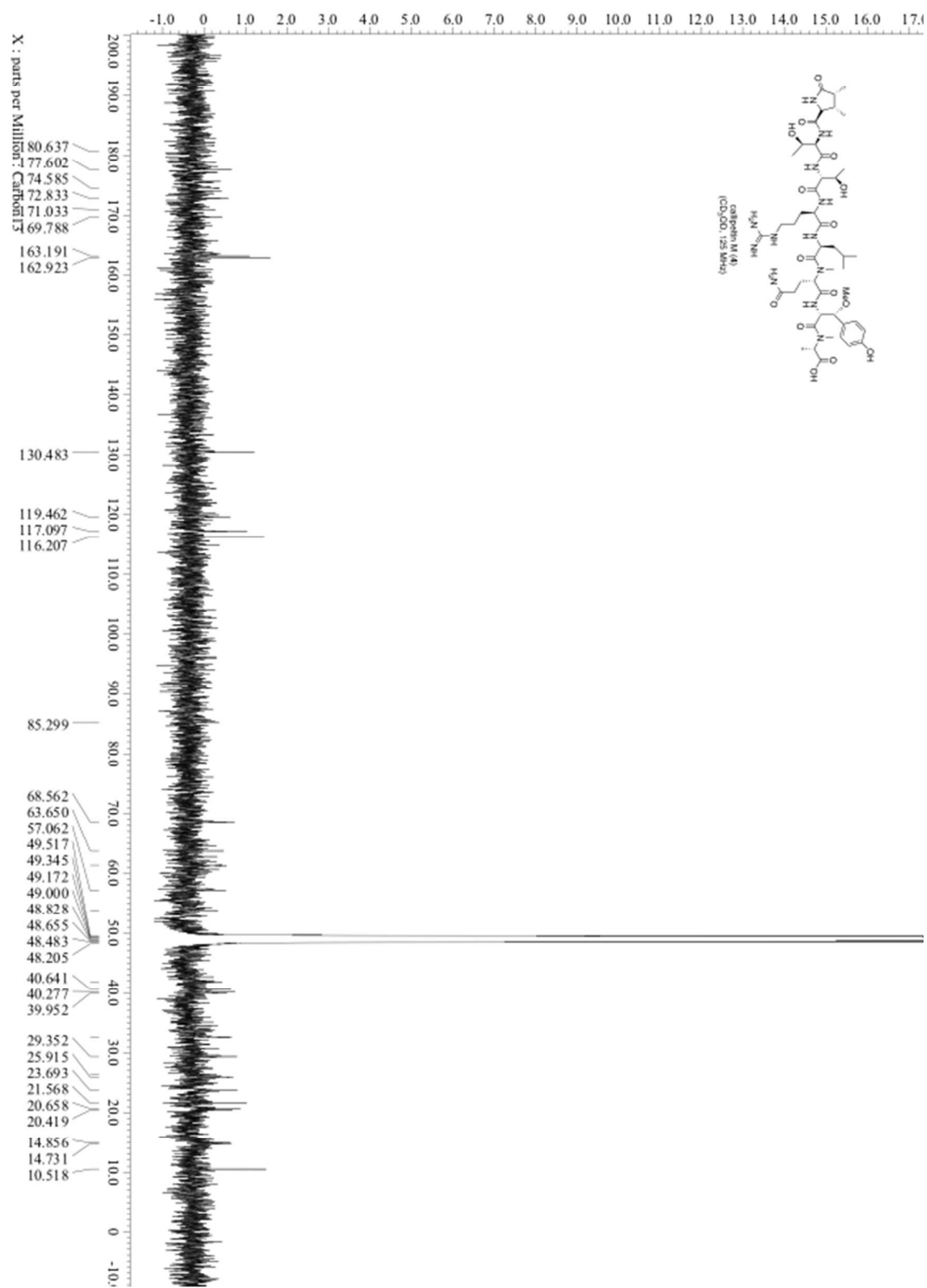
10% to 90% MeCN).

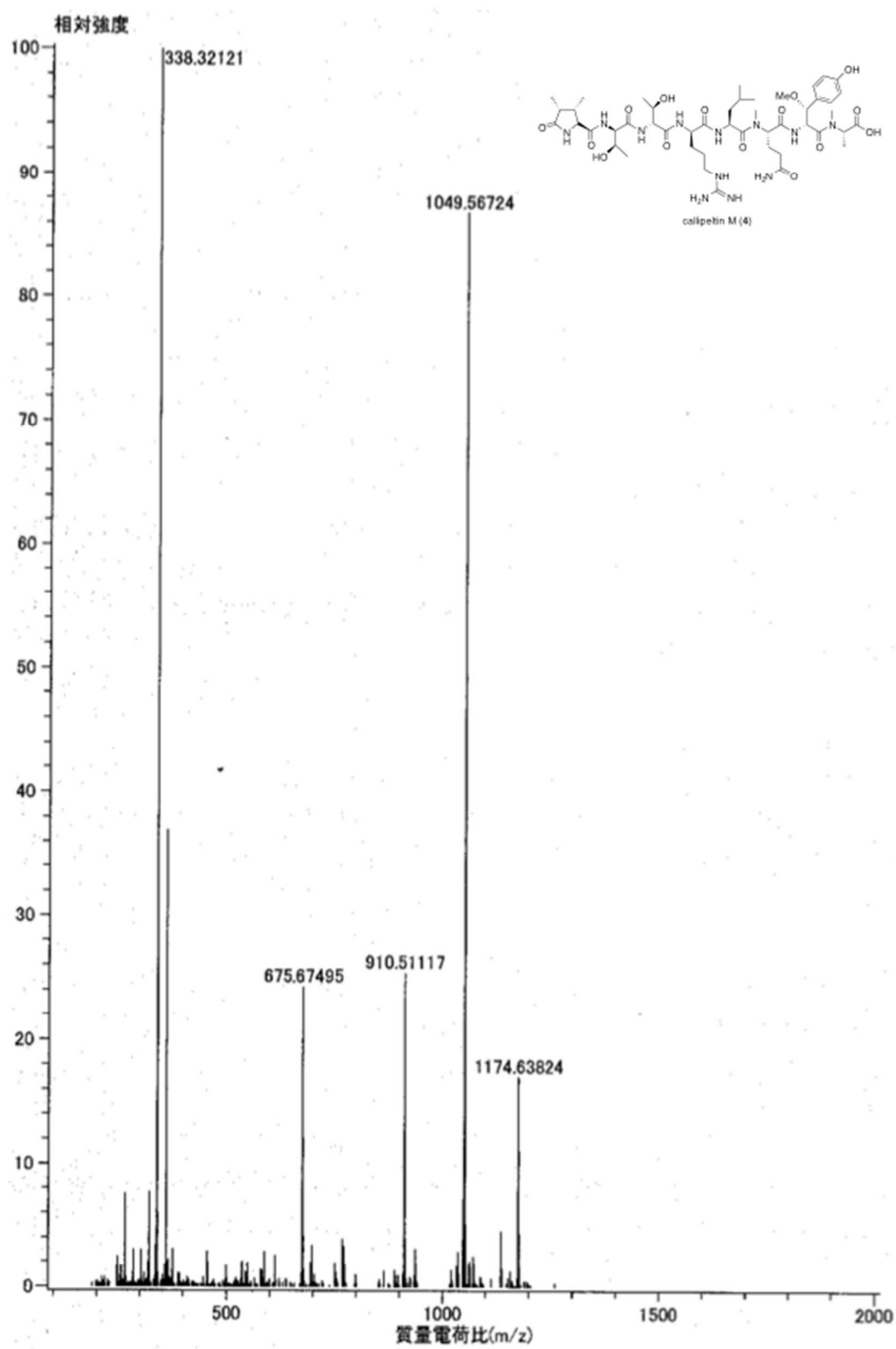


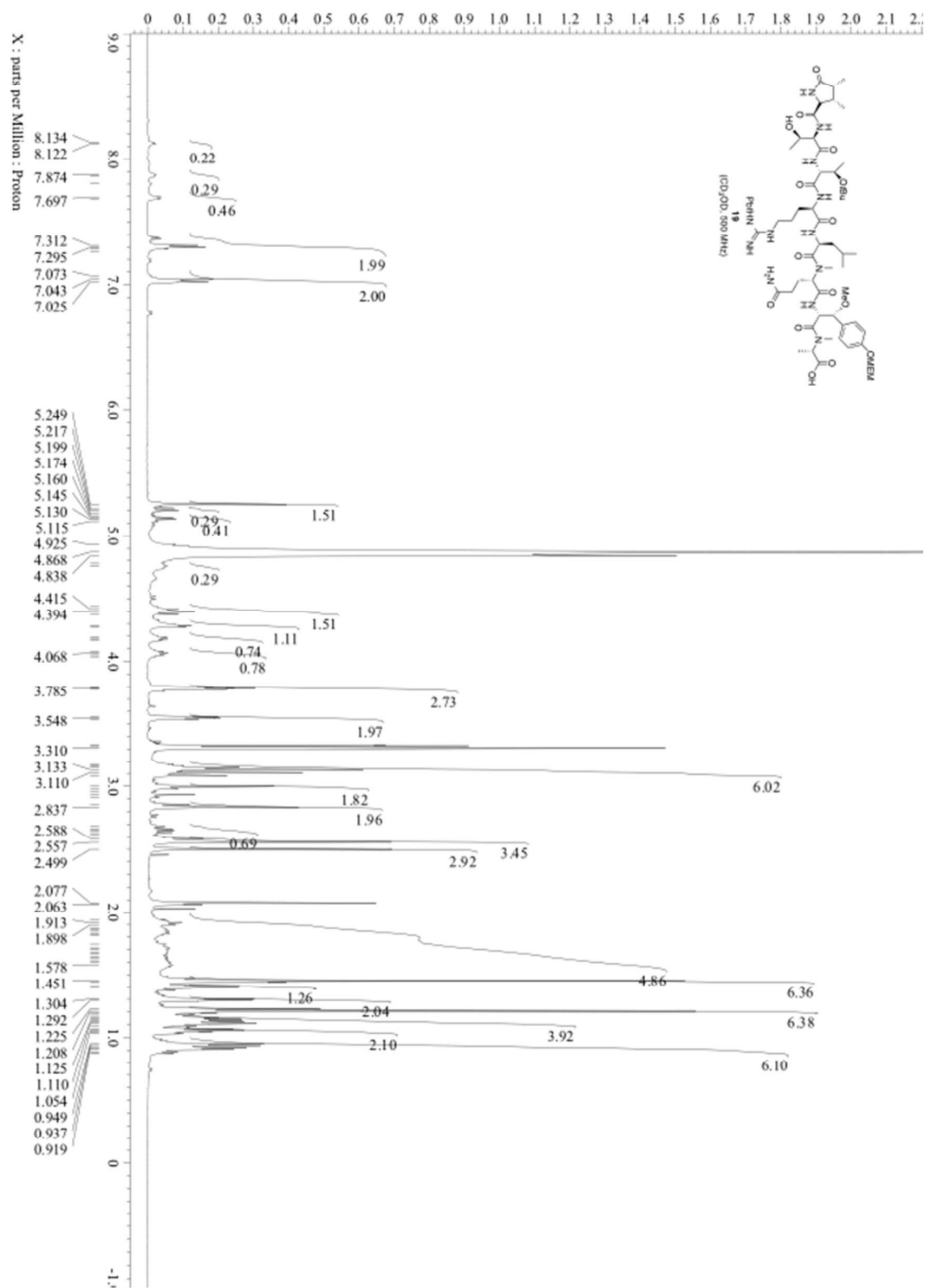


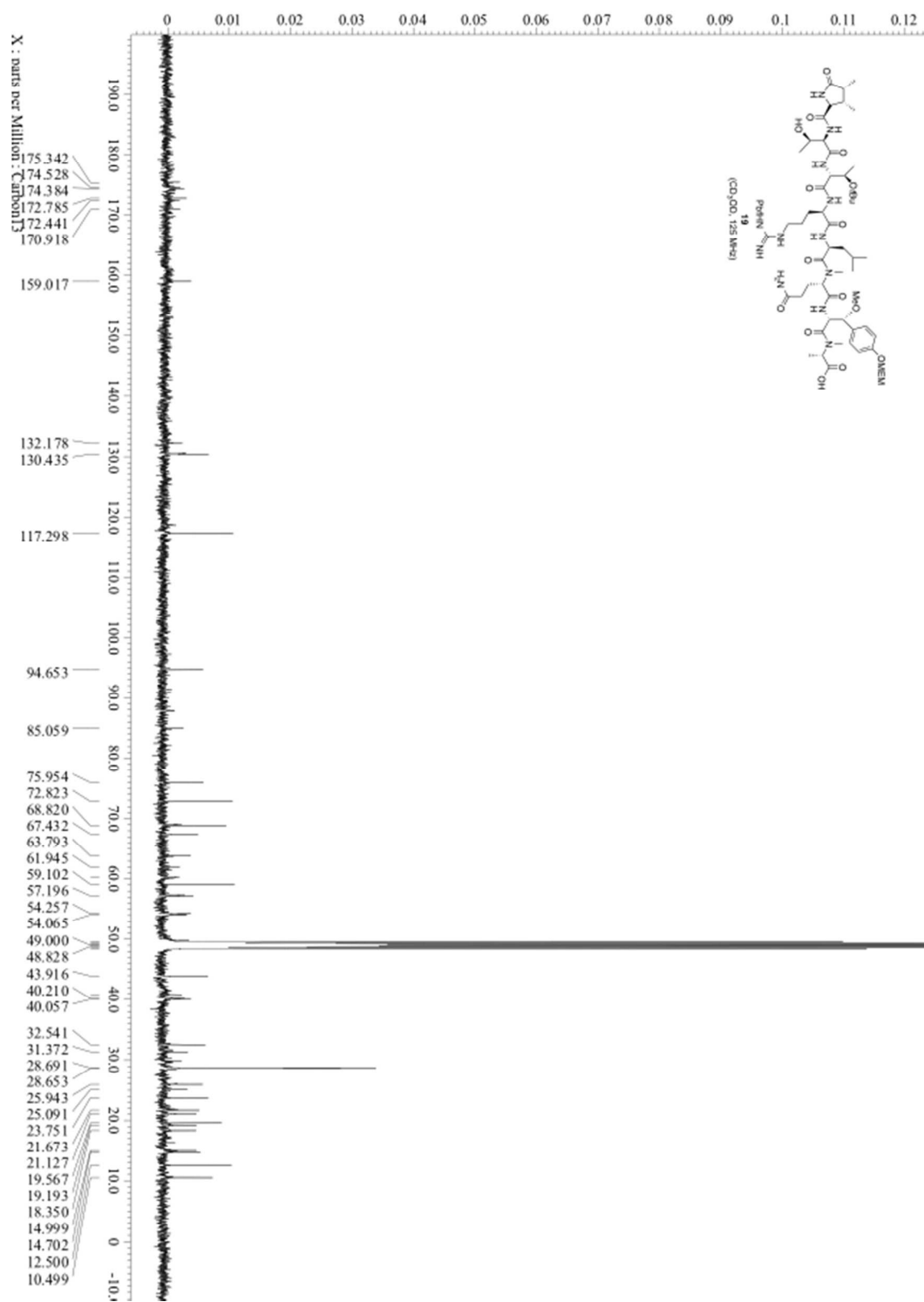


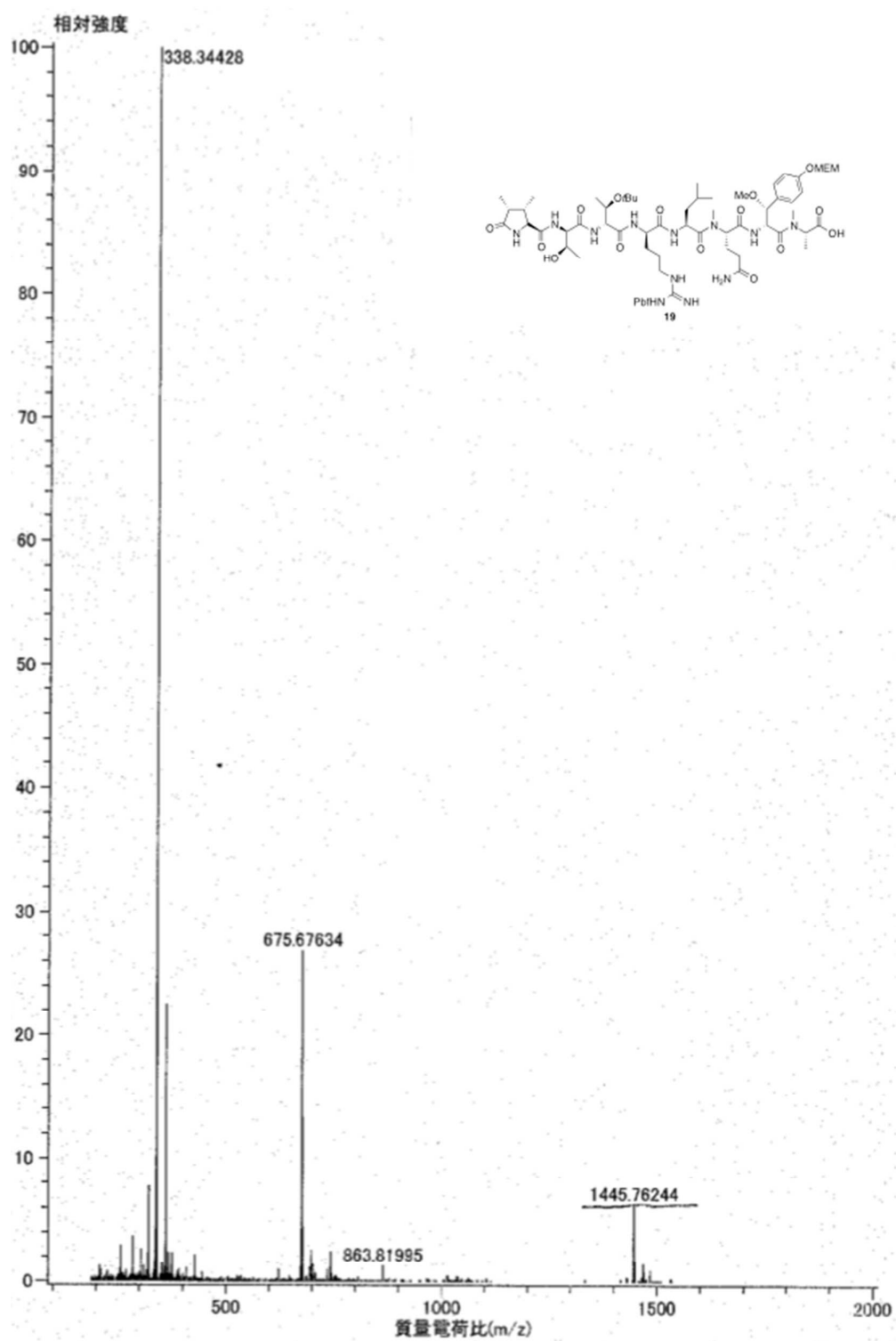


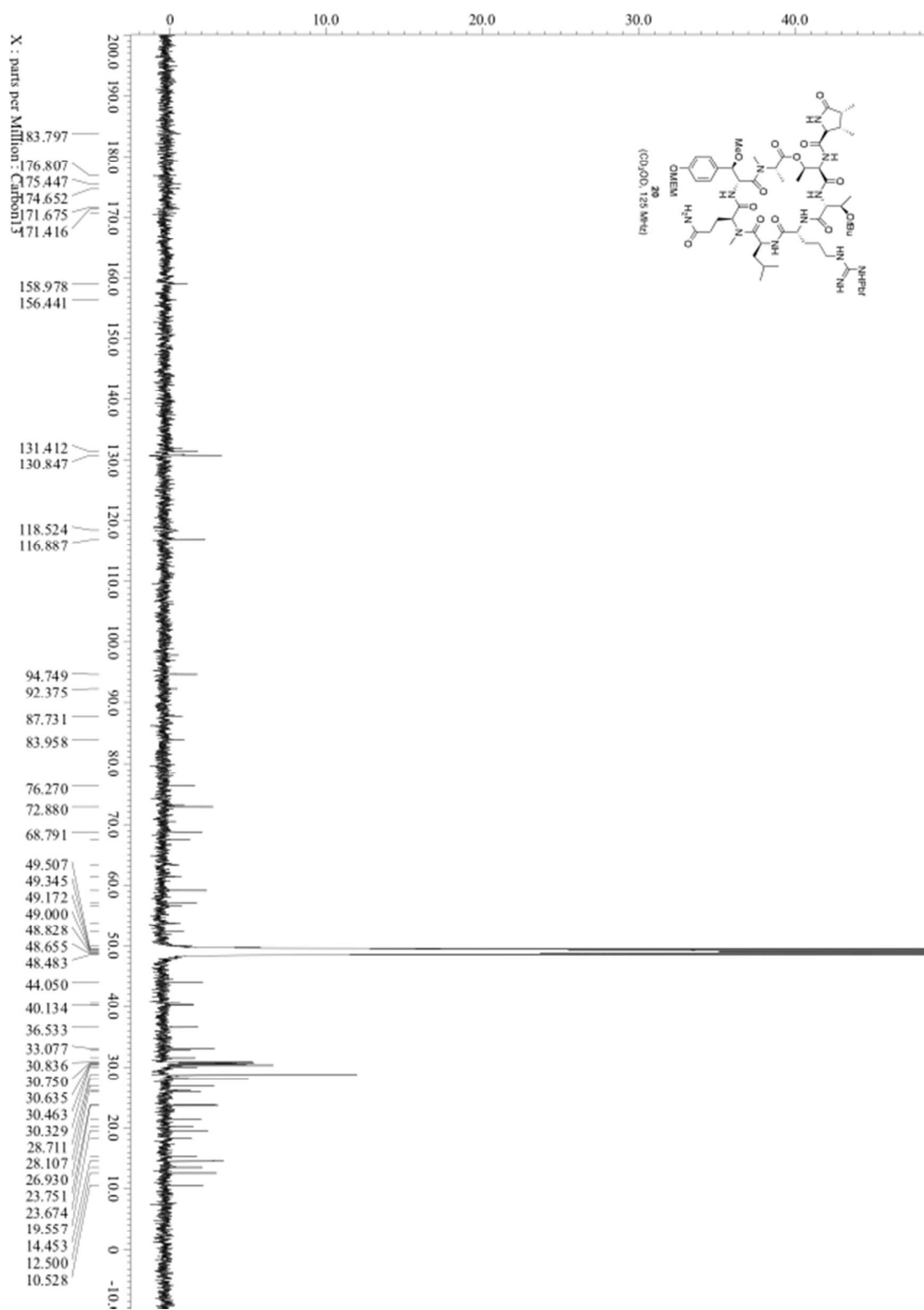


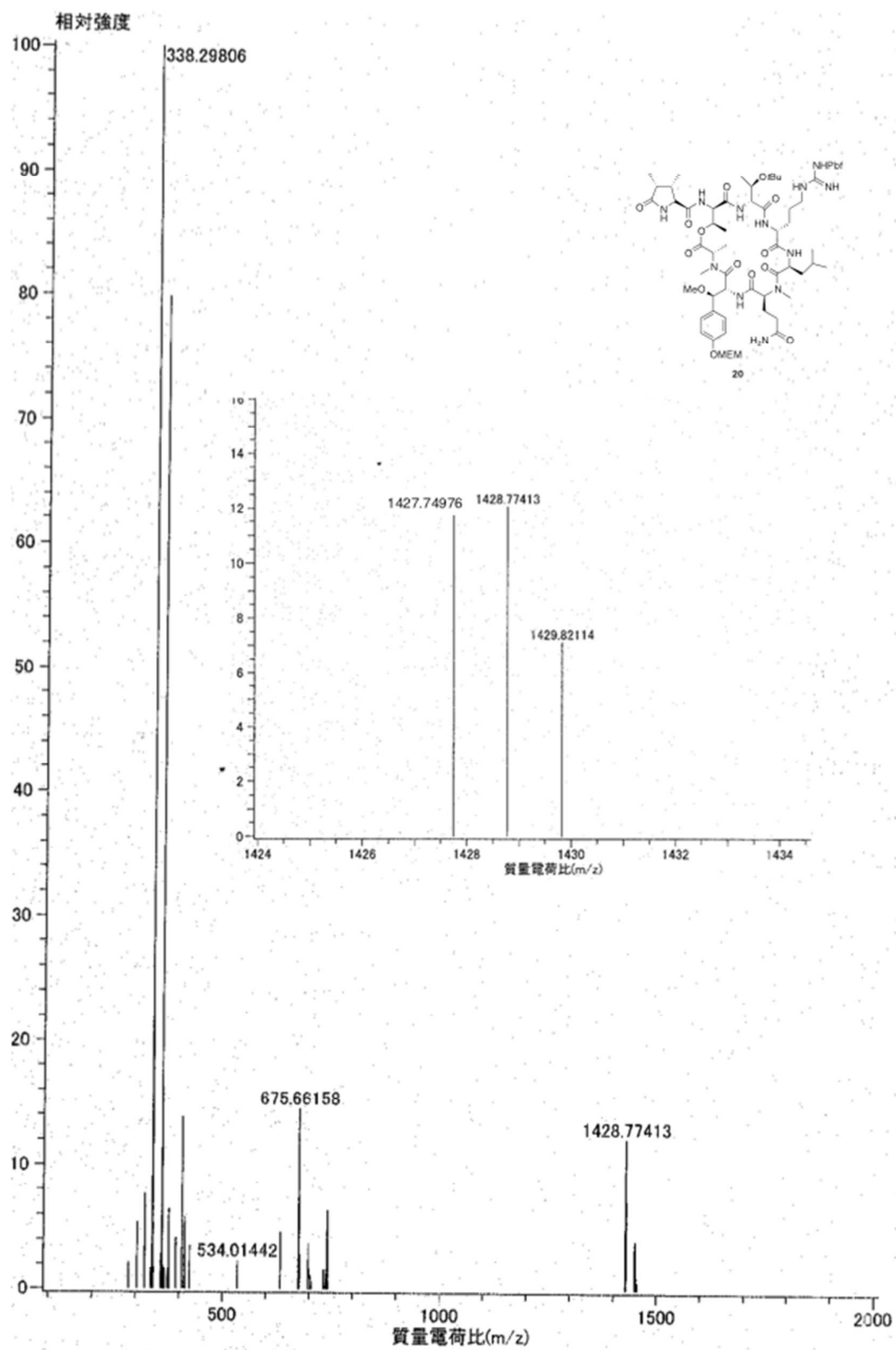


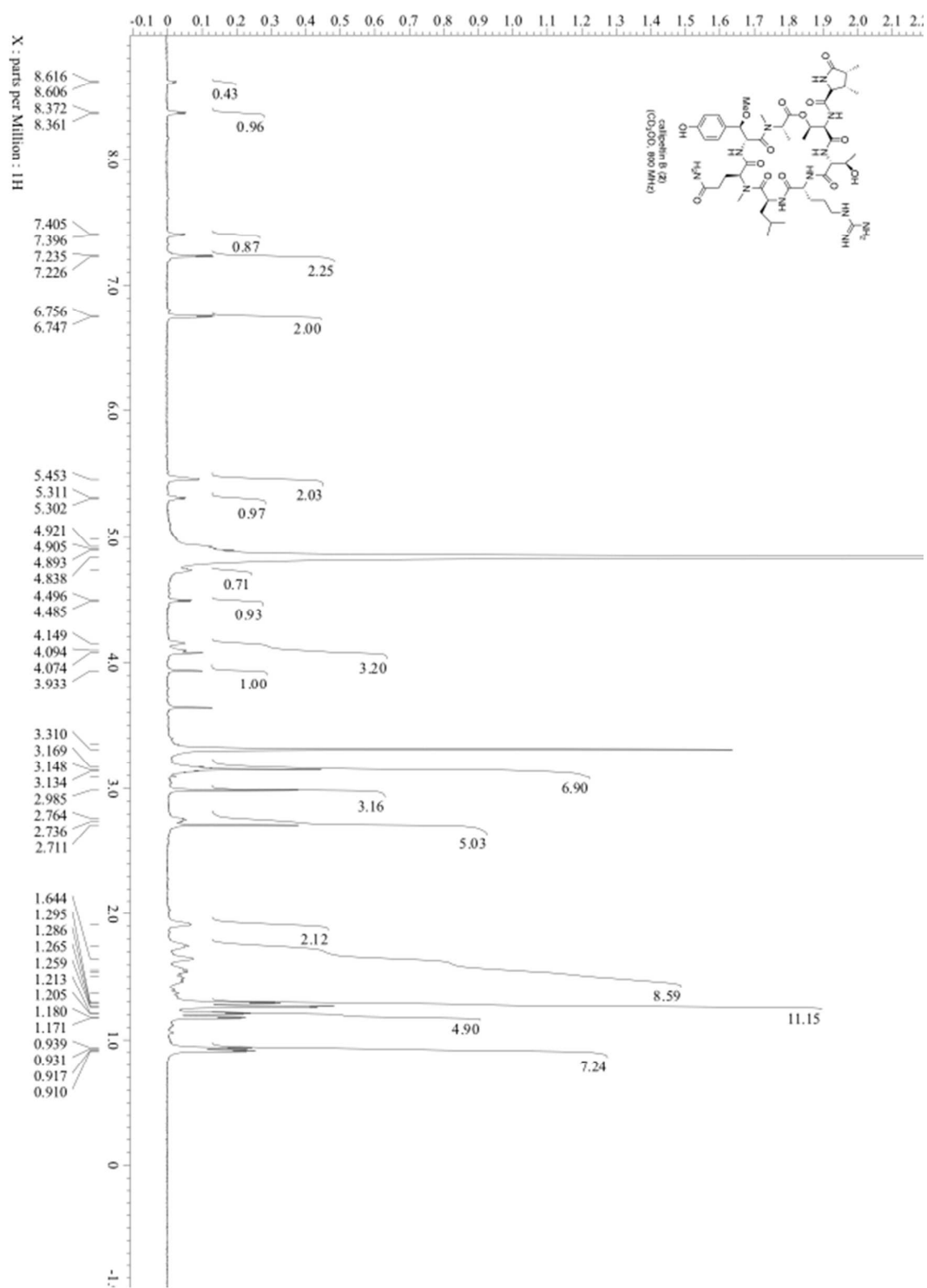


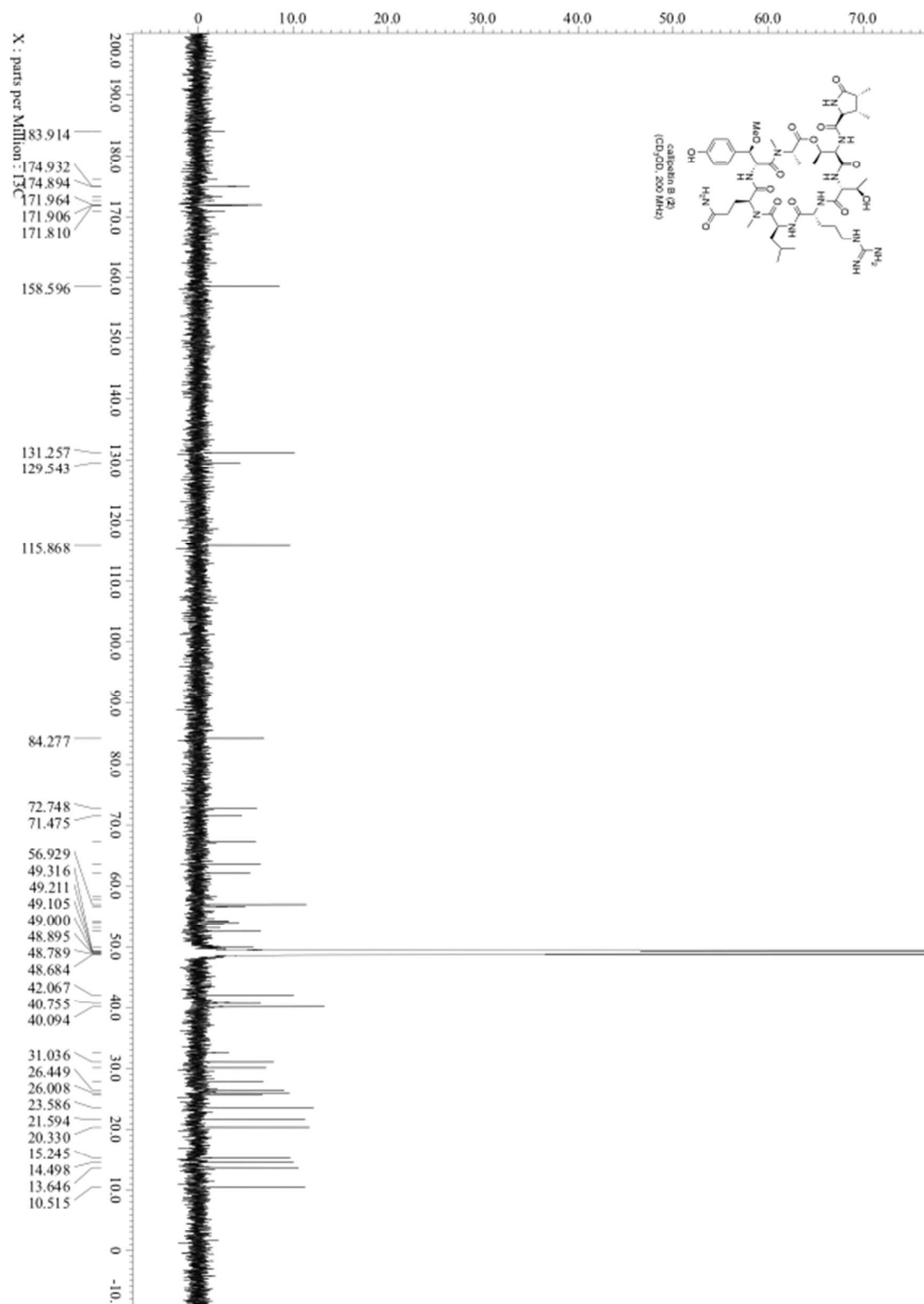












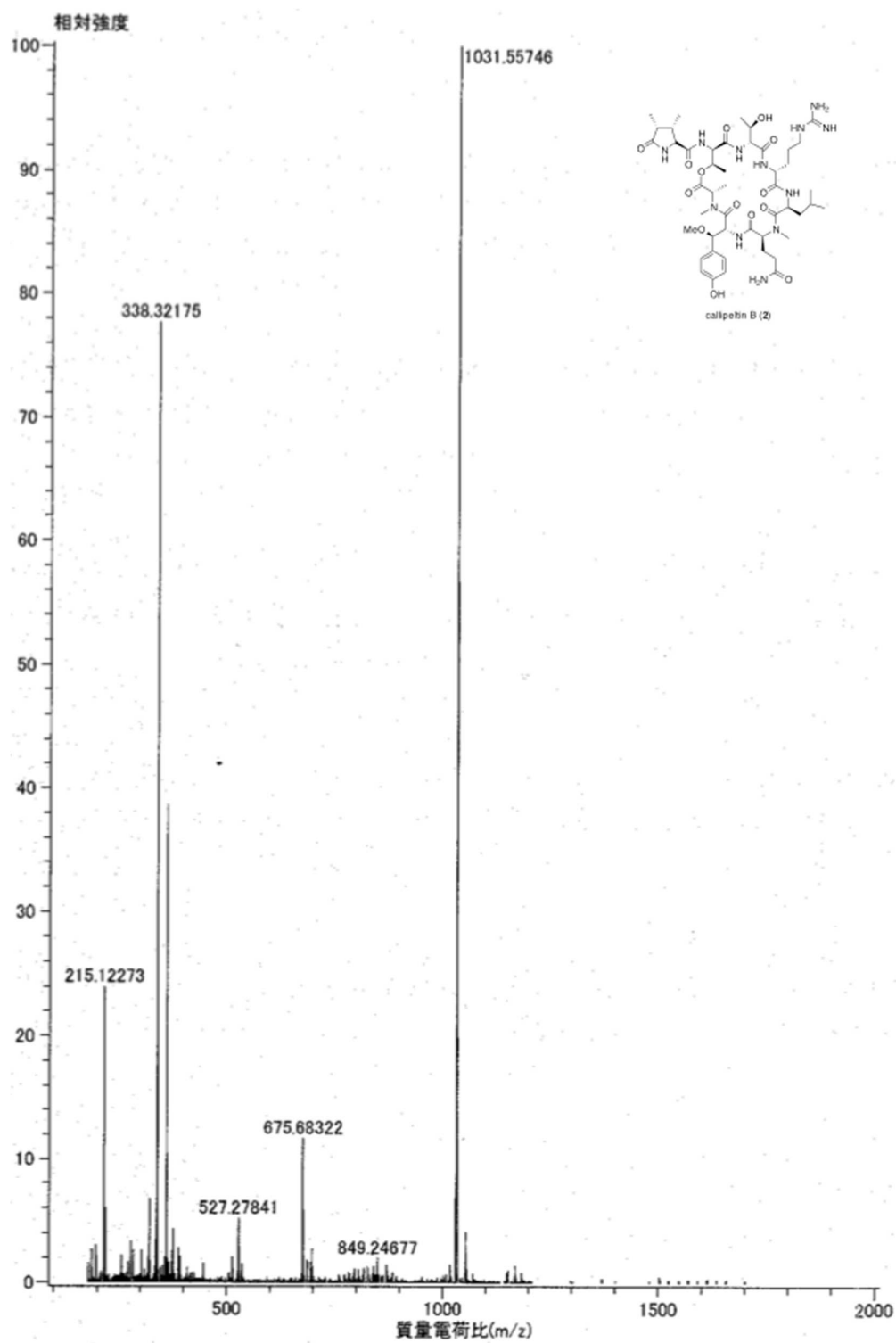


Table S1. NMR spectroscopic data for 4

synthetic 4		natural callipeltin M	
$\delta_{\text{H}}^{\text{a}}$	$\delta_{\text{C}}^{\text{b}}$	$\delta_{\text{H}}^{\text{a}}$	$\delta_{\text{C}}^{\text{b}}$
0.92 (d, $J = 6.5$ Hz, 3H)	10.5	0.94 (d, $J = 7.4$ Hz, 3H)	10.0
0.95 (d, $J = 6.0$ Hz, 3H)	14.7	0.94 (d, $J = 7.4$ Hz, 3H)	14.7
1.07 (d, $J = 7.5$ Hz, 3H)	14.9	1.07 (d, $J = 7.3$ Hz, 3H)	14.7
1.12 (d, $J = 7.0$ Hz, 3H)	20.4	1.12 (d, $J = 7.2$ Hz, 3H)	20.1
1.28 (m, 6H)	20.7	1.26 (d, overlapped, 3H)	20.1
	21.6	1.26 (d, overlapped, 3H)	21.6
1.40 (d, $J = 7.5$ Hz, 3H)	23.7	1.42 (d, $J = 6.5$ Hz, 3H)	23.9
1.47 (m, 1H)	24.9	1.40, 1.70 (m, 2H)	24.6
1.61-1.72 (m, 7H)	25.9	1.67 (m, 2H)	25.7
1.88-1.99 (m, 3H)	26.2	1.68, 1.93 (m, 2H)	26.2
	29.4	1.70 (m, 1H)	29.5
	30.7	1.70, 1.94 (m, 2H)	31.3
	32.5	1.94 (m, 2H)	32.4
2.57 (m, 1H)	32.6	2.59 (m, 1H)	32.6
2.66 (m, 1H)	39.9	2.68 (m, 1H)	39.7
2.83 (s, 3H)	40.3	2.80 (s, 3H)	39.7
3.10 (s, 3H)	40.6	3.10 (s, 3H)	40.6
3.13 (s, 3H)	41.8		42.2
3.16 (m, 2H)	48.2	3.18 (m, 2H)	48.9
3.81 (d, $J = 4.0$ Hz, 1H)	53.8	3.80 (d, $J = 3.3$ Hz, 1H)	53.9
4.01-4.09 (m, 2H)		4.02 (m, 1H)	53.9
	54.2	4.08 (m, 1H)	54.1
4.22 (d, $J = 6.5$ Hz, 1H)	57.1	4.22 (d, $J = 7.0$ Hz, 1H)	56.9
4.32-4.36 (m, 3H)	57.4	4.32 (d, $J = 8.1$ Hz, 1H)	
	61.2	4.33 (d, $J = 6.7$ Hz, 1H)	60.4
	61.3	4.35 (overlapped, 1H)	60.4
4.73 (m, 1H)	63.9	4.76 (m, 1H)	63.4
4.88 (overlapped, 1H)	68.5	4.90 (dd, $J = 10.1, 4.7$ Hz, 1H)	68.0
5.13 (q, $J = 7.5$ Hz, 1H)	68.6	5.14 (q, $J = 6.5$ Hz, 1H)	68.0
5.21 (t, $J = 9.0$ Hz, 1H)	85.3	5.23 (d, $J = 8.1$ Hz, 1H)	85.0
6.78 (d, $J = 8.5$ Hz, 2H)	116.2	6.79 (d, $J = 8.1$ Hz, 2H)	116.1
7.19 (d, $J = 8.5$ Hz, 2H)	130.5	7.21 (d, $J = 8.1$ Hz, 2H)	128.7
			129.6
	162.9		158.5
	163.2		
	169.8		169.7
	171.0		171.6
	172.8		172.1
	172.8		172.1
	173.6		173.3
	174.0		173.8
	174.6		174.5
	177.6		
	180.6		182.0

^aRecorded in CD₃OD at 500 MHz. ^bRecorded in CD₃OD at 125 MHz.

Table S2. NMR spectroscopic data for 2

synthetic 2		natural callipeltin B	
$\delta_{\text{H}}^{\text{a}}$	$\delta_{\text{C}}^{\text{b}}$	$\delta_{\text{H}}^{\text{a}}$	$\delta_{\text{C}}^{\text{b}}$
0.91 (d, $J = 5.6$ Hz, 3H)	10.5	0.92 (d, $J = 7.2$ Hz, 3H)	10.5
0.94 (d, $J = 6.4$ Hz, 3H)	13.6	0.94 (d, $J = 6.4$ Hz, 3H)	13.7
1.18 (d, $J = 7.2$ Hz, 3H)	14.5	1.18 (d, $J = 8.0$ Hz, 3H)	14.5
1.21 (d, $J = 6.4$ Hz, 3H)	15.2	1.21 (d, $J = 7.2$ Hz, 3H)	15.2
1.26 (brs, 6H)	20.3	1.26 (t, $J = 5.6$ Hz, 6H)	20.3
1.29 (d, $J = 7.2$ Hz, 3H)	21.6	1.30 (d, $J = 7.0$ Hz, 3H)	21.6
1.37 (m, 1H)	23.6	1.39 (m, 1H)	23.6
1.44-1.58 (m, 4H)	25.7	1.45-1.58 (m, 4H)	25.8
1.62-1.78 (m, 4H)	26.0	1.62-1.77 (m, 4H)	26.0
1.88-1.96 (m, 2H)	26.4	1.90-1.94 (m, 2H)	26.4
2.71 (s, 3H)	27.8	2.72 (s, 3H)	27.8
2.76 (m, 1H)	30.1	2.77 (m, 1H)	30.1
2.99 (s, 3H)	31.0	2.99 (s, 3H)	31.0
3.15 (s, 3H)	32.5	3.15 (s, 3H)	32.5
3.17 (overlapped, 3H)	40.1	3.17 (m, 3H)	40.1
3.93 (brs, 1H)		3.95 (d, $J = 3.2$ Hz, 1H)	40.1
4.07 (brs, 1H)	40.8	4.07 (d, $J = 5.6$ Hz, 1H)	40.7
4.09 (m, 1H)	42.1	4.10 (m, 1H)	42.1
4.15 (m, 1H)	49.9	4.16 (m, 1H)	49.9
4.49 (d, $J = 8.8$ Hz, 1H)	52.5	4.49 (d, $J = 10.4$, 1H)	52.5
4.73 (m, 1H)	53.8	4.73 (dd, $J = 11.2, 3.2$ Hz, 1H)	53.8
4.92 (overlapped, 2H)	53.9	4.91 (overlapped, 2H)	
5.31 (q, $J = 7.2$ Hz, 1H)	54.1	5.31 (q, $J = 7.2$ Hz, 1H)	54.1
5.45 (m, 2H)	54.2	5.46-5.49 (m, 2H)	
6.75 (d, $J = 7.2$ Hz, 2H)	56.5	6.76 (d, $J = 8.0$ Hz, 2H)	56.5
7.23 (d, $J = 7.2$ Hz, 2H)	56.9	7.23 (d, $J = 8.0$ Hz, 2H)	56.9
7.40 (d, $J = 7.2$ Hz, 1H)	62.0		62.1
8.37 (d, $J = 8.8$ Hz, 1H)	63.5		63.5
8.61 (d, $J = 8.0$ Hz, 1H)	67.2		67.2
	72.7		72.8
	84.2		84.2
	115.9		115.9
	129.5		129.5
	131.3		131.3
	158.6		158.6
	170.9		170.9
	171.8		171.8
	171.9		
	172.0		172.0
	173.3		173.4
	174.9		174.9
	176.0		175.9
	177.2		177.2
	183.3		183.3
	183.9		183.9

^aRecorded in CD₃OD at 800 MHz. ^bRecorded in CD₃OD at 200 MHz.