An Efficient and Highly Diastereoselective Synthesis of GSK1265744, a Potent HIV Integrase Inhibitor

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Supporting information

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Experimental Details

General. Unless otherwise indicated, solvents and reagents were obtained from commercial sources and used without further purification. All reactions were conducted under a nitrogen atmosphere. All reactions were not optimized. Silica gel chromatographic purifications were performed using RediSep® silica gel-packed columns on a Teledyne Isco CombiFlash® system. Low resolution mass spectra were recorded on an Agilent 1100 series (SL) LC/MSD instrument. ¹H NMR, ¹³C and ¹⁹F NMR spectra were recorded at 25 °C on a Bruker AVANCE 300 system at 300, 75 and 282 MHz respectively. Chemical shifts are reported as δ in ppm relative to chloroform-*d*. Elemental analyses were performed by Intertek QTI Laboratory in Whitehouse, New Jersey. High resolution mass spectra (HRLC-MS) were recorded using a Waters Q-TOF premier quadrupole orthogonal acceleration time-of-flight mass spectrometer coupled to an Agilent 1100 LC system. High performance liquid chromatographic (HPLC) analyses were carried out using an Agilent 1100 series HPLC instrument.

Preparation of (E)-methyl 2-((benzylamino)methylene)-3-oxobutanoate (6a): Methyl 2-(methoxymethylene)-3-oxobutanoate¹ (826 mg, 5.22 mmol) was dissolved in CH₂Cl₂ (8 mL) and benzylamine (630 µL, 5.76 mmol, 1.1 equiv) was added. The mixture was stirred at rt overnight, concentrated and the product was purified by chromatography (eluted with 10%-40% EtOAc in hexanes, 1.106 g, 4.74 mmol, 91%). The product was a pale yellow solid. ES-MS: m/z 234 (M+H⁺). ¹H NMR showed two isomers. The minor isomer was estimated to be ~4% from doublets at 8.10 ppm (major isomer) and 8.27 ppm (minor isomer) as well as singlets at 3.74 ppm (major) and 3.80 ppm (minor). The double bond geometry is inconsequential in the subsequent cyclization as the two isomers should be readily interconverting via an imineenamine equilibrium, but the major isomer was assigned the (E)-configuration based on the assumption of intramolecular H bond between the ketone and the N-H.² The ¹H NMR data are also in agreement with reported values.³ The major isomer: ¹H NMR (CDCl₃, 300 MHz) δ 11.31 (br. s., 1 H), 8.10 (d, J = 13.6 Hz, 1 H), 7.42-7.25 (m, 5 H), 4.53 (d, J = 6.1 Hz, 2 H), 3.74 (s, 3 H), 2.50 (s, 3 H). ¹³C NMR (CDCl₃, 75 MHz) δ 199.1, 167.3, 159.8, 136.1, 128.7, 127.9, 127.0, 100.0, 53.4, 50.6, 30.6. Anal calcd for C₁₃H₁₅NO₃: C, 66.94; H, 6.48; N, 6.00; found: C, 66.81; H, 6.51; N, 5.85.

¹ Crombie, L.; Games, D. E.; James, A. W. G. J. Chem. Soc., Perkin Trans. 1, 1979, 464.

² Michalik, M. J. Prakt. Chem **1985**, 327, 908.

³ Ganem, B. J. Am. Chem. Soc. **1976**, 98, 224.

Preparation of (E)-methyl 3-oxo-2-((phenylamino)methylene)butanoate (6b): Methyl 2-(methoxymethylene)-3-oxobutanoate (405 mg, 2.56 mmol) was dissolved in CH₂Cl₂ (4 mL) and aniline (260 μL, 2.85 mmol, 1.1 equiv) was added. The mixture was stirred at rt overnight, concentrated and the product was purified by chromatography (eluted with 10%-30% EtOAc in hexanes, 558.2 mg, 2.55 mmol, 99%). The product was a pale yellow solid. ES-MS: m/z 220 (M+H⁺). ¹H NMR (CDCl₃, 300 MHz) δ 8.52 (d, J = 13.2 Hz, 1 H), 7.43-7.37 (m, 2 H), 7.23-7.18 (m, 3 H), 3.79 (s, 3 H), 2.57 (s, 3 H). ¹³C NMR (CDCl₃, 75 MHz) δ 200.0, 167.1, 151.8, 138.8, 129.7, 125.4, 117.6, 102.1, 51.0, 31.0. Anal calcd for C₁₂H₁₃NO₃: C, 65.74; H, 5.98; N, 6.39; found: C, 65.57; H, 5.90; N, 6.34. The ¹H NMR data are in agreement with reported values.³

Preparation of (E)-methyl 2-((isopropylamino)methylene)-3-oxobutanoate (6c): Methyl 2-(methoxymethylene)-3-oxobutanoate (1.098 g, 6.94 mmol) was dissolved in CH₂Cl₂ (10 mL) and isopropylamine (650 μL, 7.63 mmol, 1.1 equiv) was added. The mixture was stirred at rt overnight, concentrated and the product was purified by chromatography (eluted with 15%-30% EtOAc in hexanes, 1.216 g, 6.56 mmol, 94%). The product was a clear oil but solidified into a white solid. ES-MS: m/z 186 (M+H⁺). ¹H NMR showed two isomers. The minor isomer was estimated to be ~10% from doublets at 8.02 ppm (major isomer) and 8.20 ppm (minor isomer) as well as singlets at 3.71 ppm (major) and 3.77 ppm (minor). The major isomer was assigned the (E)-configuration based on the assumption of intramolecular H bond between the ketone and the N-H. Major isomer: ¹H NMR (CDCl₃, 300 MHz) δ 11.04 (br. s., 1 H), 8.02 (d, J = 13.8 Hz, 1 H), 3.71 (s, 3 H), 3.64-3.51 (m, 1 H), 2.45 (s, 3 H), 1.29 (d, J = 6.6 Hz, 6 H). ¹³C NMR (CDCl₃, 75 MHz) δ 198.7, 167.3, 157.5, 99.0, 51.2, 50.3, 30.4, 23.0. HRMS (ES) calcd for C₉H₁₅NO₃ [M+H]⁺: 186.1130, found 186.1135.

Preparation of (Z)-methyl 2-((dimethylamino)methylene)-4-methoxy-3-oxobutanoate (**4b**): Methyl 4-methoxy-3-oxobutanoate (**8**) (3.9 mL, 30.2 mmol, 1 equiv) was dissolved in 20 mL of toluene and DMF DMA (6.4 mL, 48.2 mmol, 1.6 equiv) was added. The mixture was heated to reflux for 4 h, and concentrated under vacuum. The product was purified by Kugelrohr distillation to give a light orange oil (5.5 g, 27.3 mmol, 90%). ES-MS: m/z 202 (M+H⁺). ¹H NMR (CDCl₃, 300 MHz) δ 7.75 (s, 1 H), 4.38 (s, 2 H), 3.73 (s, 3 H), 3.40 (s, 3 H), 3.26 (br. s., 3

H), 2.88 (br. s., 3 H). 13 C NMR (CDCl₃, 75 MHz) δ 192.7, 166.7, 157.3, 98.2, 76.3, 57.9, 49.9, 46.8, 41.4. HRMS (ES) calcd for $C_9H_{15}NO_4$ [M+H] $^+$: 202.1079, found 202.1085.

Preparation of (E)-methyl 2-(((2,2-dimethoxyethyl)amino)methylene)-4-methoxy-3-oxobutanoate (6d): The tertiary vinylogous amide 4b (292.6 mg, 1.45 mmol) was dissolved in 3 mL of CH₂Cl₂, and aminoacetaldehyde dimethyl acetal (170 μL, 1.58 mmol, 1.1 equiv) was added. The mixture was stirred at rt overnight, concentrated and the product was purified by chromatography (eluted with 0-60% EtOAc in hexanes, 371.6 mmol, 1.42 mmol, 98%). The product was an off-white solid. ES-MS: m/z 262 (M+H⁺). ¹H NMR showed two isomers. The minor isomer was estimated to be ~8% from doublets at 7.96 ppm (major isomer) and 8.18 ppm (minor isomer). The major isomer was assigned the (E)-configuration based on the assumption of intramolecular H bond between the ketone and the N-H. The major isomer: ¹H NMR (CDCl₃, 300 MHz) δ 10.97 (br. s., 1 H), 7.96 (d, J = 13.7 Hz, 1 H), 4.58 (s, 2 H), 4.42 (t, J = 5.3 Hz, 1 H), 3.72 (s, 3 H), 3.47 (s, 3 H), 3.43 (s, 6 H), 3.46-3.41 (m, 2 H). ¹³C NMR (CDCl₃, 75 MHz) δ 196.3, 166.4, 160.5, 102.3, 97.8, 76.7, 58.6, 54.4, 51.3, 50.3. HRMS (ES) calcd for C₁₁H₁₉NO₆ [M+H]⁺: 262.1291, found 262.1290.

Preparation of 4-(benzylamino)-3-methylbut-3-en-2-one (6e): Sodium 2-methyl-3-oxobut-1-en-1-olate was prepared following literature procedure⁴: methyl formate (12.3 mL, 199 mmol, 2.5 equiv) was added to NaOMe (8.53 g, 158 mmol, 2 equiv) in ether (60 mL) at 0 °C. Butanone (7.0 mL, 78.1 mmol, 1 equiv) was added slowly over 5 min. The mixture was warmed to rt and stirred for 8 h. The solid product was collected by filtration and dried under vacuum at 50 °C (11.34 g). The sodium salt thus prepared (2.156 g, 17.6 mmol) was suspended in 20 mL of MeOH. HOAc (1.2 mL, 20.9 mmol, 1.2 equiv) and benzylamine (2.2 mL, 20.1 mmol, 1.1 equiv) were added. The mixture was stirred at rt overnight and the product purified by chromatography (eluted with gradient from hexanes to EtOAc, 1.896 g, 10.0 mmol, 68% over 2 steps). The product was a yellow solid. ES-MS: m/z 190 (M+H⁺). ¹H NMR showed that the product was a mixture of two isomers in a 1.3:1 ratio. The major isomer was assigned as the Z-isomer based on the more downfield chemical shift of the N-H signal due to intramolecular hydrogen bonding. The assignments of the ¹H and ¹³C NMR signals were based on ¹H signal integration, GASPE and correlation experiments (HMQC and HMBC).⁵ Major isomer: ¹H NMR (CDCl₃, 300 MHz) δ 9.98 (br. s., 1 H), 7.39-7.22 (m, 5 H), 6.62 (d, J = 12.3 Hz, 1 H), 4.32 (d, J = 6.1 Hz, 2 H), 2.09 (s, 3 H), 1.82 (s, 3 H). ¹³C NMR (CDCl₃, 75 MHz) δ 198.1, 151.7, 99.9, 52.0, 27.9, 17.1. Minor isomer: 1 H NMR (CDCl₃, 300 MHz) δ 7.39-7.22 (m, 6 H), 5.08 (br. s., 1 H), 4.41 (d, J = 5.8 Hz, 2 H), 2.16 (s, 3 H), 1.70 (s, 3 H). ¹³C NMR (CDCl₃, 75 MHz) δ 194.7, 148.7, 107.6, 51.8, 24.1, 8.4. The ¹³C NMR signals for the phenyl protons for both isomers: δ 138.45, 138.42, 128.6, 128.5, 127.5, 127.3, 126.9, 126.8. Anal calcd for C₁₂H₁₅NO: C, 76.16; H, 7.99; N, 7.40; found: C, 75.77; H, 8.15; N, 7.22.

⁴ Hjelmgaard, T.; Søtofte, I.; Tanner, D. J. Org. Chem. 2005, 70, 5688.

⁵ The NMR data as well as the assignments for the two isomers are in agreement with the literature: Zhuo, J.; *Magn. Reson. Chem.* **1998**, *36*, 565.

Preparation of 1-(benzylamino)-2-methylpent-1-en-3-one (6f): Sodium 2-methyl-3oxopent-1-en-1-olate⁶ (1.158 g, 8.51 mmol) was suspended in 12 mL of MeOH. HOAc (0.60 mL, 10.5 mmol, 1.2 equiv) and benzylamine (1.00 mL, 9.15 mmol, 1.1 equiv) were added. The mixture was stirred at rt overnight. MeOH was removed and the residue was diluted with 15 mL of CH₂Cl₂, washed with 20 mL of water. The aqueous layer was back extracted with CH₂Cl₂ (15 mL×2). The combined organic layers were dried and concentrated. The product was purified by chromatography (eluted with gradient from hexanes to EtOAc, 0.794 g, 3.91 mmol, 46%). The product was a light yellow solid. ES-MS: m/z 204 (M+H⁺). ¹H NMR showed the product was a mixture of two isomers in a 2.7:1 ratio. The major isomer was assigned as the Z-isomer based on the more downfield chemical shift of the N-H signal due to intramolecular hydrogen bonding. The assignments of the ¹H and ¹³C NMR signals were based on ¹H signal integration, GASPE and correlation experiments (HMOC and HMBC). Major isomer: ¹H NMR (CDCl₃, 300 MHz) δ 9.95 (br. s., 1 H), 7.42-7.24 (m, 5 H), 6.62 (d, J = 12.3 Hz, 1 H), 4.34 (d, J = 6.1 Hz, 2 H), 2.41 (q, J = 7.3 Hz, 2 H), 1.83 (d, J = 0.6 Hz, 3 H), 1.10 (t, J = 7.3 Hz, 3 H). ¹³C NMR (CDCl₃, 75) MHz) δ 201.0, 151.5, 99.6, 52.2, 32.8, 16.6, 8.3. Minor isomer: ¹H NMR (CDCl₃, 300 MHz) δ 7.42-7.24 (m, 6 H), 4.51 (br. s., 1 H), 4.43 (d, J = 5.7 Hz, 2 H), 2.53 (q, J = 7.4 Hz, 2 H), 1.71 (d, J = 0.9 Hz, 3 H), 1.12 (t, J = 7.4 Hz, 3 H). ¹³C NMR (CDCl₃, 75 MHz) δ 198.4, 147.4, 106.9, 52.0, 29.4, 10.1, 8.7. The 13 C NMR signals for the phenyl protons for both isomers: δ 138.6, 138.5, 128.8, 128.6, 127.8, 127.4, 127.1. Anal calcd for C₁₃H₁₇NO: C, 76.81; H, 8.43; N, 6.89; found: C, 76.74; H, 8.77; N, 6.76.

6g

Preparation of (Z)-4-(benzylamino)-3-phenylbut-3-en-2-one (6g): 4-(dimethylamino)-3-phenylbut-3-en-2-one ⁸ (637.4 mg, 3.37 mmol) was dissolved in 7 mL of MeOH and benzylamine (0.410 mL, 3.75 mmol, 1.1 equiv) was added. The mixture was heated to reflux for 22 h. Another portion of benzylamine (0.200 mL) was added and the reflux was continued for another 12 h. The mixture was cooled to rt, concentrated and the product was isolated by chromatography (eluted with 0-15% EtOAc in hexanes, 686.9 mg, 2.73 mmol, 81%). The product was a light orange oil. ES-MS: m/z 252 (M+H⁺). ¹H NMR showed the product was a mixture of two isomers. A ratio of 9:1 was estimated from doublets at 6.86 ppm (major isomer)

⁶ Myles, D. C.; Bigham, M. H. Org. Synth. **1992**, 70, 231.

⁷ The NMR data as well as the assignments for the two isomers are in agreement with the literature: Zhuo, J.; *Magn. Reson. Chem.* **1998**, *36*, 565.

⁸ Kozmin, S. A.; Iwama, T.; Huang, Y.; Rawal, V. H. J. Am. Chem. Soc. 2002, 124, 4628.

and 7.70 ppm (minor isomer). The major isomer was assigned as the Z-isomer based on the downfield chemical shift of the N-*H* signal: 1 H NMR (CDCl₃, 300 MHz) δ 10.55 (br. s., 1 H), 7.42-7.19 (m, 10 H), 6.86 (d, J = 12.6 Hz, 1 H), 4.44 (d, J = 6.1 Hz, 2 H), 2.09 (s, 3 H). 13 C NMR (CDCl₃, 75 MHz) δ 196.5, 153.2, 140.0, 137.7, 130.0, 128.4, 127.9, 127.3, 126.8, 125.7, 110.3, 52.2, 28.2. HRMS (ES) calcd for $C_{17}H_{17}NO$ [M+H] $^{+}$: 252.1388, found 252.1387.

General procedure of oxalate cyclization in Table 2. Enamine 6a (346.8 mg, 1.49 mmol) and dimethyl oxalate (617.6 mg, 5.23 mmol, 3.5 equiv) were dissolved in MeOH (1.1 mL) and a MeOH solution of NaOMe (25 wt%, 0.850 mL, 3.72 mmol, 2.5 equiv) was added. The mixture was heated to 30 °C for 36 h. The reaction was cooled to 0 °C and quenched with 1 N HCl (4.2 mL) and extracted with EtOAc (10 mL×3). The combined organic layers were dried and concentrated. The product was purified by chromatography (eluted with 40-60% acetone in TBME, 318.4 mg, 1.06 mmol, 71%) to give an off-white solid.

Dimethyl 1-benzyl-4-oxo-1,4-dihydropyridine-2,5-dicarboxylate (**7a**): ES-MS: m/z 302 (M+H⁺). 1 H NMR (CDCl₃, 300 MHz) δ 8.38 (s, 1 H), 7.41-7.34 (m, 3 H), 7.13-7.10 (m, 3 H), 5.46 (s, 2 H), 3.91 (s, 3 H), 3.78 (s, 3 H). 13 C NMR (CDCl₃, 75 MHz) δ 174.8, 165.0, 162.1, 149.4, 138.9, 134.6, 129.0, 128.6, 127.0, 126.7, 119.1, 58.2, 53.3, 52.2. Anal calcd for C₁₆H₁₅NO₅: C, 63.78,; H, 5.02; N, 4.65; found: C, 63.60,; H, 4.93; N, 4.33.

Dimethyl 4-oxo-1-phenyl-1,4-dihydropyridine-2,5-dicarboxylate (**7b**): Tan solid (182.5 mg), prepared from **6b** (201.8 mg). ES-MS: m/z 288 (M+H⁺). ¹H NMR (CDCl₃, 300 MHz) δ 8.27 (s, 1 H), 7.48-7.43 (m, 3 H), 7.26-7.20 (m, 2 H), 7.06 (s, 1 H), 3.83 (s, 3 H), 3.62 (s, 3 H). ¹³C NMR (CDCl₃, 75 MHz) δ 174.7, 164.6, 161.4, 148.5, 141.8, 139.9, 129.7, 129.4, 124.9, 124.8, 118.9, 53.2, 52.2. HRMS (ES) calcd for $C_{15}H_{13}NO_{5}$ [M+H]⁺: 288.0872, found 288.0861.

Dimethyl 1-isopropyl-4-oxo-1,4-dihydropyridine-2,5-dicarboxylate (**7c**): Off-white solid (195.3 mg), prepared from **6c** (203.1 mg). ES-MS: m/z 254 (M+H⁺). ¹H NMR (CDCl₃, 300 MHz) δ 8.27 (s, 1 H), 6.80 (s, 1 H), 4.77 (septet, J = 6.7 Hz, 1 H), 3.84 (s, 3 H), 3.78 (s, 3 H), 1.40 (d, J = 6.7 Hz, 6 H). ¹³C NMR (CDCl₃, 75 MHz) δ 174.3, 165.0, 162.2, 143.2, 140.0, 125.3, 119.3, 54.4, 53.3, 52.0, 22.7. HRMS (ES) calcd for $C_{12}H_{15}NO_5$ [M+H]⁺: 254.1028, found 254.1037.

Dimethyl 1-(2,2-dimethoxyethyl)-3-methoxy-4-oxo-1,4-dihydropyridine-2,5-dicarboxylate (7d): prepared as an orange oil starting from 11.3 g of methyl 4-methoxyacetoacetate (see below for the synthesis of 6d). ES-MS: m/z 330 (M+H⁺). ¹H NMR (CDCl₃, 300 MHz) δ 8.14 (s, 1 H), 4.51 (t, J = 4.9 Hz, 1 H), 3.99 (s, 3 H), 3.98 (s, 3 H), 3.94 (d, J = 4.9 Hz, 2 H), 3.91 (s, 3 H), 3.41 (s, 6 H). ¹³C NMR (CDCl₃, 75 MHz) δ 170.5, 164.6, 161.8, 149.6, 145.7, 133.6, 117.6, 102.2, 59.9, 56.0, 55.2, 52.8, 51.6. HRMS (ES) calcd for C₁₄H₁₉NO₈ [M+H]⁺: 330.1189, found 330.1184.

Methyl 1-benzyl-5-methyl-4-oxo-1,4-dihydropyridine-2-carboxylate (**7e**): light orange solid (203.5mg), prepared from **6e** (223.3 mg). ES-MS: m/z 258 (M+H $^+$). 1 H NMR (CDCl₃, 300 MHz) δ 7.39-7.31 (m, 4 H), 7.11-7.08 (m, 2 H), 6.95 (s, 1 H), 5.37 (s, 2 H), 3.79 (s, 3 H), 2.06 (s, 3 H). 13 C NMR (CDCl₃, 75 MHz) δ 178.3, 162.7, 140.8, 138.6, 135.7, 128.9, 128.7, 128.0, 126.6, 119.8, 57.0, 52.9, 13.6. Anal calcd for C₁₅H₁₅NO₃: C, 70.02; H, 5.88; N, 5.44; found: C, 69.73; H, 5.97; N, 5.31.

Methyl 1-benzyl-3,5-dimethyl-4-oxo-1,4-dihydropyridine-2-carboxylate (**7f**): yellow oil (178.5 mg), prepared from **6f** (223.2 mg). ES-MS: m/z 272 (M+H⁺). 1 H NMR (CDCl₃, 300 MHz) δ 7.41-7.34 (m, 3 H), 7.28 (m, 1 H), 7.15-7.12 (m, 2 H), 4.99 (s, 2 H), 3.73 (s, 3 H), 2.05 (d, J = 0.9 Hz, 3 H), 2.03 (s, 3H). 13 C NMR (CDCl₃, 75 MHz) δ 177.8, 163.9, 139.2, 137.8, 134.9, 128.9, 128.5, 127.0, 125.2, 123.3, 57.8, 52.6, 13.9, 12.1. HRMS (ES) calcd for C₁₆H₁₇NO₃ [M+H]⁺: 272.1287, found 272.1293.

Methyl 1-benzyl-4-oxo-5-phenyl-1,4-dihydropyridine-2-carboxylate (7g): off-white solid (170.3 mg), prepared from 6g (225.2 mg). ES-MS: m/z 320 (M+H $^+$). 1 H NMR (CDCl₃, 300 MHz) δ 7.64-7.61 (m, 3 H), 7.44-7.32 (m, 6 H), 7.17-7.14 (m, 3 H), 5.48 (s, 2 H), 3.83 (s, 3 H). 13 C NMR (CDCl₃, 75 MHz) δ 176.1, 162.5, 142.0, 138.1, 135.5, 133.7, 130.7, 128.8, 128.2, 128.1, 127.7, 126.7, 122.8, 57.3, 53.0. HRMS (ES) calcd for $C_{20}H_{17}NO_3$ [M+H] $^+$: 320.1287, found 320.1282.

Methyl 1-benzyl-4-oxo-1,4-dihydropyridine-2-carboxylate (**7h**): light yellow solid (76.7 mg), prepared from 4-(benzylamino)-but-3-en-2-one (**6h**, 241.6 mg)⁹: ES-MS: m/z 244 (M+H⁺). ¹H NMR (CDCl₃, 300 MHz) δ 7.44 (d, J = 7.6 Hz, 1 H), 7.39-7.30 (m, 3 H), 7.10 (m, 2 H), 6.94 (d, J = 2.7 Hz, 1 H), 6.47 (dd, J = 7.6, 2.7 Hz, 1 H), 5.36 (s, 2 H), 3.79 (s, 3 H). ¹³C NMR (CDCl₃, 75 MHz) δ 178.2, 162.3, 143.4, 139.6, 135.2, 128.6, 128.0, 126.6, 121.8, 118.9, 57.1, 52.9. HRMS (ES) calcd for $C_{14}H_{13}NO_{3}$ [M+H]⁺: 244.0974, found 244.0963.

One pot synthesis of 1-(2,2-dimethoxyethyl)-5-methoxy-6-(methoxycarbonyl)-4-oxo-1,4-dihydropyridine-3-carboxylic acid (9): A round-bottom flask was charged with methyl 4-methoxyacetoacetate (8, 12.0 mL, 92.9 mmol, 1 equiv) and DMF-DMA (18.5 mL, 139 mmol,

⁹ (a) Murphy, J. P.; Hadden, M.; Stevenson, P. J. *Tetrahedron* 1997, 53, 11827. (b) You, L.; Hsung, R. P.; Bedermann, A. A.; Kurdyumov, A. V.; Tang, Y.; Buchanan, G. S.; Cole, K. P. *Adv. Synth. Catal.* **2008**, *350*, 2885.

1.5 equiv). The mixture was stirred at room temperature overnight and concentrated to remove excess DMF-DMA. The crude product oil was dissolved in MeOH (70 mL) and the resultant solution was cooled to ~5 °C. Aminoacetaldehyde dimethyl acetal (10 mL, 92.7 mmol, 1.0 equiv) was added while maintaining temperature below 10 °C. The mixture was warmed to 20 °C for 2 h, and concentrated. Dimethyl oxalate (38.35 g, 324 mmol, 3.5 equiv) was added followed by LiOMe (2.2 M solution in MeOH, 63 mL, 139 mmol, 1.5 equiv). The mixture was heated to 46 °C for 20.5 h and cooled to -5 °C. LiOH (13.33 g, 557 mmol, 6.0 equiv) was added while maintaining internal temperature below 0 °C. The mixture was stirred at -2 °C for 4.5 h, and quenched with aqueous HCl (2.0 M, 313 mL, 626 mmol, 6.75 equiv) while maintaining internal temperature below 0 °C. When the addition was complete, the mixture was warmed to 10 °C and EtOAc (280 mL) was added. The mixture was filtered and the organic layer was separated. Water (130 mL) was added to the EtOAc layer and mixture was concentrated to remove EtOAc. The resulting slurry was filtered. The product was washed with water, and dried in a vacuum oven at 55 °C to give 9 as a white solid (17.70 g, 56.1 mmol, 61%). ES-MS: m/z 316 (M+H⁺). ¹H NMR (CDCl₃, 300 MHz) δ 8.41 (s, 1 H), 4.53 (t, J = 4.6 Hz, 1 H), 4.11 (d, J = 4.6 Hz, 2 H), 4.03 (s, 3 H), 4.02 (s, 3 H), 3.42 (s, 6 H). ¹³C NMR (CDCl₃, 75 MHz) δ 174.7, 165.8, 161.4, 148.5, 145.3, 136.4, 116.4, 102.2, 60.8, 57.1, 55.8, 53.6. Anal calcd for C₁₃H₁₇NO₈: C, 49.52; H, 5.43; N, 4.44; found: C, 49.50; H 5.47; N 4.32.

Preparation of 1-(2,2-dimethoxyethyl)-6-(ethoxycarbonyl)-5-methoxy-4-oxo-1,4-dihydropyridine-3-carboxylic acid (10): LiH (206.1 mg, 25.9 mmol, 1.3 equiv) was dissolved in ethanol (60 mL). The methyl ester **9** (6.40 g, 20.3 mmol) was added after the off-gassing had subsided. The mixture was stirred at rt for 3 h. A second portion of LiH (28.4 mg) was added and the stirring was continued for another 2 h. TBME (100mL) was added followed by HCl (1 N, 100 mL). The aqueous layer was extracted a second time with 100 mL of TBME. The combined organic layers were concentrated. The product precipitated out of residual water (~8 mL). TBME (15 mL) was added and the mixture was stirred at rt for 30 min, filtered and dried (5.435 g, 16.5 mmol, 81%) to give a white solid. ES-MS: m/z 330 (M+H⁺). ¹H NMR (CDCl₃, 300 MHz) δ 8.40 (s, 1 H), 4.55 (t, J = 4.7 Hz, 1 H), 4.49 (q, J = 7.2 Hz, 2 H), 4.10 (d, J = 4.6 Hz, 2 H), 4.03 (s, 3 H), 3.41 (s, 6 H), 1.44 (t, J = 7.2 Hz, 3 H). ¹³C NMR (CDCl₃, 75 MHz) δ 174.8, 165.8, 160.9, 148.4, 145.2, 136.8, 116.4, 102.2, 63.3, 60.7, 57.1, 55.7, 14.0. Anal calcd for C₁₄H₁₉NO₈: C, 51.06; H, 5.82; N, 4.25; found: C, 51.02; H 5.94; N 4.17.

Preparation of 1-(2,2-dimethoxyethyl)-6-(isopropoxycarbonyl)-5-methoxy-4-oxo-1,4-dihydropyridine-3-carboxylic acid (11): NaH (505.6 mg, 60 wt%, 12.6 mmol, 1.6 equiv) was

added to anhydrous IPA (20 mL) and the mixture was stirred at rt for 1 h. The methyl ester **9** (2.49 g, 7.90 mmol) was added. The mixture was stirred at rt overnight and a second portion of NaH (523.0 mg) was added. The mixture was stirred at rt for 5 h. The methyl ester was consumed, however, formation of a large amount of the 2,5-diacid was observed. The reaction was quenched with 2 N HCl and extracted with iPAC (×2). The combined organic layers were washed with water until the 2,5-diacid was completely removed. The organic layer was dried, concentrated and the product crystallized from iPAC/TBME as a white solid (370.3 mg, 1.08 mmol, 14%). ES-MS: m/z 344 (M+H⁺). ¹H NMR (CDCl₃, 300 MHz) δ 8.39 (s, 1 H), 5.35 (spt, J = 6.3 Hz, 1 H), 4.57 (t, J = 4.9 Hz, 1 H), 4.08 (d, J = 4.9 Hz, 2 H), 4.02 (s, 3 H), 3.42 (s, 6 H), 1.43 (d, 6 H). ¹³C NMR (CDCl₃, 75 MHz) δ 174.8, 165.9, 160.4, 148.2, 145.0, 137.1, 116.5, 102.2, 71.9, 60.7, 57.1, 55.7, 21.6. HRMS (ES) calcd for C₁₅H₂₁NO₈ [M+H]⁺: 344.1345, found 344.1353.

Preparation of (3S,11aR)-3-methyl-6-(methyloxy)-5,7-dioxo-2,3,5,7,11,11a-hexahydro[1,3]oxazolo[3,2-a]pyrido[1,2-d]pyrazine-8-carboxylic acid (12): Acetal 9 (22.54 g, 71.5 mmol) was dissolved in 220 mL of CH₃CN. HOAc (20 mL, 349 mmol, 4.9 equiv) and CH₃SO₃H (1.4 mL, 21.6 mmol, 0.3 equiv) were added at rt and the mixture was heated to 58-65 °C for 19.5 h. Alaninol (7.511 g, 100 mmol, 1.4 equiv) in CH₃CN (15 mL) was added slowly and the resultant mixture was stirred at 64 °C for 18.5 h. The mixture was concentrated, and the residue was redissolved in CH₂Cl₂ (170 mL). HCl (1 N, 170 mL) was added and the layers were separated. The aqueous layer was extracted with CH₂Cl₂ (170 mL×2) and the organic layers were combined and concentrated. MeOH (50 mL) was added and the resultant mixture was again concentrated. MeOH (80 mL) was added and the resultant mixture was heated at reflux for 4 h, gradually cooled to 20 °C and held at 20 °C for 15 h. The product was collected by filtration and dried under vacuum (15.67 g, 74% yield, 95.2% de) to give a white solid.

The major isomer: (3S,11aR)-3-methyl-6-(methyloxy)-5,7-dioxo-2,3,5,7,11,11a-hexahydro[1,3]oxazolo[3,2-a]pyrido[1,2-d]pyrazine-8-carboxylic acid (12a). ES-MS: m/z 295 (M+H $^+$). 1 H NMR (CDCl₃, 300 MHz) δ 8.44 (s, 1 H), 5.40 (dd, J = 9.9, 3.7 Hz, 1 H), 4.55-4.39 (m, 3 H), 4.12 (s, 3 H), 3.98 (dd, J = 12.1, 9.9 Hz, 1 H), 3.74 (dd, J = 8.2, 6.5 Hz, 1 H), 1.46 (d, J = 6.1 Hz, 3 H). 13 C NMR (CDCl₃, 75 MHz) δ 176.2, 165.7, 153.8, 152.6, 143.0, 131.5, 116.1, 82.4, 73.2, 61.6, 55.8, 52.2, 17.0. Anal calcd for $C_{13}H_{14}N_2O_6$: C, 53.06; H, 4.80; N, 9.52; found: C, 53.02; H, 4.74; N, 9.45.

The minor isomer (3S,11aS)-6-methoxy-3-methyl-5,7-dioxo-2,3,5,7,11,11a-hexahydrooxazolo[3,2-a]pyrido[1,2-d]pyrazine-8-carboxylic acid (12b) was isolated from reaction mixtures by prep HPLC (Chiralpak IC 25×1 cm, 20 μm dp, with CH₃CN as the mobile phase at 3 mL/min, rt). An off-white solid was obtained. ES-MS: m/z 295 (M+H⁺). ¹H NMR (CDCl₃, 300MHz) δ 8.48 (s, 1 H), 5.26 (dd, J = 3.0, 10.1 Hz, 1 H), 4.76 (dd, J = 3.0, 12.2 Hz, 1 H), 4.33 (m, 1 H), 4.16-4.11 (m, 1 H), 4.04 (s, 3 H), 4.07-4.00 (m, 2 H), 1.49 (d, J = 6.4 Hz, 3 H). ¹³C NMR (CDCl₃, 75 MHz) δ 176.0, 165.8, 152.9, 152.8, 143.6, 131.6, 115.7, 82.8, 73.6, 61.4, 57.2, 52.6, 18.1. HRMS (ES) calcd for C₁₃H₁₄N₂O₆ [M+H]⁺: 295.0930, found 295.0927.

Preparation of (3*S*,11a*R*)-*N*-[(2,4-difluorophenyl)methyl]-3-methyl-6-(methyloxy)-5,7-dioxo-2,3,5,7,11,11a-hexahydro[1,3]oxazolo[3,2-a]pyrido[1,2-d]pyrazine-8-carboxamide (13): Acid 12 (3.00 g, 10.2 mmol) and CDI (2.15 g, 13.2 mmol, 1.3 equiv) were slurried in DME (30 mL). The mixture was heated to 80 °C for 1 h. The resulting solution was cooled to 20 °C, then treated with 2,4-difluorobenzylamine (1.45 mL, 12.2 mmol, 1.2 equiv). After stirring for 1 h, the mixture was quenched with water (30 mL) and DME was removed under reduced pressure. The product was collected by filtration and dried in a vacuum oven overnight at 50 °C. Amide 13 was obtained as a white solid (4.06 g, 9.68 mmol, 95%).

The major isomer (3*S*,11a*R*)-*N*-[(2,4-difluorophenyl)methyl]-3-methyl-6-(methyloxy)-5,7-dioxo-2,3,5,7,11,11a-hexahydro[1,3]oxazolo[3,2-a]pyrido[1,2-d]pyrazine-8-carboxamide (13a): ES-MS: m/z 420 (M+H⁺). ¹H NMR (CDCl₃, 300 MHz) δ 10.33 (t, J = 5.7 Hz, 1 H), 8.41 (s, 1 H), 7.36 (td, J = 8.7, 6.4 Hz, 1 H), 6.86-6.77 (m, 2 H), 5.32 (dd, J = 9.9, 3.7 Hz, 1 H), 4.70-4.56 (m, 2 H), 4.49-4.33 (m, 3 H), 4.07 (s, 3 H), 3.90 (dd, J = 12.3, 9.8 Hz, 1 H), 3.71 (dd, J = 8.5, 6.8 Hz, 1 H), 1.45 (d, 3 H). ¹³C NMR (CDCl₃, 75 MHz) δ 174.3, 163.8, 162.2 (dd, J = 248, 12 Hz), 160.7 (dd, J = 249, 12 Hz), 154.6, 153.2, 142.4, 130.5 (dd, J = 9.7, 5.8 Hz), 130.0, 121.3 (dd, J = 15, 3.9 Hz), 118.9, 111.1 (dd, J = 21, 3.6 Hz), 103.7 (t, J = 25 Hz), 82.4, 73.0, 61.3, 55.2, 52.0, 36.5, 17.1. ¹⁹F NMR (CDCl₃, 282 MHz) δ -111.5 (d, J = 6.9 Hz), -114.3 (d, J = 6.9 Hz). Anal calcd for $C_{20}H_{19}F_2N_3O_5$: C, 57.28; H, 4.57; N, 10.02; found: C, 57.37; H, 4.48; N, 9.92.

The minor isomer (3S,11aS)-N-(2,4-difluorobenzyl)-6-methoxy-3-methyl-5,7-dioxo-2,3,5,7,11,11a-hexahydrooxazolo[3,2-a]pyrido[1,2-d]pyrazine-8-carboxamide (13b) was prepared analogously from the corresponding acid 12b (47.7 mg of 12b gave 33.8 mg of 13b in 50% yield). ES-MS: m/z 420 (M+H⁺). ¹H NMR (CDCl₃, 300 MHz) δ 10.33 (t, J = 5.8 Hz, 1 H), 8.44 (s, 1 H), 7.37-7.25 (m, 1 H), 6.86-6.74 (m, 2 H), 5.12 (dd, J = 3.1, 10.0 Hz, 1 H), 4.67-4.51 (m, 3 H), 4.31 (m, 1 H), 4.11-3.85 (m, 3 H), 4.03 (s, 3 H), 1.49 (d, J = 6.3 Hz, 3 H). ¹³C NMR (CDCl₃, 75 MHz) δ 174.3, 163.9, 162.2 (dd, J = 248, 12 Hz), 160.7 (dd, J = 249, 12 Hz), 153.8, 153.6, 142.6, 130.5 (dd, J = 9.6, 5.9 Hz), 129.9, 121.3 (dd, J = 15.3, 3.7 Hz), 118.8, 111.1 (dd, J = 21, 3.6 Hz), 103.8 (t, J = 25.3 Hz), 83.0, 73.6, 61.2, 56.9, 52.5, 36.5 (d, J = 3.6 Hz), 18.2. ¹⁹F NMR (CDCl₃, 282 MHz) δ -111.5 (d, J = 7.6 Hz), -114.3 (d, J = 6.9 Hz). HRMS (ES) calcd for $C_{20}H_{19}F_2N_3O_5$ [M+H]⁺: 420.1371, found 420.1362.

(S)-N-(2,4-difluorobenzyl)-2-(1-hydroxypropan-2-yl)-9-methoxy-1,8-dioxo-2,8-dihydro-1H-pyrido[1,2-a]pyrazine-7-carboxamide (14) was isolated as a yellow solid from reaction mixtures and characterized: ES-MS: m/z 420 (M+H⁺). ¹H NMR (CDCl₃, 300 MHz) δ 10.55 (t, J = 5.9 Hz, 1 H), 8.57 (s, 1 H), 7.42-7.34 (m, 1 H), 6.88-6.78 (m, 2 H), 6.73 (d, J = 6.4 Hz, 1 H), 6.59 (d, J = 6.3 Hz, 1 H), 5.04-4.93 (m, 1 H), 4.63 (d, J = 5.8 Hz, 2 H), 3.98 (s, 3 H), 3.92-3.87 (m, 1 H), 3.80-3.72 (m, 1 H), 3.25 (br. s., 1 H), 1.37 (d, J = 7.1 Hz, 3 H). ¹³C NMR (CDCl₃, 75 MHz) δ 172.0, 163.5, 162.0 (dd, J = 248, 12 Hz), 160.2 (dd, J = 249, 12 Hz), 154.1, 152.8, 137.7, 130.4 (dd, J = 9.7, 5.8 Hz), 129.1, 120.9 (dd, J = 15, 3.9 Hz), 120.0, 115.8, 111.1 (dd, J = 21, 3.5 Hz), 110.8, 103.5 (t, J = 25 Hz), 63.5, 60.8, 52.2, 36.4 (d, J = 2.8 Hz), 14.8. ¹⁹F NMR (CDCl₃, 282 MHz) δ -111.1 (d, J = 7.6 Hz), -114.2 (d, J = 6.9 Hz). HRMS (ES) calcd for C₂₀H₁₉F₂N₃O₅ [M+H]⁺: 420.1371, found 420.1374.

GSK1265744 Enol ether **13** (1.324 g, 3.16 mmol) was dissolved in CH₃CN (10 mL) at 50 °C and MgBr₂ (1.168 g, 6.34 mmol, 2 equiv) was added. The mixture was heated at 50 °C for 2.5 h, cooled to rt and quenched with HCl (1 N, 10 mL). The resultant slurry was stirred at rt overnight. The product was collected by filtration and dried to give a white solid (1.154 g, 2.85 mmol, 90%).

The demethylation was better accomplished with LiBr on large scale. A representative procedure is as following: Enol ether **13** (4.609 g, 11.0 mmol) was dissolved in THF (45 mL). LiBr (2.10 g, 24.2 mmol, 2.2 equiv) and water (0.26 g, 14.4 mmol, 1.3 equiv) were added. The mixture was heated at reflux overnight, and cooled to rt. HOAc (1.98 g, 33.0 mmol, 3 equiv) in

water (38 mL) was added slowly. The resultant suspension was stirred at rt for 2.5 h, filtered and reslurried in aq THF (25 mL/25 mL) at rt for 2 h. The product was collected by filtration and dried to give a white solid (4.14 g, 2.85 mmol, 10.2 mmol, 93%).

GSK1265744 (**1a**): ES-MS: m/z 406 (M+H⁺). ¹H NMR (DMSO-d₆, 300 MHz) δ 11.47 (s, 1 H), 10.31 (t, J = 5.9 Hz, 1 H), 8.47 (s, 1 H), 7.40 (td, J = 8.6, 6.8 Hz, 1 H), 7.25 (ddd, J = 10.4, 9.5, 2.6 Hz, 1 H), 7.06 (td, J = 8.5, 2.4 Hz, 1 H), 5.39 (dd, J = 10.1, 4.1 Hz, 1 H), 4.89 (dd, J = 12.3, 4.1 Hz, 1 H), 4.54 (d, J = 5.8 Hz, 2 H), 4.40 (dd, J = 8.4, 6.8 Hz, 1 H), 4.29 (sxt, J = 6.4 Hz, 1 H), 4.00 (dd, J = 11.9, 10.2 Hz, 1 H), 3.66 (dd, J = 8.4, 6.6 Hz, 1 H), 1.34 (d, J = 6.2 Hz, 3 H). ¹³C NMR (DMSO-d₆, 75 MHz) δ 170.4, 163.6, 161.5 (dd, J = 245, 12 Hz), 160.2 (dd, J = 247, 12 Hz), 158.9, 153.1, 141.5, 130.8 (dd, J = 9.8, 6.2 Hz), 122.4 (dd, J = 15.2, 3.6 Hz), 117.4, 114.9, 111.4 (dd, J = 21, 3.4 Hz), 103.8 (t, J = 26 Hz), 81.7, 72.4, 52.9, 51.4, 35.8 (d, J = 3.0 Hz), 16.6. ¹⁹F NMR (DMSO-d₆, 282 MHz) δ -111.9 (d, J = 7.6 Hz), -114.5 (d, J = 6.9 Hz). Anal calcd for C₁₉H₁₇F₂N₃O₅: C, 56.30; H, 4.23; N, 10.37; found: C, 55.99; H, 4.12; N, 10.21.

The minor diastereomer (3S,11aS)-N-(2,4-difluorobenzyl)-6-hydroxy-3-methyl-5,7-dioxo-2,3,5,7,11,11a-hexahydrooxazolo[3,2-a]pyrido[1,2-d]pyrazine-8-carboxamide (1b) was prepared analogously from the corresponding methyl ether 13b (71.2 mg): solid (38.0 mg, 55%). ES-MS: m/z 406 (M+H⁺). ¹H NMR (DMSO-d₆, 300 MHz) δ 11.86 (br. s., 1 H), 10.33 (t, J = 5.9 Hz, 1 H), 8.45 (s, 1 H), 7.39 (td, J = 8.6, 6.8 Hz, 1 H), 7.23 (ddd, J = 10.5, 9.4, 2.5 Hz, 1 H), 7.06 (tdd, J = 8.5, 2.5, 0.9 Hz, 1 H), 5.21 (dd, J = 10.3, 3.4 Hz, 1 H), 4.96 (dd, J = 12.0, 3.4 Hz, 1 H), 4.54 (d, J = 5.9 Hz, 2 H), 4.30-4.22 (m, 1 H), 4.12 - 3.99 (m, 3 H), 1.36 (d, J = 6.3 Hz, 3 H). ¹³C NMR (DMSO-d₆, 75 MHz) δ 170.4, 163.7, 161.5 (dd, J = 245, 12 Hz), 160.2 (dd, J = 247, 12 Hz), 159.2, 153.9, 141.6, 130.8 (dd, J = 9.7, 6.1 Hz), 122.4 (dd, J = 15.2, 3.6 Hz), 117.1, 114.9, 111.4 (dd, J = 21.0, 3.3 Hz), 103.8 (t, J = 25.8 Hz), 83.0, 73.0, 54.1, 51.4, 35.8 (d, J = 3.0 Hz), 17.9. ¹⁹F NMR (DMSO-d₆, 282 MHz) δ -111.9 (d, J = 6.9 Hz), -114.5 (d, J = 6.9 Hz). HRMS (ES) calcd for C₁₉H₁₇F₂N₃O₅ [M+H]⁺: 406.1215, found 406.1215.

Preparation of methyl 5-((2,4-difluorobenzyl)carbamoyl)-1-(2,2-dimethoxyethyl)-3-methoxy-4-oxo-1,4-dihydropyridine-2-carboxylate (19): The 5-acid **9** (3.059 g, 9.70 mmol) and CDI (1.904 g, 11.7 mmol, 1.2 equiv) were dissolved in 30 mL of THF and the resultant mixture was heated to reflux for 2.5 h. A second portion of CDI (516 mg) was added and the

reflux was continued for another 2 h. The mixture was cooled to rt and 2,4-difluorobenzylamine (1.3 mL, 10.9 mmol, 1.1 equiv) was added. The mixture was stirred at rt overnight. The reaction mixture was diluted with TBME (30 mL), and washed with HCl (1 N, 30 mL), NaOH (1 N, 30 mL) and brine (30 mL). The organic layer was concentrated and used directly in the next step. A pure sample of the amide-acetal can be obtained by chromatography: ES-MS: m/z 441 (M+H⁺). H NMR (CDCl₃, 300 MHz) δ 10.38 (t, J = 5.7 Hz, 1 H), 8.41 (s, 1 H), 7.41-7.33 (m, 1 H), 6.85-6.75 (m, 2 H), 4.61 (d, J = 5.9 Hz, 2 H), 4.49 (t, J = 4.8 Hz, 1 H), 4.04 (d, J = 4.8 Hz, 2 H), 3.98 (s, 3 H), 3.95 (s, 3H), 3.38 (s, 6 H). CDCl₃, 75 MHz) δ 173.0, 163.9, 162.0 (dd, J = 248, 12 Hz), 162.0, 160.6 (dd, J = 249, 12 Hz), 149.2, 144.4, 134.8, 130.5 (dd, J = 9.5, 5.9 Hz), 121.4 (dd, J = 15, 3.6 Hz), 119.1, 110.9 (dd, J = 21, 3.6 Hz), 103.5 (t, J = 25 Hz), 102.5, 60.5, 56.5, 55.4, 53.2, 36.2 (d, J = 3.0 Hz). NMR (CDCl₃, 282 MHz) δ -111.8 (d, J = 6.9 Hz), -114.4 (d, J = 6.9 Hz). HRMS (ES) calcd for $C_{20}H_{22}F_{2}N_{2}O_{7}$ [M+H]⁺: 441.1473, found 441.1475.

5-((2,4-difluorobenzyl)carbamoyl)-3-methoxy-4-oxo-1-(2-**Preparation** methyl oxoethyl)-1,4-dihydropyridine-2-carboxylate (15): The crude amide-acetal (19) prepared above was dissolved in 25 mL of HCOOH and the mixture was heated to 60 °C for 3 h. The solvent was removed. The residue was diluted with NaH₂PO₄ (10%, 25 mL) and extracted with CH₂Cl₂ (25 mL×2). The combined organic layer was concentrated and reslurried in 15 mL of TBME overnight. The product was isolated by filtration. The product was shown to contain 70% hydrate (3.541 g, 8.70 mmol, 90%). The hydrate does not affect the subsequent reactions, although a pure sample of the aldehyde can be obtained by chromatography: white solid. ES-MS: m/z 413 (M+H₂O+H⁺), 395 (M+H⁺). ¹H NMR (CDCl₃, 300MHz) δ 10.35 (t, J = 5.7 Hz, 1 H), 9.62 (s, 1 H), 8.42 (s, 1 H), 7.39-7.31 (m, 1 H), 6.87-6.77 (m, 2 H), 4.95 (s, 2 H), 4.62 (d, J = 5.9Hz, 2 H), 4.00 (s, 3 H), 3.94 (s, 3 H). ¹³C NMR (CDCl₃, 75 MHz) δ 192.3, 173.5, 164.0, 162.3, 162.2 (dd, J = 248, 12 Hz), 160.7 (dd, J = 249, 11 Hz), 150.4, 144.7, 133.7, 130.4 (dd, J = 9.3, 12.2)6.5 Hz), 121.2 (dd, J = 15, 3.6 Hz), 119.4, 111.2 (dd, J = 21, 3.4 Hz), 103.8 (t, J = 25 Hz), 63.6, 60.8, 53.6, 36.5 (d, J = 3.3 Hz). ¹⁹F NMR (CDCl₃, 282 MHz) δ -111.5 (d, J = 6.9 Hz), -114.3 (d, J = 7.6 Hz). HRMS (ES) calcd for $C_{18}H_{16}F_2N_2O_6 [M+H]^+$: 395.1055, found 395.1067.

One pot synthesis of 1 from 15 by $\mathrm{Mg^{2+}}$ mediated cyclization and demethylation: A mixture of aldehyde 15 (1.3136 g, contained ~70% hydrate, 3.23 mmol), $\mathrm{Mg(OTf)_2}$ (1.157 g, 3.59 mmol, 1.1 equiv) and l-alaninol (0.280 mL, 3.60 mmol, 1.1 equiv) in $\mathrm{CH_3CN}$ (15 mL) was heated to 60 °C for 21 h, and NaBr (457 mg, 4.44 mmol, 1.4 equiv) was added. The mixture was heated at 60 °C for another 15 h and cooled to rt. $\mathrm{H_2SO_4}$ (0.5 M, 15 mL) was added and the mixture as stirred at rt for 7 h. The product was collected by filtration and dried to give a white solid (681.4 mg, 1.68 mmol, 52% yield, 99.3% de).

Diastereomeric excess values for the pairs 12a/b, 13a/b and 1a/b were measured by integrations of HPLC peaks. An analytically pure sample of minor isomer 12b was separated from crude reaction mixture by prep HPLC. Analytically pure samples of minor isomers 13b and 1b were synthesized from 12b using the standard procedure. The de's measured by HPLC ratio were also consistent with those observed by ¹H NMR.

The diastereomeric excess for the pair 12a/12b was measured by integrations of HPLC peaks. HPLC: Agilent 1100 series; column: Zorbax Bonus-RP (3.5 μm, 4.6×150 mm); flow rate: 1 mL/min; solvent: 0.05% TFA in water - 0.05% TFA in CH₃CN (gradient, from 88:12 to 5:95). t_R: 12a 5.1 min, 12b 4.8 min.

The HPLC traces of the reaction mixtures for the following transformation (formation of **12** from **9-11**, Scheme 2) are included on page S15 to page S17.

Data File C:\CHEM32\2\DATA\13JUL2012\GSK1265744 IPM 2012-07-13 13-55-31\040-0401.D Sample Name: HW212905-049R4

Acq. Operator Seq. Line :

Location: Vial 40

Injection Date : 13-Jul-12, 15:36:28 Inj : 1

Inj Volume : 2 μl

: GSK1265744 IPM.M Acq. Method

Analysis Method: C:\CHEM32\2\DATA\GSK1265744 IPM 2012-07-10 14-17-06\11JUL2012\

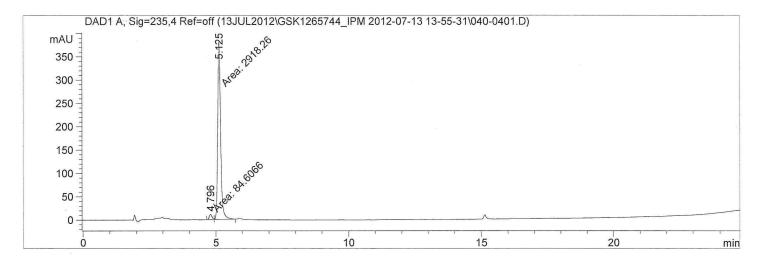
GSK1265744 IPM 2012-07-11 10-04-58\GSK1265744 IPM.M

: 7/11/2012 1:05:27 PM Last changed

(modified after loading)

Method Info : GSK1265744 IPM method using Bonus RP column, 150 x 4.6mm, 3.5um

Sample Info : Me 50C 17h



Area Percent Report

Sorted By Signal

Multiplier: 1.0000 Dilution: 1.0000 Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=235,4 Ref=off

Peak	RetTime	Type	Width	Area	Height	Area
#	[min]		[min]	[mAU*s]	[mAU]	ଚ
1	4.796	MF	0.1252	84.60658	11.26268	2.8175
2	5.125	FM	0.1274	2918.26392	381.63705	97.1825

3002.87050 392.89973 Totals :

Data File C:\CHEM32\2\DATA\13JUL2012\GSK1265744_IPM 2012-07-13 17-19-11\041-0101.D Sample Name: HW212905-049R5

Acq. Operator : Seq. Line : 1
Acq. Instrument : Instrument 2 Location : Vial 41
Injection Date : 7/13/2012 5:25:22 PM Inj : 1

Inj Volume : 2 μl

Acq. Method : C:\CHEM32\2\DATA\13JUL2012\GSK1265744_IPM 2012-07-13 17-19-11\

GSK1265744 IPM.M

Last changed : 5/16/2012 2:45:41 PM by sg

Analysis Method : C:\CHEM32\2\DATA\GSK1265744 IPM 2012-07-10 14-17-06\11JUL2012\

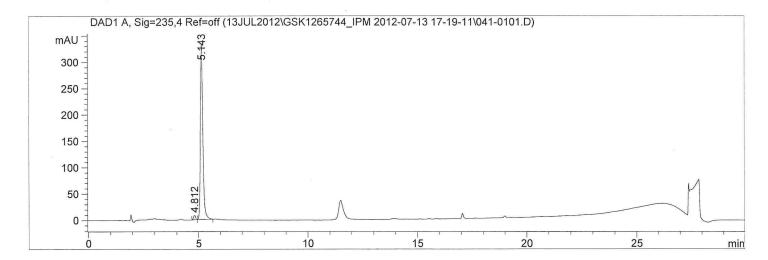
GSK1265744 IPM 2012-07-11 10-04-58\GSK1265744 IPM.M

Last changed : 7/11/2012 1:05:27 PM

(modified after loading)

Method Info : GSK1265744 IPM method using Bonus RP column, 150 x 4.6mm, 3.5um

Sample Info : Et 50C 17h



Area Percent Report

Sorted By : Signal

Multiplier: : 1.0000
Dilution: : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=235,4 Ref=off

Peak	RetTime	Type	Width	Area	Height	Area	
#	[min]		[min]	[mAU*s]	[mAU]	%	
				[
1	4.812	BV	0.1130	60.25024	8.26978	2.2895	
2	5.143	VB	0.1172	2571.37158	336.38025	97.7105	

Totals: 2631.62182 344.65003

Data File C:\CHEM32\2\DATA\13JUL2012\GSK1265744_IPM 2012-07-13 17-19-11\042-0201.D Sample Name: HW212905-049R6

Acq. Operator : Seq. Line : 2

Acq. Instrument : Instrument 2 Location : Vial 42

Injection Date : 7/13/2012 5:56:59 PM Inj : 1

Inj Volume : 2 μl

Acq. Method : C:\CHEM32\2\DATA\13JUL2012\GSK1265744_IPM 2012-07-13 17-19-11\

GSK1265744 IPM.M

Last changed : 5/16/2012 2:45:41 PM by sg

Analysis Method: C:\CHEM32\2\DATA\GSK1265744 IPM 2012-07-10 14-17-06\11JUL2012\

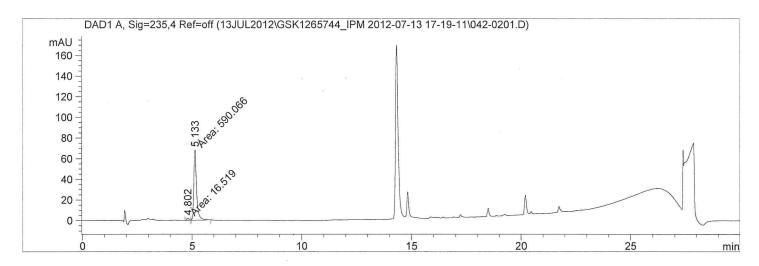
GSK1265744 IPM 2012-07-11 10-04-58\GSK1265744 IPM.M

Last changed : 7/11/2012 1:05:27 PM

(modified after loading)

Method Info : GSK1265744 IPM method using Bonus RP column, 150 x 4.6mm, 3.5um

Sample Info : iPr 50C 17h



Area Percent Report

Sorted By : Signal

Multiplier: : 1.0000
Dilution: : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=235,4 Ref=off

Peak	RetTime	Type	Width	Area	Height	Area
#	[min]		[min]	[mAU*s]	[mAU]	90
1	4.802	MF	0.1358	16.51898	2.02695	2.7233
2	5.133	FM	0.1445	590.06592	68.03947	97.2767

Totals: 606.58489 70.06643

The diastereomeric excess for the pair 13a/13b was measured by integrations of HPLC peaks. HPLC: Agilent 1100 series; column: Waters XTerra MS C_{18} (3.5 μ m, 4.6×150 mm); flow rate: 1 mL/min; solvent: 0.1% aqueous NH₄OH-CH₃CN (gradient, from 7:93 to 95:5). t_R : 13a 12.3 min, 13b: 12.0 min.

The HPLC traces of the reaction mixtures for the following transformation (formation of **13** from **15**, Table 3) are included on page S19 to page S26.

Data File C:\CHEM32\1\DATA\12JUL2011\HIVINTEGRASEAMIDE 2011-07-12 16-00-48\011-0401.D Sample Name: HW212835-011R1

Acq. Operator : HW Seq. Line : 4
Acq. Instrument : HP1100-4 Location : Vial 11
Injection Date : 7/12/2011 5:52:33 PM Inj : 1
Inj Volume : 2 µl

Acq. Method : C:\CHEM32\1\DATA\12JUL2011\HIVINTEGRASEAMIDE 2011-07-12 16-00-48\

HIVINTEGRASEAMIDE.M

Last changed : 12/23/2009 2:44:03 PM

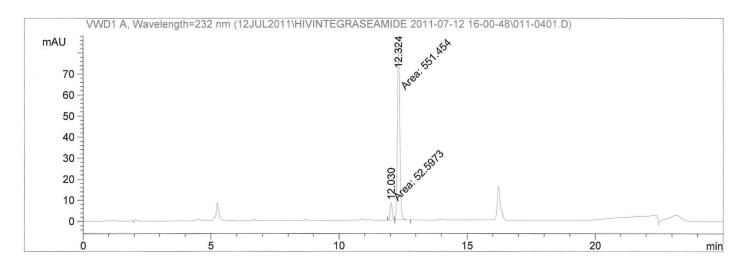
Analysis Method: C:\CHEM32\1\DATA\12JUL2011\HIVINTEGRASEAMIDE 2011-07-12 16-00-48\

011-0401.D\DA.M (HIVINTEGRASEAMIDE.M)

Last changed : 12/23/2009 2:44:03 PM

Method Info : for HIV Integrase amide isomers

Sample Info : no additive 60C 16.5h



Area Percent Report

Sorted By : Signal

Multiplier: : 1.0000
Dilution: : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: VWD1 A, Wavelength=232 nm

Peak	RetTime	Type	Width	Ar	ea	Hei	ght	Area	
#	[min]		[min]	mAU	*s	[mAU]	90	
1	12.030	MF	0.1062	52.	59728	8.2	25123	8.7074	
2	12.324	FM	0.1103	551.	45410	83.3	32913	91.2926	

Totals: 604.05138 91.58036

Data File C:\CHEM32\2\DATA\18SEP09\HIVINTEGRASE2 2009-09-18 13-26-33\012-0401.D Sample Name: HW211473-001R2

 Acq. Operator
 : CY
 Seq. Line : 4

 Acq. Instrument
 : UM28-009
 Location : Vial 12

 Injection Date
 : 9/18/2009 2:47:07 PM
 Inj : 1

Inj Volume : 2 µl

Acq. Method : C:\CHEM32\2\DATA\18SEP09\HIVINTEGRASE2 2009-09-18 13-26-33\

HIVINTEGRASE2.M

Last changed : 9/18/2009 11:23:52 AM by CY

Analysis Method: C:\CHEM32\2\DATA\18SEP09\HIVINTEGRASE2 2009-09-18 13-26-33\001-0101.

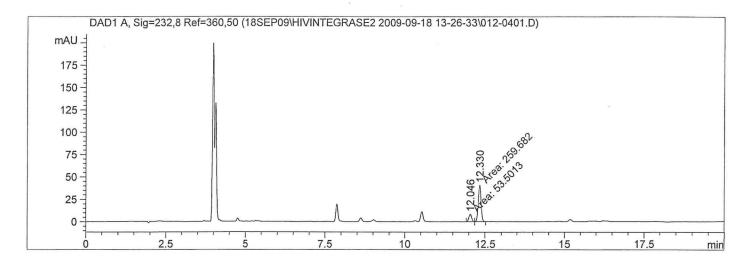
D\DA.M (HIVINTEGRASE2.M)

Last changed : 9/19/2009 8:24:10 AM by CY

(modified after loading)

Method Info : HIV Integrasr _OMe isomers IPM

Sample Info : 12h MsOH



Area Percent Report

Sorted By : Signal

Multiplier: : 1.0000 Dilution: : 1.0000

Do not use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=232,8 Ref=360,50

Peak	RetTime	Type	Width	Area	Height	Area	
#	[min]		[min]	[mAU*s]	[mAU]	90	
							ĺ
1	12.046	MF	0.1056	53.50127	8.44706	17.0830	
2	12.330	FM	0.1057	259.68240	40.95332	82.9170	

Totals: 313.18367 49.40037

Data File C:\CHEM32\1\DATA\12JUL2011\HIVINTEGRASEAMIDE 2011-07-12 16-00-48\020-1301.D Sample Name: HW212835-011R10

Acq. Operator : HW Seq. Line : 13
Acq. Instrument : HP1100-4 Location : Vial 20
Injection Date : 7/12/2011 9:49:26 PM Inj : 1
Inj Volume : 2 µl

Acq. Method : C:\CHEM32\1\DATA\12JUL2011\HIVINTEGRASEAMIDE 2011-07-12 16-00-48\

HIVINTEGRASEAMIDE.M

Last changed : 12/23/2009 2:44:03 PM

Analysis Method: C:\CHEM32\1\DATA\12JUL2011\HIVINTEGRASEAMIDE 2011-07-12 16-00-48\

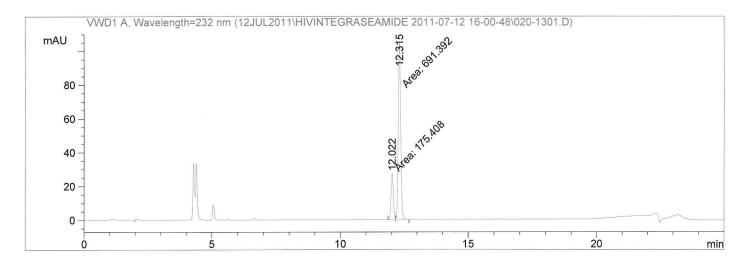
020-1301.D\DA.M (HIVINTEGRASEAMIDE.M)

Last changed : 6/7/2012 8:26:05 AM by LMO

(modified after loading)

Method Info : for HIV Integrase amide isomers

Sample Info : tfa 60C 16.5h



Area Percent Report

Sorted By : Signal

Multiplier: : 1.0000
Dilution: : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: VWD1 A, Wavelength=232 nm

Peak	RetTime	Type	Width	Ar	rea	Hei	ght	Area
#	[min]		[min]	mAU	*s	[mAU]	િ
1	12.022	MF	0.1080	175.	40779	27.	07558	20.2363
2	12.315	FM	0.1101	691.	39172	104.	67611	79.7637

Totals: 866.79951 131.75169

Data File C:\CHEM32\1\DATA\12JUL2011\HIVINTEGRASEAMIDE 2011-07-12 16-00-48\019-1201.D Sample Name: HW212835-011R9

Acq. Operator : HW Seq. Line : 12
Acq. Instrument : HP1100-4 Location : Vial 19
Injection Date : 7/12/2011 9:23:07 PM Inj : 1

Inj Volume : 2 μl

Acq. Method : C:\CHEM32\1\DATA\12JUL2011\HIVINTEGRASEAMIDE 2011-07-12 16-00-48\

HIVINTEGRASEAMIDE.M

Last changed : 12/23/2009 2:44:03 PM

Analysis Method: C:\CHEM32\1\DATA\12JUL2011\HIVINTEGRASEAMIDE 2011-07-12 16-00-48\

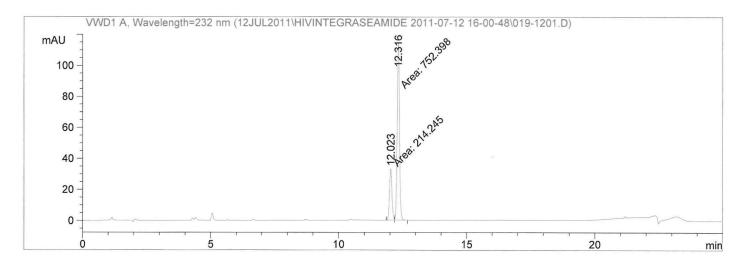
019-1201.D\DA.M (HIVINTEGRASEAMIDE.M)

Last changed : 6/7/2012 9:13:04 AM by LMO

(modified after loading)

Method Info : for HIV Integrase amide isomers

Sample Info : CHCl2COOH 60C 16.5h



Sorted By : Signal

Multiplier: : 1.0000
Dilution: : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: VWD1 A, Wavelength=232 nm

Peak	RetTime	Type	Width	A	rea	Heig	ght	Area	
#	[min]		[min]	mAU	*s	[mAU]	%	
1	12.023	MF	0.1077	214	.24475	33.3	14441	22.1638	
2	12.316	FM	0.1100	752	.39801	113.9	94874	77.8362	

Totals: 966.64276 147.09314

Data File C:\CHEM32\1\DATA\12JUL2011\HIVINTEGRASEAMIDE 2011-07-12 16-00-48\017-1001.D Sample Name: HW212835-011R7

Acq. Operator : HW Seq. Line : 10
Acq. Instrument : HP1100-4 Location : Vial 17
Injection Date : 7/12/2011 8:30:25 PM Inj : 1
Inj Volume : 2 µl

Acq. Method : C:\CHEM32\1\DATA\12JUL2011\HIVINTEGRASEAMIDE 2011-07-12 16-00-48\

HIVINTEGRASEAMIDE.M

Last changed : 12/23/2009 2:44:03 PM

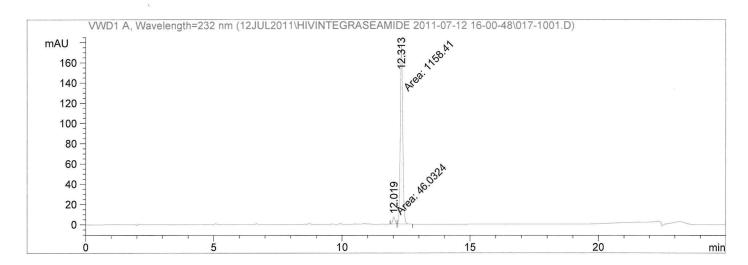
Analysis Method: C:\CHEM32\1\DATA\12JUL2011\HIVINTEGRASEAMIDE 2011-07-12 16-00-48\

017-1001.D\DA.M (HIVINTEGRASEAMIDE.M)

Last changed : 12/23/2009 2:44:03 PM

Method Info : for HIV Integrase amide isomers

Sample Info : HOAc 60C 16.5h



Area Percent Report

Sorted By : Signal

Multiplier: : 1.0000
Dilution: : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: VWD1 A, Wavelength=232 nm

Peak	RetTime	Type	Width	Ar	ea	Heig	ght	Area	
#	[min]		[min]	mAU	*s	[mAU]	90	
									1
1	12.019	MF	0.1063	46.	03238	7.2	1466	3.8219	
2	12.313	FM	0.1097	1158.	41455	175.9	4485	96.1781	

Totals: 1204.44693 183.15952

Data File C:\CHEM32\1\DATA\12JUL2011\HIVINTEGRASEAMIDE 2011-07-12 16-00-48\016-0901.D Sample Name: HW212835-011R6

Acq. Operator : HW Seq. Line : 9
Acq. Instrument : HP1100-4 Location : Vial 16
Injection Date : 7/12/2011 8:04:05 PM Inj : 1
Inj Volume : 2 µl

Acq. Method : C:\CHEM32\1\DATA\12JUL2011\HIVINTEGRASEAMIDE 2011-07-12 16-00-48\

HIVINTEGRASEAMIDE.M

Last changed : 12/23/2009 2:44:03 PM

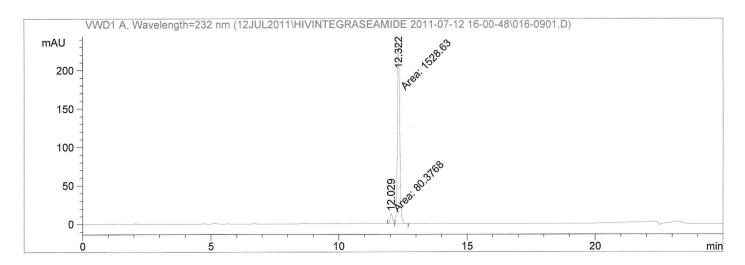
Analysis Method: C:\CHEM32\1\DATA\12JUL2011\HIVINTEGRASEAMIDE 2011-07-12 16-00-48\

016-0901.D\DA.M (HIVINTEGRASEAMIDE.M)

Last changed : 12/23/2009 2:44:03 PM

Method Info : for HIV Integrase amide isomers

Sample Info : IBA 60C 16.5h



Area Percent Report

Sorted By : Signal

Multiplier: : 1.0000
Dilution: : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: VWD1 A, Wavelength=232 nm

Peak	RetTime	Type	Width	Ar	cea	Hei	ght	Area
#	[min]		[min]	mAU	*s	[mAU]	90
								[
1	12.029	MF	0.1065	80.	.37677	12.	57614	4.9954
2	12.322	FM	0.1097	1528.	.63416	232.	32300	95.0046

Totals: 1609.01093 244.89914

Data File C:\CHEM32\1\DATA\12JUL2011\HIVINTEGRASEAMIDE 2011-07-12 16-00-48\012-0501.D Sample Name: HW212835-011R2

Acq. Operator : HW Seq. Line : 5
Acq. Instrument : HP1100-4 Location : Vial 12
Injection Date : 7/12/2011 6:18:52 PM Inj : 1
Inj Volume : 2 µl

Acq. Method : C:\CHEM32\1\DATA\12JUL2011\HIVINTEGRASEAMIDE 2011-07-12 16-00-48\

HIVINTEGRASEAMIDE.M

Last changed : 12/23/2009 2:44:03 PM

Analysis Method: C:\CHEM32\1\DATA\12JUL2011\HIVINTEGRASEAMIDE 2011-07-12 16-00-48\

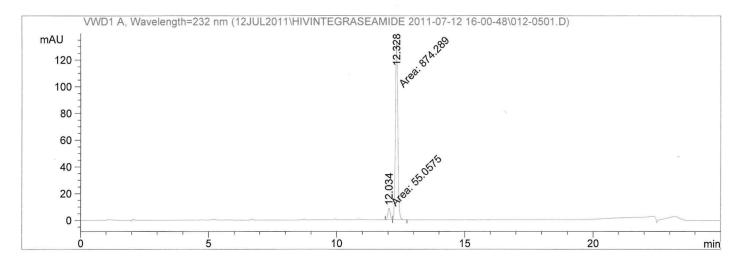
012-0501.D\DA.M (HIVINTEGRASEAMIDE.M)

Last changed : 6/7/2012 9:18:47 AM by LMO

(modified after loading)

Method Info : for HIV Integrase amide isomers

Sample Info : pivalic acid 60C 16.5h



Area Percent Report

Sorted By : Signal

Multiplier: : 1.0000
Dilution: : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: VWD1 A, Wavelength=232 nm

Peak	RetTime	Type	Width	Ar	ea	Heig	ght	Area
#	[min]		[min]	mAU	*s	[mAU]	8
1	12.034	MF	0.1066	55.	05747	8.6	50756	5.9243
2	12.328	FM	0.1097	874.	28882	132.8	31303	94.0757

Totals: 929.34629 141.42060

Data File C:\CHEM32\1\DATA\12JUL2011\HIVINTEGRASEAMIDE 2011-07-12 16-00-48\015-0801.D Sample Name: HW212835-011R5

Seq. Line: 8 Acq. Operator : HW Acq. Instrument: HP1100-4 Location : Vial 15

HIVINTEGRASEAMIDE.M

Injection Date : 7/12/2011 7:37:47 PM

Inj: 1 Inj Volume : 2 μl

Acq. Method : C:\CHEM32\1\DATA\12JUL2011\HIVINTEGRASEAMIDE 2011-07-12 16-00-48\

Last changed : 12/23/2009 2:44:03 PM

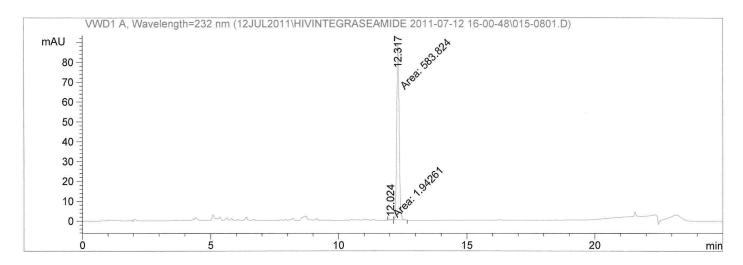
Analysis Method: C:\CHEM32\1\DATA\12JUL2011\HIVINTEGRASEAMIDE 2011-07-12 16-00-48\

015-0801.D\DA.M (HIVINTEGRASEAMIDE.M)

Last changed : 12/23/2009 2:44:03 PM

Method Info : for HIV Integrase amide isomers

Sample Info : Mg(OTf)2 60C 16.5h



Area Percent Report

Sorted By Signal

Multiplier: : 1.0000 Dilution: Use Multiplier & Dilution Factor with ISTDs

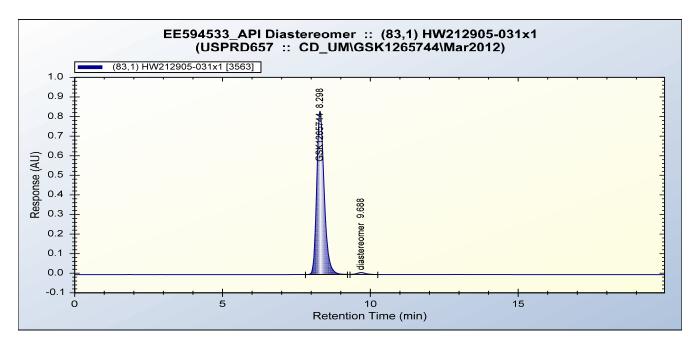
Signal 1: VWD1 A, Wavelength=232 nm

	RetTime		Width	Area	Height	Area
#	[min]		[min]	mAU *s	[mAU]	90
1	12.024	MF	0.1030	1.94261	3.14408e-1	0.3316
2	12.317	FM	0.1098	583.82373	88.58716	99.6684

Totals: 585.76634 88.90157

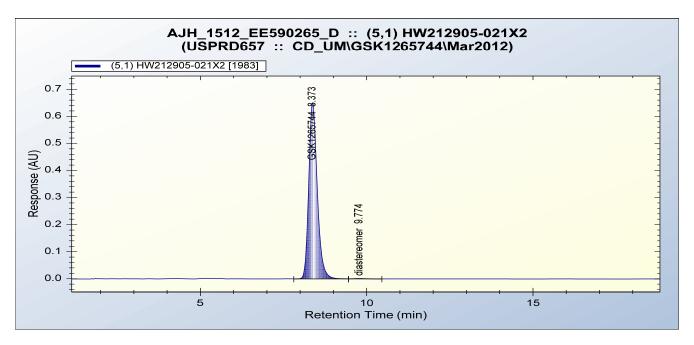
The diastereomeric excess for the pair 1a/1b was measured by integrations of HPLC peaks. HPLC: Agilent 1100 series; column: Chiralcel OD-RH; flow rate: 1 mL/min; solvent: 0.1% TFA in water - CH₃CN (isocratic, 60:40). t_R: 1a 8.3 min, 1b 9.7 min.

The HPLC trace of a typical batch of **1a** prepared by the sequence in Scheme 2:



RT	Name	Height	Area	Area %
8.298	GSK1265744	833004.089	14704851.135	98.705
9.688	diastereomer	9600.641	192879.920	1.295

The HPLC trace of a typical batch of **1a** prepared by the sequence in Scheme 4:



RT	Name	Height	Area	Area %
8.373	GSK1265744	650993.420	11650529.173	99.664
9.774	diastereomer	1538.878	39264.200	0.336

Computational Methods:

To calculate various energetic states of the final product 13 and the isomers of intermediate 16, the bound bioactive conformation of related drug Dolutegravir, as determined by crystallographic structure 3S3M (PDB code), was used as a strarting template conformation. The Jaguar software suite distributed with Maestro v9.3.5 was used to run *ab initio* density functional energy calculations. The B3LYP/lacvp** basis set was used for full structural optimization. The energy difference between the two diastereomers of 13 by DFT calculation (B3LYP/lacvp**) is 0.7 kcal/mol favoring the desired trans isomer (13a); and, the cis isomer 16b is more stable by 1.0 kcal/mol by DFT calculation (B3LYP/lacvp**).

Jaguar input command file:

&gen basis=lacvp** igeopt=1 dftname=b3lyp & &zmat

Optimized XYZ coordinate files:

16a_trans

3D

Structure	written by	MMmdl.						
55 57 0	0 1 0	9	99 7	<i>7</i> 2000				
-1.5377	-1.4712	2.1446	С	0 0	0	0	0	0
-1.3380	-2.1879	0.9123	0	0 0	0	0	0	0
-2.2933	-3.0722	0.5276	С	0 0	0	0	0	0
-1.8670	-4.3220	0.1465	С	0 0	0	0	0	0
-0.3901	-4.6251	0.1578	С	0 0	0	0	0	0
-0.0798	-5.5717	1.0606	0	0 0	0	0	0	0
1.3067	-5.9763	1.0645	С	0 0	0	0	0	0
0.4116	-4.0748	-0.5614	0	0 0	0	0	0	0
-0.5393	-6.2710	-2.5141	N	0 0	0	0	0	0
-0.1364	-6.9931	-3.7313	С	0 0	2	0	0	0
-0.5271	-6.3096	-5.0495	С	0 0	0	0	0	0
-0.8805	-8.3138	-3.4869	С	0 0	0	0	0	0
-2.0657	-7.9464	-2.7647	0	0 0	0	0	0	0
-1.9399	-6.6127	-2.2940	C	0 0	1	0	0	0
-2.3431	-6.6054	-0.8064	С	0 0	0	0	0	0
-2.7630	-5.2783	-0.3124	N	0 0	0	0	0	0
-4.0779	-4.9542	-0.3919	C	0 0	0	0	0	0
-4.5852	-3.7317	-0.0390	C	0 0	0	0	0	0
-6.0716	-3.5828	-0.2341	C	0 0	0	0	0	0
-6.7565	-4.5257	-0.6388	0	0 0	0	0	0	0
-6.5678	-2.3524	0.0570	N	0 0	0	0	0	0
-7.9690	-2.0343	-0.1136	С	0 0	0	0	0	0
-8.7193	-1.8300	1.1924	C	0 0	0	0	0	0
-8.2390	-2.2501	2.4353	С	0 0	0	0	0	0
-8.9729	-2.0646	3.6082	C	0 0	0	0	0	0
-10.2123	-1.4446	3.5241	C	0 0	0	0	0	0
-10.9348	-1.2580		F	0 0	0	0	0	0
-10.7382	-1.0048	2.3148	C	0 0	0	0	0	0
-9.9711	-1.2126	1.1771	C	0 0	0	0	0	0
-10.4648	-0.7943	-0.0127	F	0 0	0	0	0	0
-3.7075	-2.6784	0.110	C	0 0	0	0	0	0
-4.0827	-1.5264	0.7542	0	0 0	0	0	0	0
-0.6201	-0.8998	2.2940	H	0 0	0	0	0	0

```
0
                                                             0
  -1.6666
              -2.1729
                            2.9779 H
                                          0
                                              0
                                                  0
                                                     0
                                              0
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                                                      0
                                                         0
  -2.4006
              -0.8102
                            2.0761 H
                                          0
                                                             0
   1.3851
               -6.7368
                            1.8406 H
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               -5.1233
                            1.2894 H
                                              0
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                                                     0
                                                         0
                                                             0
                                          0
   1.5736
               -6.3847
                            0.0870 H
                                              0
                                                      0
                                                         0
                                                             0
  -0.3712
               -5.2700
                           -2.5655 H
                                          0
                                              0
                                                  0
                                                      0
                                                         0
                                                             0
   0.9483
              -7.1497
                           -3.7105 H
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                                                  0
                                                     0
                                                         0
                                                             0
  -1.6129
              -6.1961
                           -5.1362 H
                                          0
                                              0
                                                  0
                                                     0
                                                         0
                                                             0
  -0.1820
               -6.8951
                           -5.9088 H
                                          0
                                              0
                                                  0
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                                                         0
                                                             0
               -5.3155
                           -5.1212 H
                                              0
                                                  0
                                                     0
                                                         0
                                                             0
  -0.0727
                                          0
  -0.2814
              -9.0059
                           -2.8821 H
                                              0
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                                          0
  -1.1774
              -8.8170
                           -4.4127 H
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                           -2.8481 H
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  -2.6497
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  -1.5222
              -6.9821
                           -0.1982 H
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  -3.1981
               -7.2731
                           -0.6906 H
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  -4.7560
               -5.7148
                           -0.7621 H
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  -5.8912
               -1.6579
                            0.3804 H
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                           -0.7300 H
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M END
> <s_j_Task>
Optimization
> <s_j_QM_Basis>
lacvp**
> <s_j_QM_Method>
DFT(b3lyp)
> <i_j_Number_of_canonical_orbitals>
562
> <r_j_Gas_Phase_Energy>
-1629.34808634685
> <r j HOMO>
-0.225303439580083
> <r_j_LUMO>
-0.0526456820245516
$$$$
16b_cis
                    3D
 Structure written by MMmdl.
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           -1.4620
                        2.2271 C
                                   0
                                      0
                                          0 0 0
                                                   0
   -1.2404
            -2.1397
                        0.9714 0
                                   0
                                      0
                                          0 0 0
                                                   0
   -2.2103
             -2.9995
                        0.5627 C
                                   0
                                      0
                                          0
                                            0 0
                                                   0
                        0.1572 C
                                      0
                                         0
                                            0 0
   -1.8007
             -4.2450
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-0.3267

-4.5651

0

0 0 0

0 0

0.1588 C

-0.0071 1.3787 0.4630		-5.4554 -5.8648 -4.0748	1.1120 1.1232 -0.6157	0 C 0	0 0 0	0 0 0	0 0 0	0 0 0	0 0 0	0 0 0
-0.5040 -0.2018		-6.2525 -6.9300	-2.5486 -3.8161	N C	0	0 0	0 1	0 0	0	0
1.2948 -0.9549		-7.0824 -8.2460	-4.0573 -3.5792	C C	0	0 0	0 0	0 0	0 0	0
-2.0628 -1.9051		-7.8979 -6.5549	-2.7364	О С	0	0 0	0 1	0 0	0	0
-2.2955		-6.5139	-2.2977 -0.8100	C	0	0	0	0	0 0	0
-2.7026 -4.0112		-5.1767 -4.8297	-0.3345 -0.4278	N C	0	0	0	0	0	0
-4.5029		-3.6068	-0.0542	С	0	0	0	0	0	0
-5.9811 -6.6708		-3.4220 -4.3289	-0.2800 -0.7526	С О	0	0	0	0	0	0 0
-6.4659		-2.2009	0.0640	N	0	0	0	0	0	0
-7.8483 -8.7127		-1.8389 -1.8340	-0.1654 1.0857	C	0	0	0	0	0	0 0
-8.3417		-2.4383	2.2890	С	0	0	0	0	0	0
-9.1813 -10.4163		-2.4321 -1.8054	3.4045 3.3029	C	0	0	0 0	0 0	0	0 0
-11.2399		-1.7907	4.3712	F	0	0	0	0	0	0
-10.8360 -9.9662		-1.1870 -1.2212	2.1304 1.0495	C	0	0	0 0	0 0	0 0	0 0
-10.3554 -3.6164		-0.6277 -2.5822	-0.1044 0.4752	F C	0	0	0	0	0	0
-3.9793		-1.4356	0.8182	Ο	0	0	0	0	0	0
-0.5144 -1.5513		-0.8920 -2.1900	2.3871 3.0389	H H	0	0	0	0	0	0 0
-2.2965		-0.8017	2.1853	Н	0	0	0	0	0	0
1.4611 2.0256		-6.5875 -5.0037	1.9341 1.3026	H H	0	0 0	0	0 0	0 0	0 0
1.6373 -0.3041		-6.3195	0.1642	Н	0	0	0	0	0	0
-0.3041 -0.6587		-5.2556 -6.4112	-2.5660 -4.6787	H H	0	0 0	0 0	0 0	0 0	0
1.7671 1.7737		-7.5944 -6.1054	-3.2133 -4.1762	H H	0	0 0	0	0 0	0	0
1.4844		-7.6573	-4.9700	Н	0	0	0	0	0	0
-0.3190 -1.3329		-8.9775 -8.6938	-3.0629 -4.5042	H H	0	0	0	0 0	0	0 0
-2.6023		-5.9053	-2.8650	Н	0	0	0	0	0	0
-1.4721 -3.1514		-6.8856 -7.1769	-0.2010 -0.6742	H H	0	0 0	0	0 0	0 0	0 0
-4.6951 -5.7850		-5.5705 -1.5330	-0.8266 0.4310	Н	0	0 0	0 0	0 0	0 0	0
-7.8889		-0.8499	-0.6337	H H	0	0	0	0	0	0
-8.2532 -7.3714		-2.5562 -2.9197	-0.8859 2.3501	H H	0	0	0	0	0	0
-8.8919		-2.9022	4.3377	Н	0	0	0	0	0	0
-11.8007 1 2 1	0	-0.6994 0 0	2.0599	Η	0	0	0	0	0	0
1 33 1 1 34 1	0	0 0 0 0								
1 35 1	0	0 0								
2 3 1 3 4 2	0	0 0								

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3 31
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  4 16
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Μ
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> <s_j_Task>
Optimization

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> <s_j_QM_Basis>
lacvp**
> <s_j_QM_Method>
DFT(b3lyp)
> <i_j_Number_of_canonical_orbitals>
562
> <r_j_Gas_Phase_Energy>
-1629.34969116351
> <r_j_HOMO>
-0.226462171532978
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> <r_j_LUMO>
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-0.0503860678562342

\$\$\$\$

13a_trans

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1.5917 H

0.0164

-1.5330

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0

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                           -0.5771 H
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                            2.4667 H
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  -7.4398
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                           -0.2758 H
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 1 32
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M END
> <s_j_Task>
Optimization
```

> <s_j_QM_Basis> lacvp**

> <s_j_QM_Method> DFT(b3lyp)

> <i_j_Number_of_canonical_orbitals> 514

- > <r_j_Gas_Phase_Energy> -1513.62603483402
- > <r_j_HOMO> -0.224289255692887
- > <r_j_LUMO> -0.0645261793384926

\$\$\$\$ 13b_cis

3D

Structure written by MMmdl. 49 52 0 0 1 0 999 V2000 -0.9530 -1.44252.0943 C 0 0 0 0 0 0 0.9682 0 -0.7156-2.3081 0 0 0 0 0 0 -1.6825 -3.1110 0.5011 C 0 0 0 0 0 0 -1.3239-4.3628 0.0304 C 0 0 0 0 0 0 0 0 0 0 0.0975 -4.8691 0.0634 C 0 0 0 0 1.0754 -4.1374 0.1304 0 0 0 0 -6.2368 0.0178 N 0 0 0 0 0 0.2145 0 -6.9524 0 0 0 1.4876 -0.1734 C 0 0 0 0 0 0 2.1963 -6.5737 -1.4772 C 0 0 0.9627 -8.4025 -0.1664 C 0 0 0 0 0 0 -0.7069 O -0.3653 -8.3063 0 0 0 0 0 2 0 0 -0.9168 -7.1471 -0.1391 C 0 0 -1.9338 -6.5078 -1.0658 C 0 0 0 0 0 0 -2.3122 -5.2077 -0.4914 N0 0 0 0 0 0 0 0 -3.6048 -4.8200-0.5277 C 0 -0.0904 C 0 0 0 0 0 -4.0342-3.5949 0

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0
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                           -0.2311 C
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                           -0.6302 O
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              -2.1001
                            0.0912 N
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  -7.3269
              -1.7121
                           -0.0084 C
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              -1.9630
                            1.2600 C
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Optimization
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> <s_j_QM_Method>
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> <r_j_Gas_Phase_Energy>

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> <r_j_HOMO>

> <r_j_LUMO>

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