

Cobalt-Catalyzed Regio- and Enantioselective Allylic Alkylation of Malononitriles

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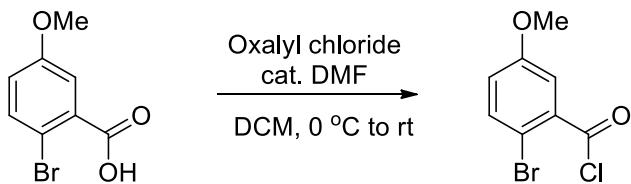
1. General information:

All air and moisture sensitive manipulations were carried out with standard Schlenk technique or in a nitrogen-filled glove box. Dried and oxygen free solvents were obtained from solvent purification system (Vigor YJC-7) and used thereafter. Column chromatography was performed on a column of silica gels (200-300 mesh) using either petroleum ether (PE)/ethyl acetate (EA) or dichloromethane (DCM)/MeOH as an eluent. The NMR spectra were recorded on a Bruker-400/500 MHz instrument and chemical shifts are reported in ppm relative to the residual deuterated solvents. High-resolution mass spectra (HRMS) were performed at Instrumental Analysis Center of Shanghai Jiao Tong University with electrospray spectrometer Waters Micromass Q-TOF Premier Mass Spectrometer. Enantiomers ratio were determined by Shimadzu HPLC-16 and Shimadzu HPLC-2030 instrument using DAICEL chiral column at 40 °C. Melting points were measured with Hanon MP100 melting point apparatus. Optical rotations were measured on an Anton Paar MCP100 automatic polarimeter using a 100 mm path-length cell at 589 nm. $\text{Co}(\text{BF}_4)_2 \cdot 6\text{H}_2\text{O}$ was purchased from Sigma-Aldrich. All other chemicals were purchased from TCI, Energy Chemical, Macklin or Alfa Aesar. Magnesium monoperoxyphthalate hexahydrate (MMPP) 80% analytical grade was obtained from Bide Pharmatech, China. Solvents were obtained from tansoole, China. Liquid substrates were distilled and degassed prior to use.

2. Synthesis of (S)-2-(2-bromo-5-methoxyphenyl)-4-methyl-4,5-dihydrooxazole (LS1):

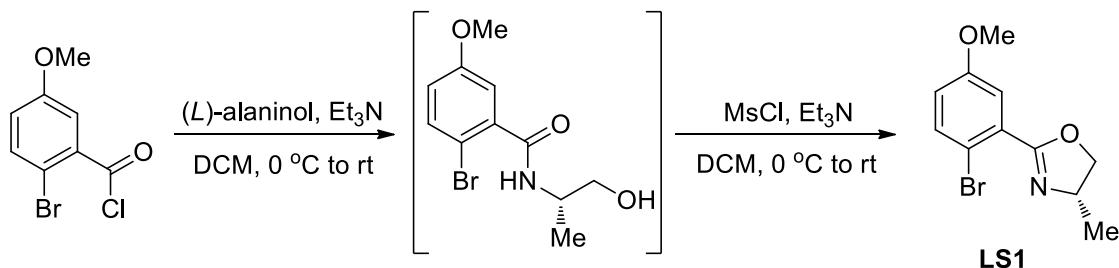
LS1 compound was prepared in two steps as below:

Step 1:



A suspension of 2-bromo-5-methoxybenzoic acid (4.620 g, 20 mmol) in DCM (40 mL) was carefully treated with oxalyl chloride (5.080 g, 40 mmol, 2 eq) in the presence of 2 drops of DMF at 0 °C under inert atmosphere. After stirring at room temperature for 4-5 hours, a clear solution was obtained. The reaction mixture was concentrated under vacuo to get the corresponding benzoyl chloride as light yellowish liquid. The crude product was directly used in the next step.

Step 2:



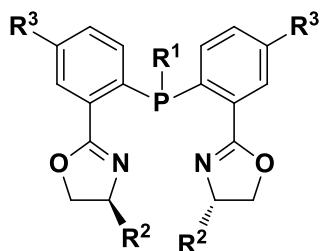
To a solution of (L)-alaninol (1.577 g, 21 mmol) and Et₃N (8.8 mL, 3 eq) in DCM (40 mL) was added 2-bromo-5-methoxybenzoyl chloride (diluted in 10 mL of DCM) dropwise at 0 °C. The reaction mixture was stirred at room temperature for 12 hours. After that, it was cooled at 0 °C. Methanesulfonyl chloride (MsCl) (3.3 mL, 2 eq) and Et₃N (8.8 mL, 3 eq) were added carefully. The reaction mixture was stirred at room temperature for 4-5 hours. Then, it was washed with water twice and with brine twice. Organic layer was dried by Na₂SO₄, and concentrated. The residue was purified by column chromatography (PE/EA, 6:1) to get **LS1** as a colorless oil (4.590 g, 85%).

¹H NMR (500 MHz, CDCl₃) δ 7.47 (d, *J* = 8.8 Hz, 1H), 7.20 (d, *J* = 3.1 Hz, 1H), 6.82 (dd, *J* = 8.8, 3.1 Hz, 1H), 4.51 (dd, *J* = 9.4, 8.0 Hz, 1H), 4.39 (tt, *J* = 13.4, 6.6 Hz, 1H), 3.97 (t, *J* = 7.8 Hz, 1H), 3.78 (s, 3H), 1.37 (d, *J* = 6.6 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 162.9, 158.7, 134.7, 130.7, 118.5, 116.2, 112.3, 74.4, 62.5, 55.8, 21.6.

HRMS (ESI): calculated *m/z* for C₁₁H₁₂BrNO₂ [M + H]⁺ = 270.0124, found 270.0131.

3. General procedure for the ligand synthesis:

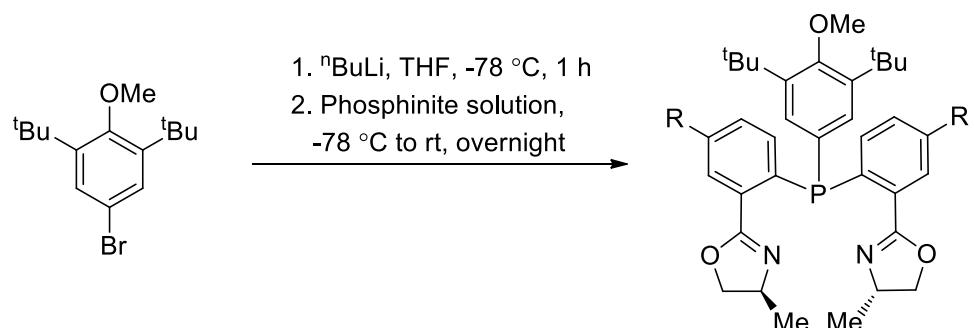
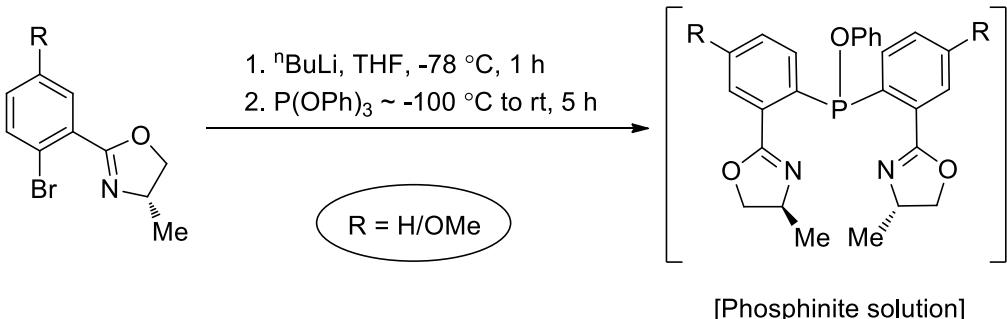


L1 [$R^1 = \text{Ph}$, $R^2 = \text{Ph}$, $R^3 = \text{H}$]; **L2** [$R^1 = \text{Ph}$, $R^2 = \text{iPr}$, $R^3 = \text{H}$]; **L3** [$R^1 = \text{Ph}$, $R^2 = \text{tBu}$, $R^3 = \text{H}$]; **L4** [$R^1 = \text{Ph}$, $R^2 = \text{Bn}$, $R^3 = \text{H}$]; **L5** [$R^1 = \text{Ph}$, $R^2 = \text{Me}$, $R^3 = \text{H}$] were synthesized according to the reported literature procedure.¹

L6 [$R^1 = 4\text{-OMe-3,5-}^{\text{t}}\text{Bu}_2\text{C}_6\text{H}_2$, $R^2 = \text{Me}$, $R^3 = \text{H}$], and **L7** [$R^1 = 4\text{-OMe-3,5-}^{\text{t}}\text{Bu}_2\text{C}_6\text{H}_2$, $R^2 = \text{Me}$, $R^3 = \text{OMe}$] were prepared according to the procedure **General Method 1**, reported in literature.¹

L8 [$R^1 = \text{Me}$, $R^2 = \text{Me}$, $R^3 = \text{H}$]; **L9** [$R^1 = \text{Me}$, $R^2 = \text{Me}$, $R^3 = \text{OMe}$] were prepared according to the procedure **General Method 2**.

General Method 1:

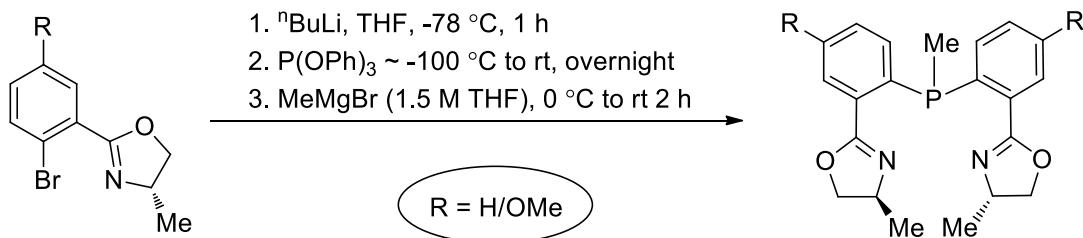


In a flame dried Schlenk tube, 2-bromooxazoline substrate (6 mmol, 2.0 eq) was dissolved in THF (12 mL) under an argon atmosphere and the reaction mixture was cooled down to $-78\text{ }^{\circ}\text{C}$. ${}^{\text{n}}\text{BuLi}$ (2.2 eq, 1.6 M in hexane) was added dropwise and the mixture was stirred for 1 hour at this temperature. The reaction mixture was further

cooled to ~ -100 °C and was added triphenylphosphite (0.931 g, 3 mmol, 1 eq) in 2 mL of THF in one portion under vigorous stirring. The reaction was slowly warmed up to room temperature and stirred for 5 hours. Phosphinite reaction mixture was used *in situ* for further substitution to get the desired ligand.

In another flame dried Schlenk tube, lithiation of the 5-bromo-1,3-di-*tert*-butyl-2-methoxybenzene (1.077 g, 3.6 mmol, 1.2 eq) was carried out using $^n\text{BuLi}$ (1.32 eq, 1.6 M in hexane) at -78 °C. Then the reaction mixture was added the phosphinite solution via syringe over 10 minutes at -78 °C. Combined reaction mixture was allowed to warm to room temperature slowly and stirred overnight. The reaction mixture was quenched with water and extracted with ethyl acetate. The solvent was removed and the residue was purified by flash column chromatography on silica gels using PE/EA as an eluent to obtain the pure product.

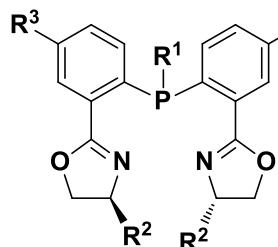
General Method 2:



In a flame dried Schlenk tube, 2-bromooxazoline substrate (6 mmol, 2 eq) was dissolved in THF (12 mL) under an argon atmosphere and cooled down to -78 °C. $^n\text{BuLi}$ (2.2 eq, 1.6 M in hexane) was added dropwise and the mixture was stirred for 1 hour at this temperature. The reaction mixture was further cooled down to ~ -100 °C and was added triphenylphosphite (0.931 g, 3 mmol, 1 eq) in 2 mL of THF in one portion under vigorous stirring. The reaction was slowly warmed up to room temperature and stirred overnight. Then, 1.5 mL of MeMgBr (3 M in THF) was added dropwise to the reaction mixture at 0 °C. The ice-bath was removed and the reaction mixture was stirred for two hours further at room temperature. The reaction mixture was quenched with 2 mL of degassed water and filtered to remove the solid. Filtrate was dried over Na_2SO_4 and solvent was removed under reduced pressure. Crude product was purified by flash column chromatography on silica gels using PE/EA as an eluent to obtain the pure product.

L6 [R¹ = 4-OMe-3,5-tBu₂C₆H₂, R² = Me, R³ = H]:

White solid; Yield 0.670 g, 39%.



R¹ = 4-OMe-3,5-tBu₂C₆H₂
R² = Me; R³ = H

¹H NMR (400 MHz, CDCl₃) δ 7.82 (dd, *J* = 5.5, 3.1 Hz, 2H), 7.34 – 7.24 (m, 6H), 7.01 – 6.93 (m, 2H), 4.34 – 4.30 (m, 1H), 4.27 – 4.15 (m, 3H), 3.75 (t, *J* = 7.6 Hz, 1H), 3.68 (s, 3H), 3.64 (t, *J* = 7.7 Hz, 1H), 1.33 (s, 18H), 1.10 (d, *J* = 6.5 Hz, 3H), 1.06 (d, *J* = 6.4 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 164.1 (d, *J* = 9.5 Hz), 160.3 (s), 143.4 (d, *J* = 8.0 Hz), 140.6 (dd, *J* = 24.9, 14.1 Hz), 133.9 (d, *J* = 5.8 Hz), 133.6 (d, *J* = 13.0 Hz), 132.4 (dd, *J* = 82.0, 22.0 Hz), 130.8 (d, *J* = 10.6 Hz), 130.3 (d, *J* = 13.7 Hz), 129.9 (dd, *J* = 16.5, 3.7 Hz), 127.8 (d, *J* = 22.0 Hz), 73.9 (d, *J* = 4.5 Hz), 64.4 (s), 62.1 (d, *J* = 18.9 Hz), 36.0 (s), 32.2 (s), 21.29 (d, *J* = 16.9 Hz).

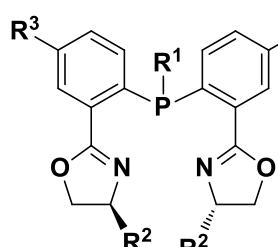
³¹P NMR (162 MHz, CDCl₃) δ -5.70.

[α]_D²⁵ = -26.0 (c 0.25, CHCl₃).

HRMS (ESI): *m/z* [M + H]⁺ Calcd for C₃₅H₄₄N₂O₃P 571.3084; found 571.3094.

L7 [R¹ = 4-OMe-3,5-tBu₂C₆H₂, R² = Me, R³ = OMe]:

White solid; Yield 0.700 g, 37%.



R¹ = 4-OMe-3,5-tBu₂C₆H₂
R² = Me; R³ = OMe

¹H NMR (500 MHz, CDCl₃) δ 7.35 (dt, *J* = 8.5, 2.6 Hz, 2H), 7.25 (d, *J* = 8.0 Hz, 2H), 6.92 – 6.83 (m, 4H), 4.34 (dd, *J* = 9.2, 8.1 Hz, 1H), 4.27 – 4.15 (m, 3H), 3.82 (s, 6H), 3.77 (t, *J* = 7.7 Hz, 1H), 3.70 – 3.67 (m, 4H), 1.33 (s, 18H), 1.15 (d, *J* = 6.6 Hz, 3H), 1.08 (d, *J* = 6.5 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 164.4 (s), 160.2 (s), 159.3 (d, *J* = 15.7 Hz), 143.4 (d, *J* = 7.8 Hz), 135.4 (d, *J* = 44.3 Hz), 134.3 – 133.4 (m), 133.3 (d, *J* = 24.1 Hz), 131.8 – 131.7 (m), 131.7 – 131.5 (m), 116.9 (d, *J* = 7.0 Hz), 114.7 (dd, *J* = 8.3, 4.4 Hz), 74.0 (d, *J* = 6.8 Hz), 64.4 (s), 62.1 (d, *J* = 15.6 Hz), 55.6 (d, *J* = 1.8 Hz),

36.0 (s), 32.3 (s), 21.3 (d, *J* = 21.2 Hz).

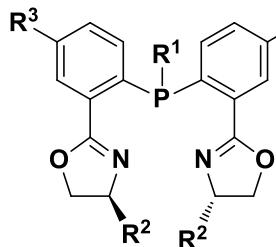
³¹P NMR (202 MHz, CDCl₃) δ -9.32.

[α]_D²⁵ = -15.6 (c 0.25, CHCl₃).

HRMS (ESI): *m/z* [M + H]⁺ Calcd for C₃₇H₄₈N₂O₅P 631.3295; found 631.3307.

L8 [R¹ = Me, R² = Me, R³ = H]:

White solid; Yield 0.418 g, 38%.



R¹ = Me; R² = Me; R³ = H

¹H NMR (400 MHz, CDCl₃) δ 7.75 – 7.71 (m, 2H), 7.38 – 7.26 (m, 6H), 4.35 – 4.29 (m, 2H), 4.24 – 4.13 (m, 2H), 3.82 (dd, *J* = 7.9, 7.0 Hz, 1H), 3.65 (t, *J* = 8.2 Hz, 1H), 1.62 (d, *J* = 5.4 Hz, 3H), 1.16 (dd, *J* = 9.2, 6.6 Hz, 6H).

¹³C NMR (101 MHz, CDCl₃) δ 164.2 (d, *J* = 16.1 Hz), 142.5 (t, *J* = 24.7 Hz), 132.3 (dd, *J* = 40.8, 22.3 Hz), 131.6 (d, *J* = 28.4 Hz), 130.6 (d, *J* = 7.9 Hz), 129.8 – 129.7 (m), 127.8 (d, *J* = 15.6 Hz), 74.0 (d, *J* = 9.5 Hz), 62.2 (d, *J* = 16.6 Hz), 21.4 (d, *J* = 13.9 Hz), 14.3 (d, *J* = 17.3 Hz).

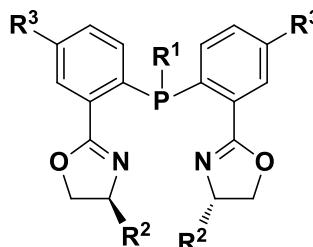
³¹P NMR (162 MHz, CDCl₃) δ –26.51.

[*α*]_D²⁵ = –25.2 (c 0.25, CHCl₃).

HRMS (ESI): *m/z* [M + H]⁺ Calcd for C₂₁H₂₄N₂O₂P = 367.1570; found 367.1575.

L9 [R¹ = Me, R² = Me, R³ = OMe]:

Colorless sticky oil; Yield 0.460 g, 36%.



R¹ = Me; R² = Me; R³ = OMe

¹H NMR (400 MHz, CDCl₃) δ 7.28 (dd, *J* = 5.4, 2.7 Hz, 2H), 7.23 – 7.17 (m, 2H), 6.92 (ddd, *J* = 8.8, 6.3, 2.7 Hz, 2H), 4.39 – 4.34 (m, 2H), 4.29 – 4.17 (m, 2H), 3.85 (t, *J* = 6.0 Hz, 1H), 3.81 (d, *J* = 0.6 Hz, 6H), 3.73 (t, *J* = 8.2 Hz, 1H), 1.57 (d, *J* = 5.5 Hz, 3H), 1.23 (d, *J* = 6.6 Hz, 3H), 1.19 (d, *J* = 6.6 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 164.3 (d, *J* = 10.9 Hz), 159.2 (d, *J* = 10.3 Hz), 134.0 – 133.2 (m), 133.2 (s), 132.9 (s), 117.0 (d, *J* = 5.6 Hz), 114.6 (dd, *J* = 9.2, 4.7 Hz), 74.1 (d, *J* = 6.6 Hz), 62.1 (d, *J* = 12.1 Hz), 55.5 (d, *J* = 1.5 Hz),

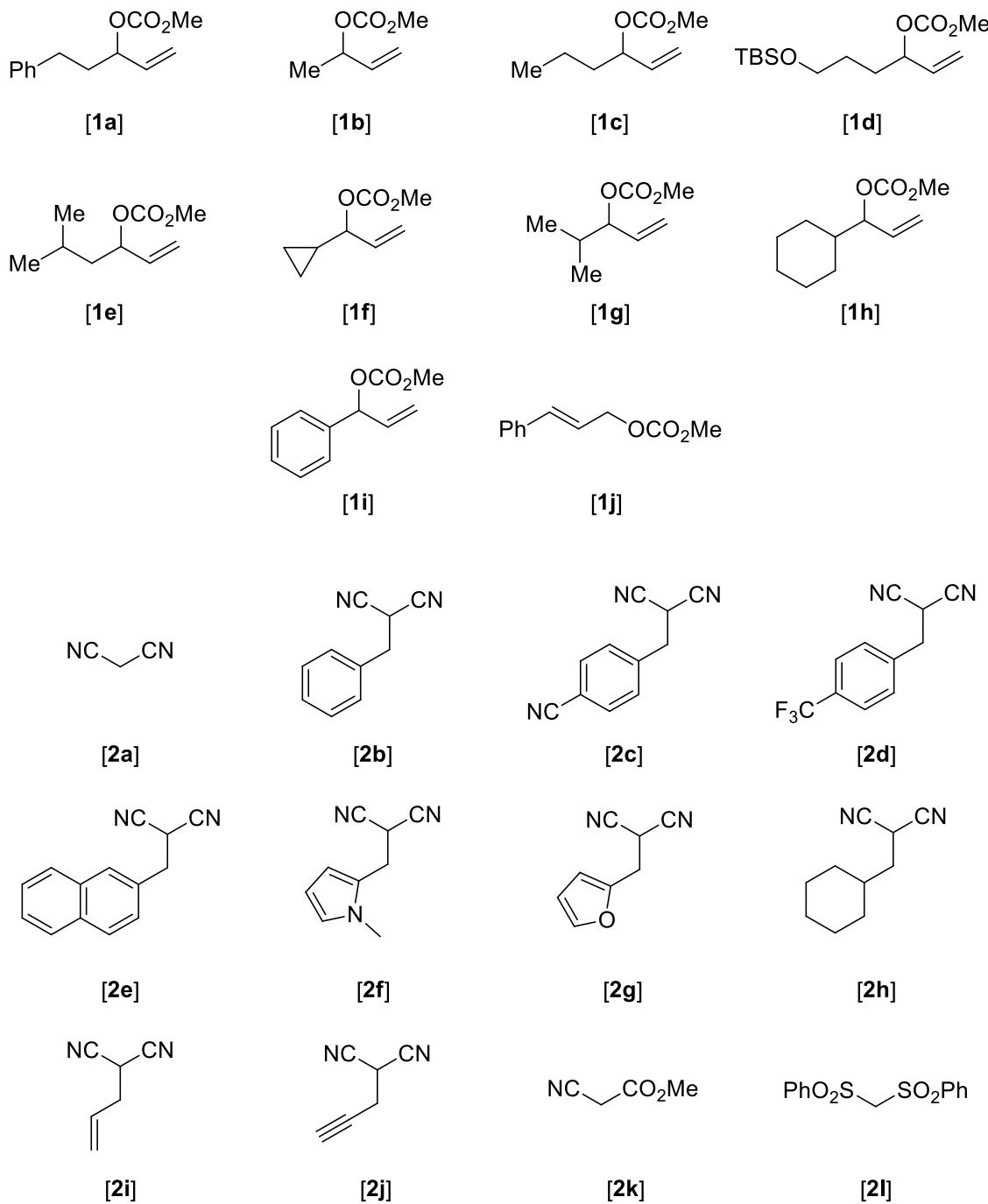
21.4 (d, *J* = 15.5 Hz), 14.5 (d, *J* = 17.0 Hz).

³¹P NMR (202 MHz, CDCl₃) δ –30.91.

[*α*]_D²⁵ = –6.4 (c 0.25, CHCl₃).

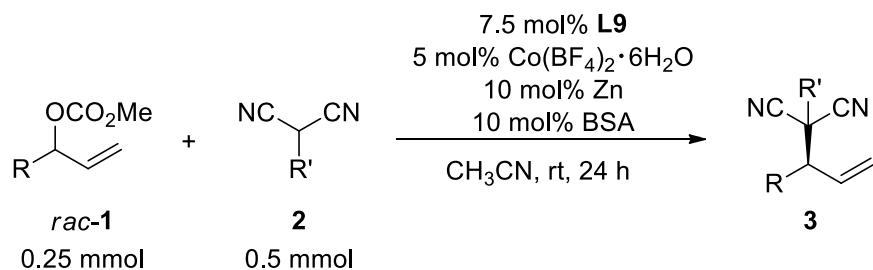
HRMS (ESI): *m/z* [M + H]⁺ Calcd for C₂₃H₂₈N₂O₄P = 427.1781; found 427.1789.

4. List of substrates:



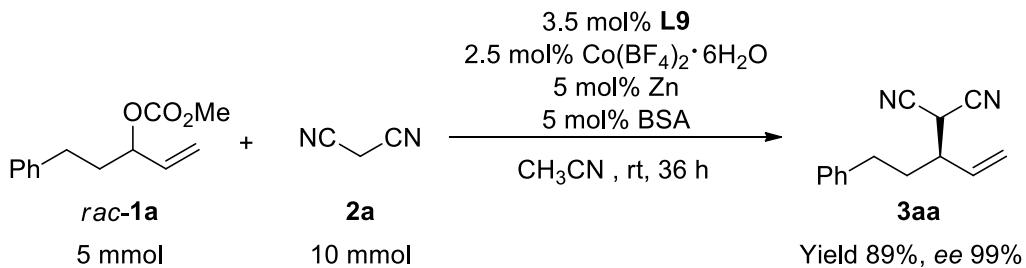
All the allylic carbonates were synthesized according to the known literature reported procedure.² **2a**, **2k** and **2l** were purchased from commercial source directly. Compounds **2b** to **2h** were also prepared accordingly to the known literature procedure.³ **2i** and **2j** were known compounds and prepared from malononitrile (2 eq) and corresponding bromide (1 eq) in the presence of Hünig's base (1 eq) in dichloromethane (0.2 M) solvent.⁴

5. Representative procedure of cobalt-catalyzed allylic alkylation of malonitrile:



In an N_2 filled glove box, **L9** (8.0 mg, 18.8×10^{-3} mmol, 7.5 mol%) and $\text{Co}(\text{BF}_4)_2 \cdot 6\text{H}_2\text{O}$ (4.3 mg, 12.6×10^{-3} mmol, 5 mol%) in 0.2 mL acetonitrile was transferred into a 10 mL reaction tube equipped with a Teflon stopcock and a stir bar. Zn dust (1.7 mg, 26.0×10^{-3} mmol, 10 mol%) was added followed by addition of 0.5 mL acetonitrile to complete transfer of zinc. The mixture was allowed to stir for 20 minutes. Afterward, allylic carbonate (0.25 mmol), malonitrile derivatives (0.5 mmol, 2 eq), bis(trimethylsilyl)acetamide (BSA) (6 μL , 25×10^{-3} mmol, 10 mol%), and 0.3 mL of acetonitrile were added sequentially to the reaction mixture. The reaction tube was sealed and brought out from the glove box. The reaction mixture was then stirred for 24 hours at room temperature. Then, solvent was removed and column chromatography was run on a column of silica gels (200-300 mesh) using PE/EA as an eluent to get the desired product.

6. Large scale synthesis of 3aa using 2.5 mol% catalyst:

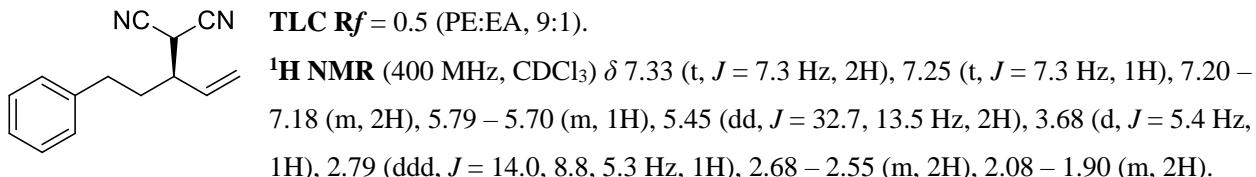


In an N_2 filled glove box, a reaction tube equipped with a Teflon stopcock and a stir bar was charged with 3.5 mol% of **L9** (75 mg, 0.18 mmol), and 2.5 mol% of $\text{Co}(\text{BF}_4)_2 \cdot 6\text{H}_2\text{O}$ (43 mg, 0.126 mmol) in 1 mL of acetonitrile. Zn dust (16.4 mg, 0.25 mmol, 5 mol%) was added followed by addition of 1 mL acetonitrile to complete transfer of zinc. The mixture was stirred for 20 minutes. Then, **rac-1a** (1.102 g, 5 mmol) and **2a** (0.660 g, 10 mmol), bis(trimethylsilyl)acetamide (BSA) (60 μL , 5 mol%) were added to the reaction mixture sequentially. Then, the reaction was monitored by TLC and stopped after 36 hours. The reaction tube was removed from the glove box and solvent was removed under reduced pressure. The residue was purified by flash column chromatography using PE/EA as an eluent. Yield: 0.930 g, 89%; *ee*: 99%.

7. Spectral data of allylic alkylation of malononitrile products:

(R)-2-(5-phenylpent-1-en-3-yl)malononitrile (3aa):^{5,6}

Clear oil; Yield: 47 mg, 89%.



¹³C NMR (101 MHz, CDCl₃) δ 140.1, 133.8, 128.9, 128.5, 126.7, 122.3, 111.9, 111.7, 44.4, 33.3, 32.8, 28.8.

HPLC (Shimadzu LC-2030) (Daicel Chiralpak OD-H Column, ⁱPrOH:*n*-Hexane = 10.0:90.0, 1.0 mL/min), Rt = 12.95 min (major) and 16.04 min (minor), 99% *ee*.

$[\alpha]_D^{25} = -21.6$ (*c* 0.05, CHCl₃).

HRMS (ESI): *m/z* [M + K]⁺ Calcd for C₁₄H₁₄N₂K = 249.0794; found 249.0710.

Assignment of absolute configuration:

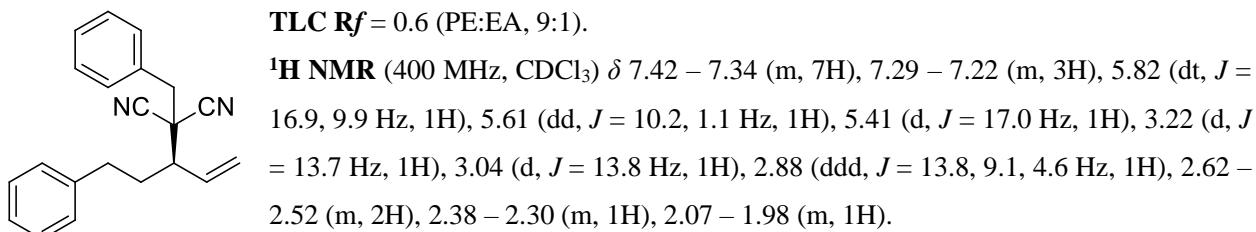
Literature data⁴ for (*R*)-enantiomer: $[\alpha]_D^{24} -8.05$ (*c* = 0.77, CHCl₃), 82% *ee*

Literature data⁵ for (*S*)-enantiomer: $[\alpha]_D^{25} +21.7$ (*c* = 1.28, CHCl₃), 88% *ee*

By comparing the literature reported data, the absolute configuration of the product **3aa** was assigned as (*R*) and all other compounds were assigned by analogy.

(R)-2-benzyl-2-(5-phenylpent-1-en-3-yl)malononitrile (3ab):⁵

White solid; Yield: 65 mg, 87%; M.P. 79-81 °C.



¹³C NMR (126 MHz, CDCl₃) δ 140.3, 133.4, 132.5, 130.3, 129.0, 128.8, 128.6, 126.5, 123.5, 115.0, 114.3, 50.2, 44.4, 41.7, 32.9, 32.4.

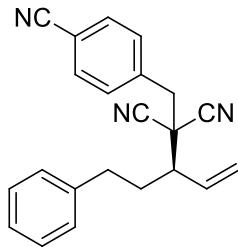
HPLC (Shimadzu LC-16) (Daicel Chiralpak IG Column, ⁱPrOH:*n*-Hexane = 1.0:99.0, 1.0 mL/min), Rt = 17.25 min (minor) and 20.31 min (major), > 99% *ee*.

$[\alpha]_D^{25} = -26.4$ (*c* 0.05, CHCl₃). Literature data⁴ for (*R*)-enantiomer: $[\alpha]_D^{26} -2.4$ (*c* = 0.5, CHCl₃).

HRMS (ESI): *m/z* [M + Na]⁺ Calcd for C₂₁H₂₀N₂Na = 323.1524; found 323.1525.

(R)-2-(4-cyanobenzyl)-2-(5-phenylpent-1-en-3-yl)malononitrile (3ac):

White solid; Yield: 71 mg, 87%; M.P. 131-132 °C.



TLC R_f = 0.3 (PE:EA, 9:1).

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.67 (d, J = 8.3 Hz, 2H), 7.46 (d, J = 8.3 Hz, 2H), 7.32 (t, J = 7.3 Hz, 2H), 7.25 – 7.22 (m, 1H), 7.18 (d, J = 7.1 Hz, 2H), 5.77 (dt, J = 16.8, 9.9 Hz, 1H), 5.61 (dd, J = 10.2, 0.9 Hz, 1H), 5.40 (d, J = 16.7 Hz, 1H), 3.23 (d, J = 13.7 Hz, 1H), 3.03 (d, J = 13.7 Hz, 1H), 2.86 (ddd, J = 13.6, 8.7, 4.6 Hz, 1H), 2.54 (dt, J = 22.3, 10.2 Hz, 2H), 2.33 – 2.25 (m, 1H), 2.05 – 1.95 (m, 1H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 140.1, 137.7, 133.0, 132.8, 131.2, 128.9, 128.6, 126.7, 124.1, 118.4, 114.5, 113.8, 113.1, 50.4, 44.0, 41.7, 32.8, 32.5.

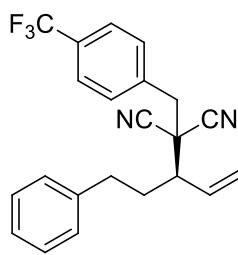
HPLC (Shimadzu LC-2030) (Daicel Chiralpak OD-H Column, $^i\text{PrOH}:n\text{-Hexane}$ = 2.0:98.0, 1.0 mL/min), R_t = 49.73 min (minor) and 53.14 min (major), > 99% *ee*.

$[\alpha]_D^{25}$ = -2.6 (c 0.05, CHCl_3).

HRMS (ESI): m/z [M + Na]⁺ Calcd for $\text{C}_{22}\text{H}_{19}\text{N}_3\text{Na}$ 348.1477; found 348.1474.

(R)-2-(5-phenylpent-1-en-3-yl)-2-(4-(trifluoromethyl)benzyl)malononitrile (3ad):

White solid; Yield: 80 mg, 87%; M.P. 120–122 °C.



TLC R_f = 0.7 (PE:EA, 9:1).

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.64 (d, J = 8.1 Hz, 2H), 7.47 (d, J = 8.1 Hz, 2H), 7.32 (t, J = 7.3 Hz, 2H), 7.25 – 7.18 (m, 3H), 5.78 (dt, J = 16.9, 9.9 Hz, 1H), 5.61 (dd, J = 10.2, 1.2 Hz, 1H), 5.39 (dd, J = 16.9, 0.6 Hz, 1H), 3.23 (d, J = 13.7 Hz, 1H), 3.04 (d, J = 13.7 Hz, 1H), 2.86 (ddd, J = 13.7, 8.8, 4.6 Hz, 1H), 2.60 – 2.48 (m, 2H), 2.34 – 2.26 (m, 1H), 2.05 – 1.95 (m, 1H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 140.2, 136.5, 133.2, 131.6 – 130.8 (m), 130.8, 128.9, 128.6, 126.7, 126.1 (q, J = 3.7 Hz), 127.3 – 120.8 (m), 124.0, 114.7, 114.0, 50.4, 44.1, 41.5, 32.8, 32.5.

HPLC (Shimadzu LC-2030) (Daicel Chiralpak OD-H Column, $^i\text{PrOH}:n\text{-Hexane}$ = 0.1:99.9, 0.5 mL/min), R_t = 57.50 min (minor) and 61.26 min (major), > 99% *ee*.

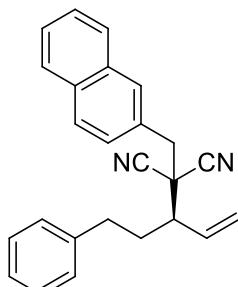
$[\alpha]_D^{25}$ = -1.0 (c 0.05, CHCl_3).

HRMS (ESI): m/z [M + Na]⁺ Calcd for $\text{C}_{22}\text{H}_{19}\text{F}_3\text{N}_2\text{Na}$ 391.1398; found 391.1387.

(R)-2-(naphthalen-2-ylmethyl)-2-(5-phenylpent-1-en-3-yl)malononitrile (3ae):

White solid; Yield: 80 mg, 91%; M.P. 103–105 °C.

TLC R_f = 0.3 (PE:EA, 9:1).



$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.84 (dd, J = 9.2, 3.8 Hz, 4H), 7.52 – 7.48 (m, 2H), 7.45 (dd, J = 8.5, 1.8 Hz, 1H), 7.31 (t, J = 7.4 Hz, 2H), 7.23 (d, J = 8.2 Hz, 1H), 7.21 – 7.18 (m, 2H), 5.81 (dt, J = 16.9, 10.0 Hz, 1H), 5.60 (dd, J = 10.2, 1.0 Hz, 1H), 5.40 (d, J =

16.9 Hz, 1H), 3.35 (d, J = 13.8 Hz, 1H), 3.17 (d, J = 13.8 Hz, 1H), 2.84 (ddd, J = 13.8, 9.2, 4.6 Hz, 1H), 2.58 – 2.52 (m, 2H), 2.35 – 2.30 (m, 1H), 2.04 – 1.97 (m, 1H).

^{13}C NMR (126 MHz, CDCl_3) δ 140.4, 133.5, 133.4, 133.3, 130.0, 129.8, 128.9 (2C), 128.6, 128.2, 127.9, 127.7, 126.8, 126.7, 126.6, 123.7, 115.1, 114.4, 50.4, 44.5, 42.0, 32.9, 32.5.

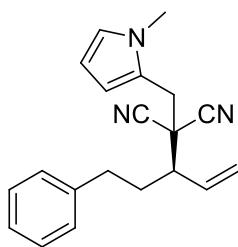
HPLC (Shimadzu LC-2030) (Daicel Chiralpak OD-H Column, $^1\text{PrOH}:n\text{-Hexane}$ = 8.0:92.0, 1.0 mL/min), R_t = 14.82 min (minor) and 24.01 min (major), 99% *ee*.

$[\alpha]_D^{25} = -2.6$ (*c* 0.05, CHCl_3).

HRMS (ESI): m/z [M + Na]⁺ Calcd for $\text{C}_{25}\text{H}_{22}\text{N}_2\text{Na}$ 373.1681; found 373.1682.

(R)-2-((1-methyl-1*H*-pyrrol-2-yl)methyl)-2-(5-phenylpent-1-en-3-yl)malononitrile (3af):

Clear oil; Yield: 65 mg, 86%.



TLC R_f = 0.4 (PE:EA, 9:1).

^1H NMR (400 MHz, CDCl_3) δ 7.31 (t, J = 7.3 Hz, 2H), 7.25 – 7.18 (m, 3H), 6.62 (dd, J = 2.5, 1.9 Hz, 1H), 6.24 (dd, J = 3.6, 1.7 Hz, 1H), 6.12 – 6.10 (m, 1H), 5.76 (dt, J = 16.9, 9.9 Hz, 1H), 5.58 (dd, J = 10.2, 1.1 Hz, 1H), 5.39 (d, J = 17.0 Hz, 1H), 3.63 (s, 3H), 3.26 (d, J = 15.2 Hz, 1H), 3.09 (d, J = 15.2 Hz, 1H), 2.83 (ddd, J = 13.9, 9.3, 4.7 Hz, 1H), 2.59 – 2.51 (m, 2H), 2.32 – 2.23 (m, 1H), 2.02 – 1.92 (m, 1H).

^{13}C NMR (126 MHz, CDCl_3) δ 140.4, 133.4, 128.9, 128.6, 126.6, 124.1, 123.7, 123.0, 115.3, 114.6, 111.1, 107.9, 49.8, 43.8, 34.3, 32.9, 32.7, 32.5.

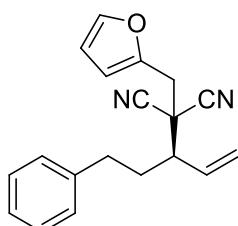
HPLC (Shimadzu LC-2030) (Daicel Chiralpak OD-H Column, $^1\text{PrOH}:n\text{-Hexane}$ = 5.0:95.0, 1.0 mL/min), R_t = 18.29 min (major) and 21.90 min (minor), 99% *ee*.

$[\alpha]_D^{25} = -9.6$ (*c* 0.05, CHCl_3).

HRMS (ESI): m/z [M + H]⁺ Calcd for $\text{C}_{20}\text{H}_{22}\text{N}_3$ 304.1808; found 304.1815.

(R)-2-(furan-2-ylmethyl)-2-(5-phenylpent-1-en-3-yl)malononitrile (3ag):

Clear oil; Yield: 60 mg, 83%.



TLC R_f = 0.5 (PE:EA, 9:1).

^1H NMR (400 MHz, CDCl_3) δ 7.41 (dd, J = 1.8, 0.7 Hz, 1H), 7.32 (t, J = 7.3 Hz, 2H), 7.25 – 7.18 (m, 3H), 6.40 – 6.37 (m, 2H), 5.74 (dt, J = 16.9, 9.9 Hz, 1H), 5.58 (dd, J = 10.2, 1.2 Hz, 1H), 5.40 (dd, J = 16.9, 0.7 Hz, 1H), 3.28 (dd, J = 46.9, 15.1 Hz, 2H), 2.83 (ddd, J = 13.9, 9.3, 4.6 Hz, 1H), 2.58 – 2.45 (m, 2H), 2.32 – 2.23 (m, 1H), 2.01 – 1.92 (m, 1H).

^{13}C NMR (126 MHz, CDCl_3) δ 146.5, 143.6, 140.4, 133.1, 128.8, 128.6, 126.5, 123.9, 114.7, 114.2, 111.0, 110.8, 49.3, 42.4, 34.7 32.9, 32.3.

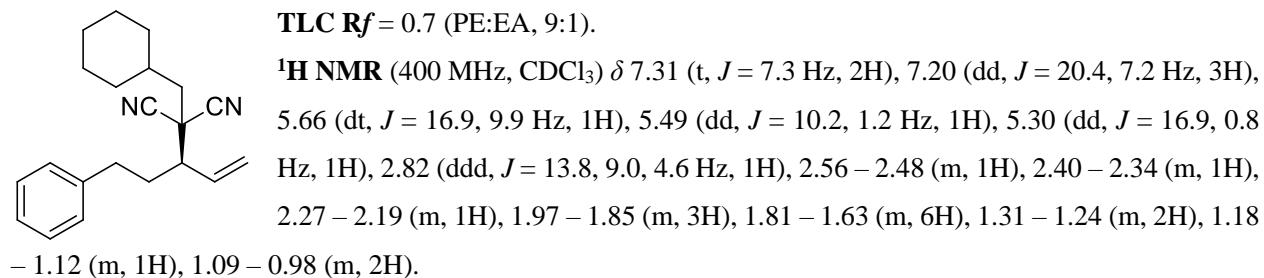
HPLC (Shimadzu LC-16) (Daicel Chiralpak IC Column, $^i\text{PrOH}:n\text{-Hexane} = 0.5:99.5$, 1.0 mL/min), $R_t = 20.69$ min (minor) and 22.48 min (major), $> 99\% ee$.

$[\alpha]_D^{25} = -6.3$ ($c 0.05$, CHCl_3).

HRMS (ESI): m/z [M + Na] $^+$ Calcd for $\text{C}_{19}\text{H}_{18}\text{N}_2\text{ONa}$ 313.1317; found 313.1321.

(R)-2-(cyclohexylmethyl)-2-(5-phenylpent-1-en-3-yl)malononitrile (3ah):

Clear oil; Yield: 67.5 mg, 88%.



$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 140.5, 133.5, 128.8, 128.6, 126.6, 123.2, 115.8, 115.3, 51.4, 43.0, 40.4, 35.9, 33.7, 33.5, 32.9, 32.1, 26.1, 26.1, 26.0.

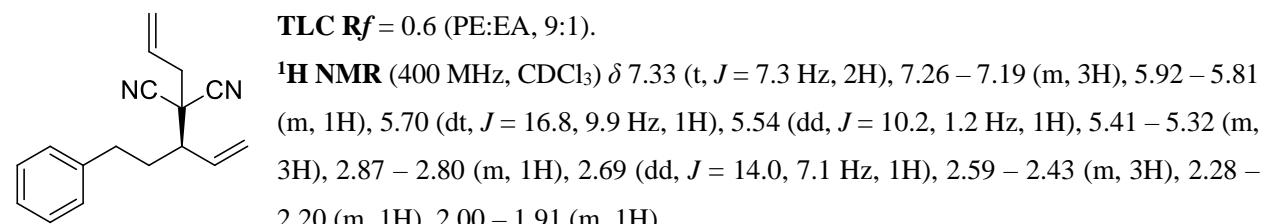
HPLC (Shimadzu LC-2030) (Daicel Chiralpak OD-H Column, $^i\text{PrOH}:n\text{-Hexane} = 1.0:99.0$, 1.0 mL/min), $R_t = 6.88$ min (major) and 7.66 min (minor), 96 % ee .

$[\alpha]_D^{25} = -14.1$ ($c 0.05$, CHCl_3).

HRMS (ESI): m/z [M + Na] $^+$ Calcd for $\text{C}_{21}\text{H}_{26}\text{N}_2\text{Na}$ 329.1994; found 329.1992.

(R)-2-allyl-2-(5-phenylpent-1-en-3-yl)malononitrile (3ai):⁵

Clear oil; Yield: 53 mg, 85%.



$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 140.3, 133.1, 128.8, 128.6, 126.5, 123.4, 123.2, 114.9, 114.3, 49.2, 42.2, 40.1, 32.8, 32.3.

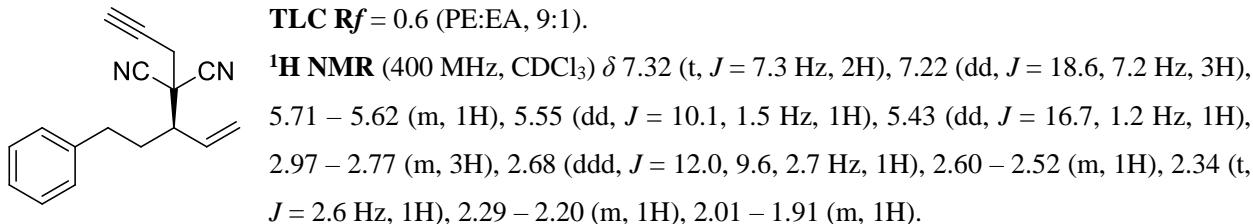
HPLC (Shimadzu LC-2030) (Daicel Chiralpak OD-H Column, $^i\text{PrOH}:n\text{-Hexane} = 5.0:95.0$, 1.0 mL/min), $R_t = 11.66$ min (minor) and 12.65 min (major), $> 99\% ee$.

$[\alpha]_D^{25} = -6.3$ ($c 0.05$, CHCl_3). Literature data⁴ for (R)-enantiomer: $[\alpha]_D^{26} = -2.78$ ($c = 0.5$, CHCl_3), 71% ee .

HRMS (ESI): m/z [M + Na] $^+$ Calcd for $\text{C}_{17}\text{H}_{18}\text{N}_2\text{Na}$ 273.1368; found 273.1370.

(R)-2-(5-phenylpent-1-en-3-yl)-2-(prop-2-yn-1-yl)malononitrile (3aj):

Clear oil; Yield: 47 mg, 76%.



$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 140.2, 132.4, 128.8, 128.6, 128.6, 126.6, 124.1, 114.3, 113.7, 75.3, 74.7, 48.6, 41.3, 32.9, 32.3, 27.3.

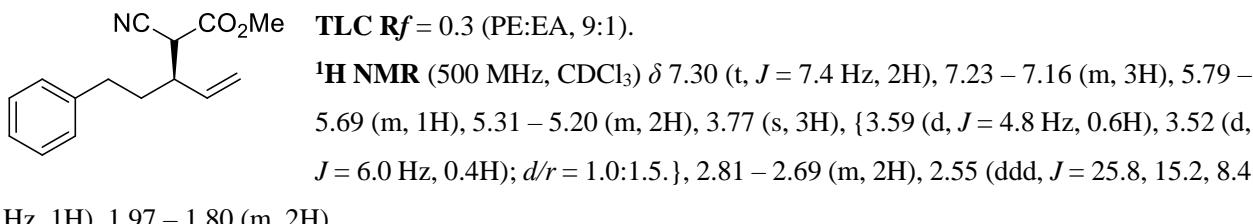
HPLC (Shimadzu LC-2030) (Daicel Chiralpak OD-H Column, $^i\text{PrOH}:n\text{-Hexane}$ = 5.0:95.0, 1.0 mL/min), R_t = 8.32 min (minor) and 10.60 min (major), > 99% *ee*.

$[\alpha]_D^{25} = -11.1$ (c 0.05, CHCl_3).

HRMS (ESI): m/z [M + Na]⁺ Calcd for $\text{C}_{17}\text{H}_{16}\text{N}_2\text{Na}$ 271.1211; found 271.1210.

(R)-methyl 2-cyano-3-phenethylpent-4-enoate (3ak):

Clear oil; Yield: 34 mg, 56%.



$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 165.9, 165.8, 141.0, 140.9, 136.3, 135.4, 128.7, 128.7, 128.5, 126.4, 126.3, 120.2, 119.7, 115.4, 115.1, 53.5, 53.5, 44.2, 44.1, 43.5, 43.5, 34.4, 33.2, 33.1, 33.0.

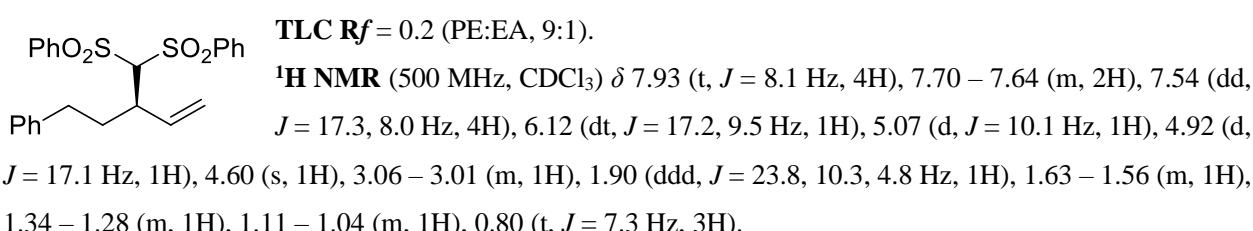
HPLC (Shimadzu LC-2030) (Daicel Chiralpak IC Column, $^i\text{PrOH}:n\text{-Hexane}$ = 0.1:99.1, 1.0 mL/min), d/r = 1.0:1.5. R_t = 40.78 min (minor) and 46.95 min (major), > 99% *ee* for the major diastereomer.

$[\alpha]_D^{25} = -5.3$ (c 0.05, CHCl_3).

HRMS (ESI): m/z [M + Na]⁺ Calcd for $\text{C}_{15}\text{H}_{17}\text{NO}_2\text{Na}$ 266.1157; found 266.1158.

(R)-(2-vinylpentane-1,1-diyldisulfonyl)dibenzene (3al):

White solid; Yield: 58 mg, 61%; M.P. 91-93 °C.



^{13}C NMR (126 MHz, CDCl_3) δ 140.0, 139.0, 137.7, 134.7, 134.4, 129.8, 129.5, 129.3, 129.2, 118.4, 88.0, 44.5, 32.7, 21.5, 13.7.

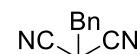
HPLC (Shimadzu LC-2030) (Daicel Chiralpak IG Column, $^i\text{PrOH}:n\text{-Hexane} = 3.0:97.0$, 0.5 mL/min), $Rt = 86.38$ min (major) and 90.37 min (minor), $> 99\% ee$.

$[\alpha]_D^{25} = +7.6$ (c 0.05, CHCl_3).

HRMS (ESI): m/z [M + Na]⁺ Calcd for $\text{C}_{19}\text{H}_{22}\text{O}_4\text{S}_2\text{Na}$ 401.0857; found 401.0856.

(R)-2-benzyl-2-(but-3-en-2-yl)malononitrile (3bb):

White solid; Yield: 49 mg, 93%; M.P. 40-42 °C.

 **TLC** $Rf = 0.4$ (PE:EA, 9:1).

^1H NMR (500 MHz, CDCl_3) δ 7.41 – 7.38 (m, 5H), 5.89 (ddd, $J = 17.0, 10.3, 8.8$ Hz, 1H), 5.40 (dd, $J = 22.5, 13.6$ Hz, 2H), 3.20 (d, $J = 13.7$ Hz, 1H), 3.08 (d, $J = 13.8$ Hz, 1H), 2.77 – 2.71 (m, 1H), 1.46 (d, $J = 6.8$ Hz, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 135.0, 132.6, 130.3, 129.1, 128.9, 121.0, 114.9, 114.4, 45.2, 44.9, 41.6, 17.2.

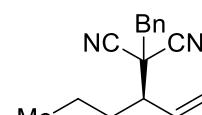
HPLC (Shimadzu LC-2030) (Daicel Chiralpak OJ-H Column, $^i\text{PrOH}:n\text{-Hexane} = 5.0:95.0$, 1.0 mL/min), $Rt = 9.75$ min (major) and 15.05 min (minor), 90 % ee .

$[\alpha]_D^{25} = -22.0$ (c 0.05, CHCl_3).

HRMS (ESI): m/z [M + Na]⁺ Calcd for $\text{C}_{14}\text{H}_{14}\text{N}_2\text{Na}$ 233.1055; found 233.1054.

(R)-2-benzyl-2-(hex-1-en-3-yl)malononitrile (3cb):

White solid; Yield: 52 mg, 87%; M.P. 42-43 °C.

 **TLC** $Rf = 0.5$ (PE:EA, 9:1).

^1H NMR (400 MHz, CDCl_3) δ 7.43 – 7.37 (m, 5H), 5.71 (dt, $J = 16.9, 9.9$ Hz, 1H), 5.49 (dd, $J = 10.2, 1.2$ Hz, 1H), 5.35 (dd, $J = 16.9, 0.6$ Hz, 1H), 3.23 (d, $J = 13.7$ Hz, 1H), 3.05 (d, $J = 13.7$ Hz, 1H), 2.54 – 2.48 (m, 1H), 1.94 – 1.86 (m, 1H), 1.71 – 1.65 (m, 1H), 1.55 – 1.44 (m, 1H), 1.34 – 1.23 (m, 1H), 0.96 (t, $J = 7.3$ Hz, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 133.7, 132.6, 130.4, 129.1, 128.8, 122.8, 115.2, 114.5, 50.9, 44.6, 41.9, 33.1, 20.3, 13.8.

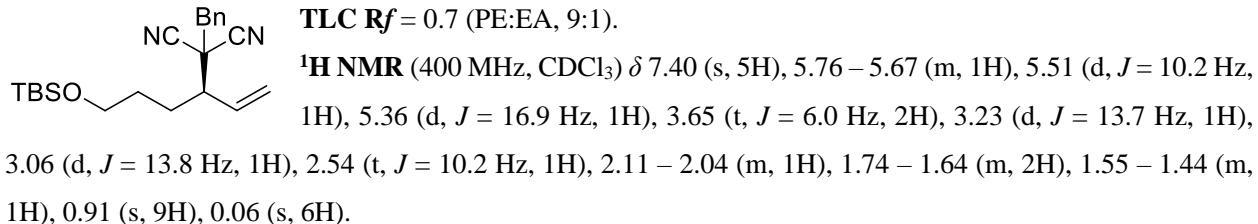
HPLC (Shimadzu LC-2030) (Daicel Chiralpak OD-H Column, $^i\text{PrOH}:n\text{-Hexane} = 5.0:95.0$, 1.0 mL/min), $Rt = 6.27$ min (minor) and 7.15 min (major), 99% ee .

$[\alpha]_D^{25} = -29.9$ (c 0.05, CHCl_3).

HRMS (ESI): m/z [M + Na]⁺ Calcd for $\text{C}_{16}\text{H}_{18}\text{N}_2\text{Na}$ 261.1368; found 261.1368.

(R)-2-benzyl-2-(6-((tert-butyldimethylsilyl)oxy)hex-1-en-3-yl)malononitrile (3db):

White solid; Yield: 68 mg, 74%; M.P. 43-44 °C.



$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 133.7, 132.6, 130.4, 129.1, 128.8, 123.0, 115.0, 114.5, 62.5, 50.8, 44.6, 41.9, 30.2, 27.5, 26.1, 18.5, -5.1.

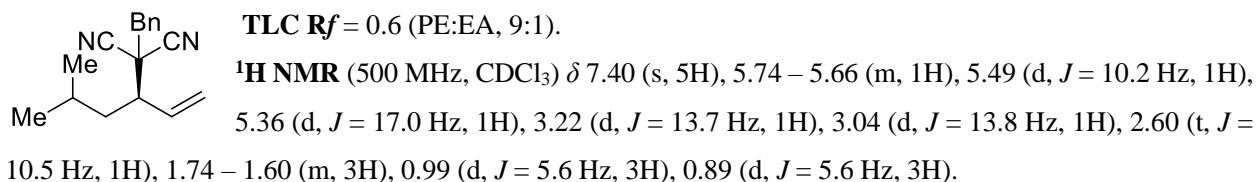
HPLC (Shimadzu LC-2030) (Daicel Chiralpak AD-H Column, $^i\text{PrOH}:n\text{-Hexane}$ = 2.0:98.0, 1.0 mL/min), R_t = 10.40 min (major) and 11.84 min (minor), 99% *ee*.

$[\alpha]_D^{25} = -15.3$ (*c* 0.05, CHCl_3).

HRMS (ESI): m/z [M + H]⁺ Calcd for $\text{C}_{22}\text{H}_{33}\text{N}_2\text{OSi}$ 369.2357; found 369.2361; and [M + Na]⁺ Calcd for $\text{C}_{22}\text{H}_{32}\text{N}_2\text{OSiNa}$ 391.2182; found 391.2168.

(R)-2-benzyl-2-(5-methylhex-1-en-3-yl)malononitrile (3eb):

White solid; Yield: 55.5 mg, 88%; M.P. 80-81 °C.



$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 133.8, 132.7, 130.4, 129.1, 128.8, 122.7, 115.2, 114.5, 49.4, 44.8, 41.9, 40.0, 25.3, 24.0, 20.7.

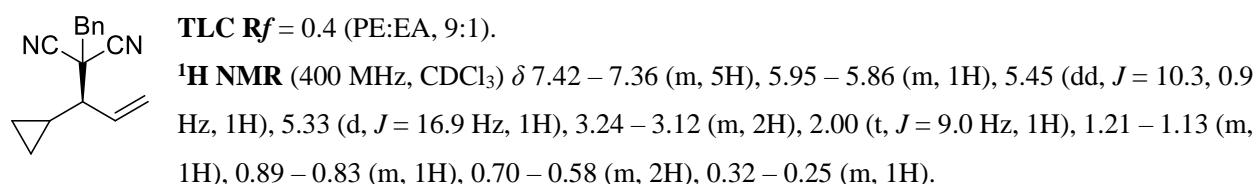
HPLC (Shimadzu LC-2030) (Daicel Chiralpak OD-H Column, $^i\text{PrOH}:n\text{-Hexane}$ = 5.0:95.0, 1.0 mL/min), R_t = 5.25 min (minor) and 6.28 min (major), > 99% *ee*.

$[\alpha]_D^{25} = -14.5$ (*c* 0.05, CHCl_3).

HRMS (ESI): m/z [M + Na]⁺ Calcd for $\text{C}_{17}\text{H}_{20}\text{N}_2\text{Na}$ 275.1524; found 275.1525.

(S)-2-benzyl-2-(1-cyclopropylallyl)malononitrile (3fb):

White solid; Yield: 56 mg, 95%; M.P. 51-53 °C.



¹³C NMR (126 MHz, CDCl₃) δ 133.1, 132.7, 130.4, 129.0, 128.8, 121.7, 114.9, 114.8, 54.8, 44.8, 42.0, 12.6, 6.7, 3.1.

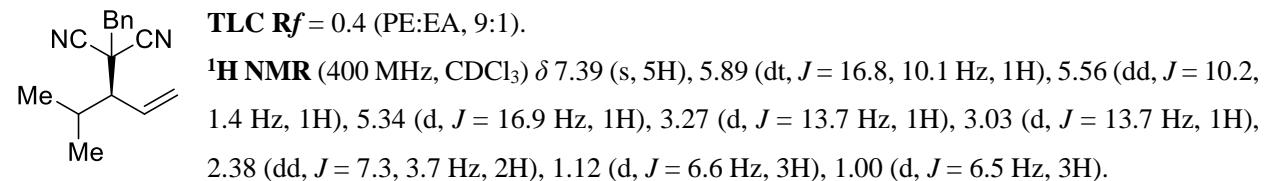
HPLC (Shimadzu LC-2030) (Daicel Chiralpak OD-H Column, ⁱPrOH:*n*-Hexane = 5.0:95.0, 1.0 mL/min), Rt = 9.04 min (minor) and 9.74 min (major), 99% *ee*.

$[\alpha]_D^{25} = -11.5$ (c 0.05, CHCl₃).

HRMS (ESI): *m/z* [M + Na]⁺ Calcd for C₁₆H₁₆N₂Na 259.1211; found 259.1211.

(R)-2-benzyl-2-(4-methylpent-1-en-3-yl)malononitrile (3gb):

White solid; Yield: 40 mg, 67%; M.P. 82-83 °C.



¹³C NMR (126 MHz, CDCl₃) δ 132.5, 130.6, 130.5, 129.0, 128.9, 124.1, 115.1, 115.0, 55.9, 43.4, 42.6, 30.4, 22.7, 17.1.

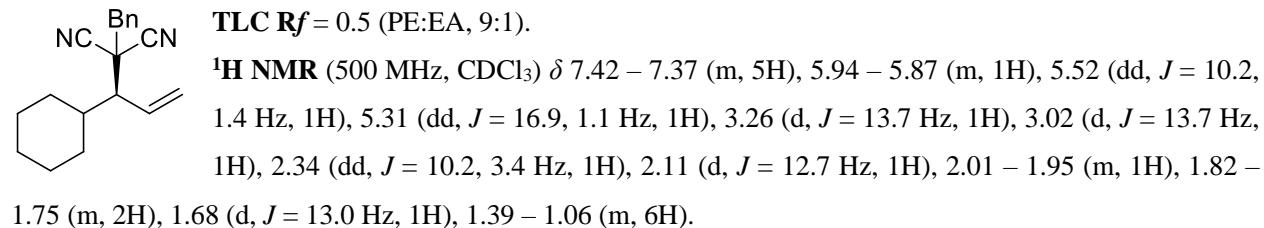
HPLC (Shimadzu LC-2030) (Daicel Chiralpak OD-H Column, ⁱPrOH:*n*-Hexane = 5.0:95.0, 1.0 mL/min), Rt = 6.07 min (minor) and 7.83 min (major), 98% *ee*.

$[\alpha]_D^{25} = -11.5$ (c 0.05, CHCl₃).

HRMS (ESI): *m/z* [M + Na]⁺ Calcd for C₁₆H₁₈N₂Na 261.1368; found 261.1363.

(S)-2-benzyl-2-(1-cyclohexylallyl)malononitrile (3hb):

White solid; Yield: 49 mg, 70%; M.P. 130-132 °C.



¹³C NMR (126 MHz, CDCl₃) δ 132.5, 131.7, 130.6, 129.0, 128.9, 123.5, 115.1, 115.1, 56.2, 43.4, 42.3, 40.5, 32.8, 28.0, 26.5, 26.3, 26.2.

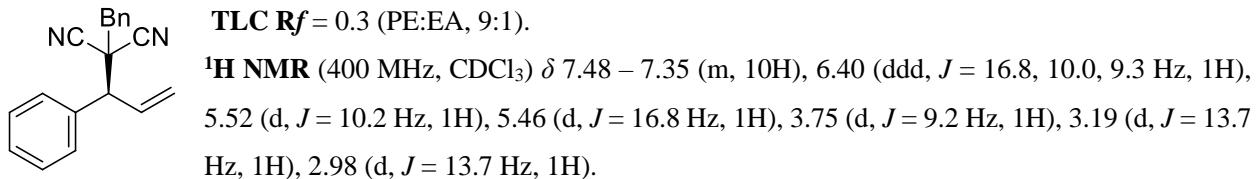
HPLC (Shimadzu LC-2030) (Daicel Chiralpak OD-H Column, ⁱPrOH:*n*-Hexane = 5.0:95.0, 1.0 mL/min), Rt = 5.68 min (minor) and 6.77 min (major), > 99% *ee*.

$[\alpha]_D^{25} = -20.3$ (c 0.05, CHCl₃).

HRMS (ESI): *m/z* [M + Na]⁺ Calcd for C₁₉H₂₂N₂Na 301.1681; found 301.1669.

(S)-2-benzyl-2-(1-phenylallyl)malononitrile (3ib):

White solid; Yield: 60.5 mg, 89%; M.P. 73-74 °C.



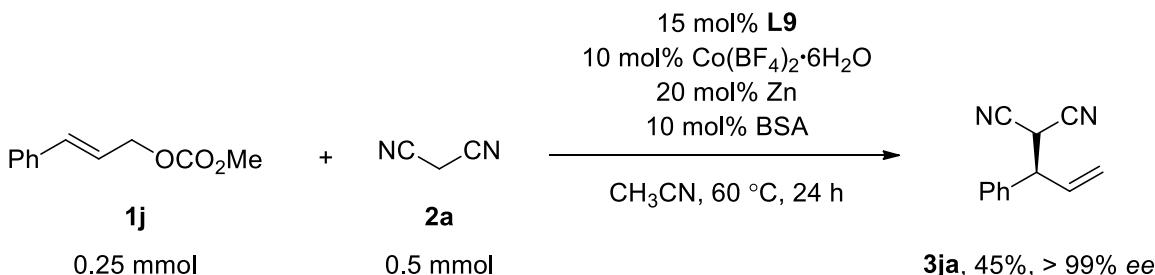
$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 136.1, 133.3, 132.4, 130.4, 129.4, 129.2, 129.1, 128.9, 122.2, 114.9, 114.5, 56.5, 45.6, 42.5.

HPLC (Shimadzu LC-2030) (Daicel Chiralpak OD-H Column, $^3\text{PrOH}:n\text{-Hexane}$ = 5.0:95.0, 1.0 mL/min), R_t = 9.23 min (minor) and 10.30 min (major), 99% *ee*.

$[\alpha]_D^{25} = +19.8$ (c 0.05, CHCl_3).

HRMS (ESI): m/z [M + Na]⁺ Calcd for $\text{C}_{19}\text{H}_{16}\text{N}_2\text{Na}$ 295.1211; found 295.1211.

8. Synthesis of (R)-2-(1-phenylallyl)malononitrile (3ja):⁶



In an N_2 filled glove box, **L9** (16 mg, 37.5×10^{-3} mmol, 15 mol%) and $\text{Co}(\text{BF}_4)_2 \cdot 6\text{H}_2\text{O}$ (8.6 mg, 25.2×10^{-3} mmol, 10 mol%) in 0.2 mL acetonitrile was taken into a 10 mL reaction tube equipped with a Teflon stopcock and a stir bar. Zn dust (3.3 mg, 50.5×10^{-3} mmol, 20 mol%) and 0.5 mL acetonitrile to complete transfer of zinc were added to it. The mixture was allowed to stir for 20 minutes. Afterward, (*E*)-cinnamyl methyl carbonate (**1j**) (48.2 mg, 0.25 mmol, 1 eq), malononitrile (**2a**) (33 mg, 0.5 mmol, 2 eq), bis(trimethylsilyl)acetamide (BSA) (6 μL , 25×10^{-3} mmol, 10 mol%), and 0.3 mL of acetonitrile were added sequentially to the reaction mixture. The reaction tube was sealed and brought out from the glove box. The reaction mixture was then stirred for 24 hours at 60 °C in oil bath. Then, it was cooled to room temperature. and solvent was removed under reduced pressure. The desired pure product was obtained by silica gels (200-300 mesh) column chromatography using PE/EA as an eluent.

Clear oil; Yield: 20.5 mg, 45%.

TLC R_f = 0.2 (PE:EA, 9:1).

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.45 – 7.32 (m, 5H), 6.17 (ddd, J = 17.0, 10.3, 7.7 Hz, 1H), 5.49 (d, J = 10.3 Hz, 1H), 5.43 (dd, J = 17.0, 0.8 Hz, 1H), 4.01 (d, J = 6.6 Hz, 1H), 3.94 (t, J = 7.2 Hz, 1H).

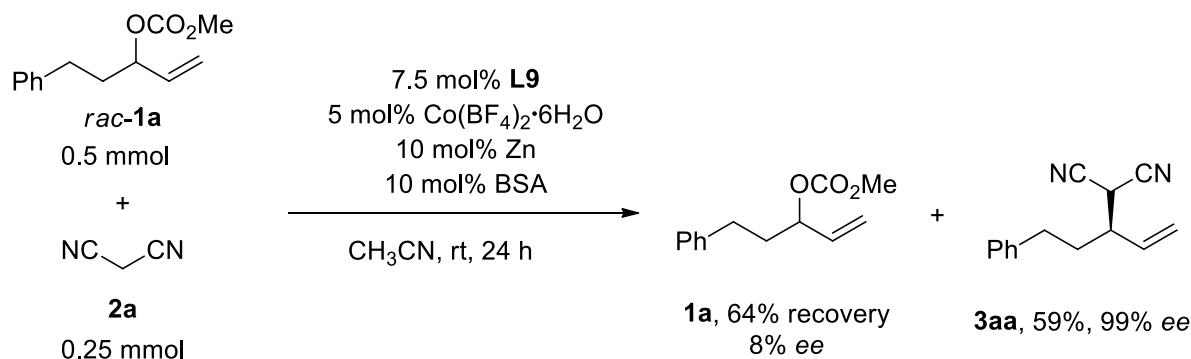
¹³C NMR (126 MHz, CDCl₃) δ 136.3, 133.3, 129.6, 129.1, 128.0, 121.1, 111.8, 111.8, 50.3, 29.9.

HPLC (Shimadzu LC-16) (Daicel Chiralpak OD-H Column, ⁱPrOH:*n*-Hexane = 10.0:90.0, 1.0 mL/min), Rt = 17.01 min (minor) and 22.55 min (major), > 99% *ee*.

[α]_D²⁵ = +6.1 (*c* 0.05, CHCl₃). Literature report: [α]_D²⁴ = +41.4 (*c* 0.54, CHCl₃), 91% *ee*.

HRMS (ESI): *m/z* [M + K]⁺ calculated for C₁₂H₁₀N₂K 221.0481; found 221.0396.

9. Kinetic resolution study:



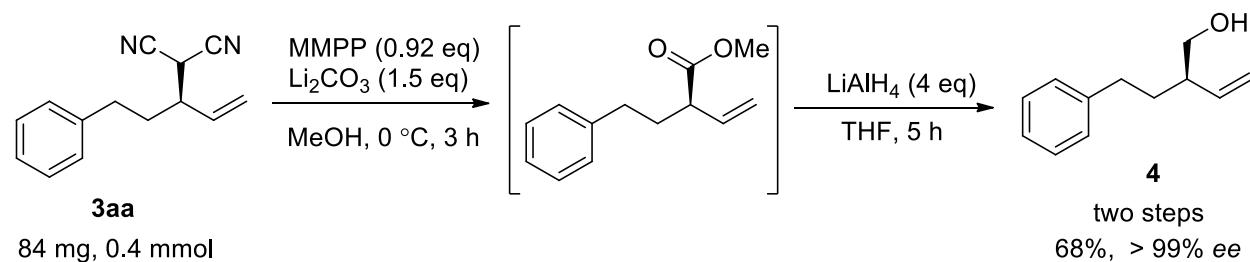
In an N₂ filled glove box, a reaction tube contains **L9** (8 mg, 18.8 × 10⁻³ mmol, 7.5 mol%) and Co(BF₄)₂·6H₂O (4.3 mg, 12.6 × 10⁻³ mmol, 5 mol%) in 0.2 mL acetonitrile was added Zn dust (1.7 mg, 26.0 × 10⁻³ mmol, 10 mol%). Additionally, 0.5 mL of acetonitrile was added to complete transfer of zinc. The mixture was allowed to stir for 20 minutes. Afterward, **rac-1a** (110 mg, 0.5 mmol, 2 eq), malonitrile (**2a**) (17 mg, 0.25 mmol, 1 eq), bis(trimethylsilyl)acetamide (BSA) (6 μL, 25 × 10⁻³ mmol, 10 mol%), and 0.3 mL of acetonitrile were added sequentially to the reaction mixture. The reaction tube was sealed with Teflon stopcock and brought out from the glove box. The reaction mixture was then stirred for 16 hours at room temperature. Then, solvent was removed and column chromatography was run on a column of silica gels (200-300 mesh) using PE/EA as an eluent to get the desired product.

The amount of recovery **1a** was 70 mg, 64% yield; *ee* 8%.

The amount of product **3aa** was 31 mg, 30% yield; *ee* 99%.

Enantiomeric excess of recovered **1a** was determined by **HPLC** (Shimadzu LC-2030) (Daicel Chiralpak OD-H Column, ⁱPrOH:*n*-Hexane = 5.0:95.0, 1.0 mL/min), Rt = 4.65 min (minor) and 4.92 min (major), 6% *ee*.

10. Synthesis of (*R*)-2-phenethylbut-3-en-1-ol (4):



Compound **3aa** (84 mg, 0.4 mmol) was dissolved in MeOH (3 mL) in a 10 mL round bottom flask at 0 °C. Li₂CO₃ (45 mg, 0.6 mmol, 1.5 eq) and Magnesium monoperoxyphthalate hexahydrate (MMPP) 80% analytical grade (144 mg, 0.92 eq) were added sequentially to it. The reaction mixture was stirred for 3 hours at 0 °C. Then, it was diluted with EA (20 mL) and filtered through a pad of celite to remove solid residues and concentrated in vacuo. The crude product was further dissolved in EA (20 mL) and filtered through a pad of celite to get the clear oily product. The crude oily product was treated with LiAlH₄ (66 mg, 4 eq) in 8 mL THF at 0 °C and stirred continue at room temperature for 5 hours. The reaction mixture was quenched with water (5 mL) carefully and 5 mL of dil. HCl (1 M) was added and extracted with EA. The organic phase was dried on Na₂SO₄ and chromatographed on a column of silica gels (PE/EA as an eluent).

Clear oil; Yield: 48 mg, 68%.

TLC R_f = 0.3 (PE:EA, 4:1).

¹H NMR (400 MHz, CDCl₃) δ 7.32 – 7.28 (m, 2H), 7.22 – 7.18 (m, 3H), 5.66 (ddd, *J* = 17.2, 10.3, 8.8 Hz, 1H), 5.22 (ddd, *J* = 18.3, 13.8, 1.4 Hz, 2H), 3.59 (dd, *J* = 10.6, 5.2 Hz, 1H), 3.47 (dd, *J* = 10.6, 8.0 Hz, 1H), 2.72 (ddd, *J* = 15.0, 10.0, 5.3 Hz, 1H), 2.58 (ddd, *J* = 13.8, 9.8, 6.9 Hz, 1H), 2.32 – 2.23 (m, 1H), 1.82 – 1.73 (m, 1H), 1.67 (s, 1H), 1.64 – 1.55 (m, 1H).

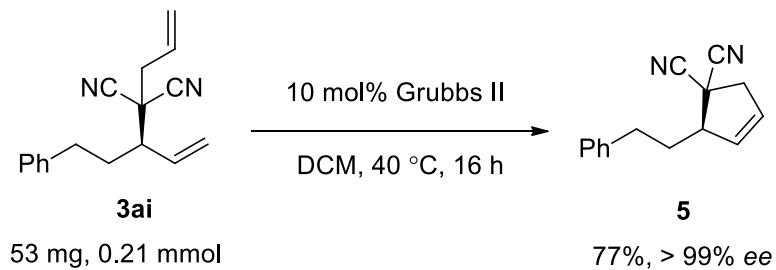
¹³C NMR (101 MHz, CDCl₃) δ 142.3, 139.8, 128.6, 128.5, 125.9, 118.0, 65.7, 46.6, 33.4, 32.6.

[*α*]_D²⁵ = +5.6 (c 0.25, CHCl₃).

HPLC (Shimadzu LC-2030) (Daicel Chiralpak OJ-H Column, iPrOH:*n*-Hexane = 1.0 : 99.0, 1.0 mL/min), *Rt* = 22.97 min (minor) and 24.04 min (major), > 99% *ee*.

HRMS (ESI): *m/z* [M + Na]⁺ Calcd for C₁₂H₁₆ONa 199.1099; found 199.1096.

11. Synthesis (*R*)-2-phenethylcyclopent-3-ene-1,1-dicarbonitrile (5):



In a Schlenk tube equipped with Teflon stop cock and stir bar, **3ai** (53 mg, 0.21 mmol) and Grubbs II (18 mg, 0.021 mmol, 10 mol%) were mixed in DCM (5 mL) under argon atmosphere. The reaction mixture was heated at 40 °C in oil bath for 16 hours. Then solvent was removed and column chromatography was performed on silica gels using PE/EA as an eluent.

Clear oil; Yield: 36 mg, 77%.

TLC R_f = 0.3 (PE:EA, 20:1).

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.33 – 7.30 (m, 2H), 7.24 – 7.21 (m, 3H), 5.82 – 5.76 (m, 2H), 3.31 (ddd, J = 8.4, 3.7, 1.8 Hz, 1H), 3.24 – 3.12 (m, 2H), 2.81 (dd, J = 11.7, 4.8 Hz, 2H), 2.15 (tt, J = 14.0, 7.2 Hz, 1H), 2.00 (ddd, J = 12.2, 7.9, 6.2 Hz, 1H).

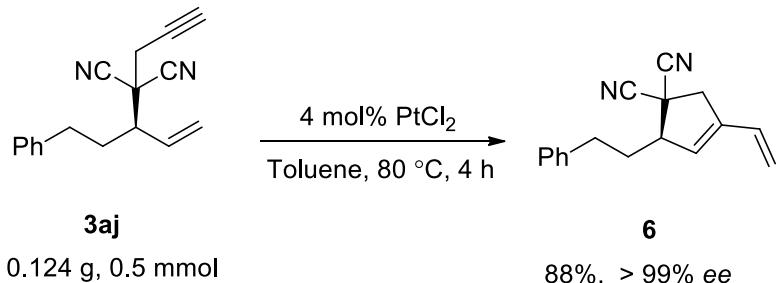
$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 140.3, 132.4, 128.9, 128.5, 127.2, 126.6, 116.9, 115.0, 54.7, 44.9, 37.9, 33.6.

HPLC (Shimadzu LC-16) (Daicel Chiralpak AD-H Column, $^3\text{PrOH}:n\text{-Hexane} = 2.0:98.0$, 0.5 mL/min), R_t = 16.92 min (minor) and 18.11 min (major), > 99% *ee*.

$[\alpha]_D^{25} = -92.8$ (*c* 0.25, CHCl_3).

HRMS (ESI): m/z [M + Na]⁺ Calcd for $\text{C}_{15}\text{H}_{14}\text{N}_2\text{Na}$ 245.1055; found 245.1059.

12. Synthesis (*R*)-2-phenethyl-4-vinylcyclopent-3-ene-1,1-dicarbonitrile (6):



Under Argon atmosphere, a Schlenk tube equipped with Teflon stop cock and stir bar was charged with **3aj** (0.124 g, 0.5 mmol) and PtCl_2 (5.5 mg, 0.02 mmol, 4 mol%) in Toluene (5 mL). The reaction mixture was heated

at 80 °C in oil bath for 4 hours. Then, solvent was removed and column chromatography was performed on silica gels using PE/EA as an eluent.

Clear oil; Yield: 109 mg, 88%.

TLC R_f = 0.3 (PE:EA, 4:1).

¹H NMR (400 MHz, CDCl₃) δ 7.33 – 7.29 (m, 2H), 7.24 – 7.21 (m, 3H), 6.47 (dd, *J* = 17.6, 10.8 Hz, 1H), 5.67 (s, 1H), 5.27 (d, *J* = 10.8 Hz, 1H), 5.15 (d, *J* = 17.6 Hz, 1H), 3.37 (t, *J* = 7.7 Hz, 1H), 3.34 – 3.19 (m, 2H), 2.81 (t, *J* = 7.9 Hz, 2H), 2.17 (dt, *J* = 14.5, 7.8 Hz, 1H), 2.00 (dq, *J* = 13.7, 8.1 Hz, 1H).

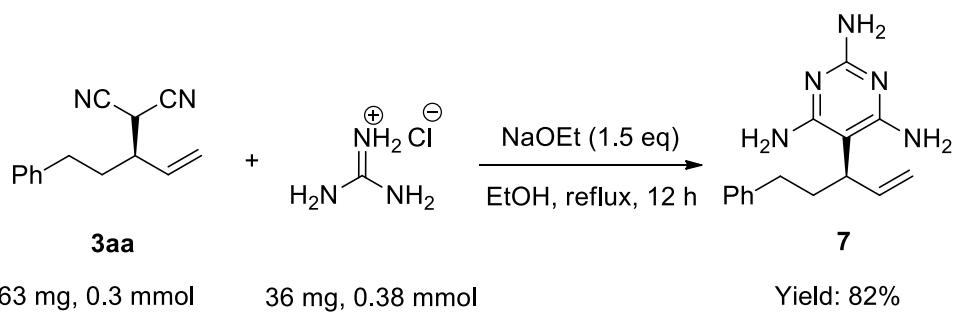
¹³C NMR (126 MHz, CDCl₃) δ 140.3, 139.1, 131.0, 129.3, 128.9, 128.5, 126.6, 118.2, 116.7, 114.9, 54.4, 43.2, 37.9, 33.7, 33.5.

[α]_D²⁵ = -133.2 (*c* 0.25, CHCl₃).

HPLC (Shimadzu LC-2030) (Daicel Chiralpak OD-H Column, *i*PrOH:*n*-Hexane = 1.0 : 99.0, 0.5 mL/min), *R_t* = 45.73 min (minor) and 47.24 min (major), > 99% *ee*

HRMS (ESI): *m/z* [M + Na]⁺ Calcd for C₁₇H₁₆N₂Na 271.1211; found 271.1215.

13. Synthesis (*R*)-5-(5-phenylpent-1-en-3-yl)pyrimidine-2,4,6-triamine (7):



To an ethanolic solution of **3aa** (63 mg, 0.3 mmol) and guanidine hydrochloride (36 mg, 0.38 mmol, 1.25 eq), was added NaOEt (32 mg, 0.45 mmol, 1.5 eq) and the reaction mixture was refluxed in oil bath for 12 hours. Then, solvent was removed and column chromatography was performed on silica gels using DCM/MeOH as an eluent.

Light yellow solid; Yield: 66 mg, 82%; M.P. 63-64 °C.

TLC R_f = 0.3 (PE:EA, 4:1).

¹H NMR (400 MHz, MeOD) δ 7.21 (dd, *J* = 10.1, 4.7 Hz, 2H), 7.14 – 7.10 (m, 3H), 6.07 – 5.99 (m, 1H), 5.17 (d, *J* = 2.3 Hz, 1H), 5.13 (dt, *J* = 3.6, 1.5 Hz, 1H), 4.93 (s, 6H), 3.46 (dtt, *J* = 11.1, 4.4, 2.3 Hz, 1H), 2.64 – 2.48 (m, 2H), 2.10 – 2.04 (m, 2H).

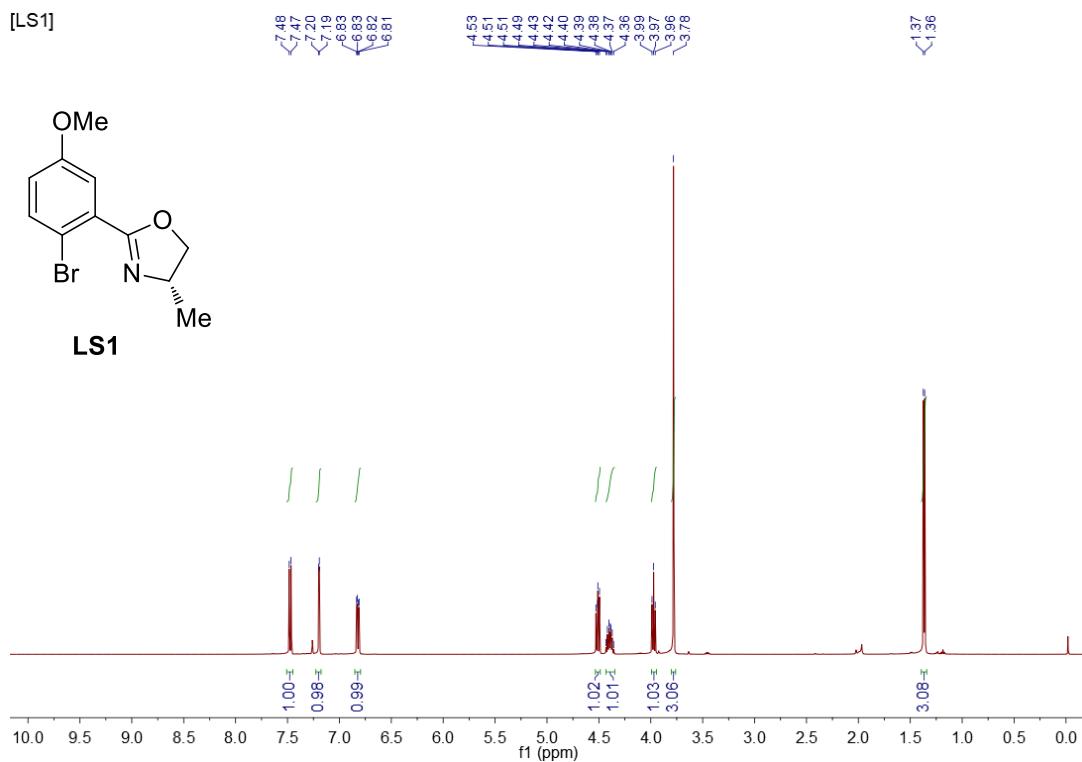
¹³C NMR (126 MHz, MeOD) δ 163.5, 161.4, 143.8, 141.0, 129.5, 129.5, 126.9, 115.1, 88.0, 39.1, 35.2, 33.8.

[α]_D²⁵ = -35.6 (*c* 0.25, MeOH).

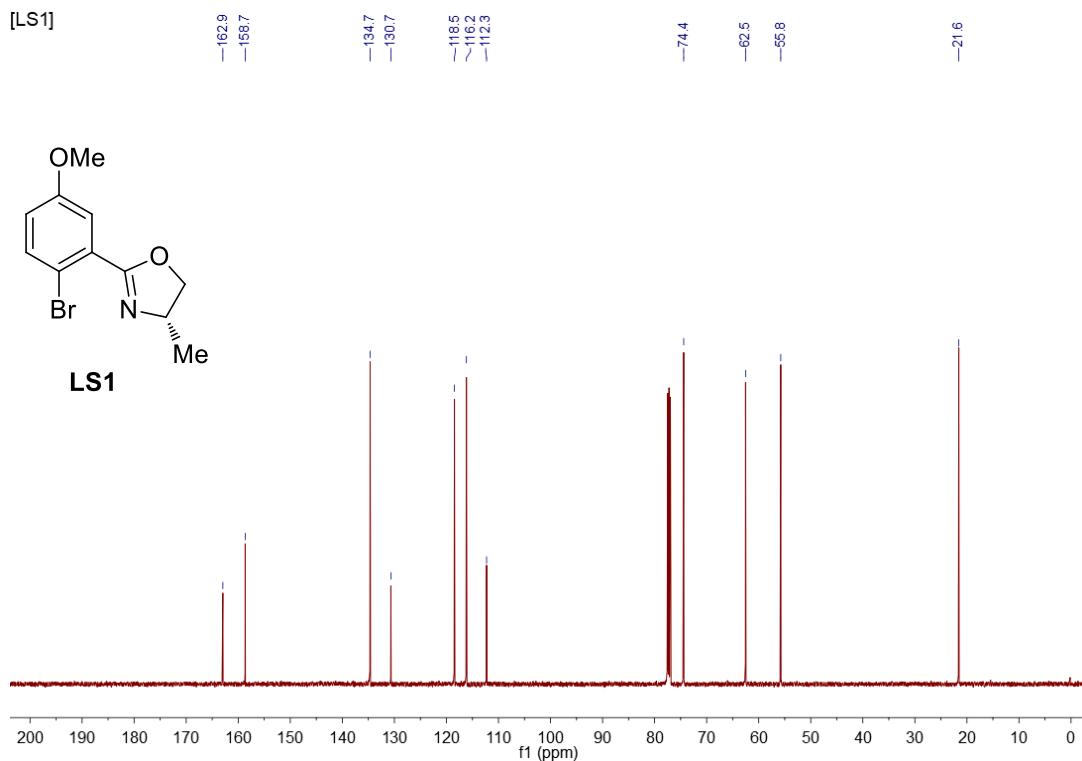
HRMS (ESI): *m/z* [M + H]⁺ Calcd for C₁₅H₂₀N₅ 270.1713; found 270.1714.

14. NMR spectra of new compounds:

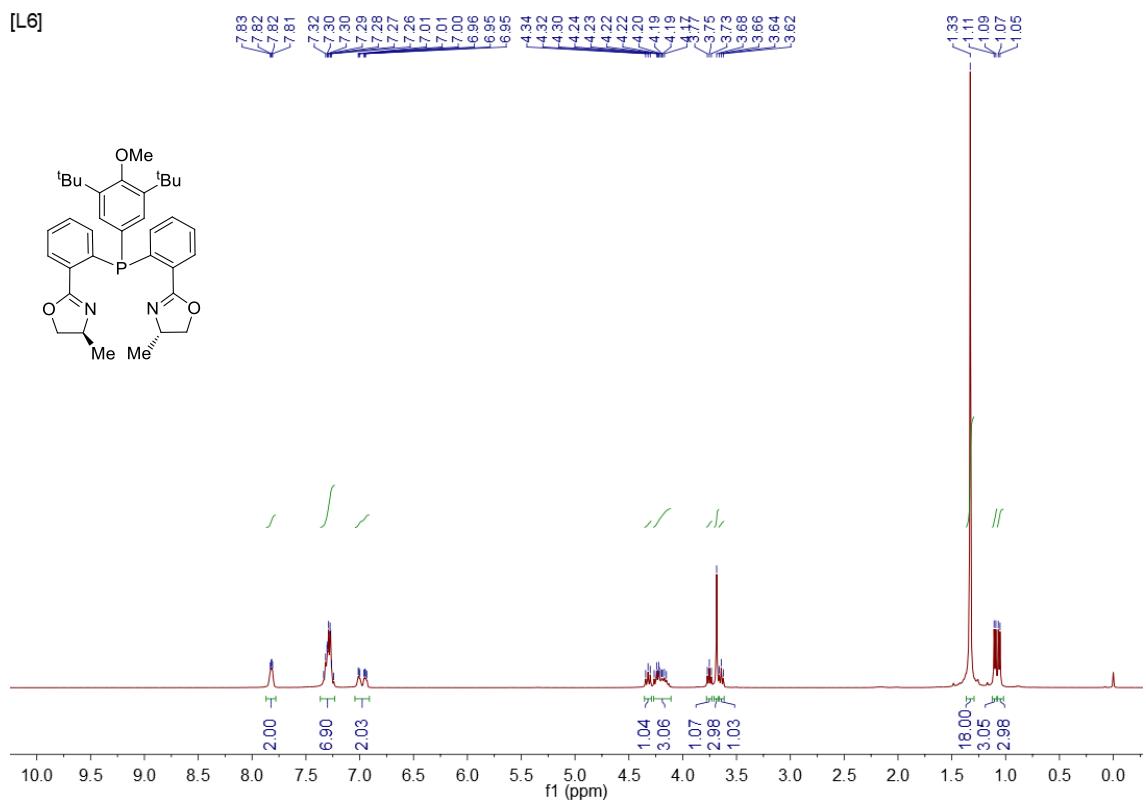
500 MHz, CDCl_3



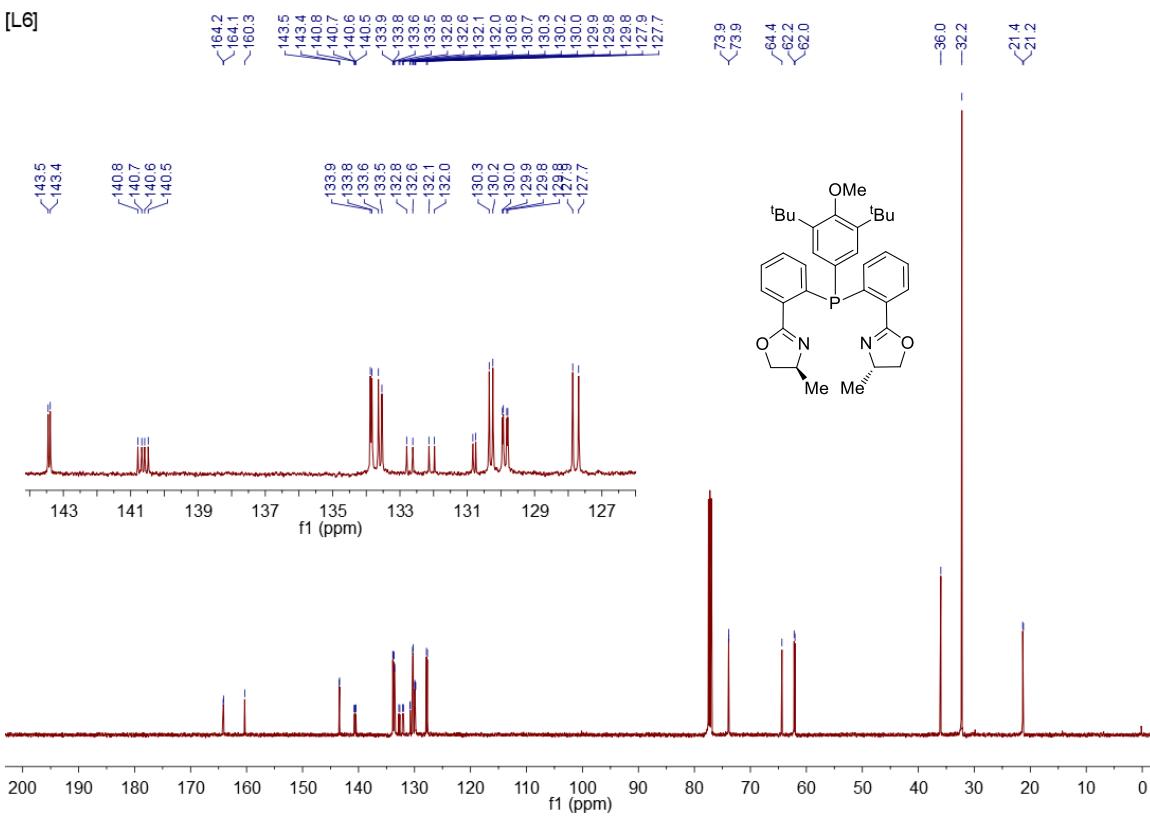
126 MHz, CDCl_3



400 MHz, CDCl₃

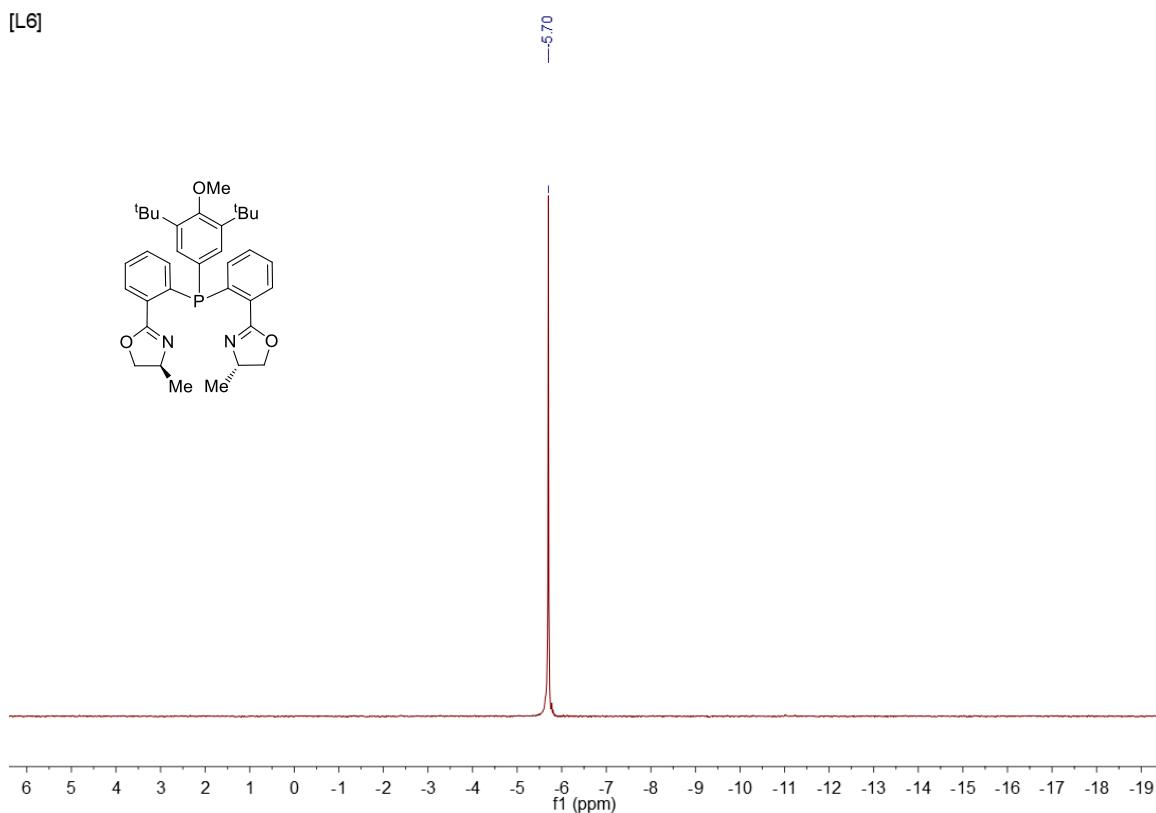


126 MHz, CDCl₃



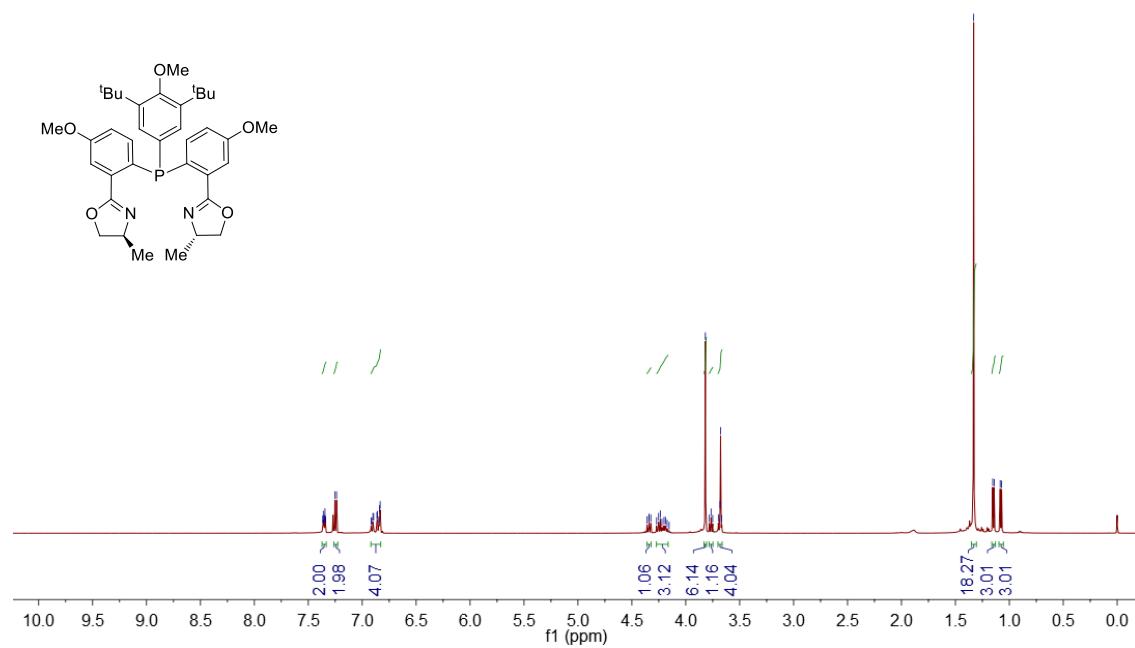
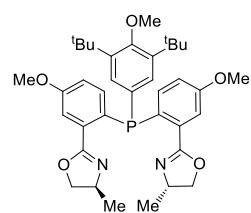
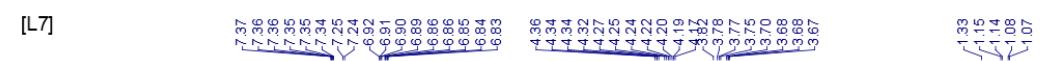
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[L6]

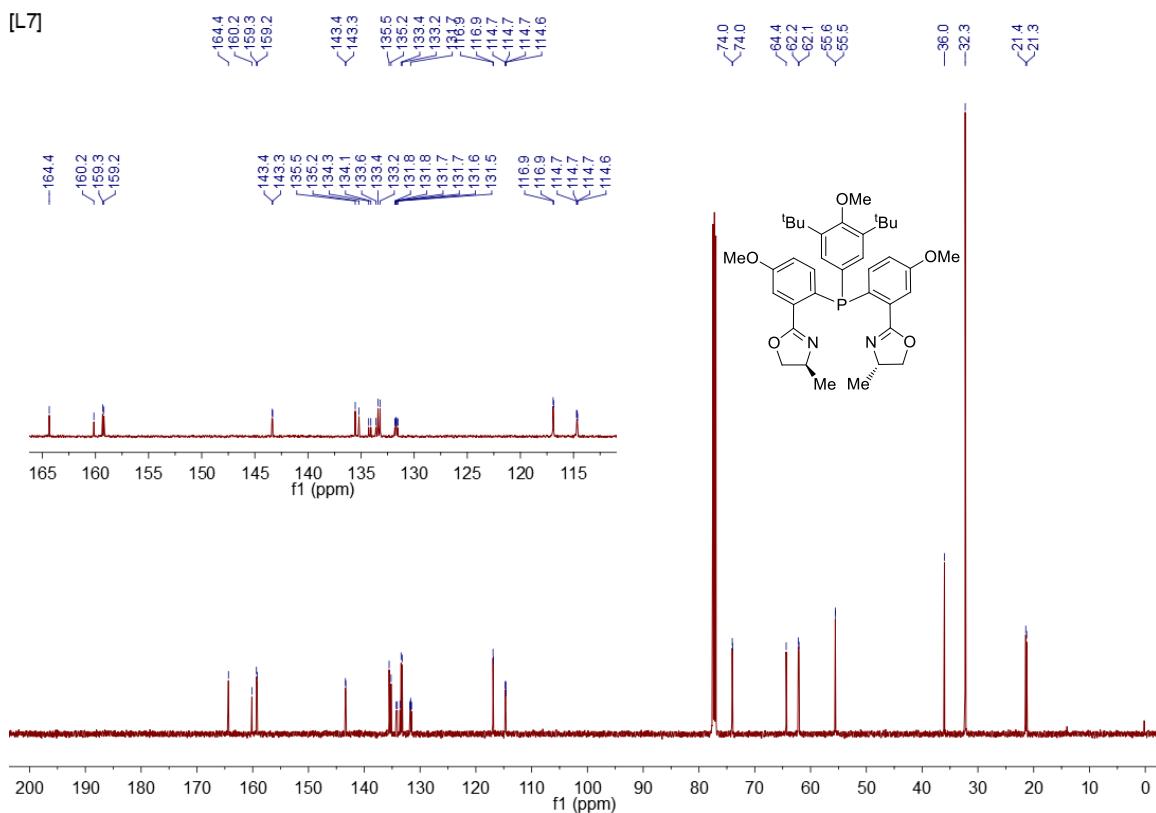


500 MHz, CDCl_3

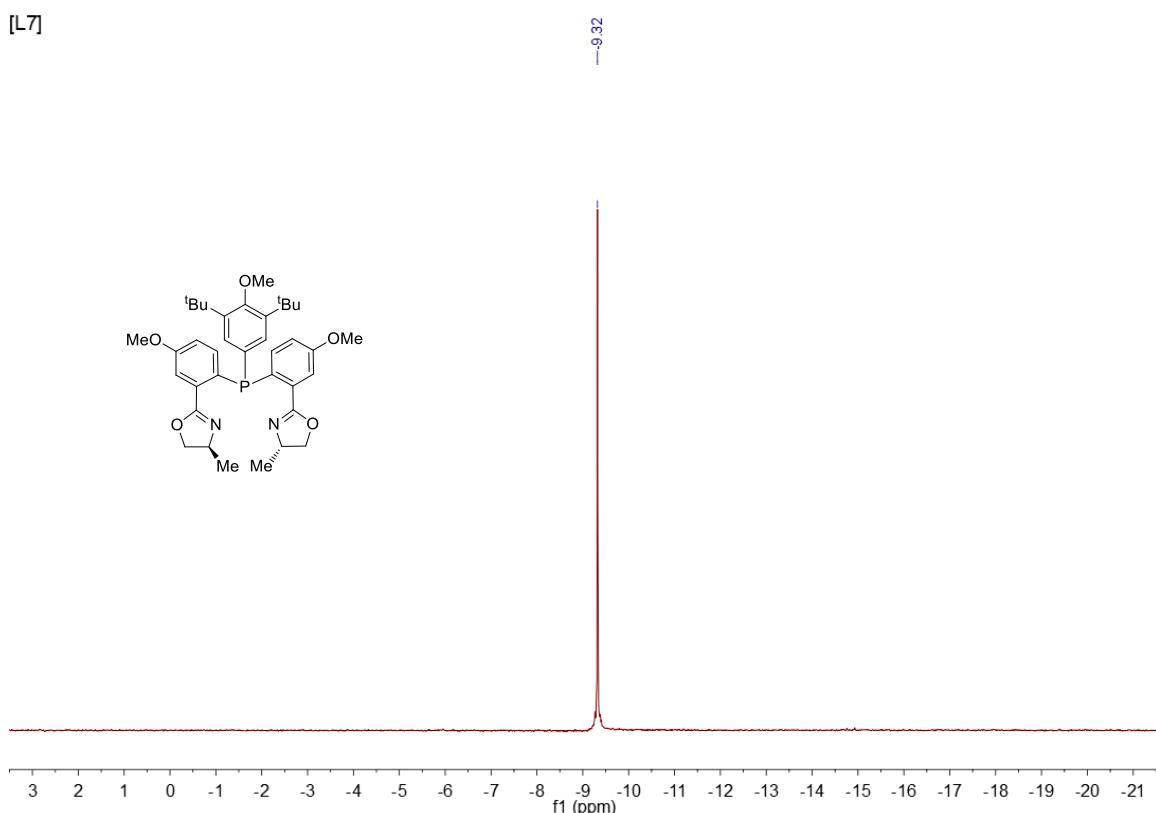
[L7]



126 MHz, CDCl_3

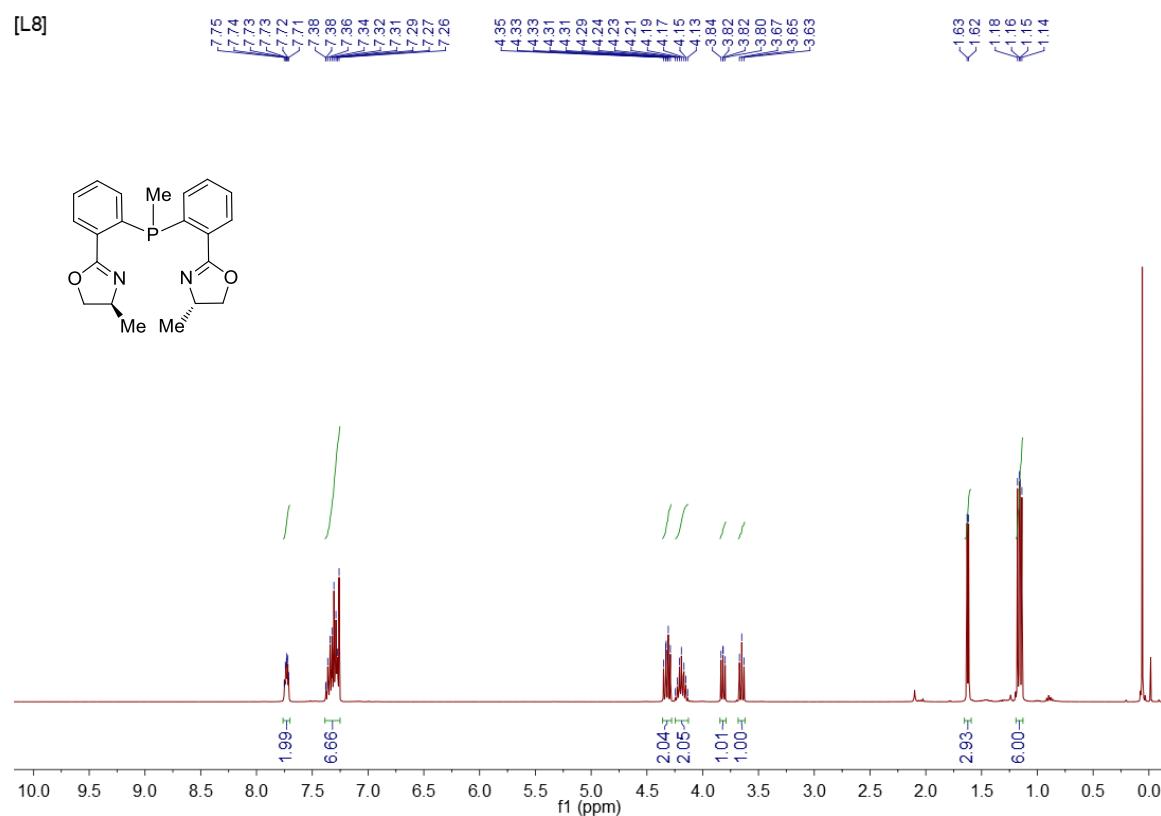


202 MHz, CDCl_3



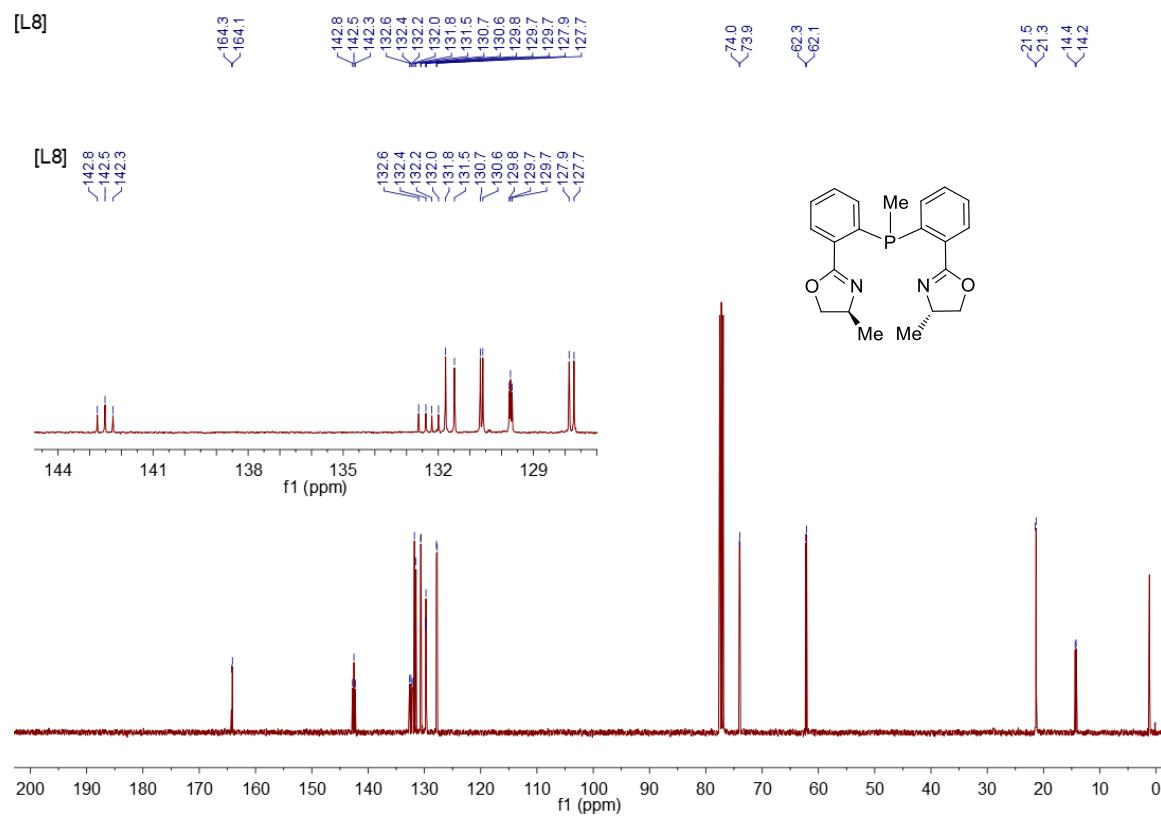
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[L8]



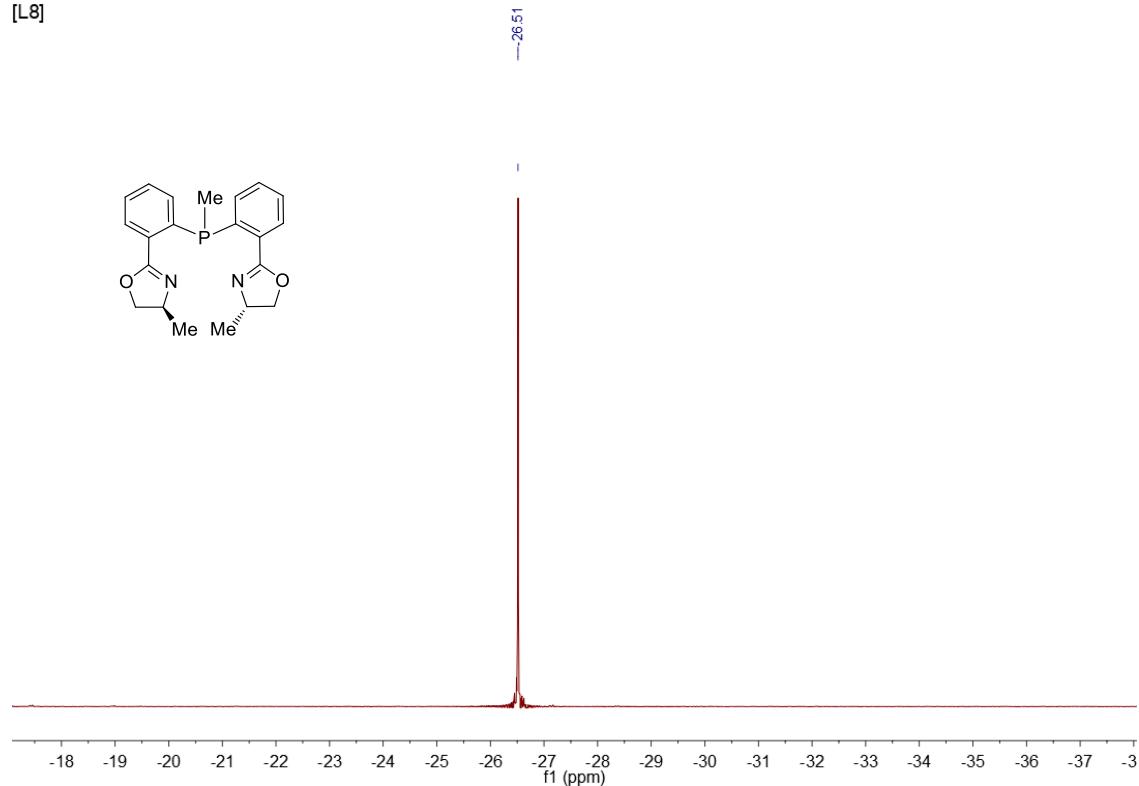
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[L8]



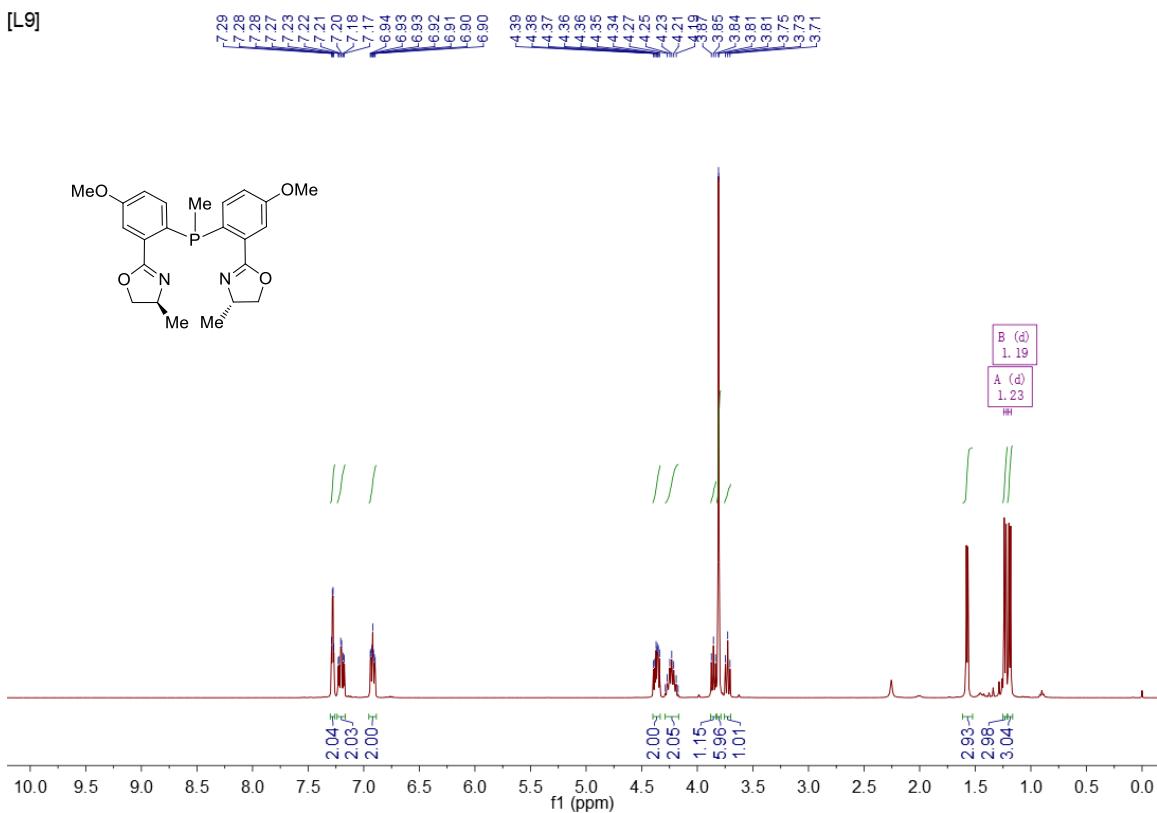
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[L8]



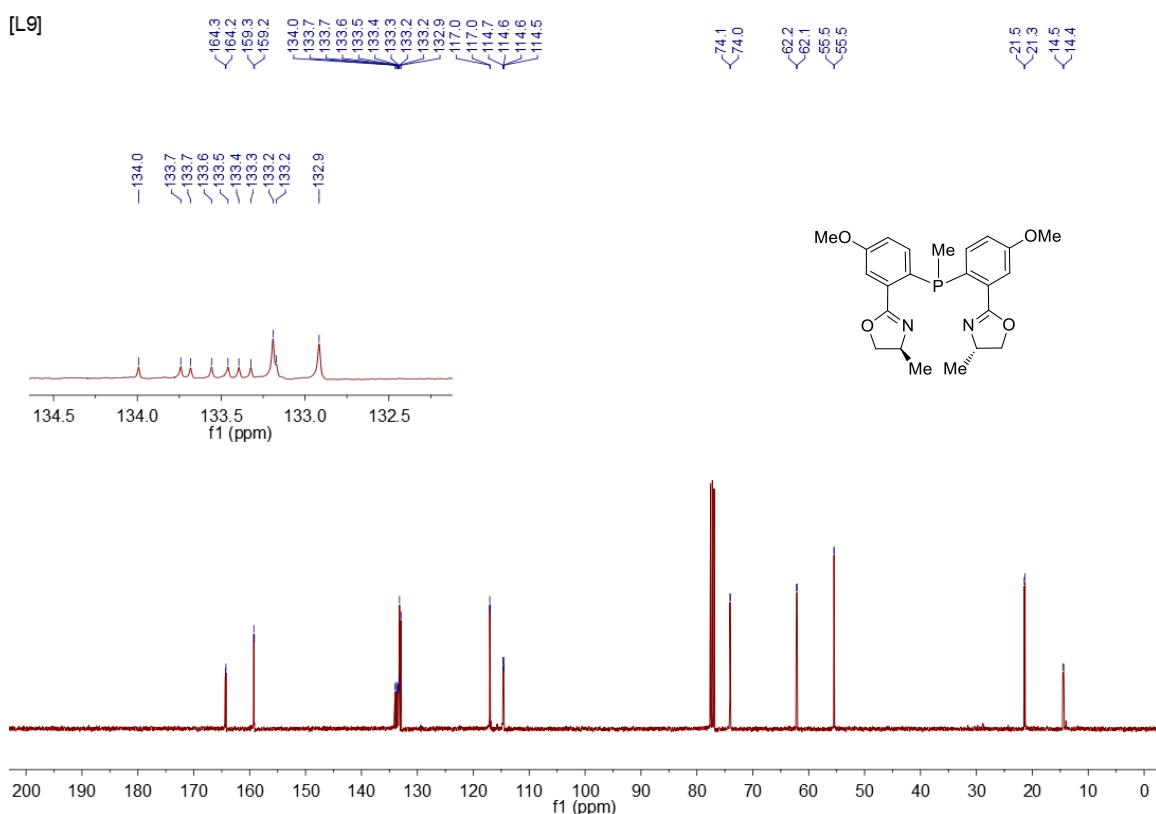
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[L9]



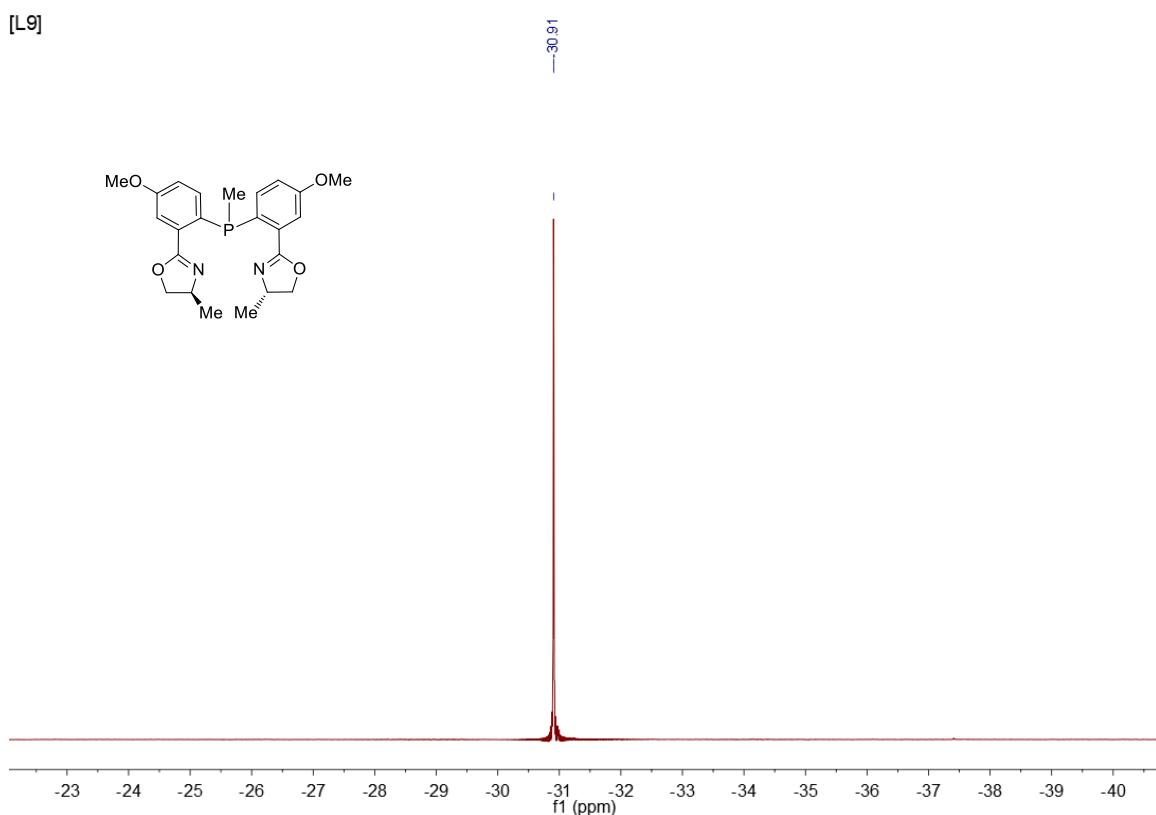
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[L9]



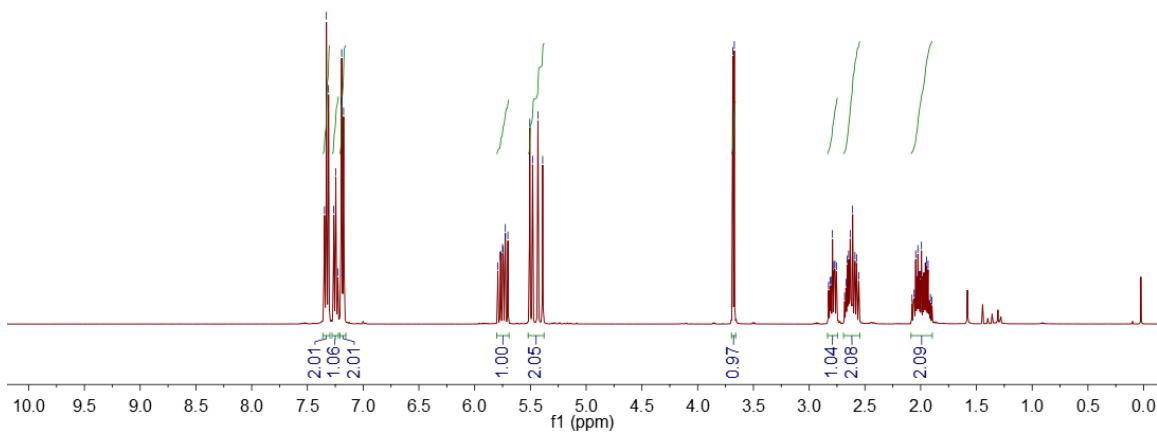
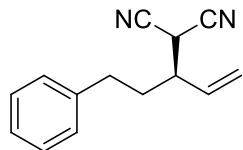
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[L9]



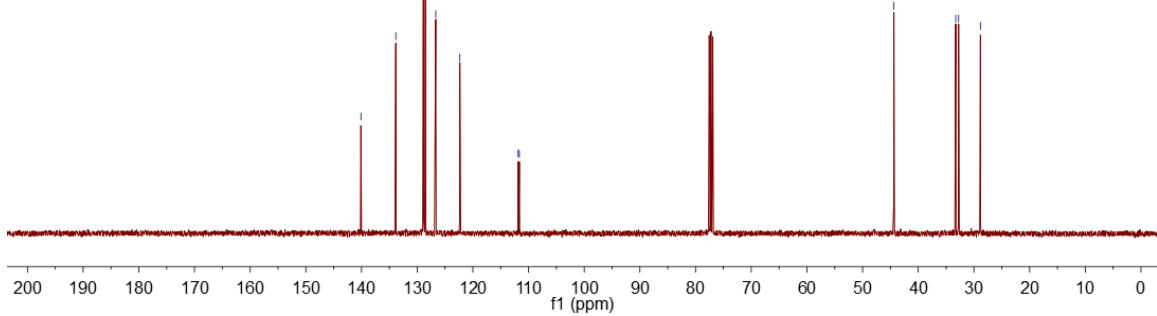
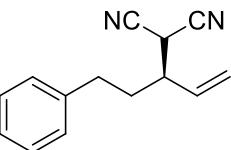
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[3aa]



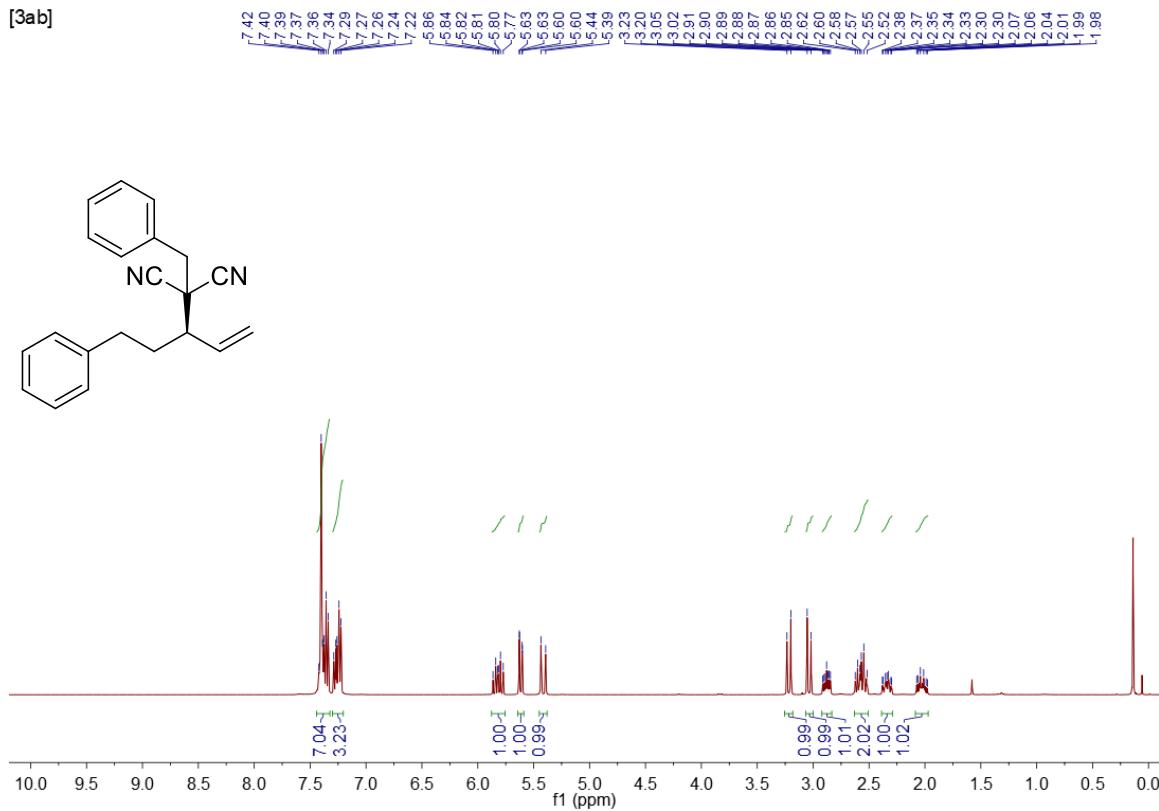
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[3aa]



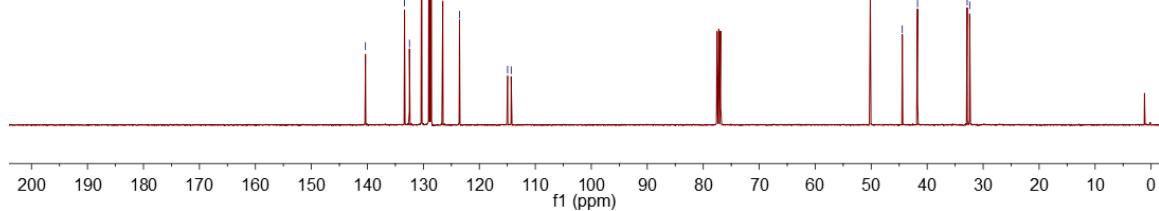
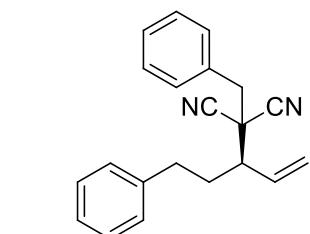
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[3ab]



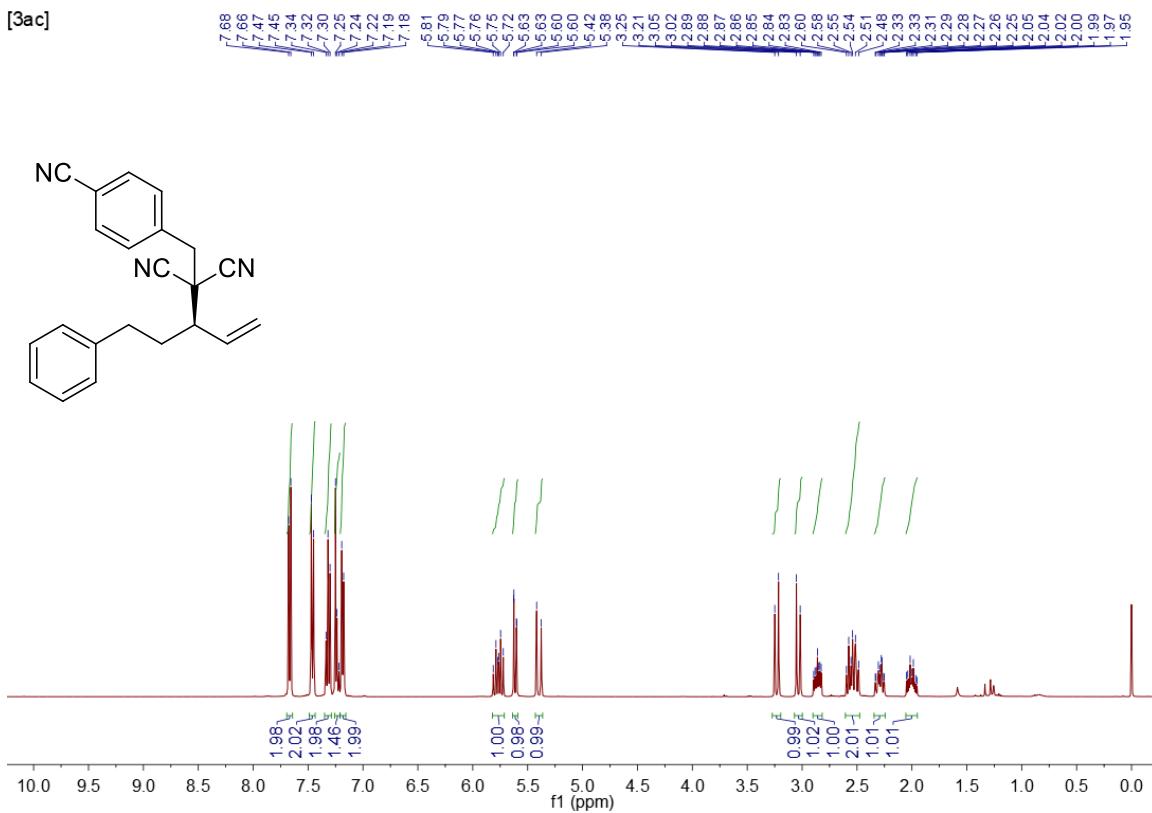
126 MHz, CDCl_3

[3ab]



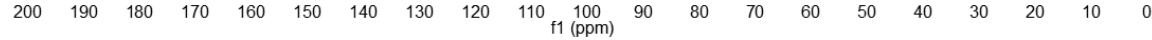
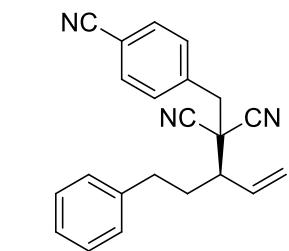
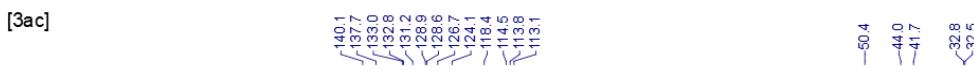
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[3ac]



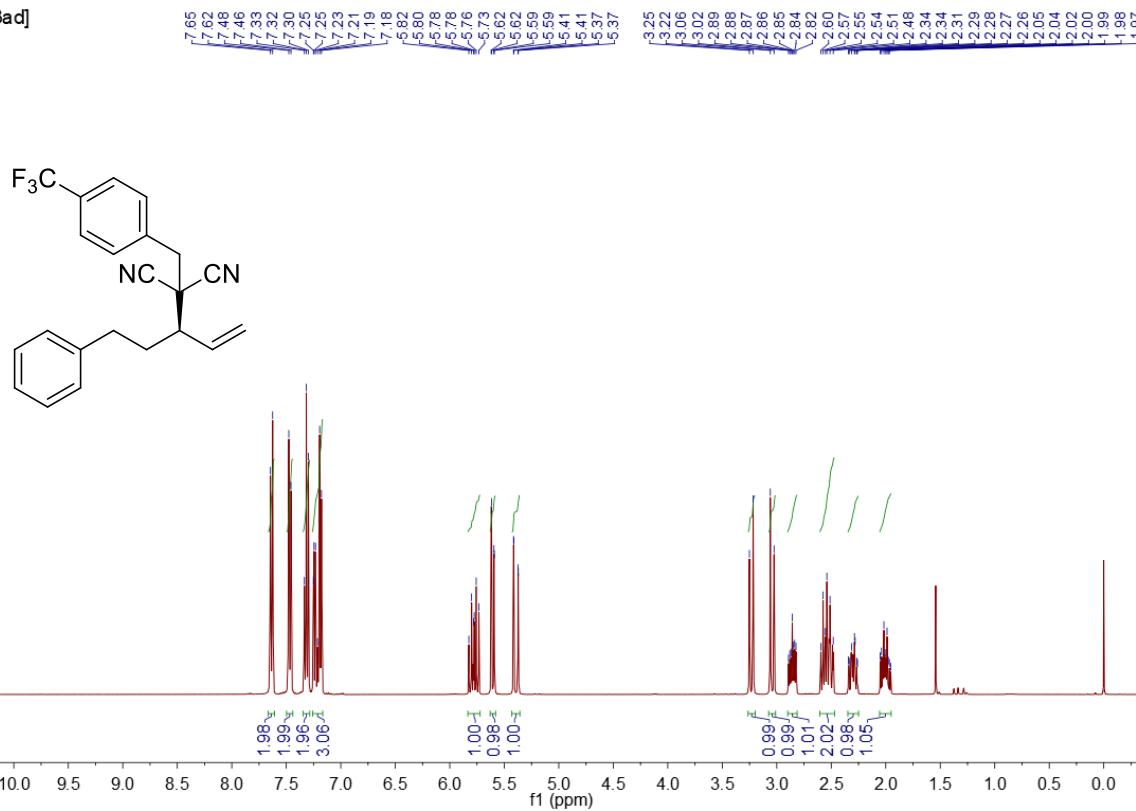
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[3ac]



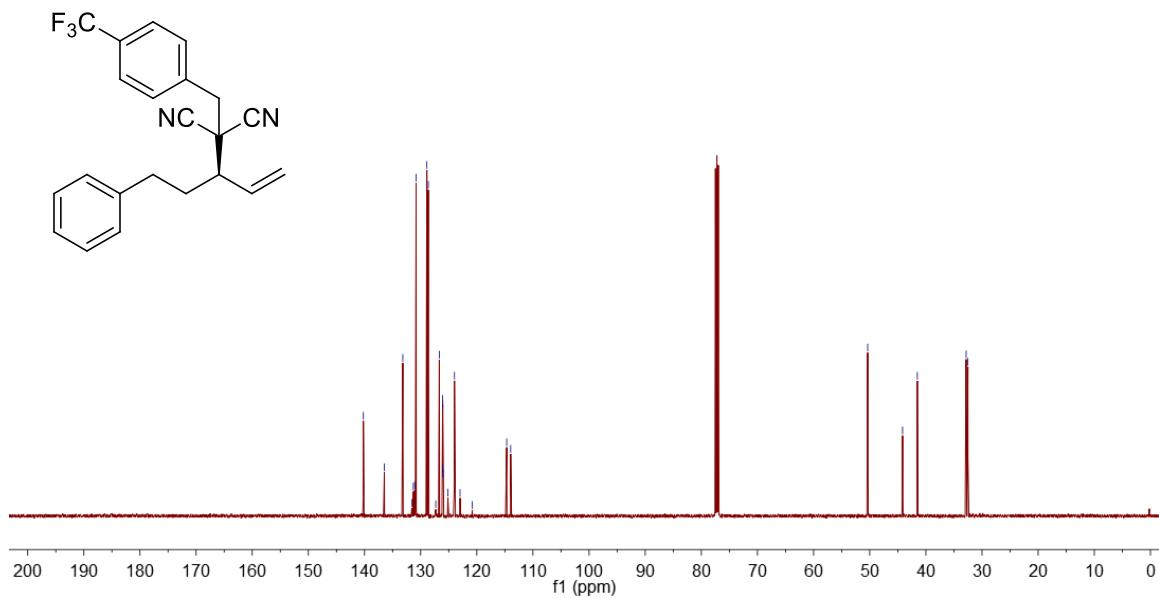
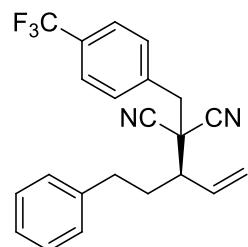
400 MHz, CDCl_3

[3ad]



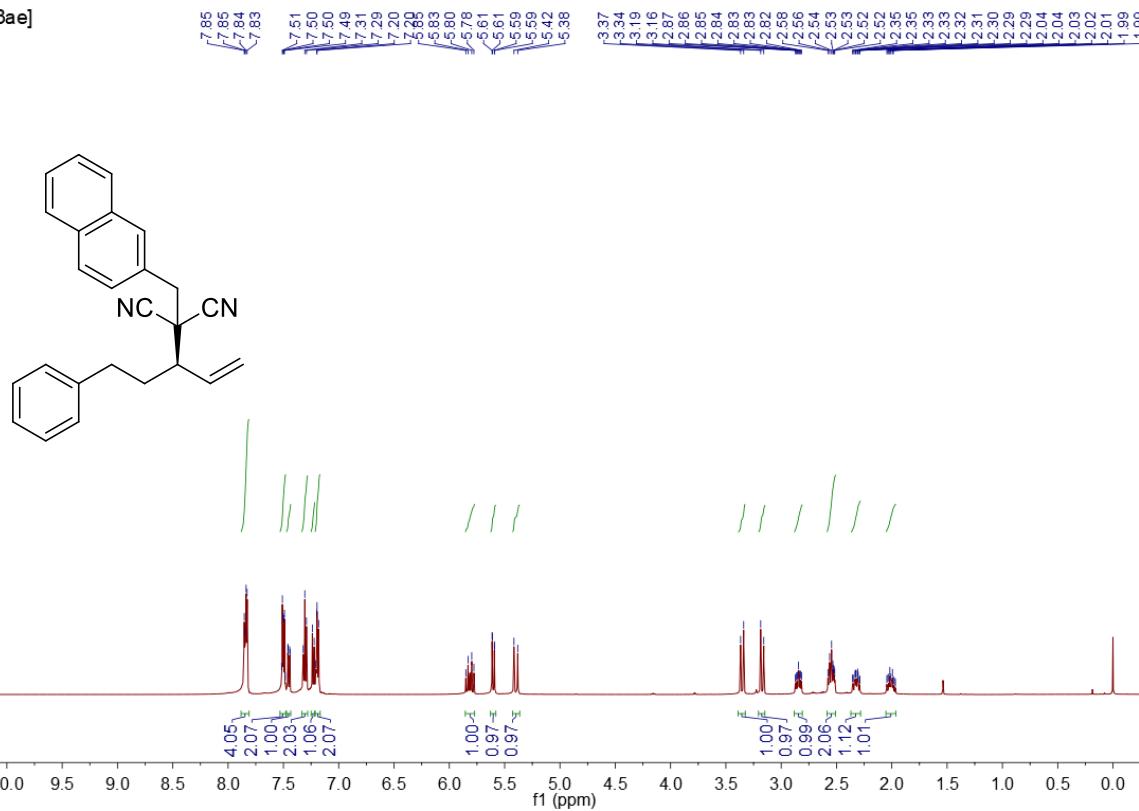
126 MHz, CDCl_3

[3ad]



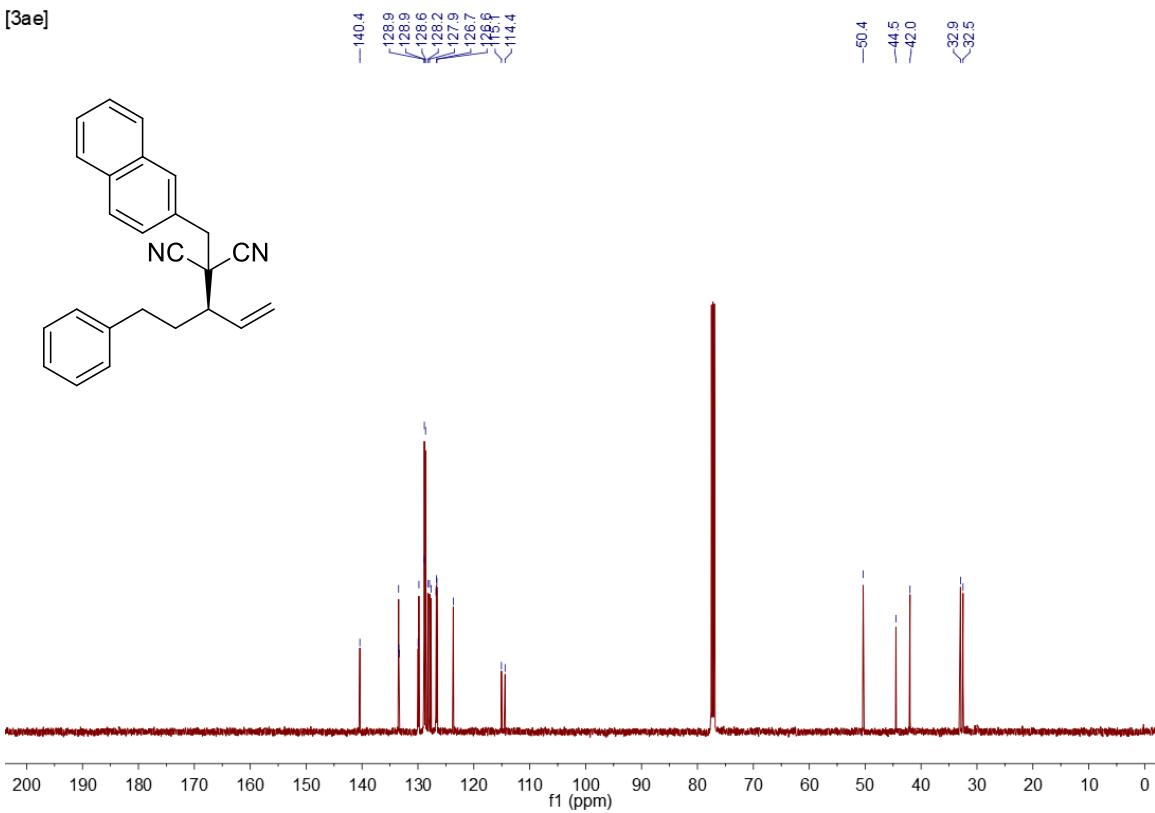
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[3ae]



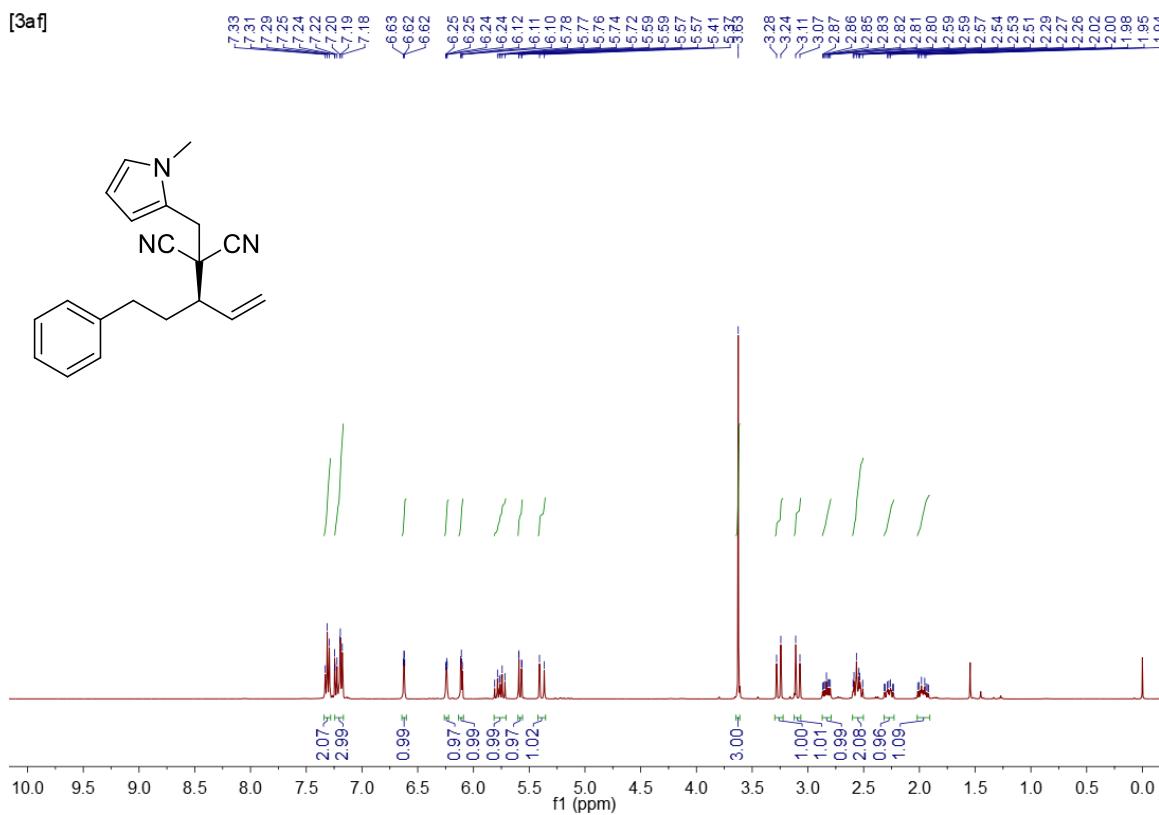
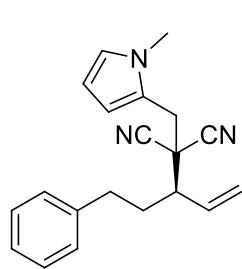
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[3aε]



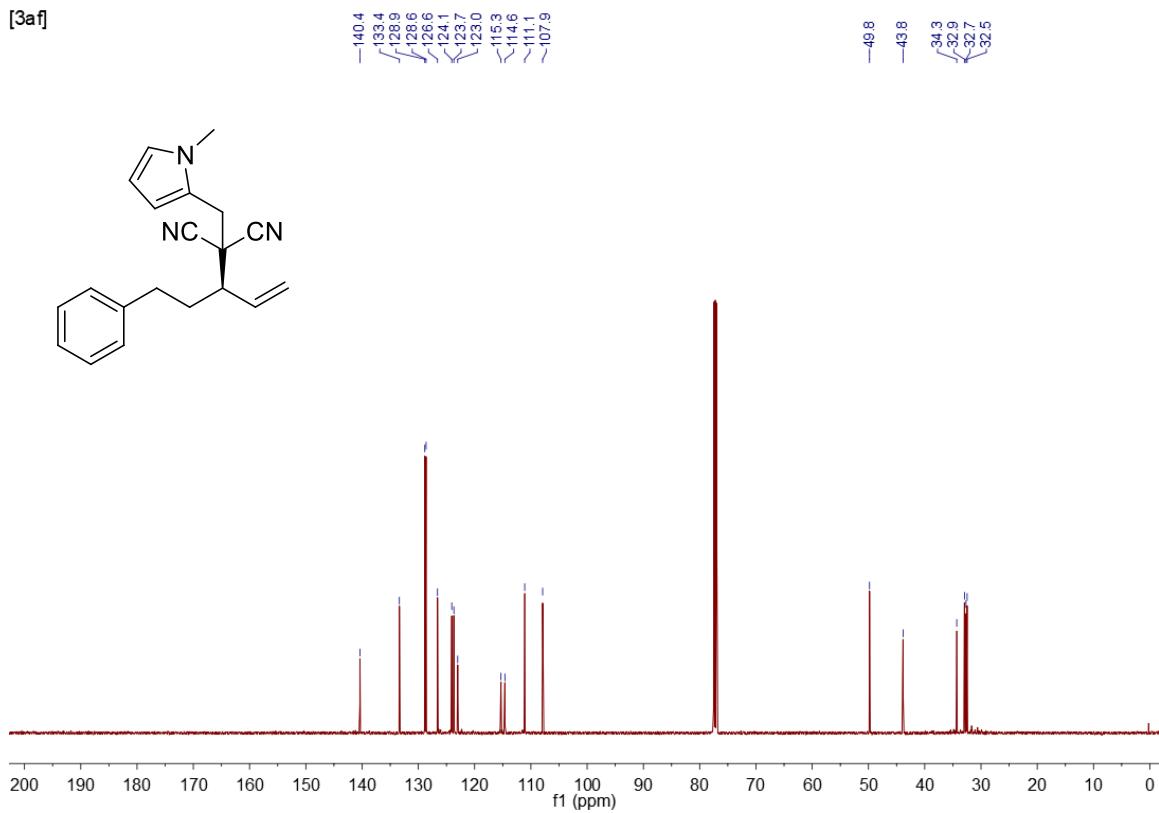
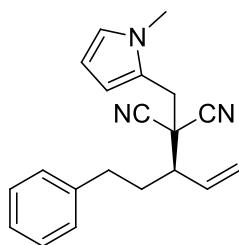
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[3af]



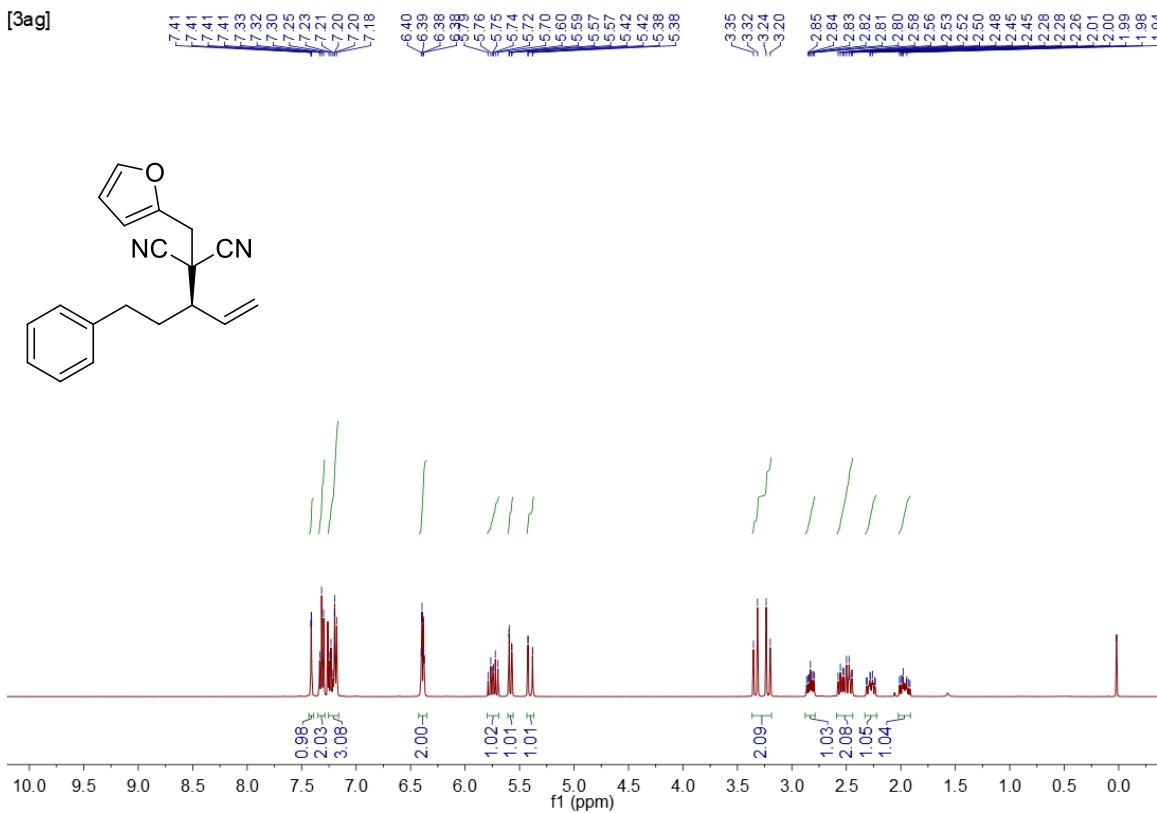
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[3af]



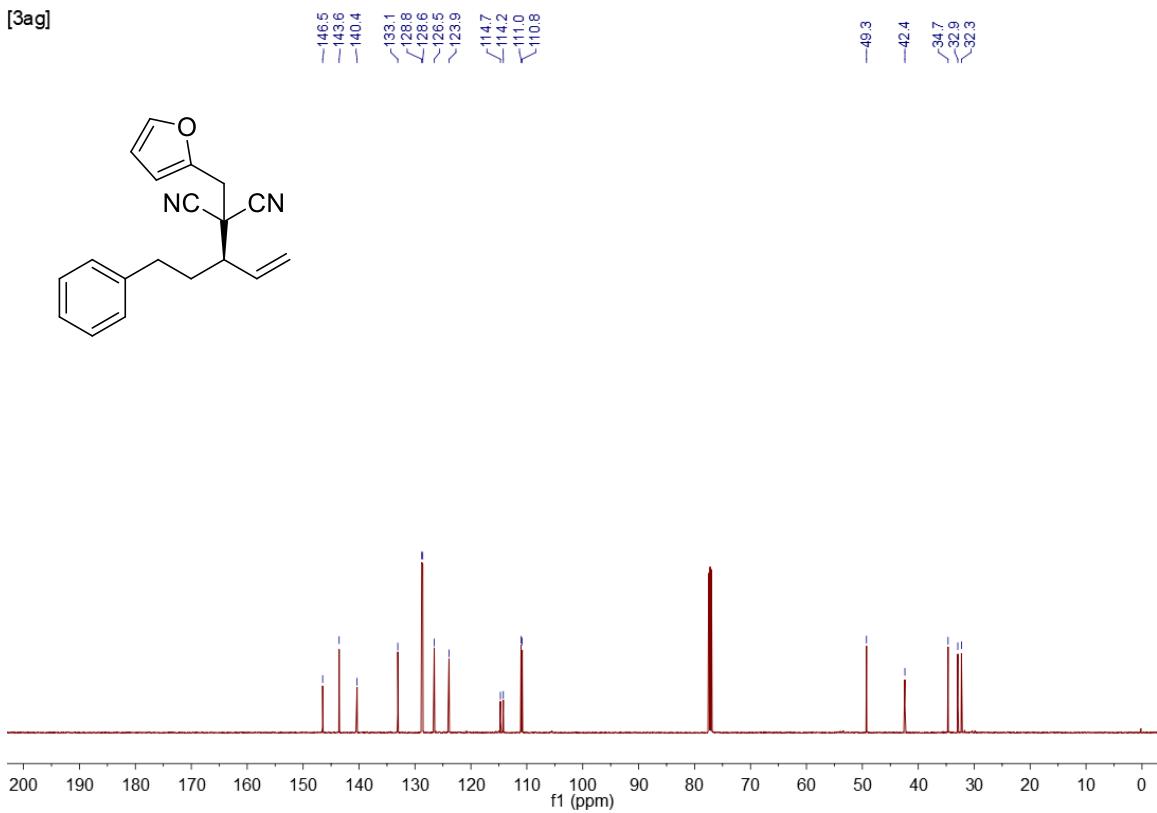
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[3ag]



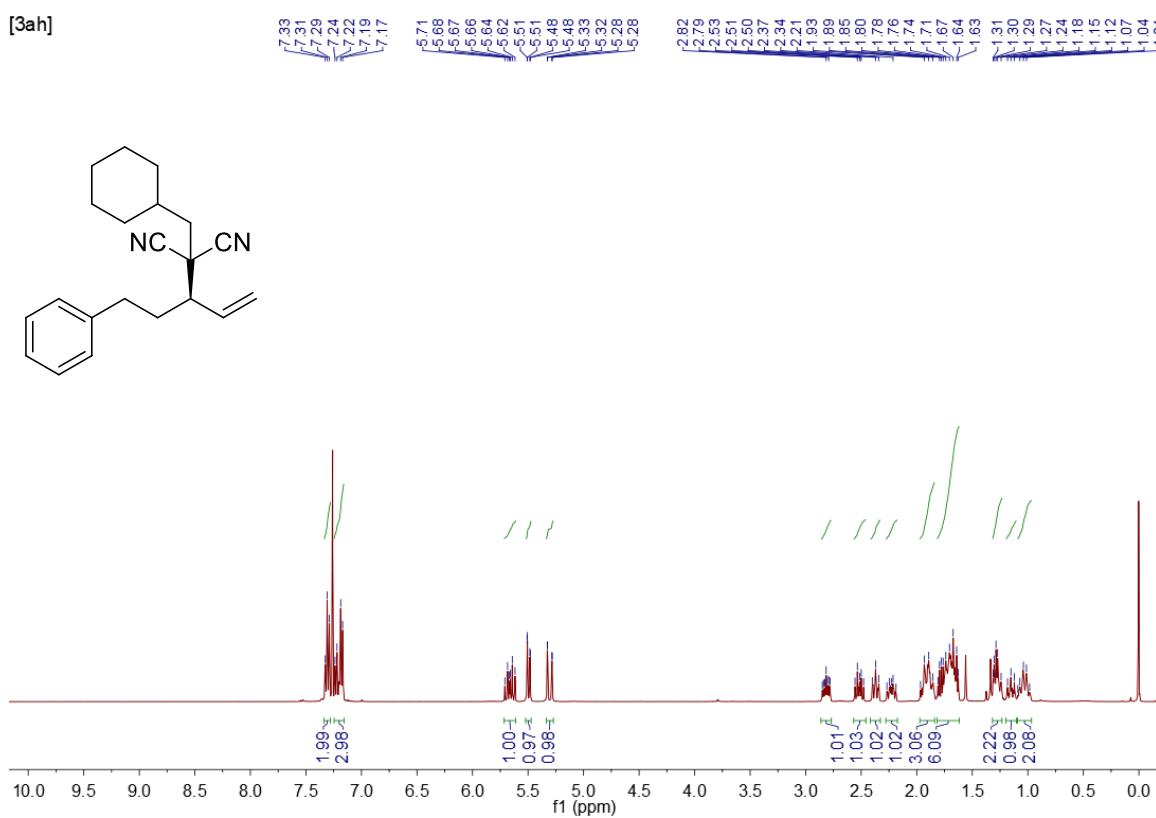
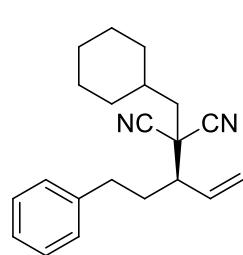
126 MHz, CDCl_3

[3ag]



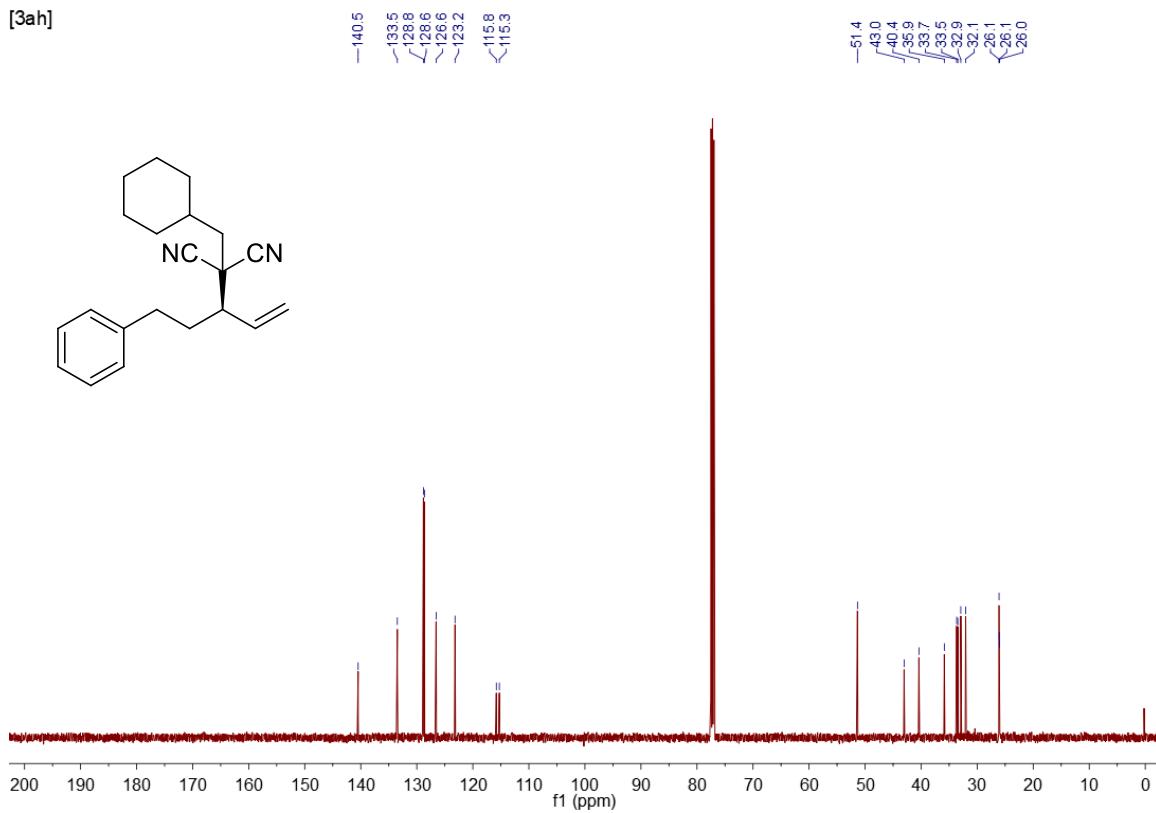
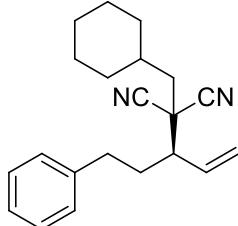
400 MHz, CDCl_3

[3ah]



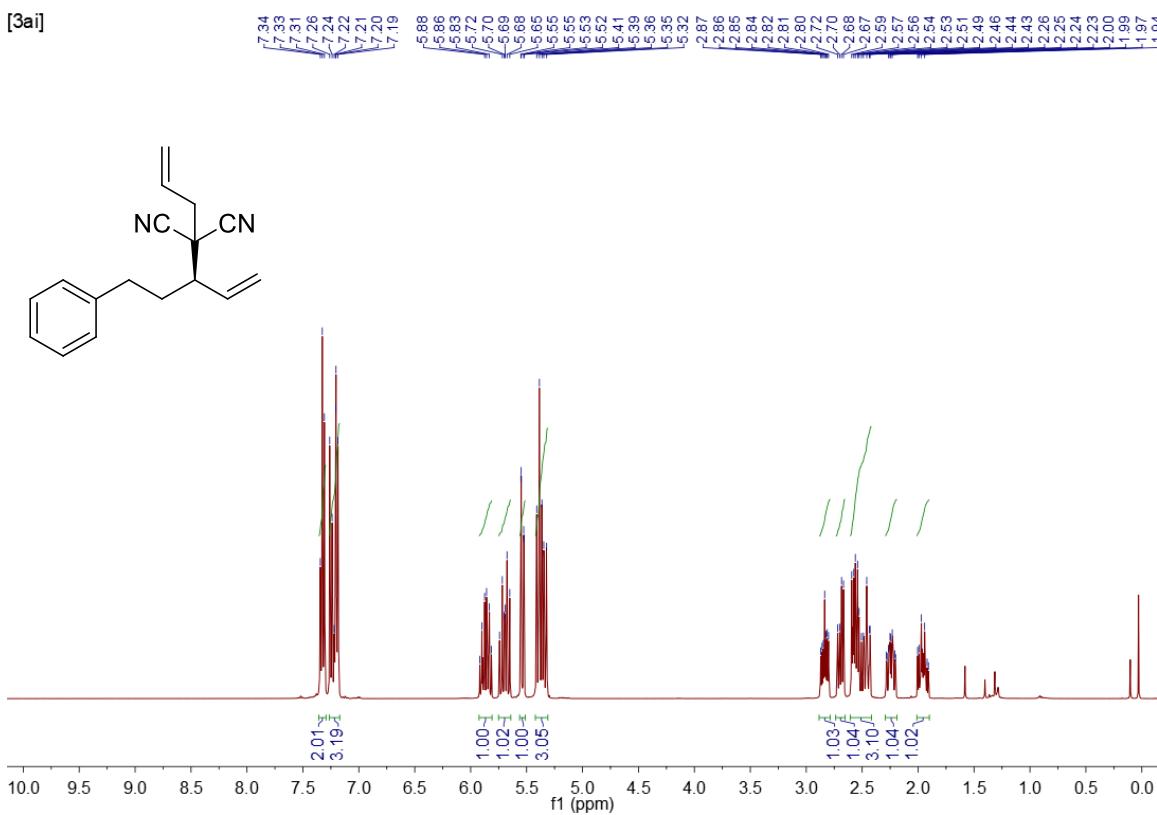
126 MHz, CDCl_3

[3ah]



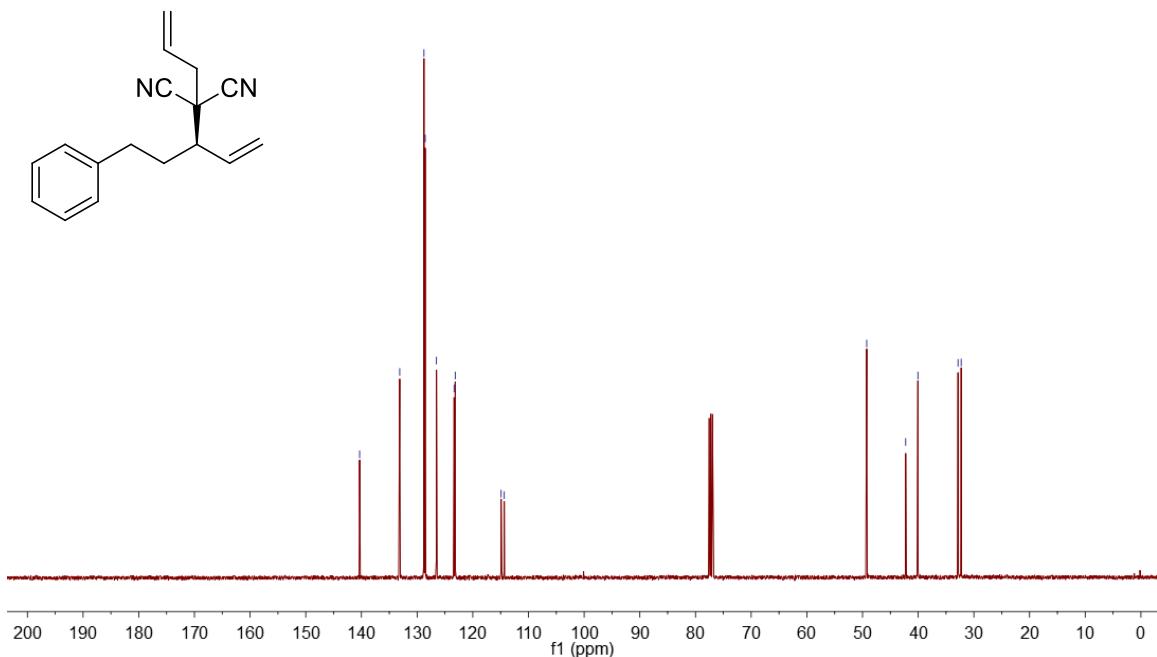
400 MHz, CDCl_3

[3ai]



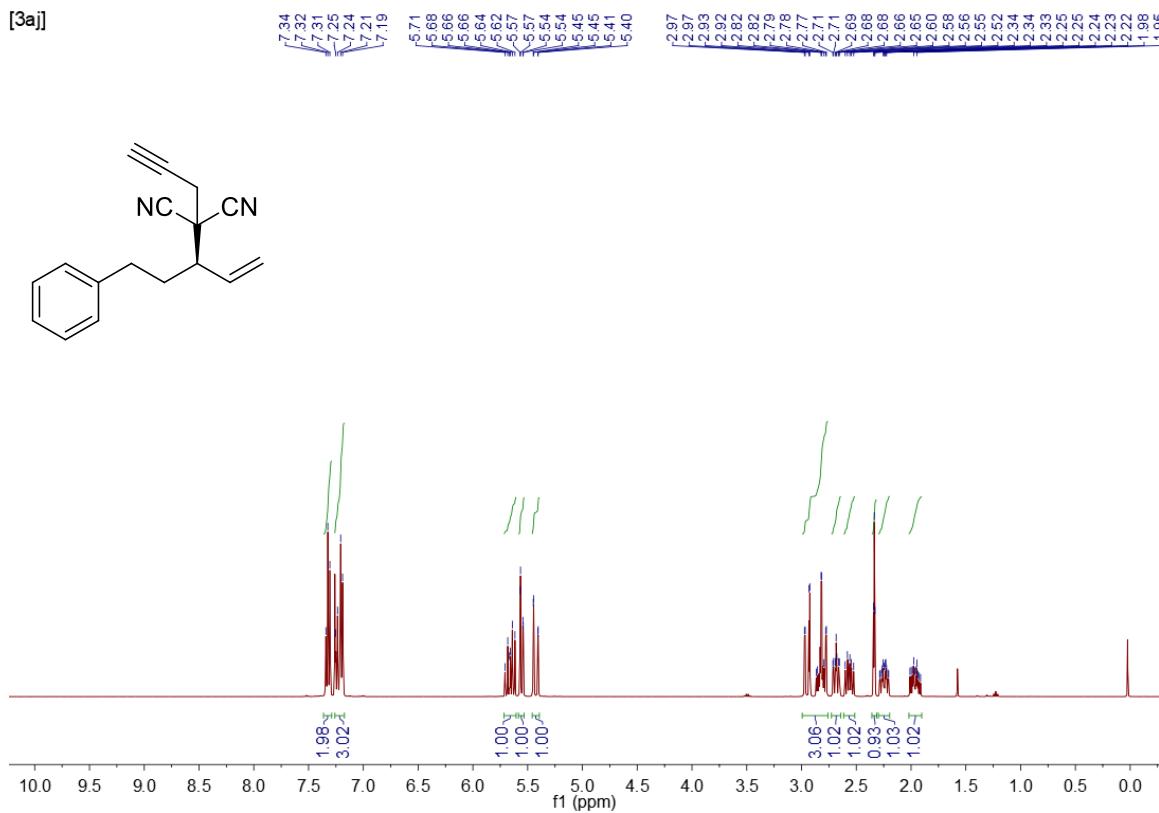
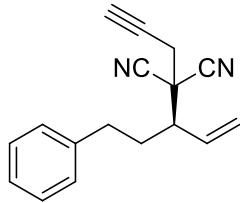
101 MHz, CDCl_3

[3ai]



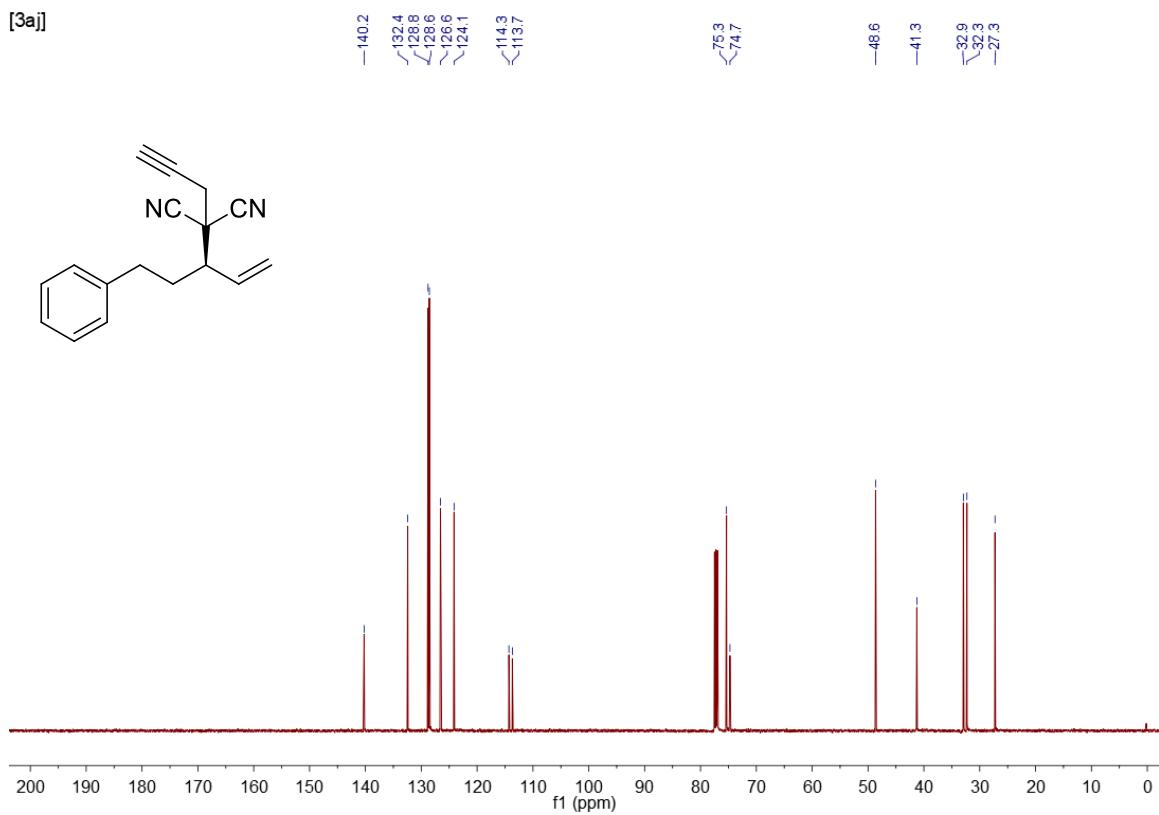
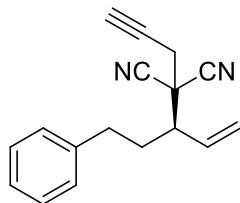
400 MHz, CDCl_3

[3aj]



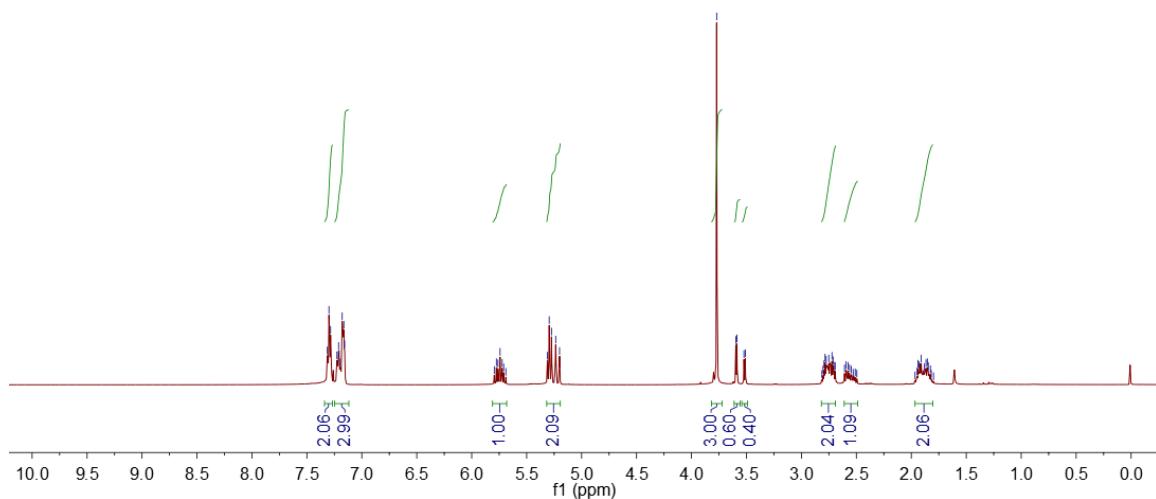
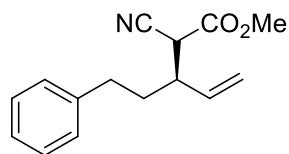
126 MHz, CDCl_3

[3aj]



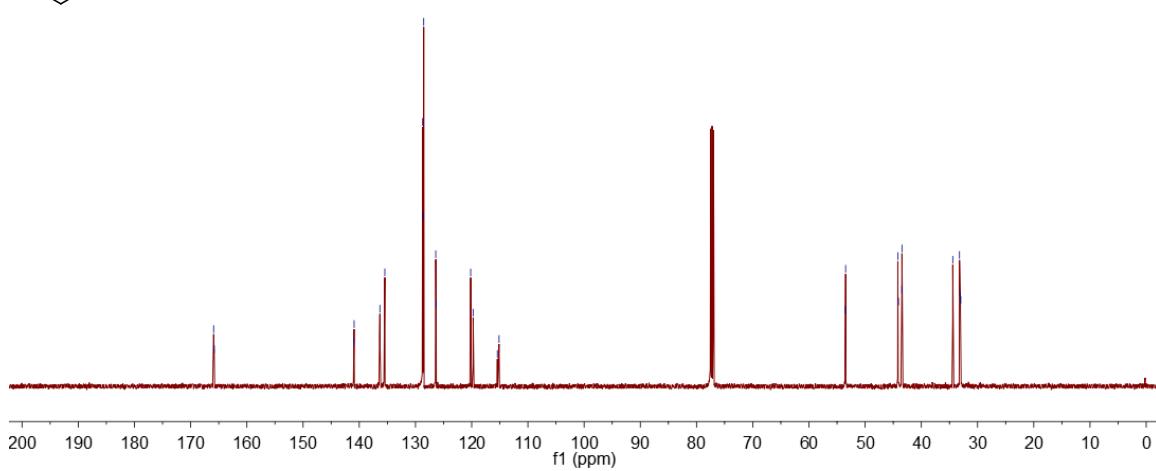
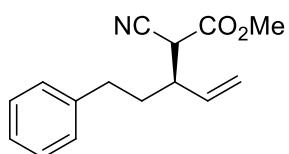
500 MHz, CDCl_3

[3ak]



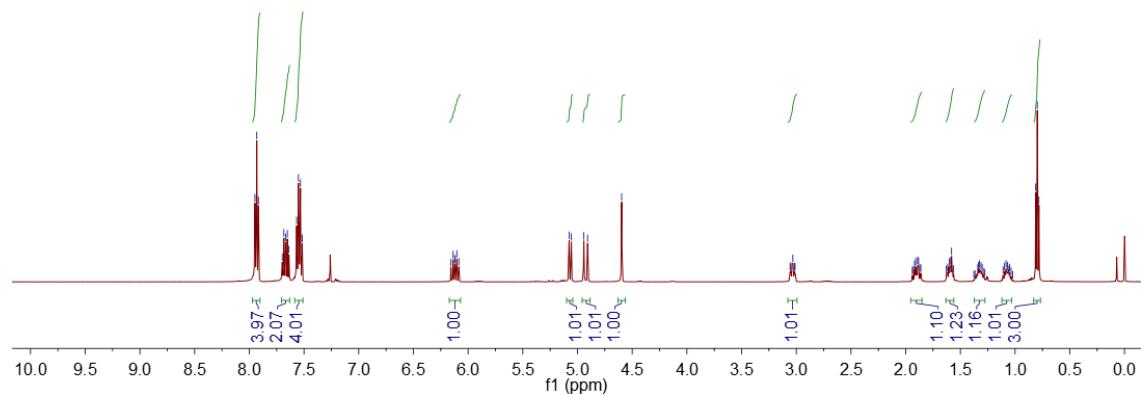
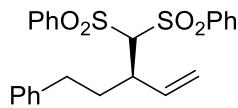
126 MHz, CDCl_3

[3ak]



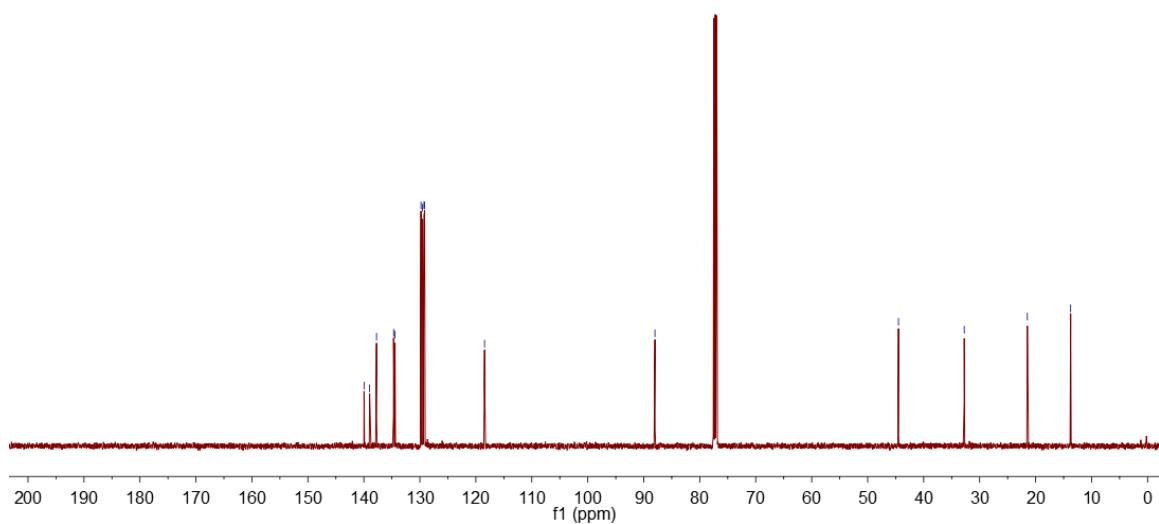
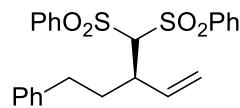
500 MHz, CDCl_3

[3a l]



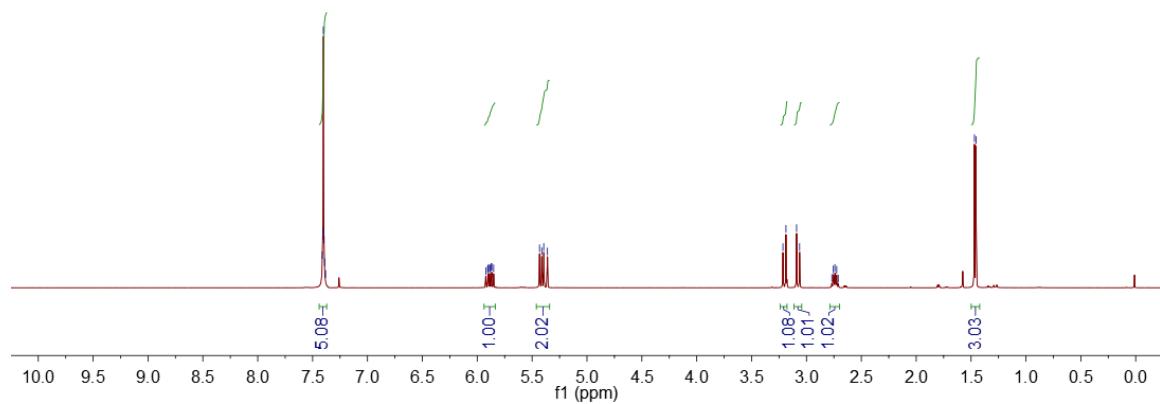
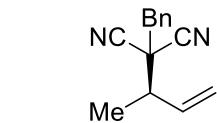
126 MHz, CDCl₃

[3aI]



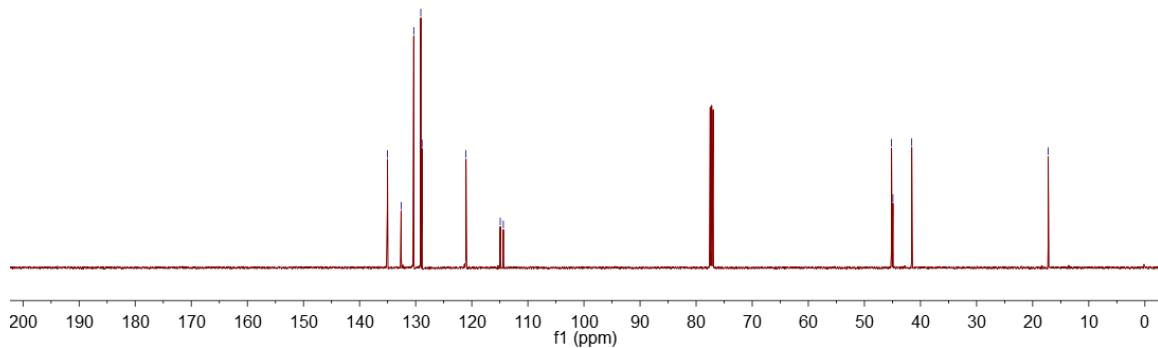
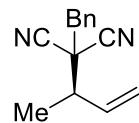
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[3bb]



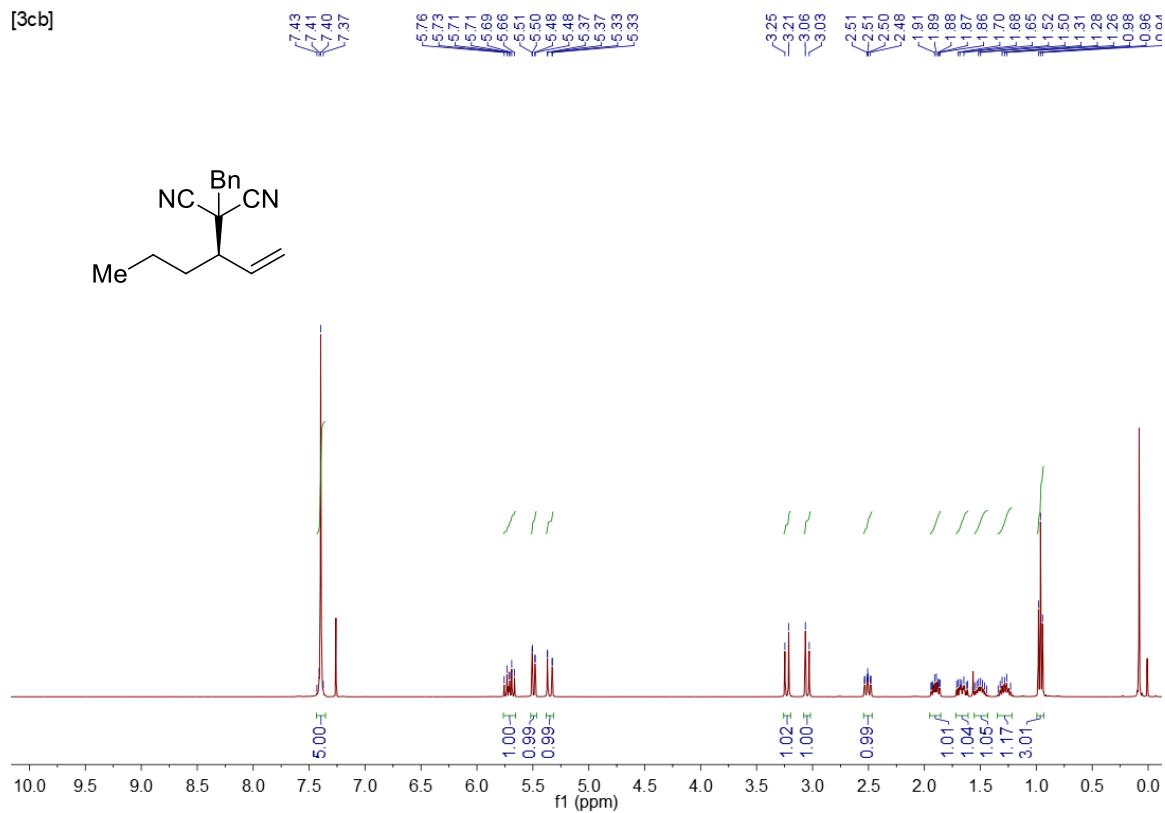
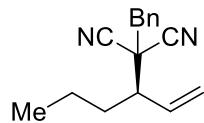
126 MHz, CDCl_3

[3bb]



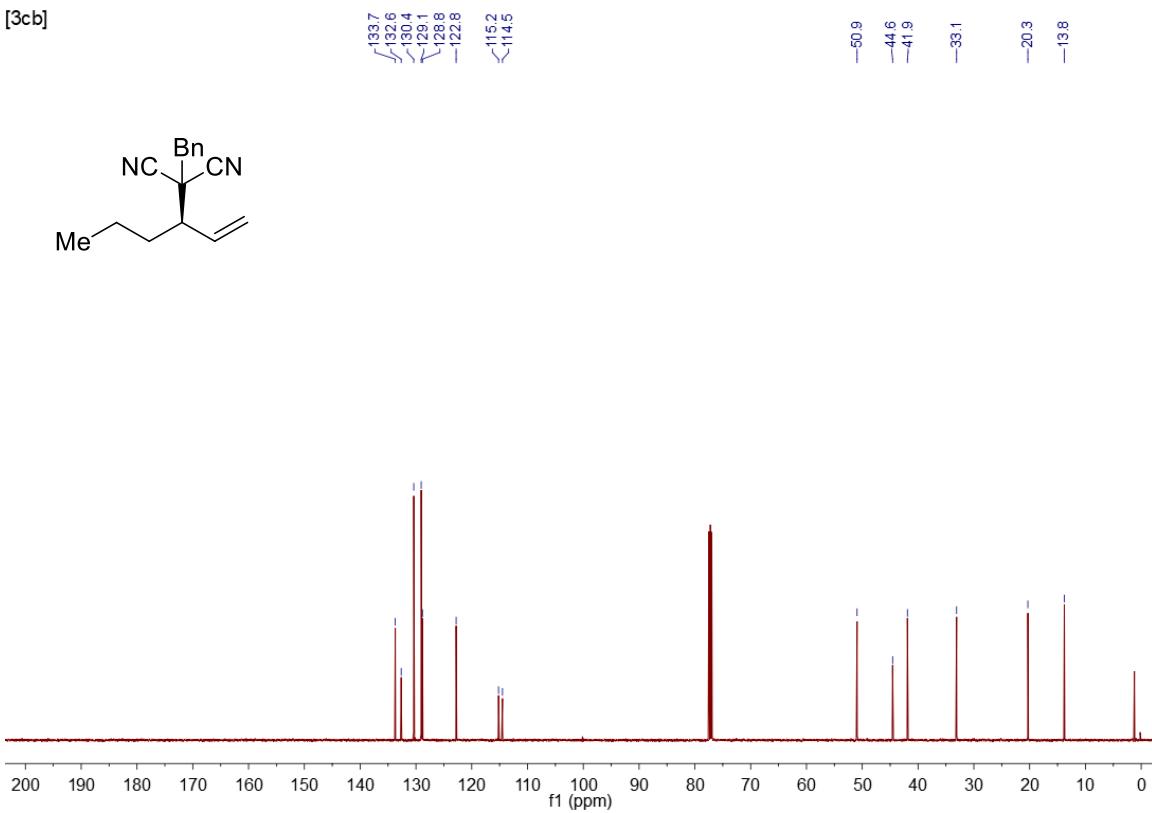
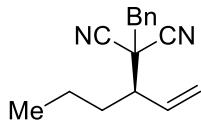
400 MHz, CDCl_3

[3cb]

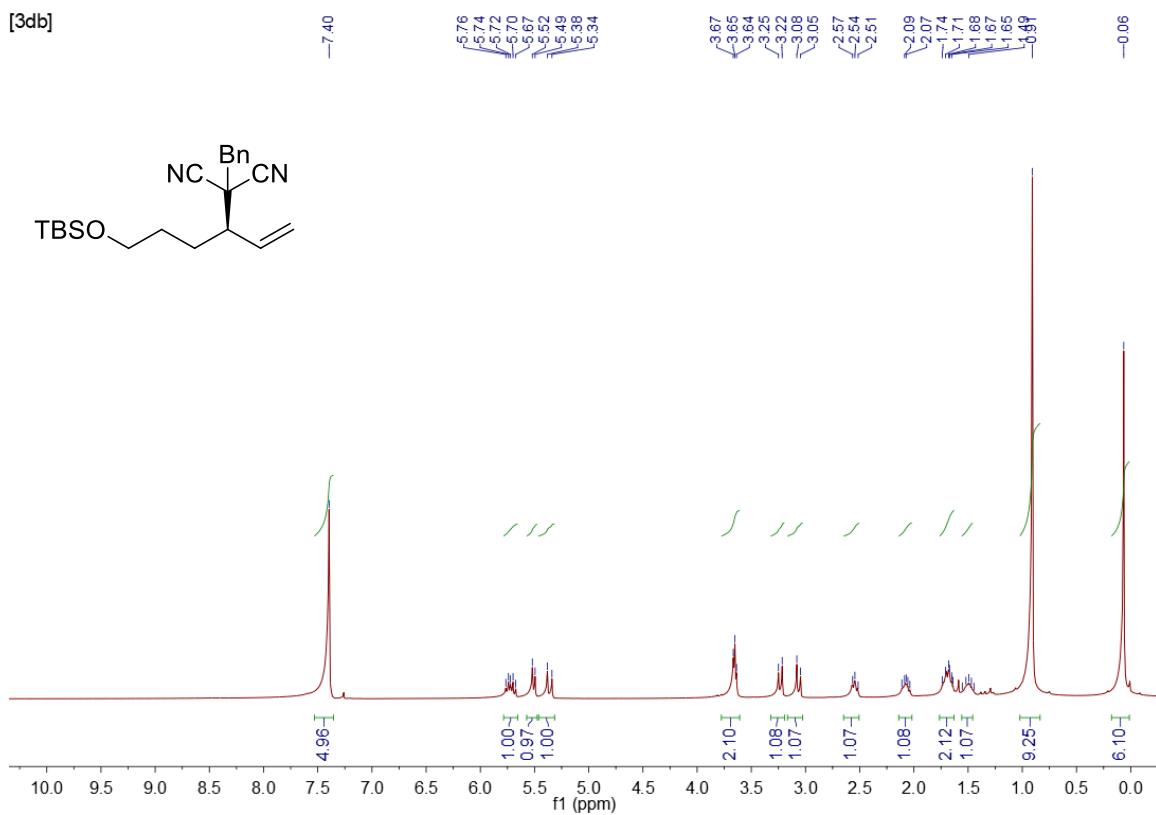


126 MHz, CDCl_3

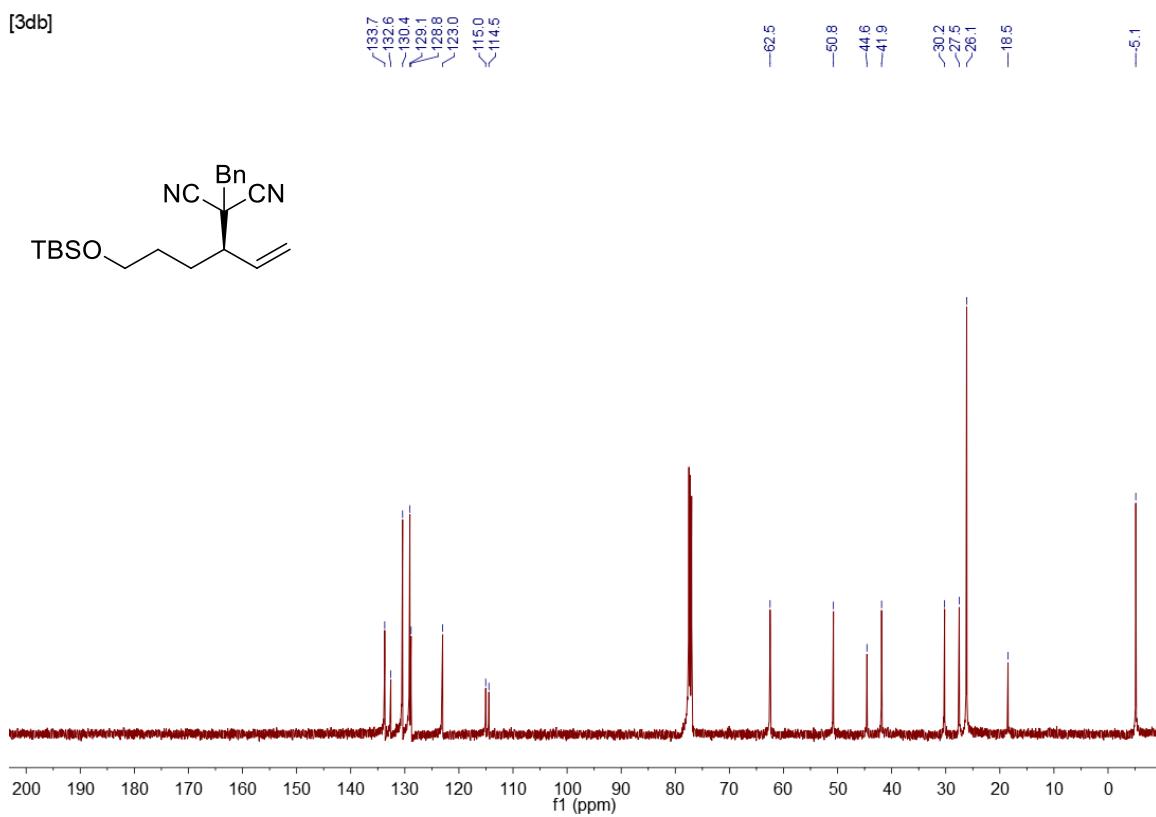
[3cb]



400 MHz, CDCl_3

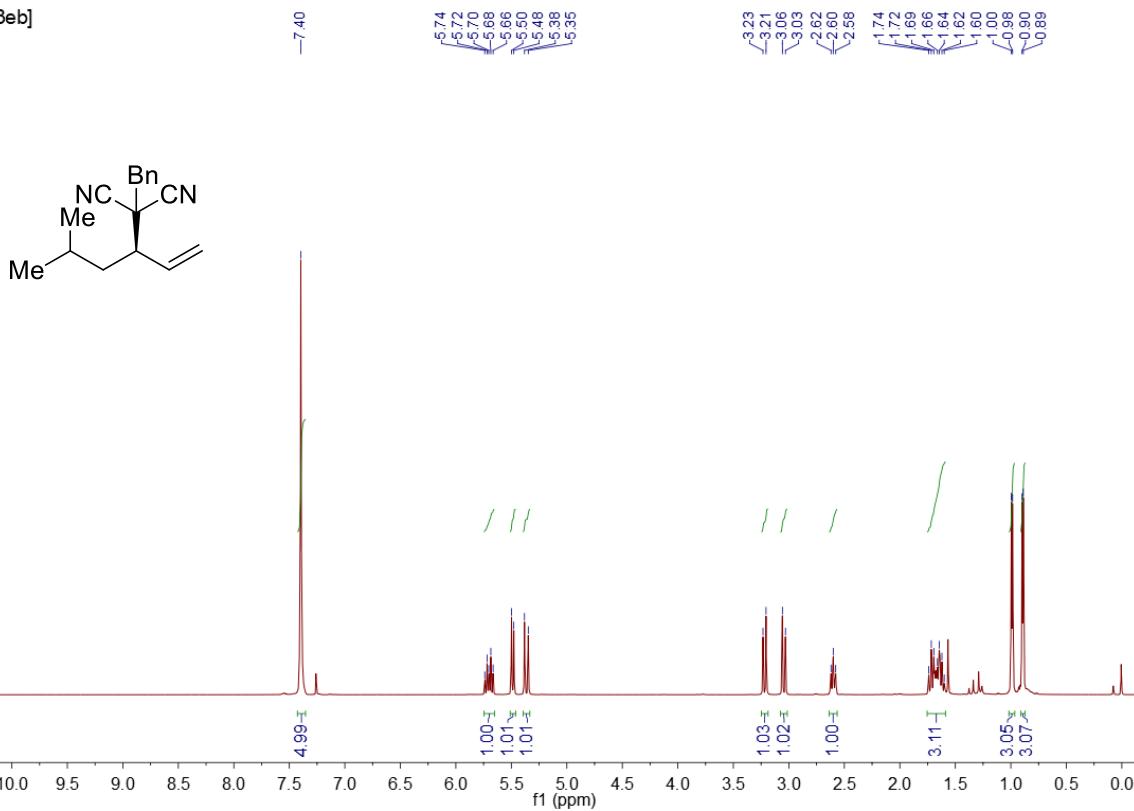


126 MHz, CDCl_3



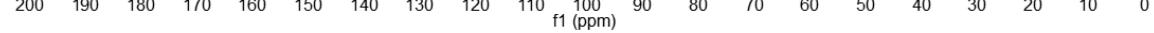
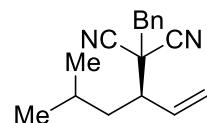
500 MHz, CDCl_3

[3eb]

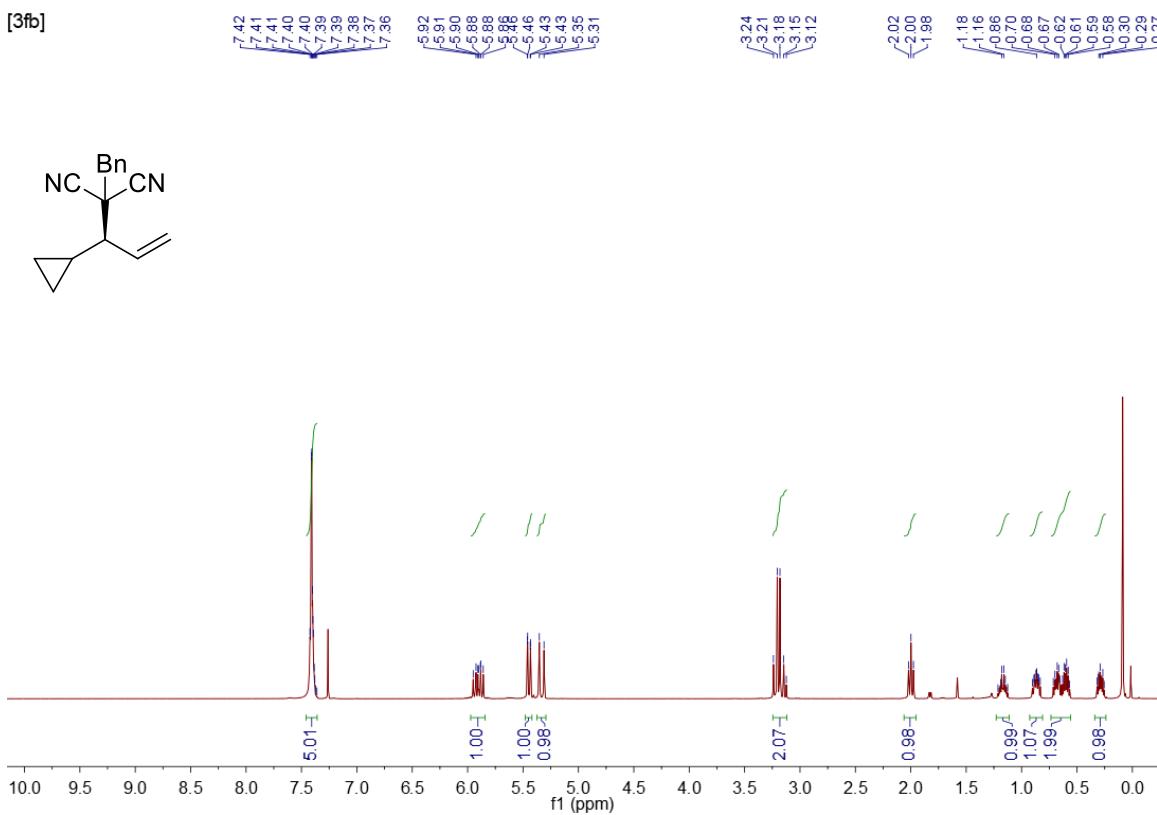


126 MHz, CDCl_3

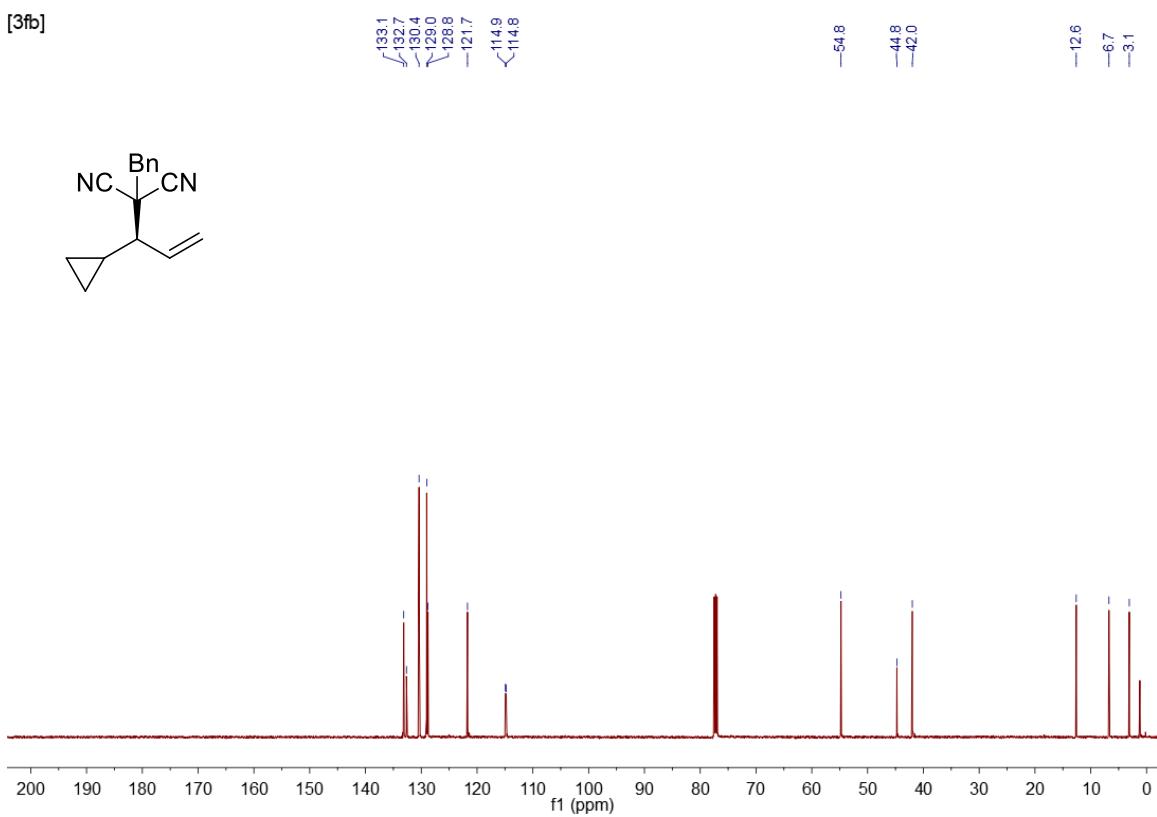
[3eb]



400 MHz, CDCl_3

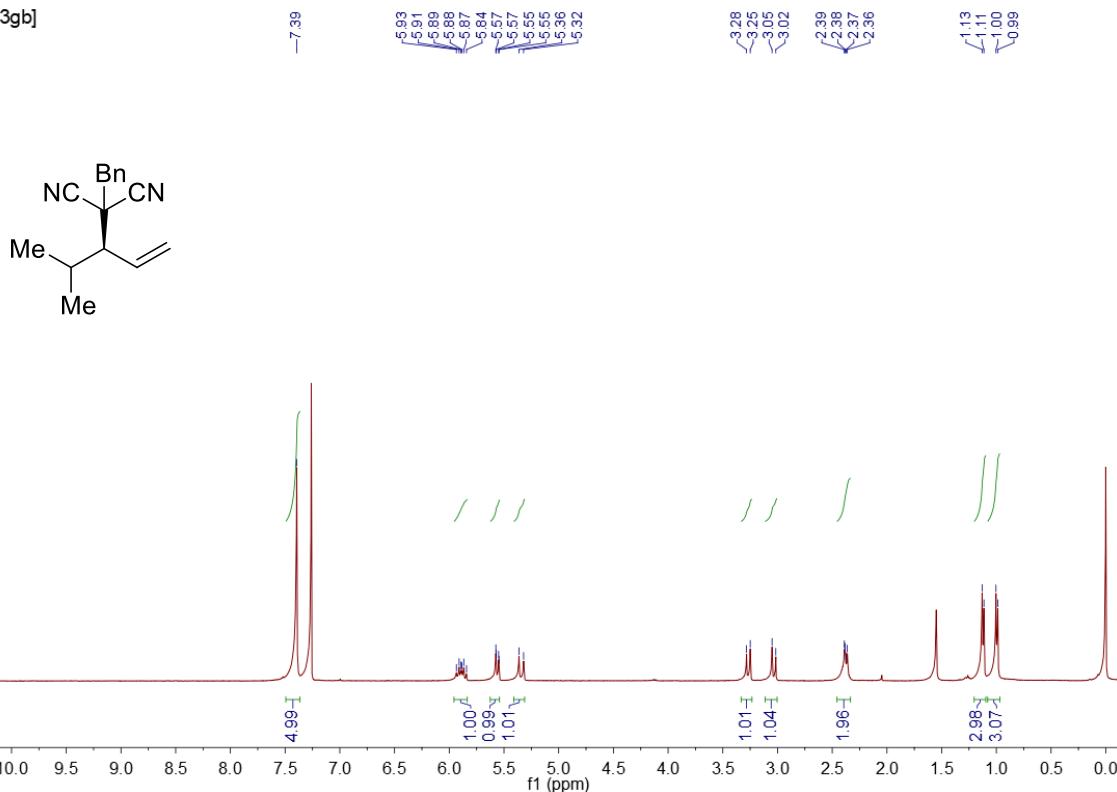


126 MHz, CDCl_3



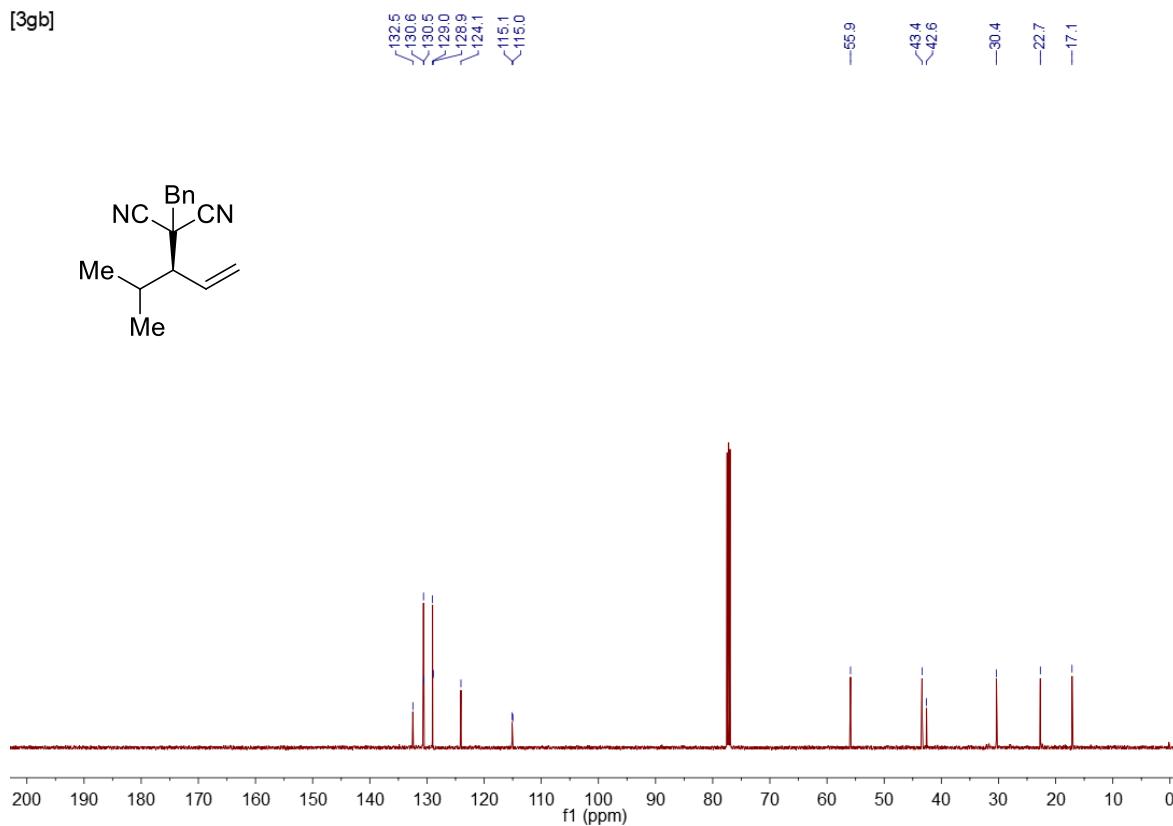
400 MHz, CDCl_3

[3gb]

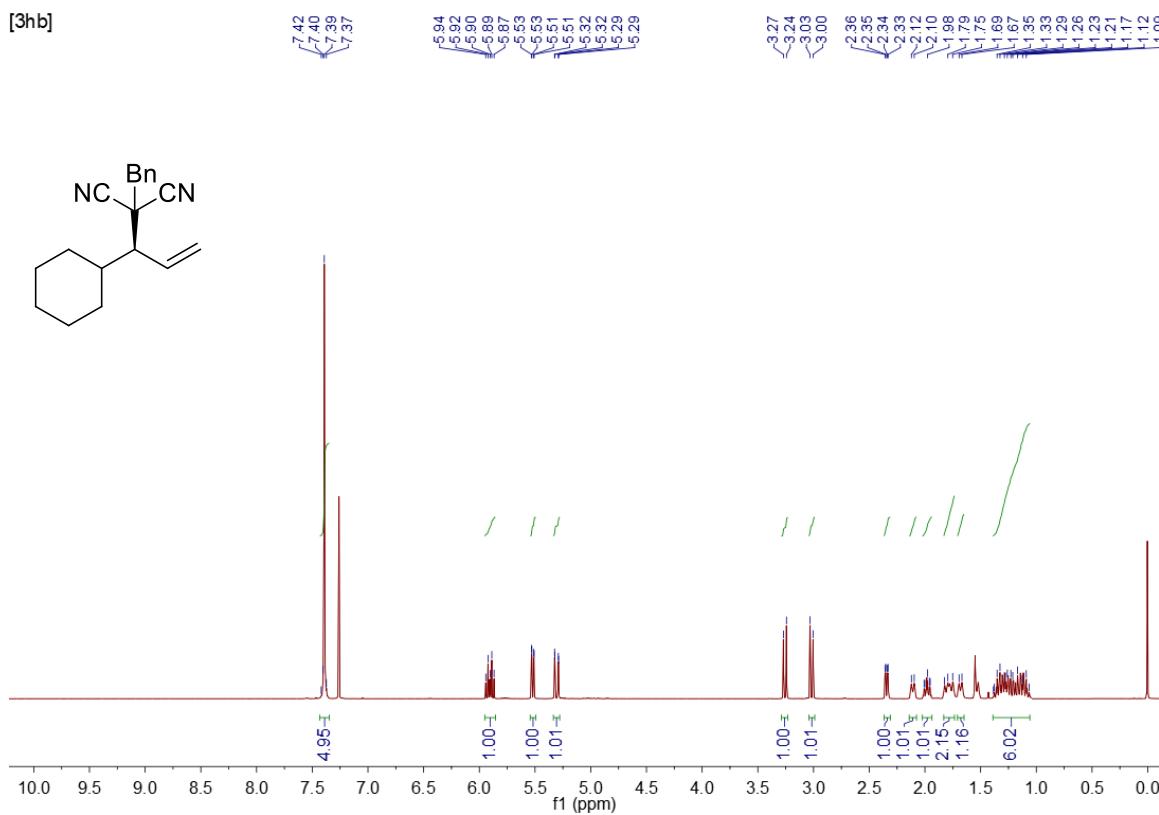


126 MHz, CDCl_3

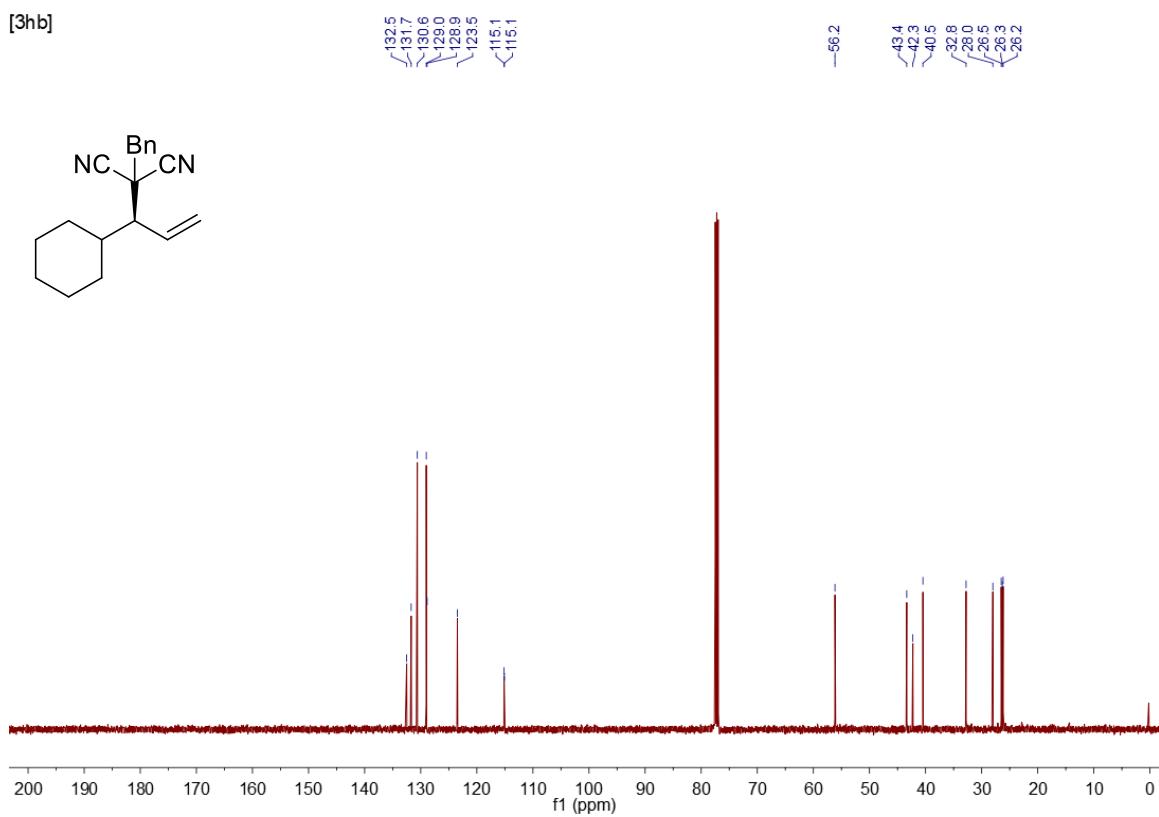
[3gb]



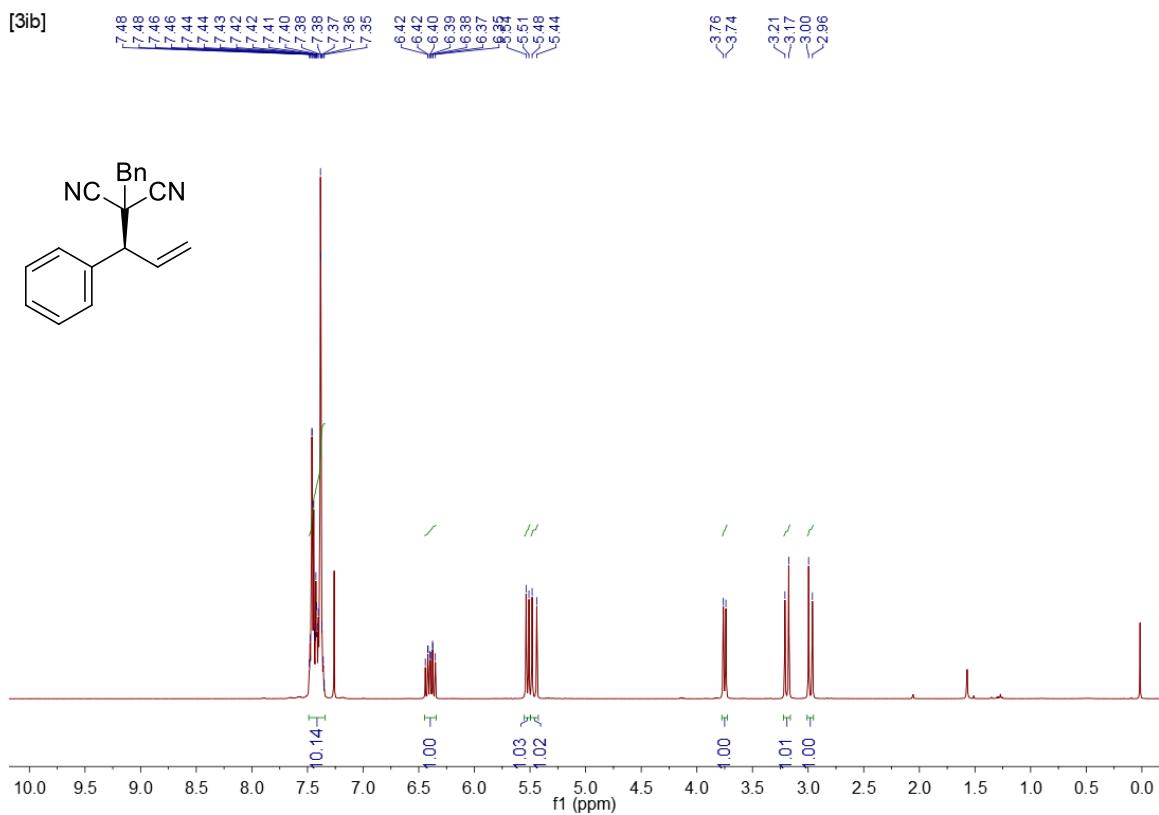
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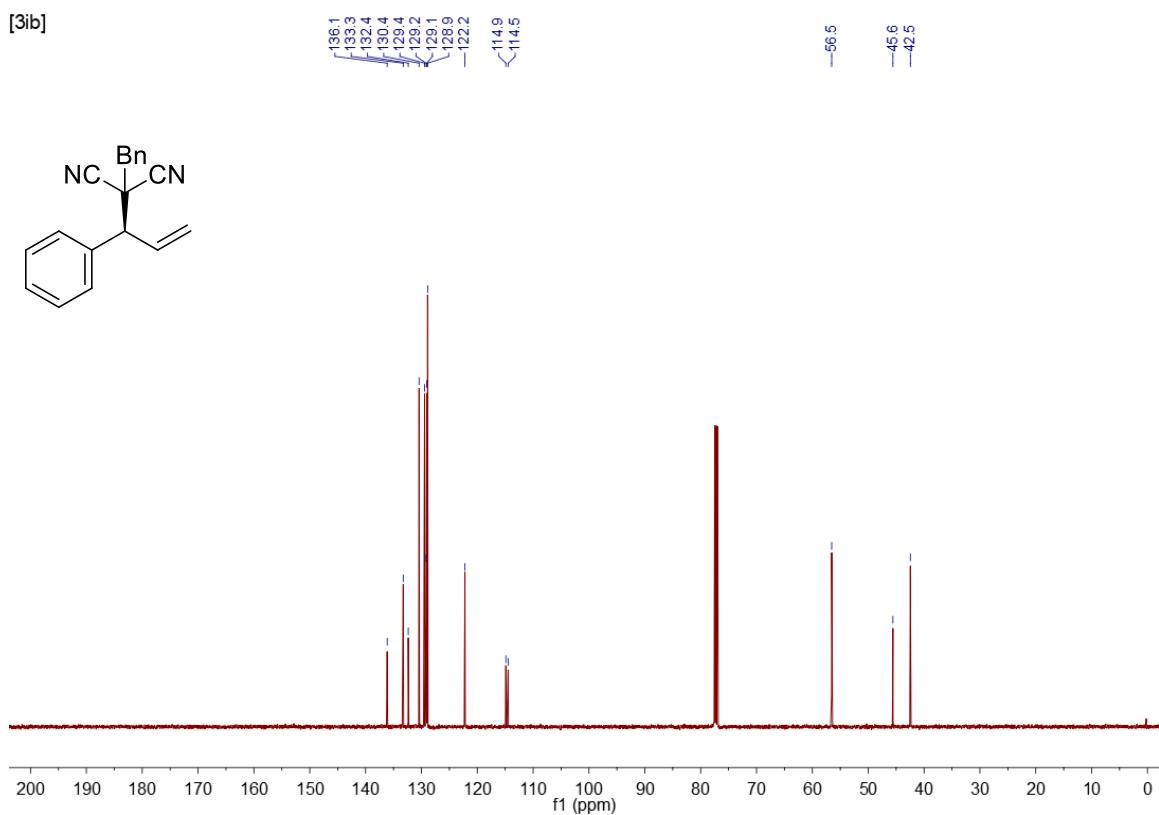
126 MHz, CDCl_3



500 MHz, CDCl_3

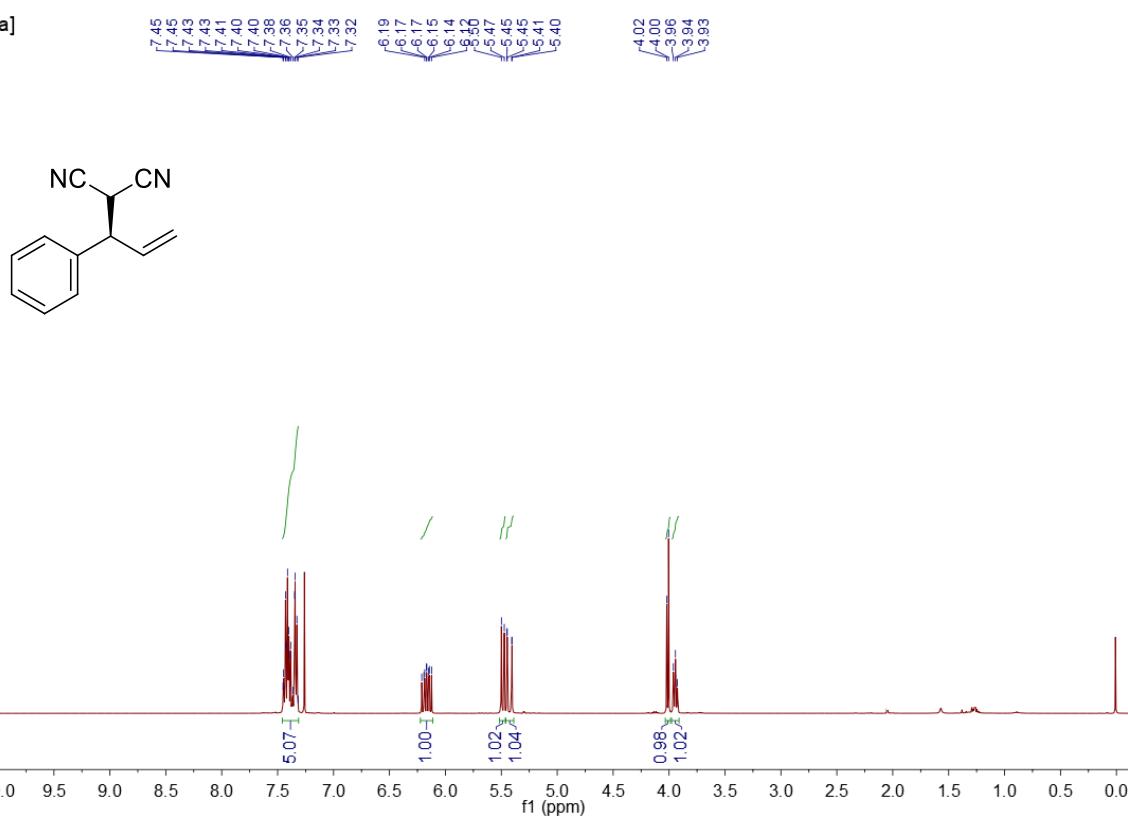


126 MHz, CDCl_3



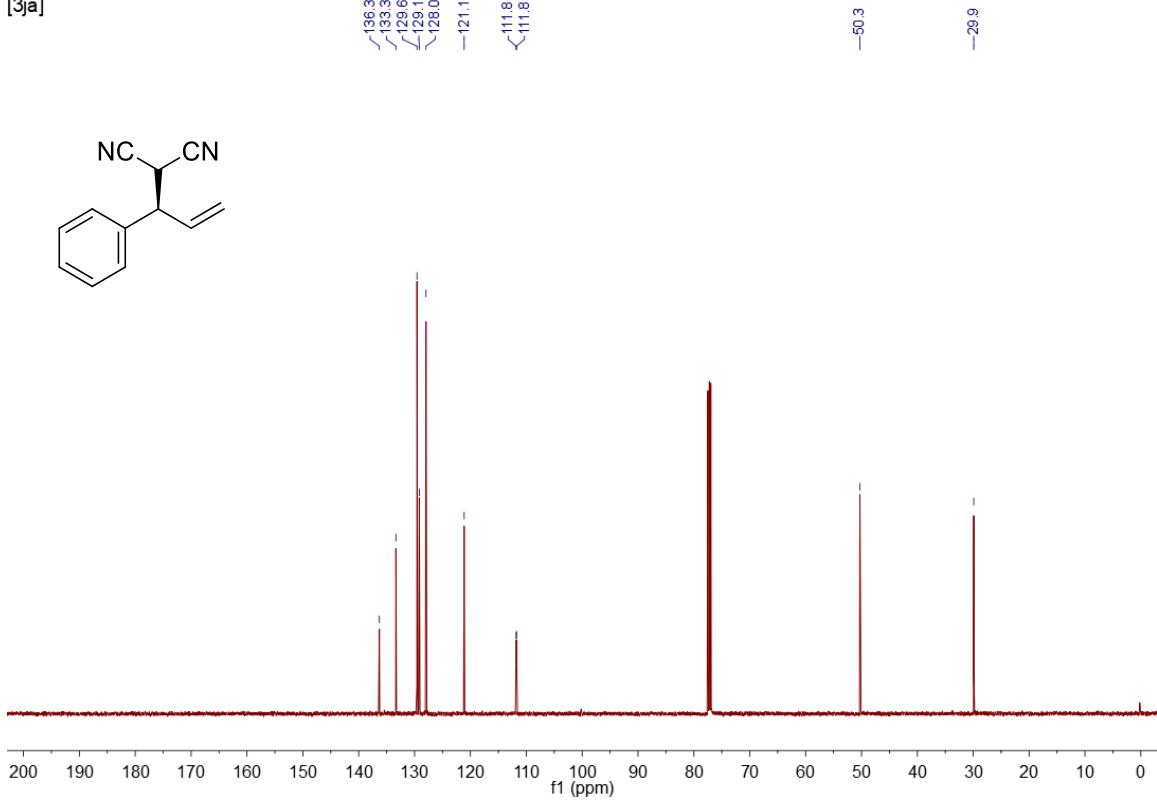
400 MHz, CDCl_3

[3ja]



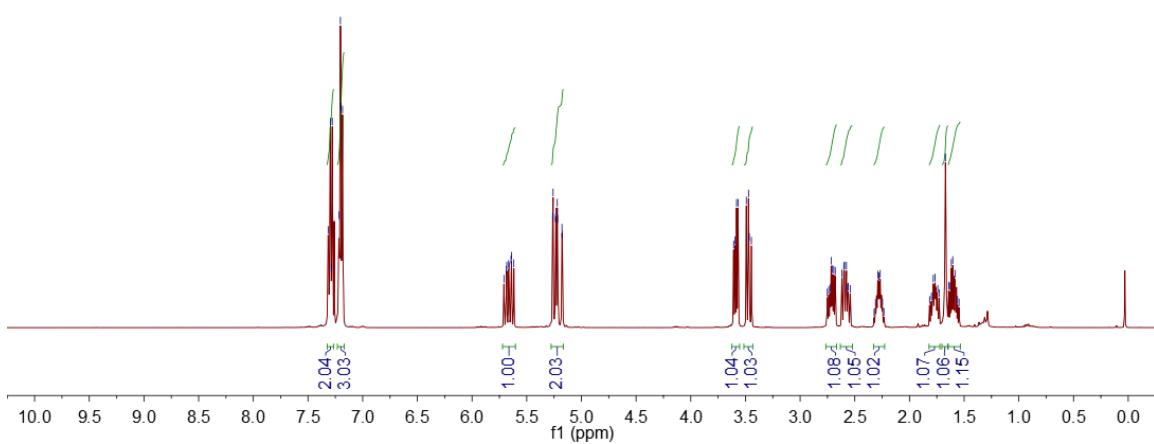
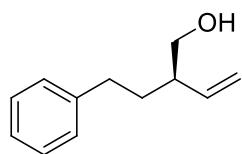
126 MHz, CDCl_3

[3ja]



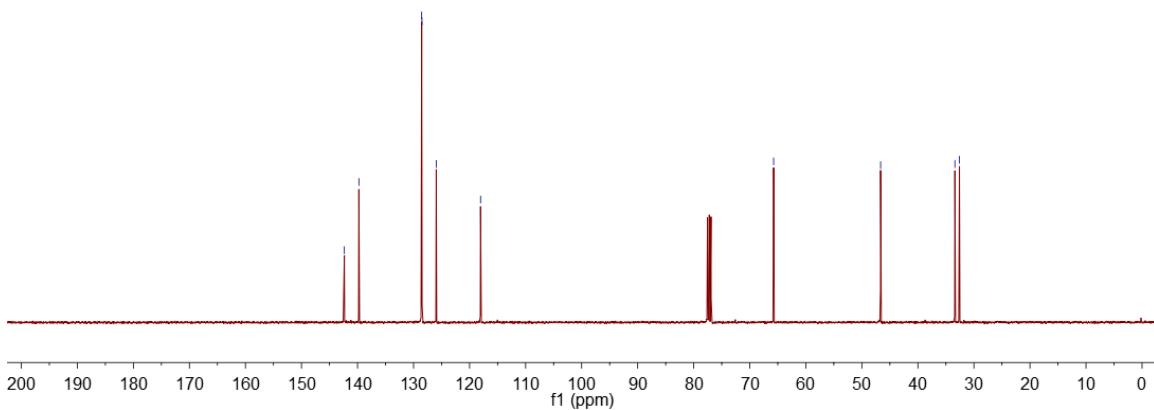
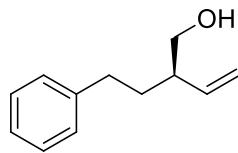
400 MHz, CDCl_3

[4]



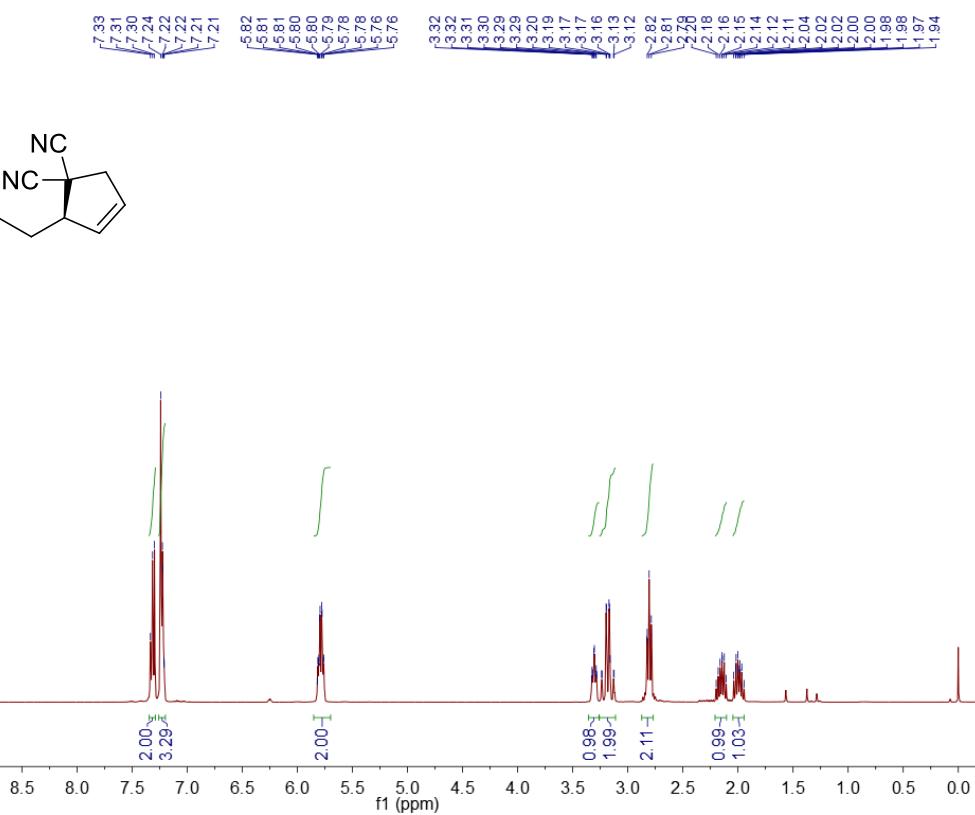
101 MHz, CDCl_3

[4]



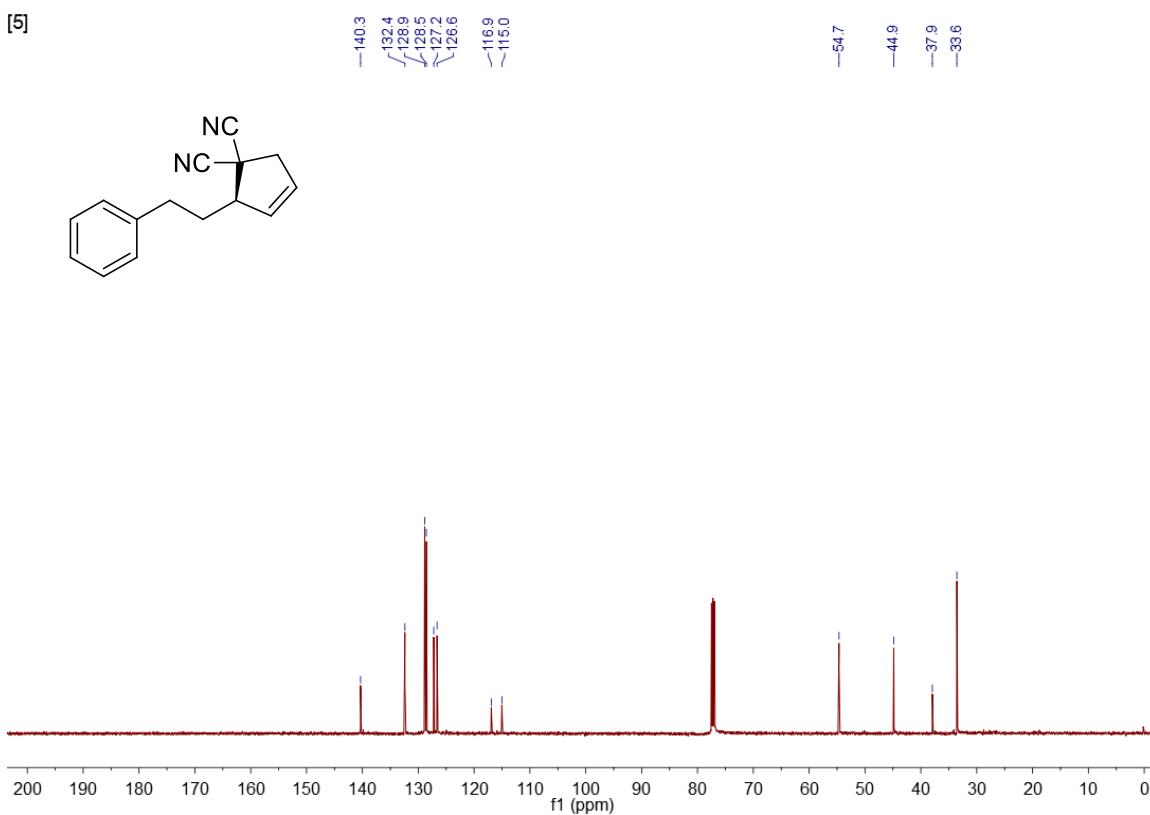
400 MHz, CDCl_3

[5]



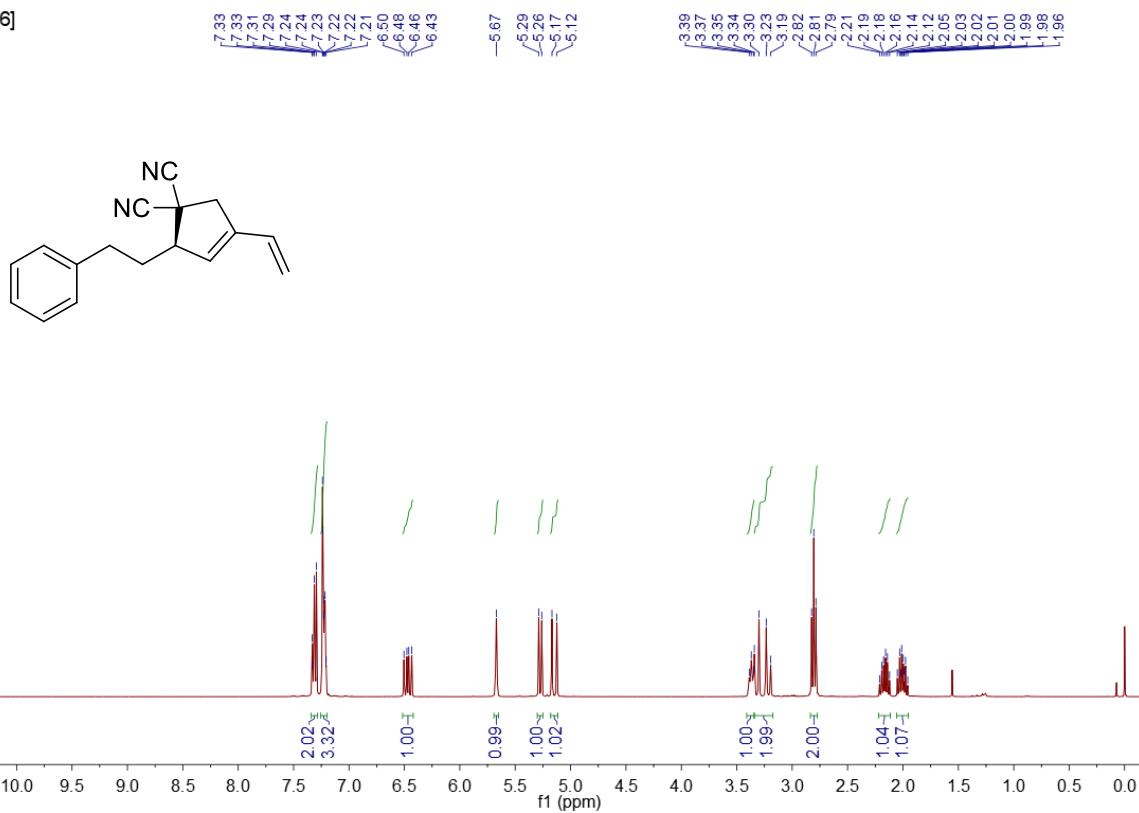
126 MHz, CDCl_3

[5]



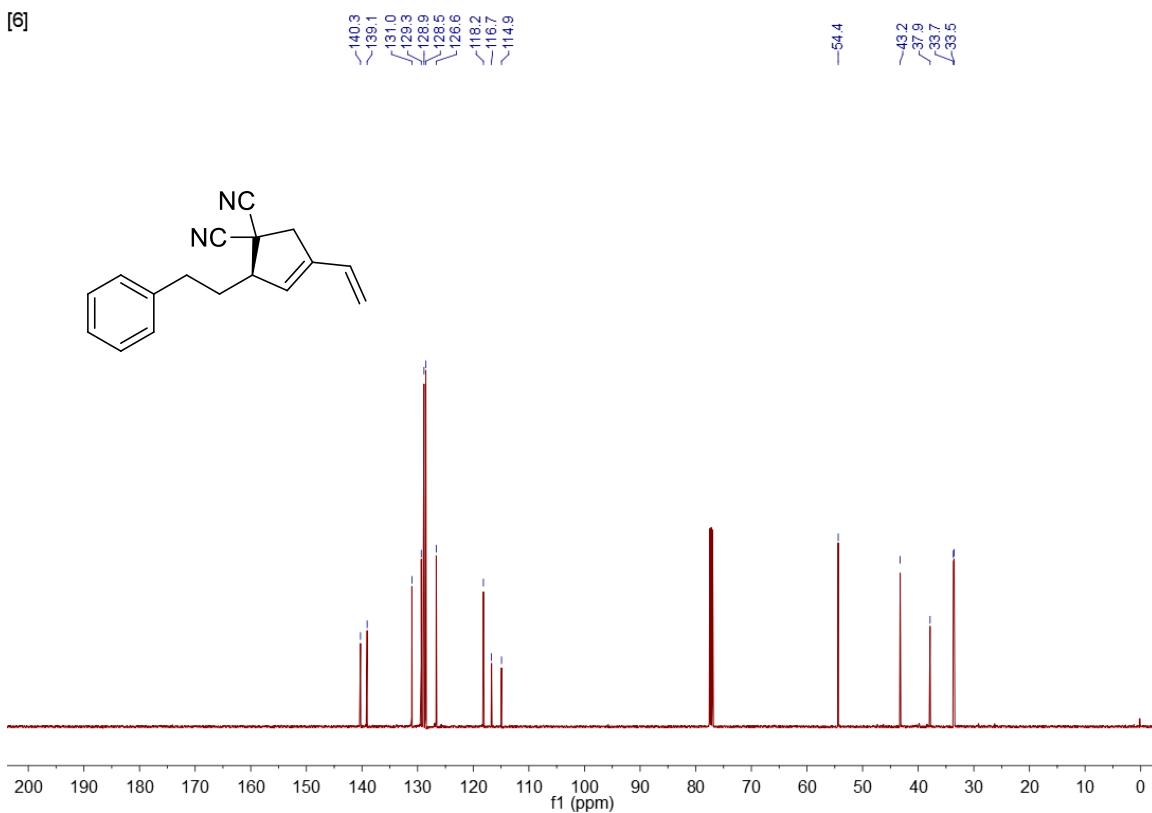
400 MHz, CDCl_3

[6]

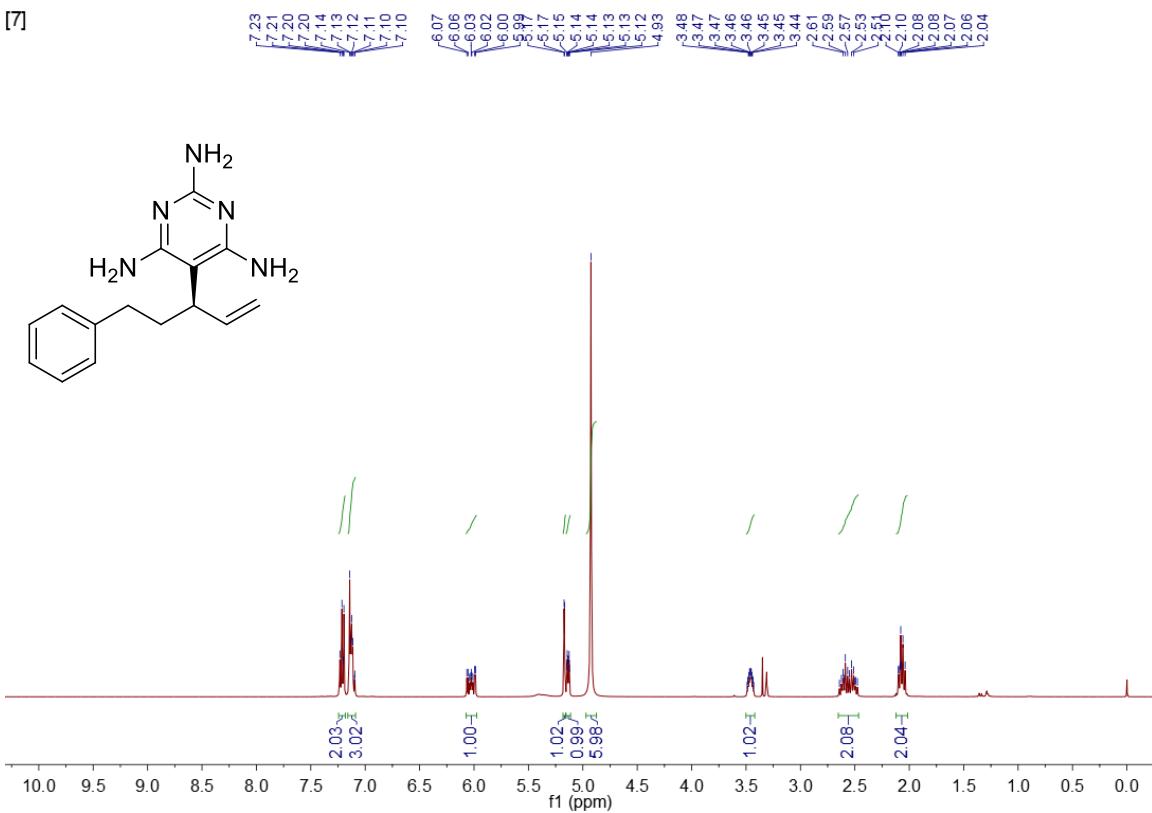


126 MHz, CDCl_3

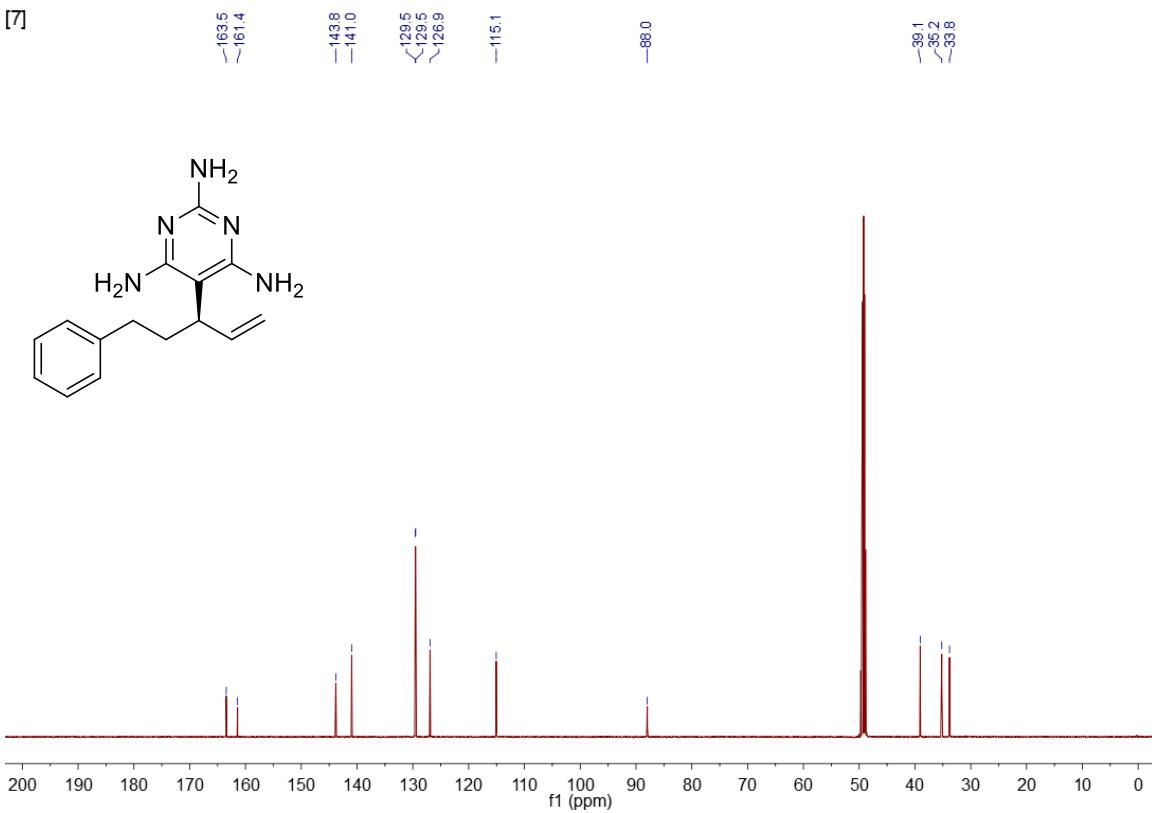
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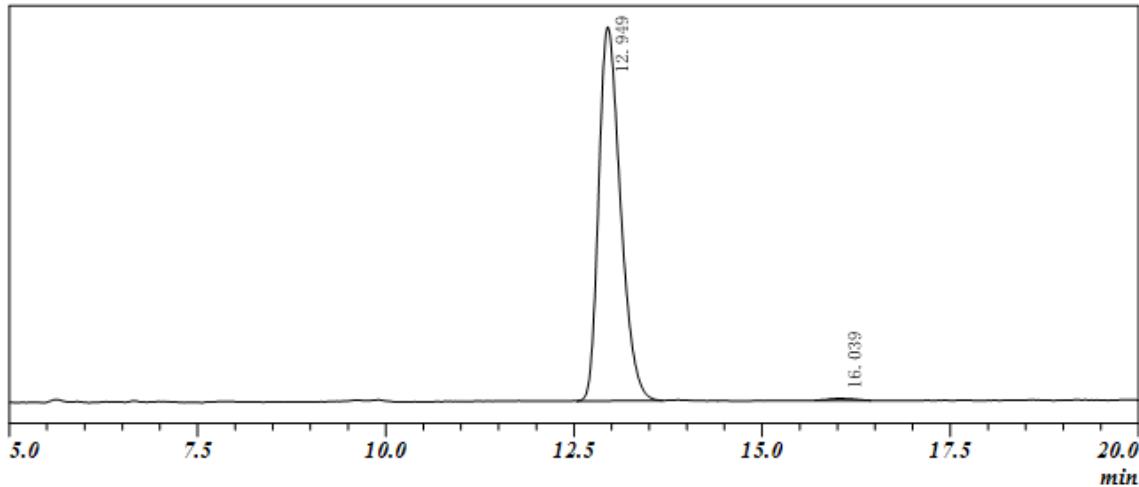
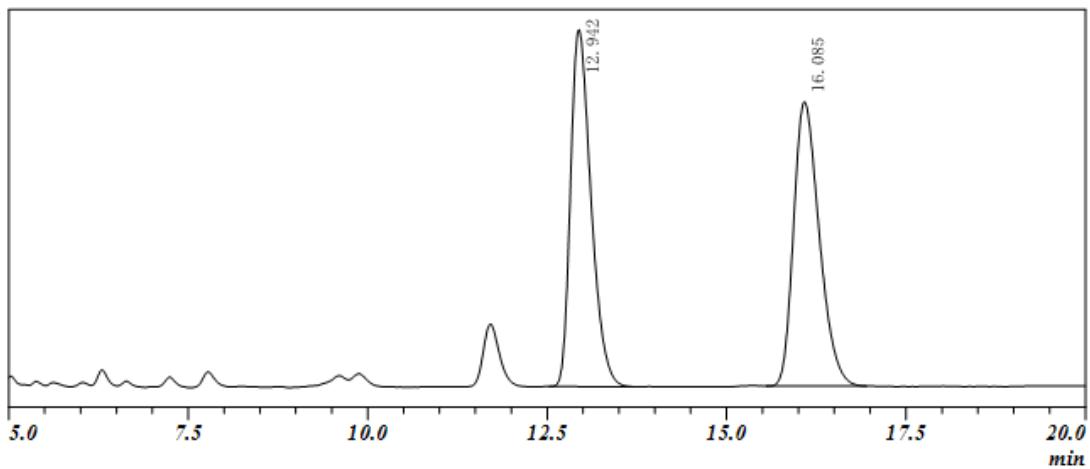
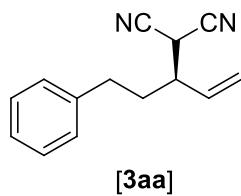
400 MHz, CD₃OD

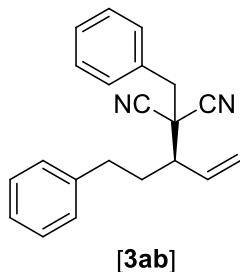


126 MHz, CD₃OD

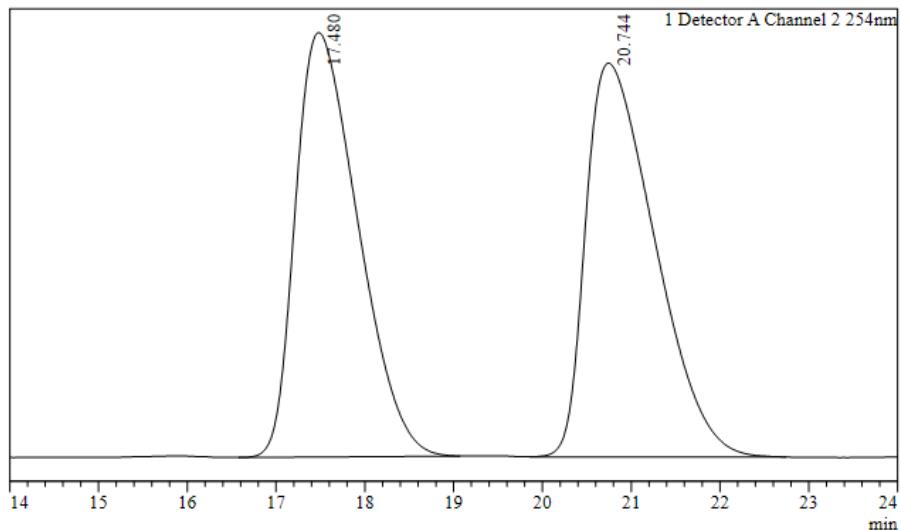


15. HPLC spectra of new compounds:



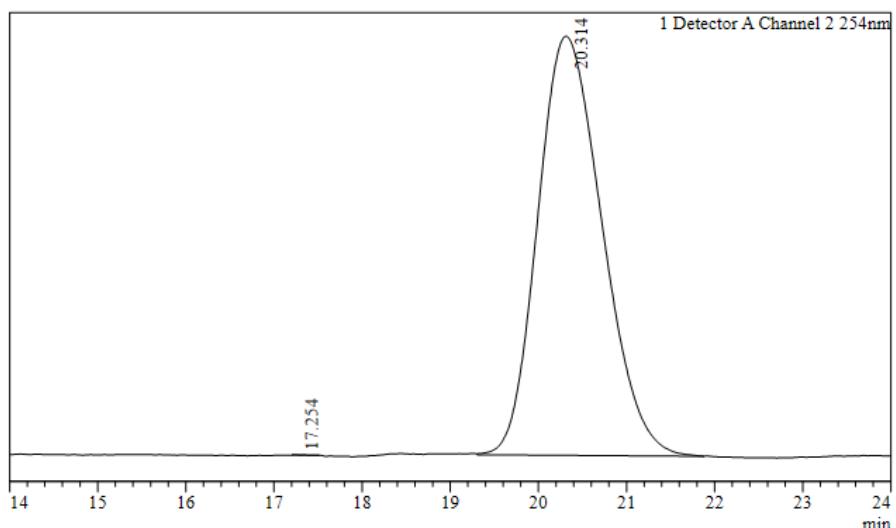


[3ab]



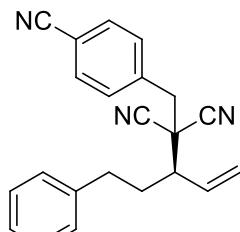
Peak Table

Detector A Channel 2 254nm					
Peak#	Ret. Time	Area	Area%	Height	Height%
1	17.480	2565596	49.413	52344	51.876
2	20.744	2626524	50.587	48559	48.124
Total		5192120	100.000	100903	100.000

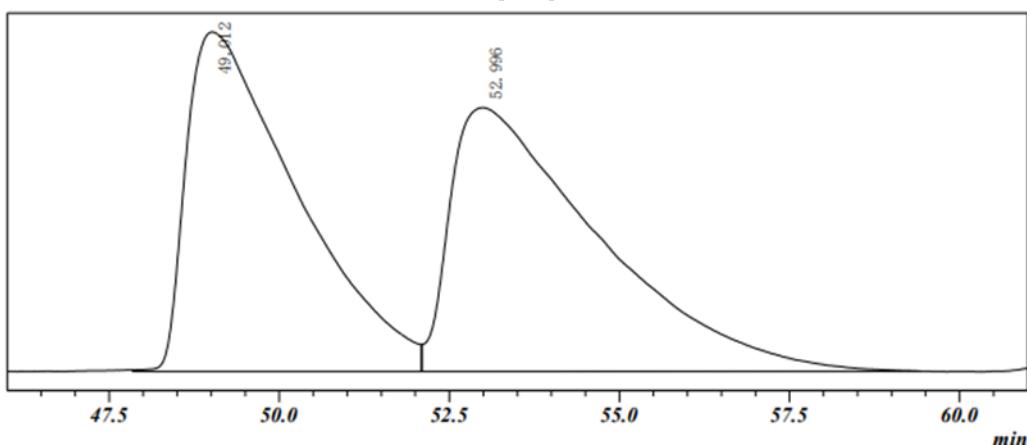


Peak Table

Detector A Channel 2 254nm					
Peak#	Ret. Time	Area	Area%	Height	Height%
1	17.254	8	0.001	2	0.016
2	20.314	704197	99.999	13856	99.984
Total		704206	100.000	13858	100.000



[3ac]

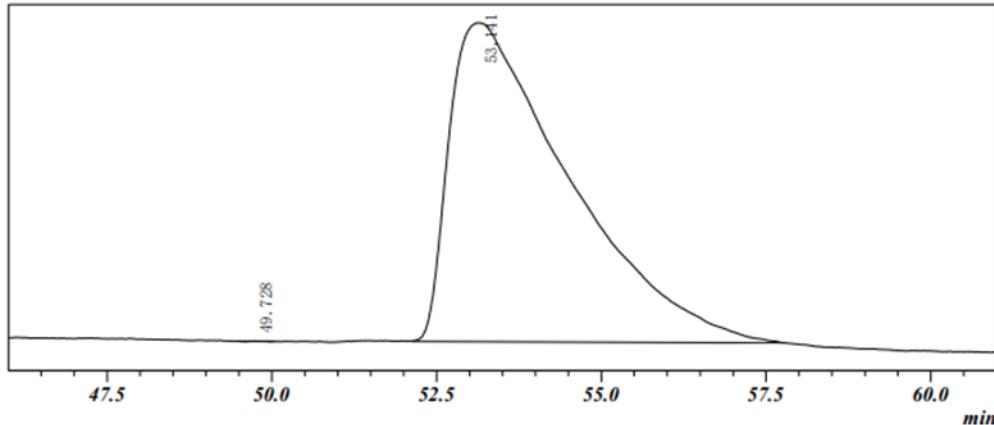


PEAK TABLE

<峰表>

檢測器A Ch1 254nm

Peak NO.	Retention time	Area	Area%	Height	Height%
1	49.012	2396442	48.859	21214	56.272
2	52.996	2508391	51.141	16485	43.728
总计		4904832	100.000	37699	100.000

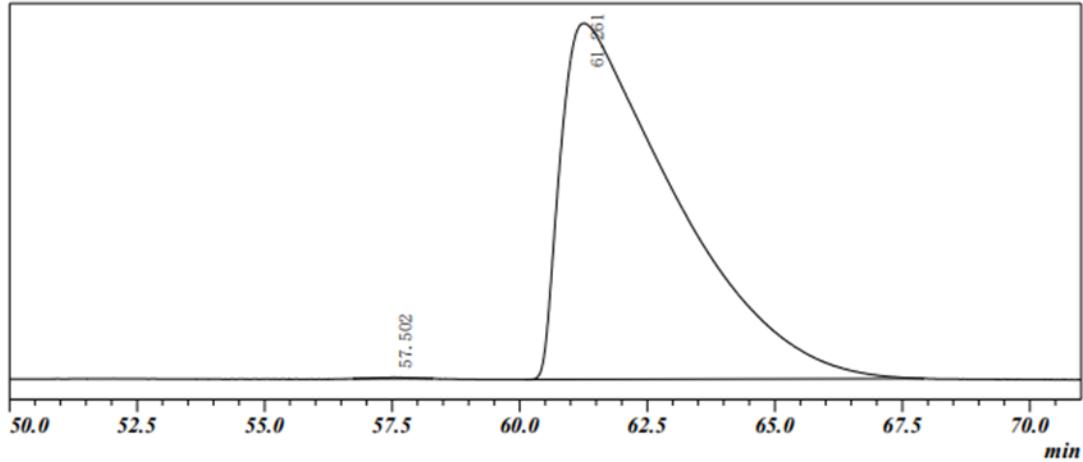
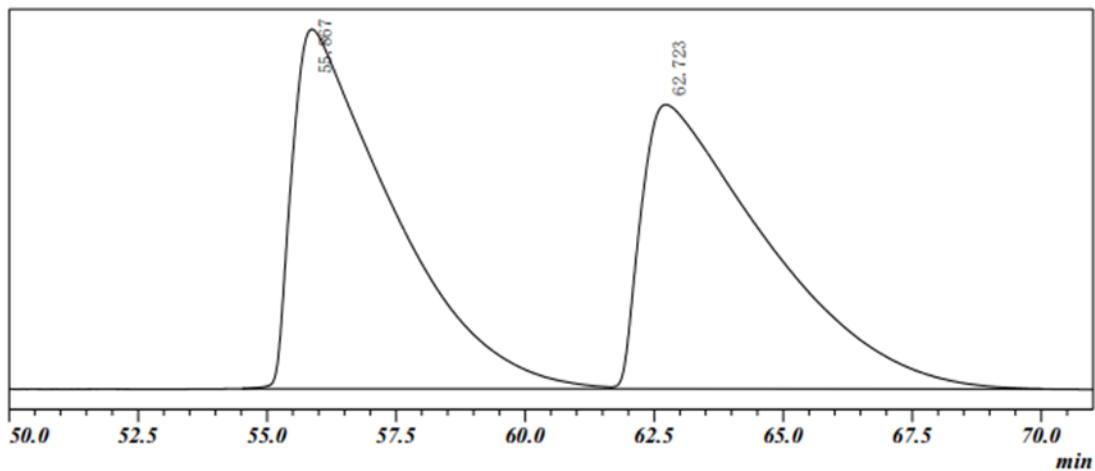
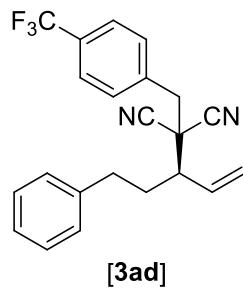


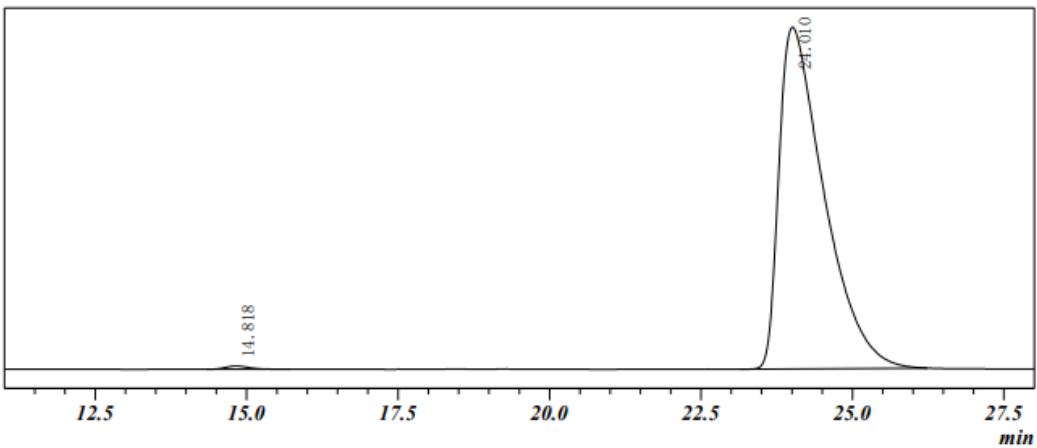
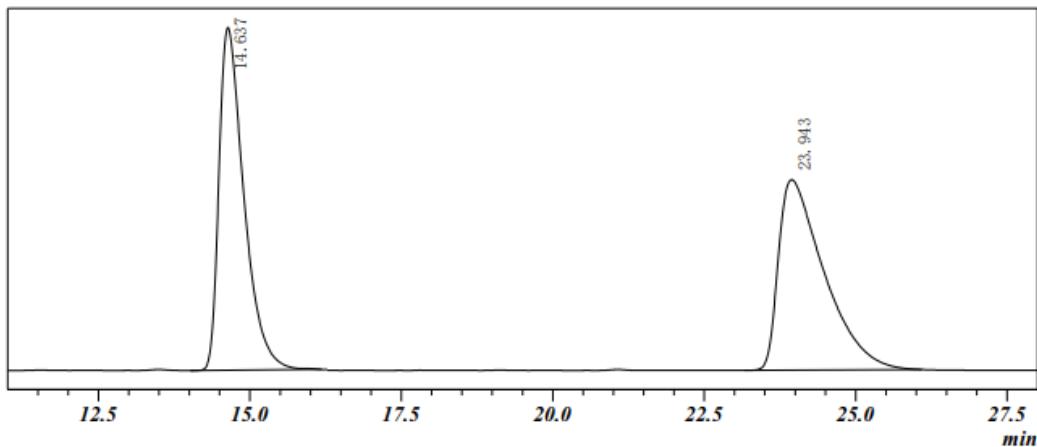
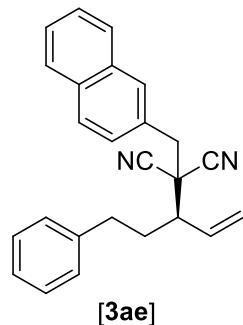
PEAK TABLE

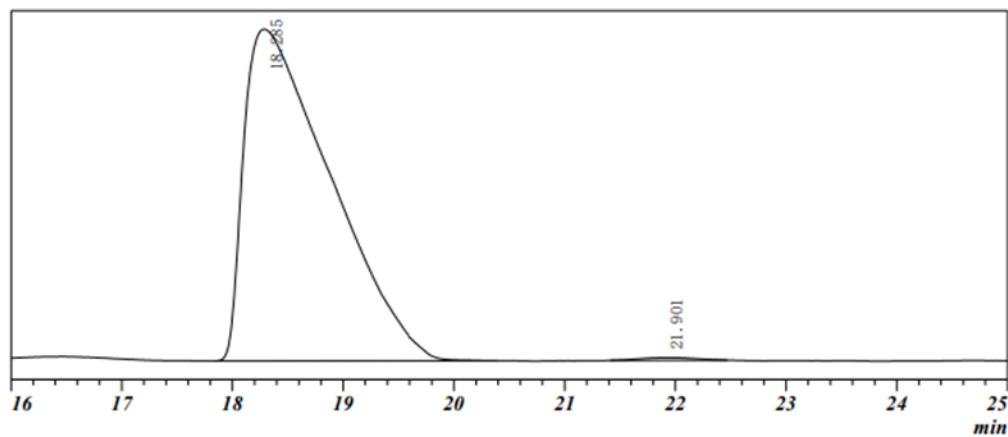
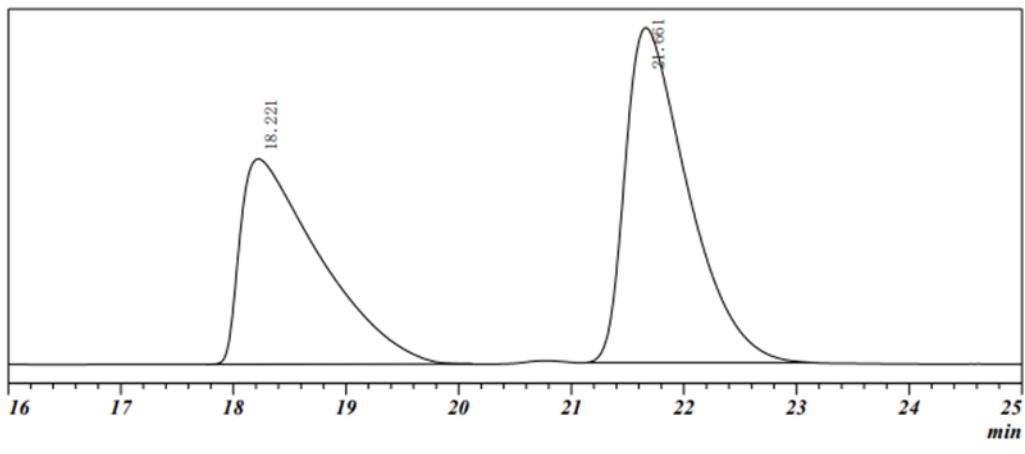
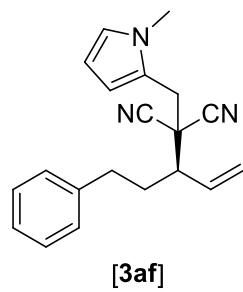
<峰表>

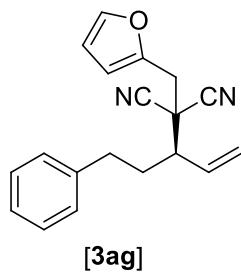
檢測器A Ch1 254nm

Peak NO.	Retention time	Area	Area%	Height	Height%
1	49.728	16	0.001	14	0.133
2	53.141	1368344	99.999	10787	99.867
总计		1368360	100.000	10801	100.000

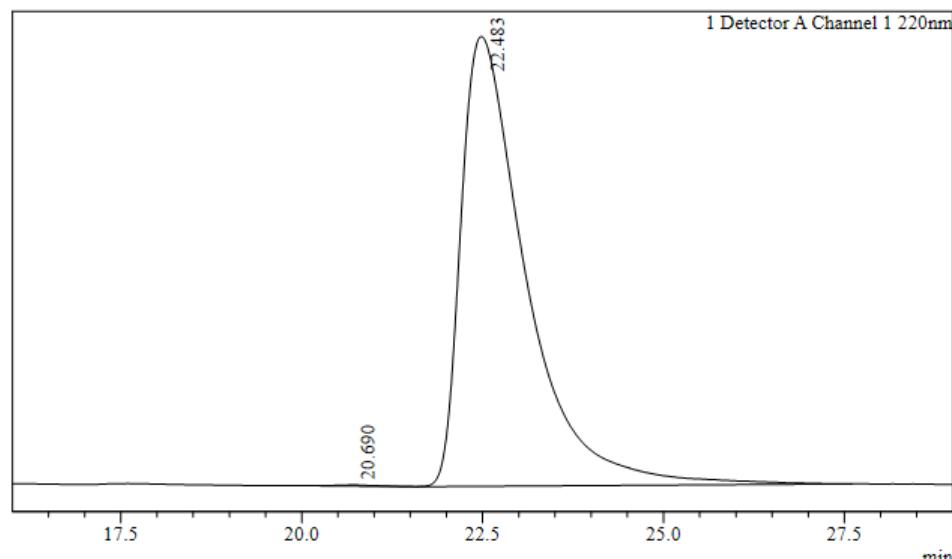
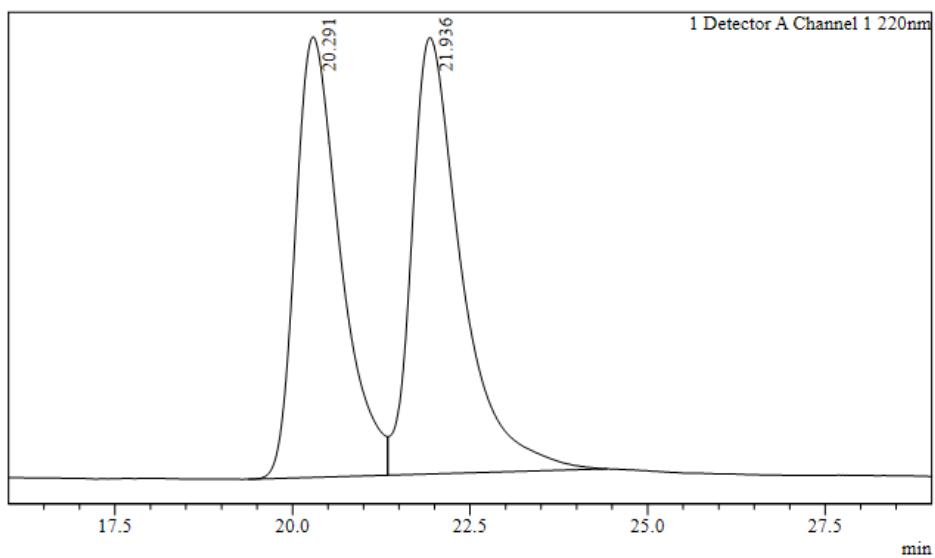


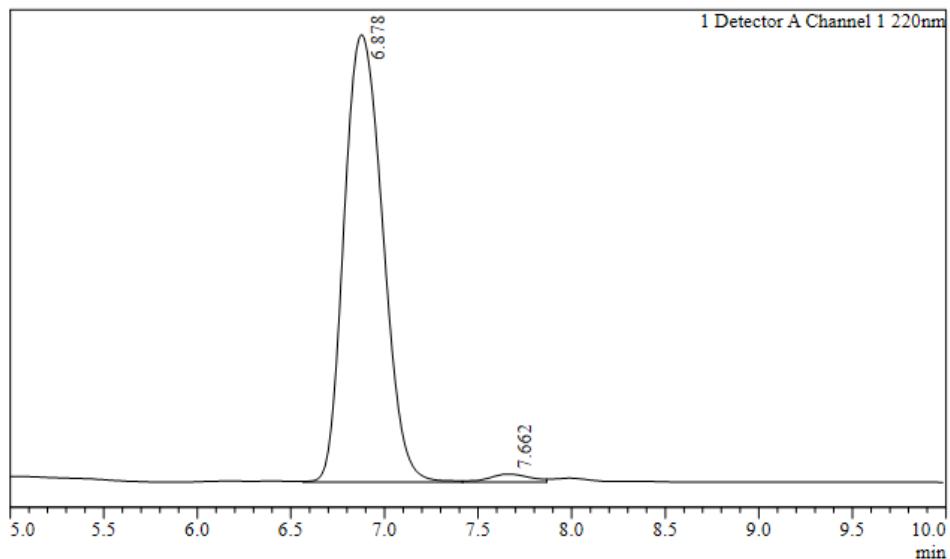
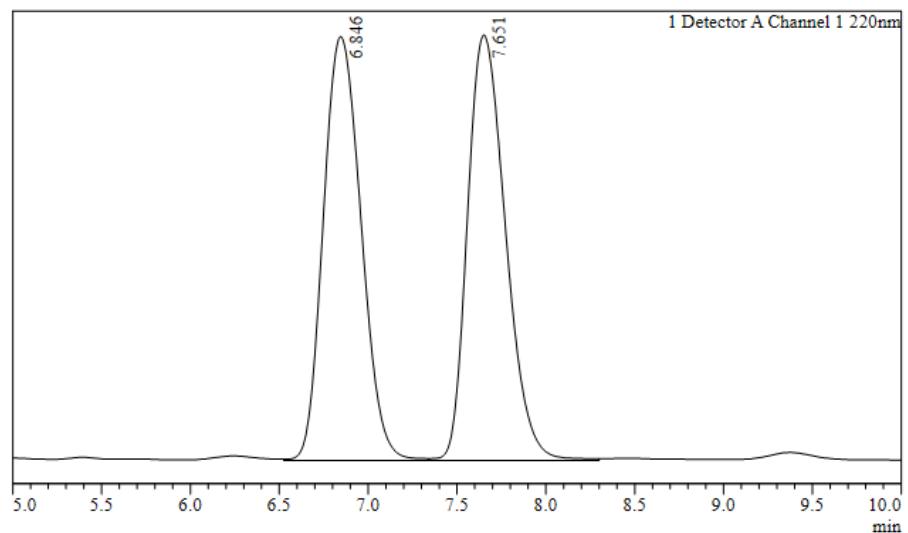
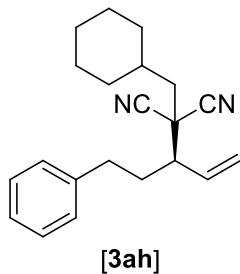


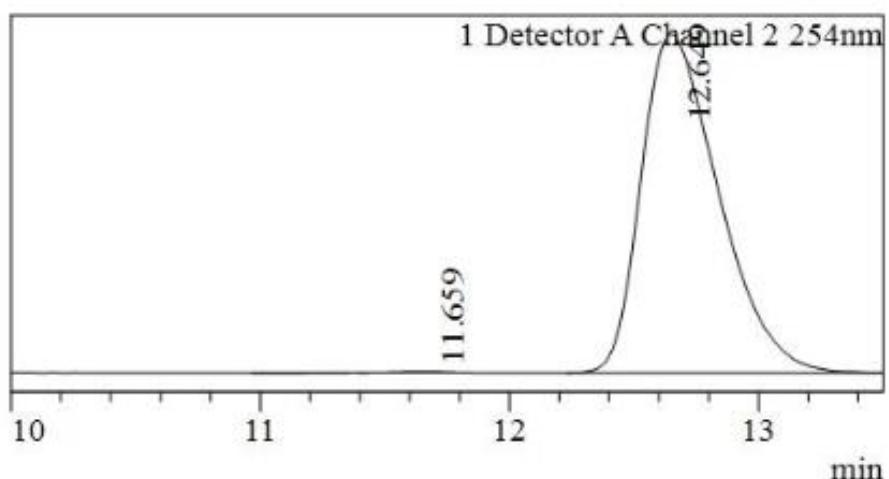
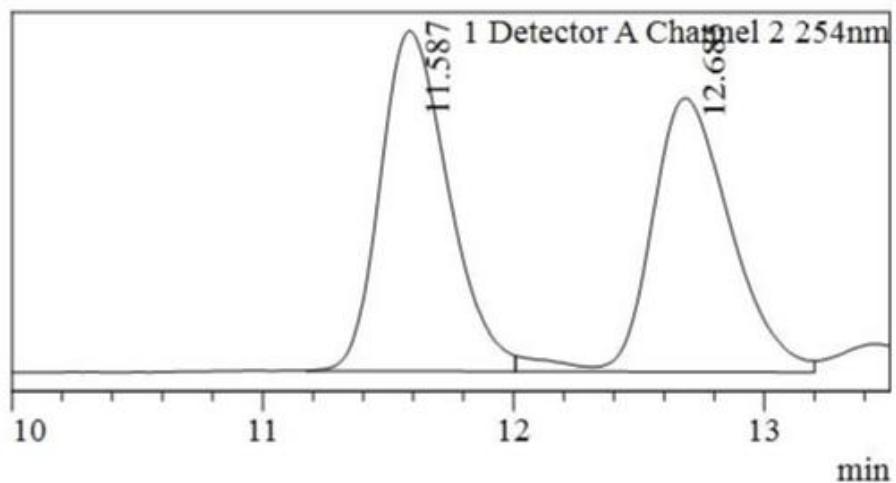
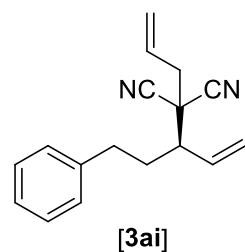


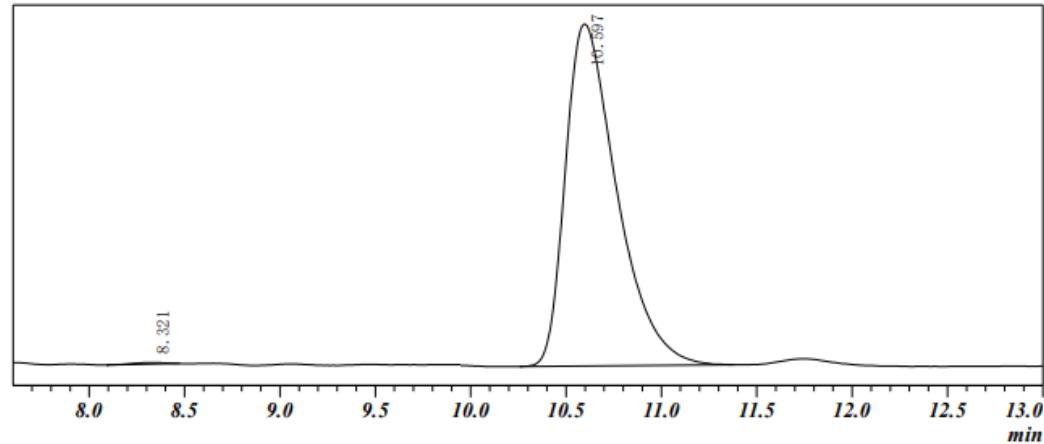
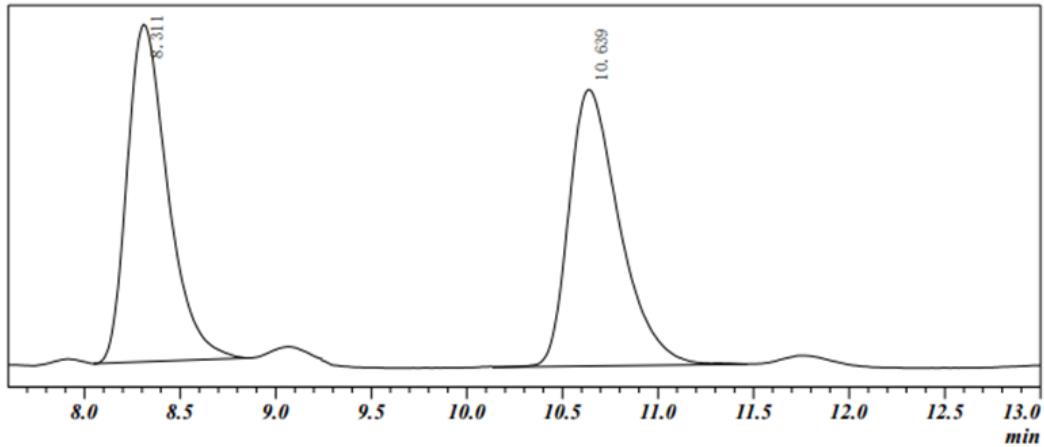
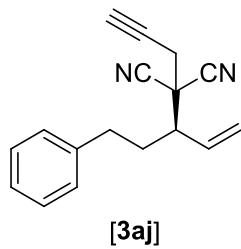


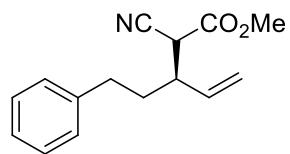
[3ag]



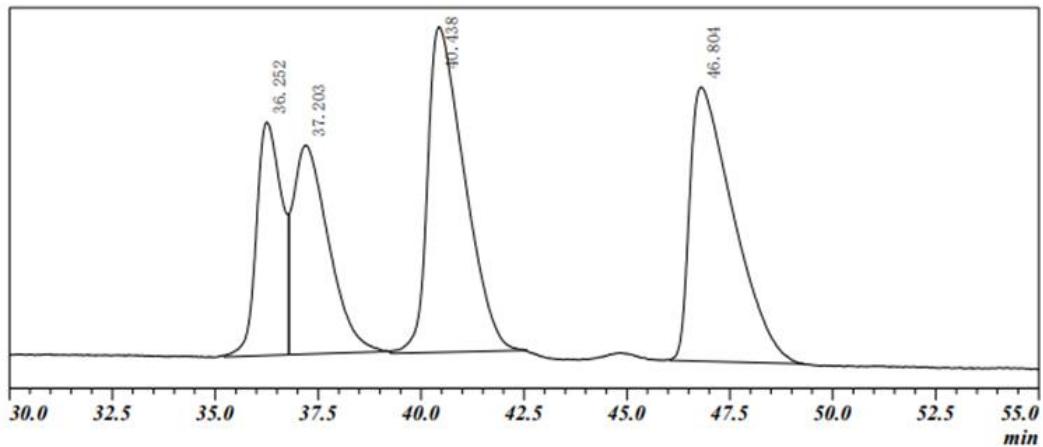








[3ak]

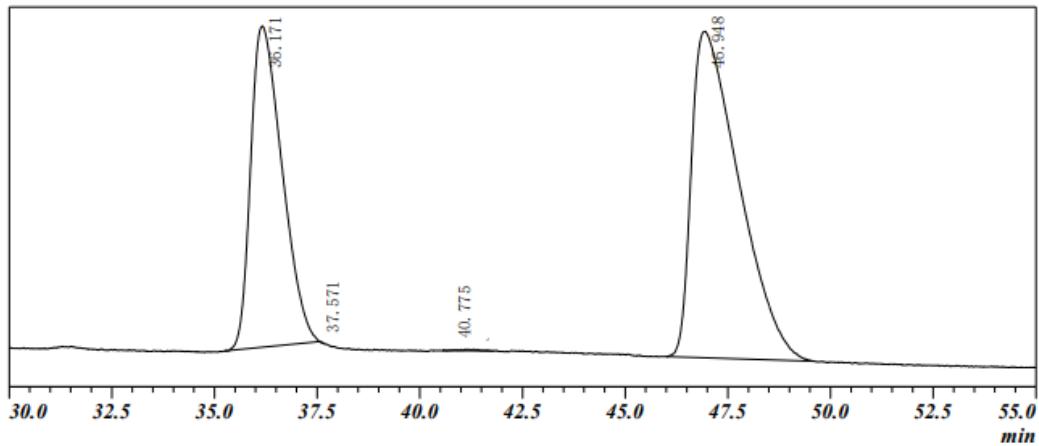


PEAK TABLE

<峰表>

檢測器A Ch1 254nm

Peak NO.	Retention time	Area	Area%	Height	Height%
1	36.252	250839	16.021	5796	22.400
2	37.203	310104	19.806	5189	20.054
3	40.438	502717	32.108	8086	31.248
4	46.804	502063	32.066	6805	26.298
总计		1565723	100.000	25876	100.000

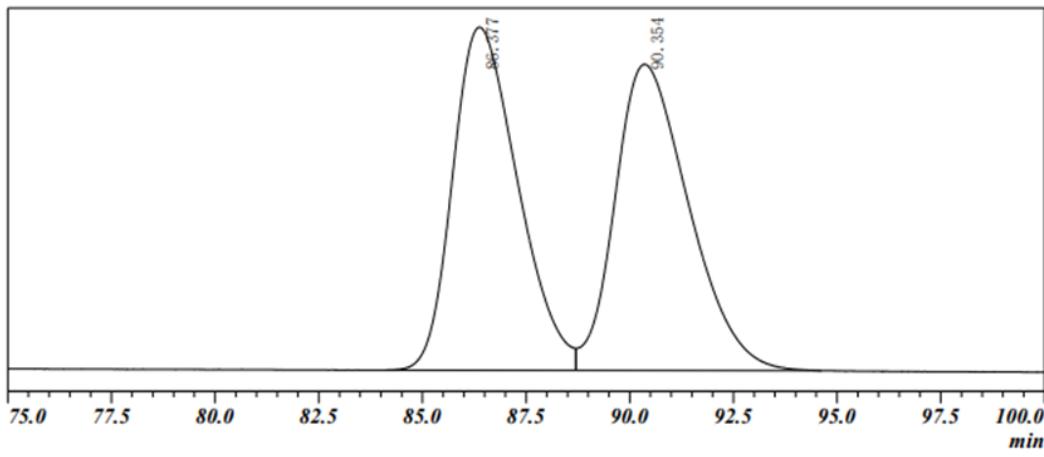
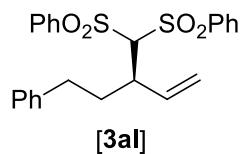


PEAK TABLE

<峰表>

檢測器A Ch1 254nm

Peak NO.	Retention time	Area	Area%	Height	Height%
1	36.171	332437	39.364	6255	49.473
2	37.571	2	0.000	0	0.001
3	40.775	290	0.034	30	0.235
4	46.948	511795	60.602	6359	50.290
总计		844525	100.000	12644	100.000

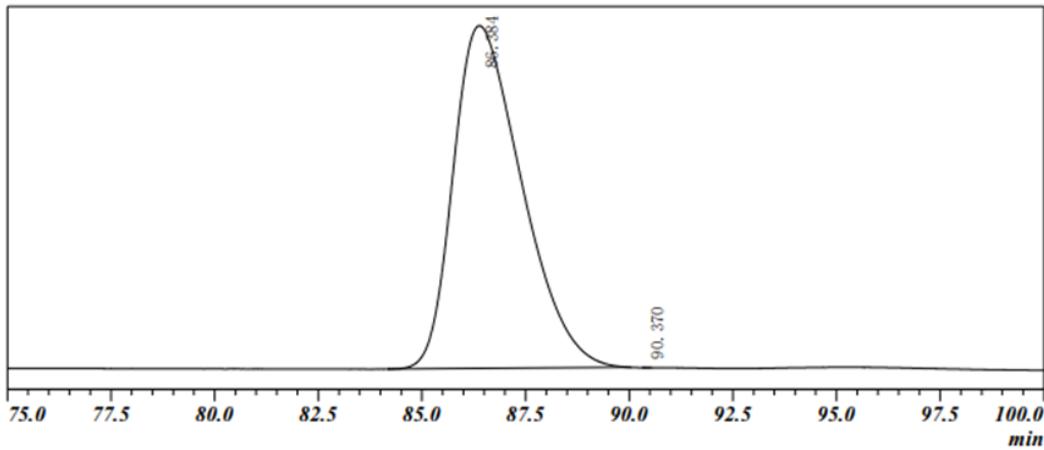


PEAK TABLE

<峰表>

検測器A Ch1 254nm

Peak NO.	Retention time	Area	Area%	Height	Height%
1	86.377	4326177	49.674	39624	52.843
2	90.354	4383030	50.326	35360	47.157
总计		8709206	100.000	74984	100.000

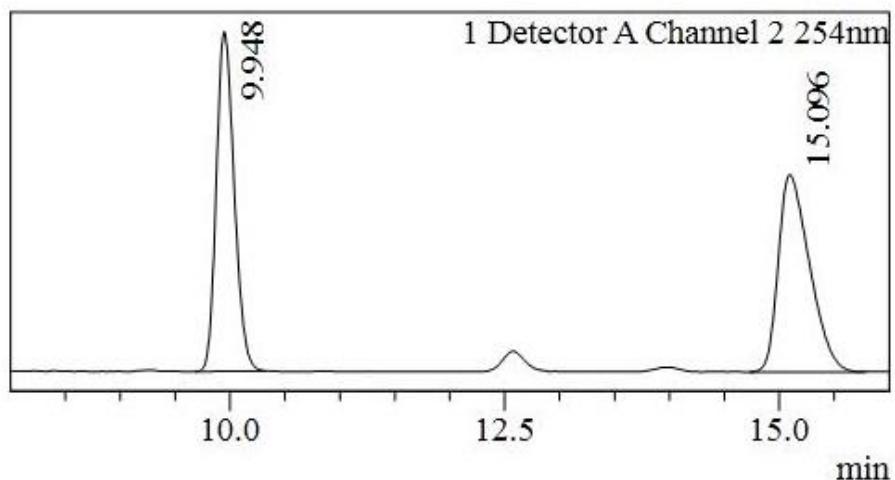
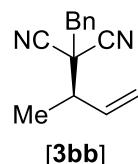


PEAK TABLE

<峰表>

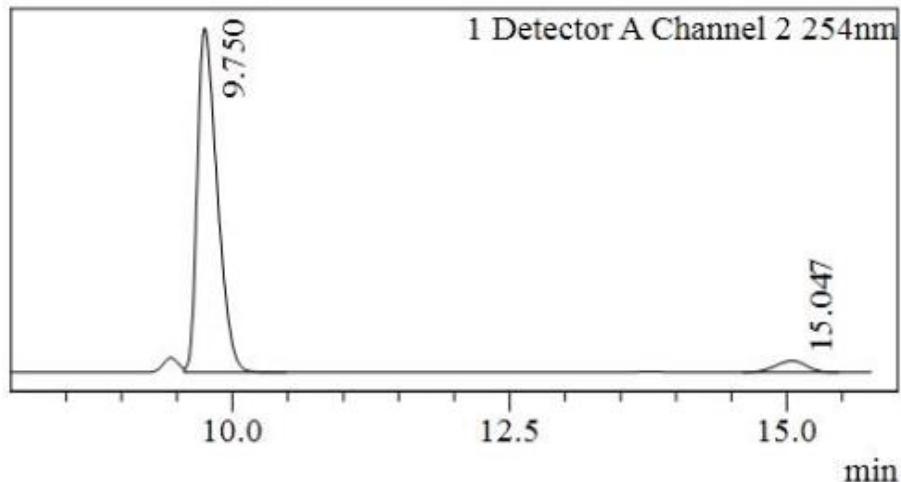
検測器A Ch1 254nm

Peak NO.	Retention time	Area	Area%	Height	Height%
1	86.384	6273413	100.000	54546	99.988
2	90.370	17	0.000	7	0.012
总计		6273429	100.000	54552	100.000



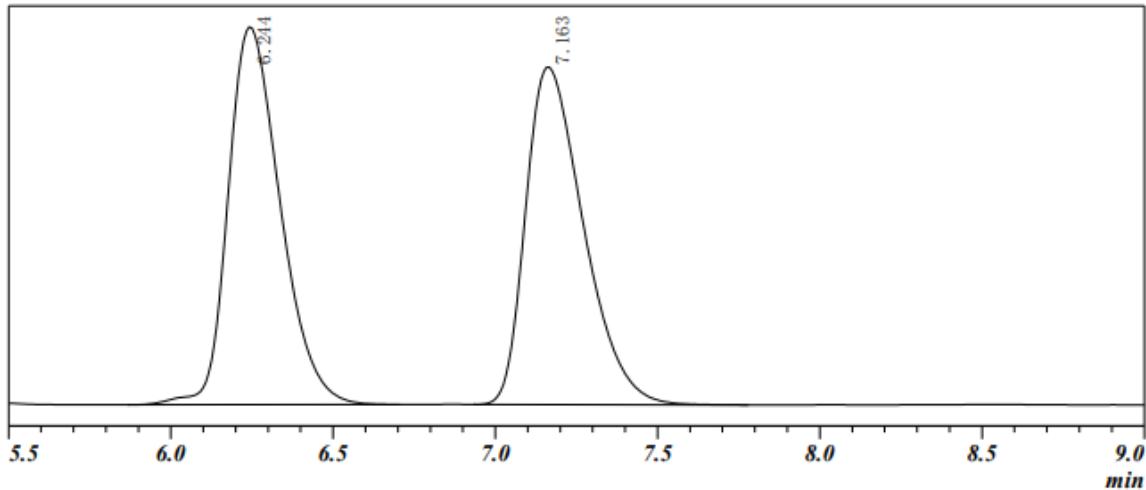
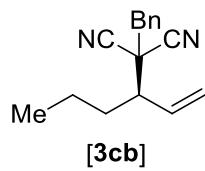
Peak Table

Detector A Channel 2 254nm						
Peak#	Ret. Time	Area	Height	Area%	Conc.	Name
1	9.948	98529	8677	49.661	49.661	
2	15.096	99874	5039	50.339	50.339	
Total		198403	13716	100.000		



Peak Table

Detector A Channel 2 254nm						
Peak#	Ret. Time	Area	Height	Area%	Conc.	Name
1	9.750	423862	33937	94.974	94.974	
2	15.047	22428	1132	5.026	5.026	
Total		446290	35069	100.000		

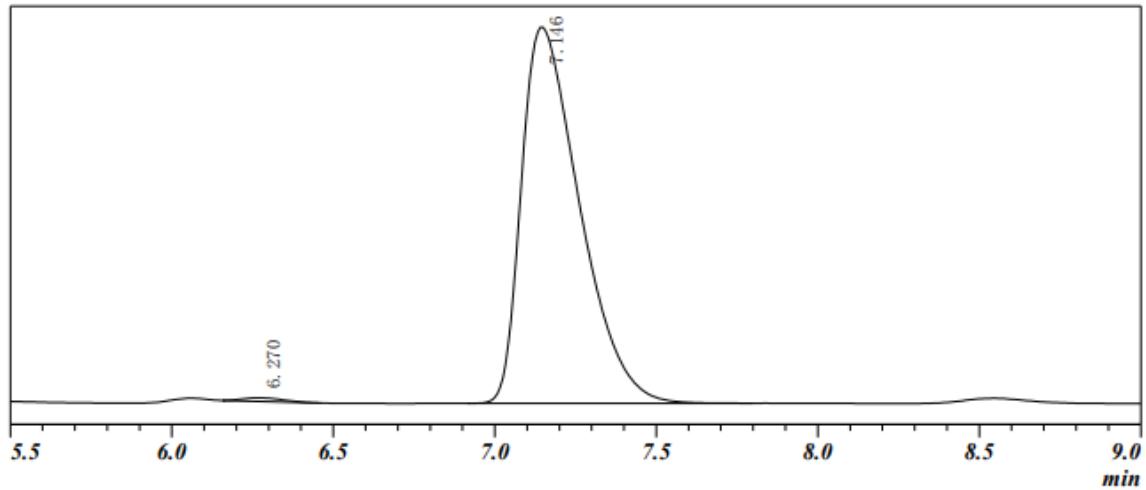


PEAK TABLE

<峰表>

检测器A Ch1 254nm

Peak NO.	Retention time	Area	Area%	Height	Height%
1	6.244	574136	50.238	51284	52.819
2	7.163	568696	49.762	45810	47.181
总计		1142832	100.000	97094	100.000

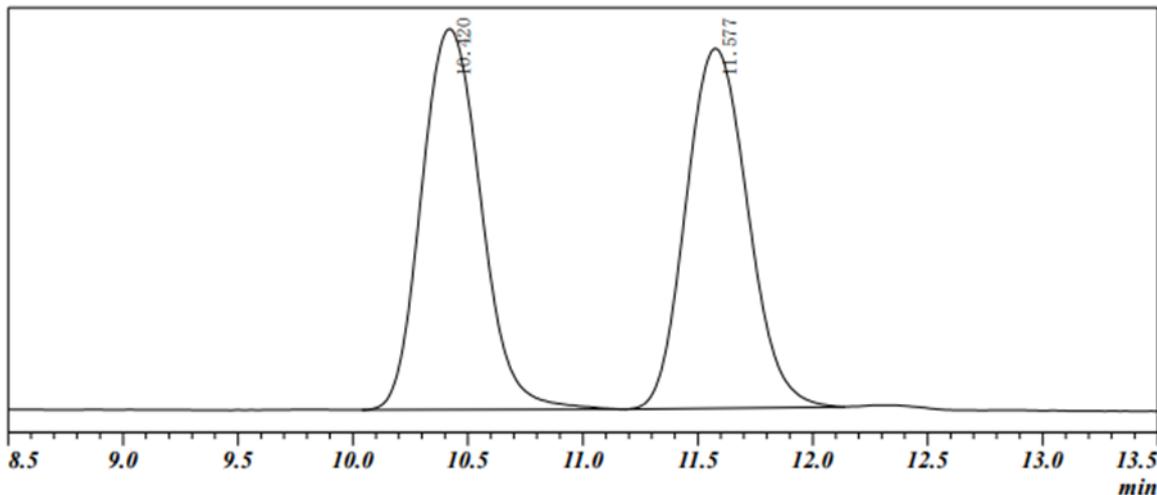
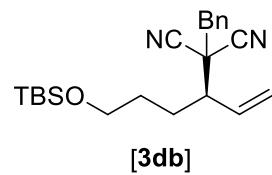


PEAK TABLE

<峰表>

检测器A Ch1 254nm

Peak NO.	Retention time	Area	Area%	Height	Height%
1	6.270	5628	0.618	679	0.932
2	7.146	904633	99.382	72211	99.068
总计		910261	100.000	72890	100.000

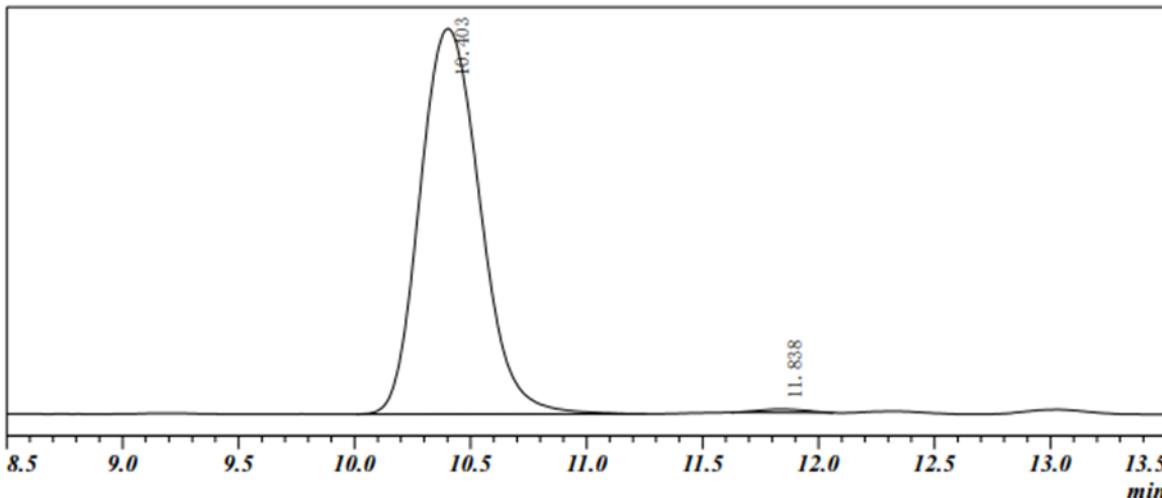


PEAK TABLE

<峰表>

檢測器A Ch1 254nm

Peak NO.	Retention time	Area	Area%	Height	Height%
1	10.420	279693	50.465	15548	51.405
2	11.577	274537	49.535	14698	48.595
总计		554230	100.000	30245	100.000

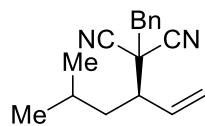


PEAK TABLE

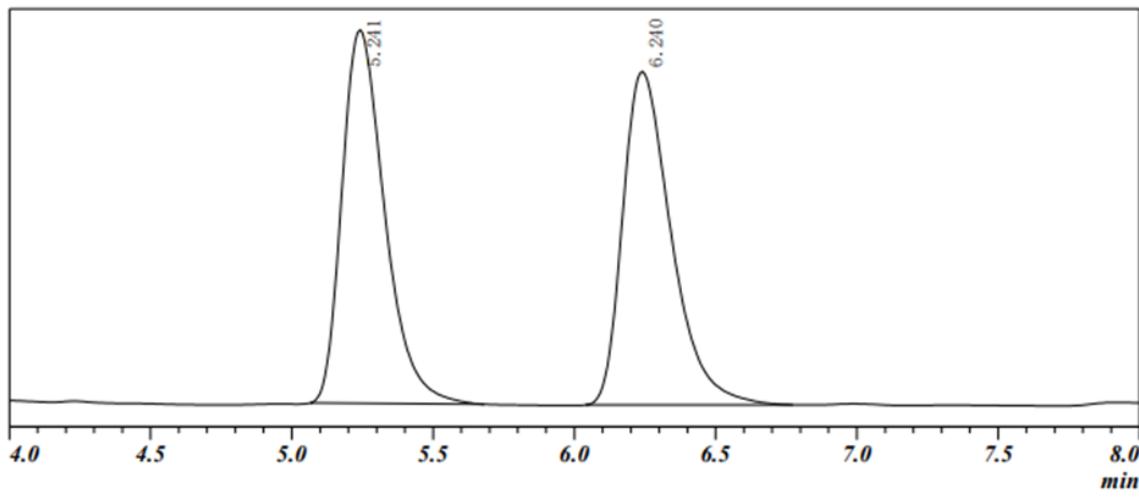
<峰表>

檢測器A Ch1 254nm

Peak NO.	Retention time	Area	Area%	Height	Height%
1	10.403	689112	99.323	38258	99.147
2	11.838	4700	0.677	329	0.853
总计		693812	100.000	38588	100.000



[3eb]

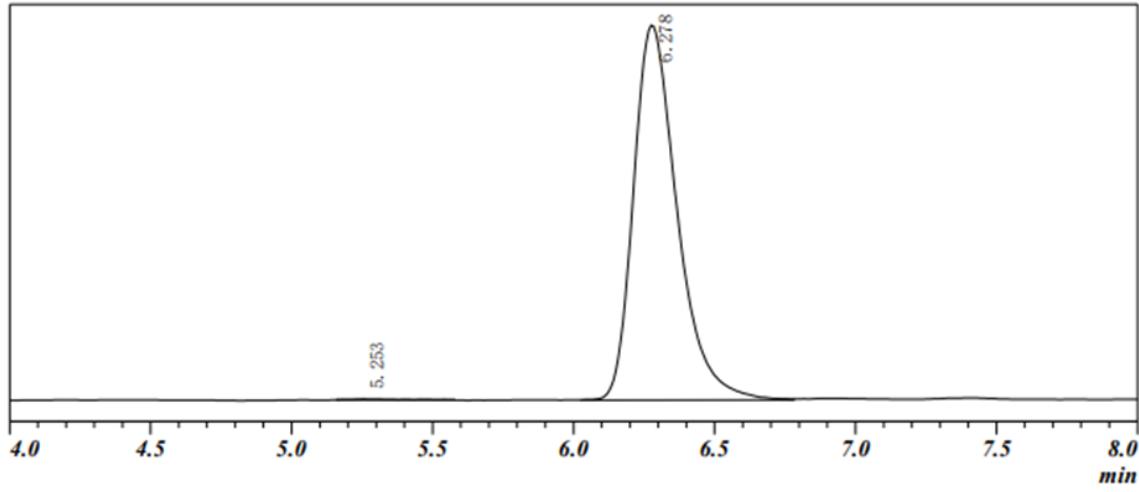


PEAK TABLE

<峰表>

檢測器A Ch1 254nm

Peak NO.	Retention time	Area	Area%	Height	Height%
1	5.241	475890	49.659	45227	52.835
2	6.240	482431	50.341	40373	47.165
总计		958321	100.000	85601	100.000

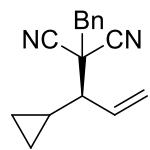


PEAK TABLE

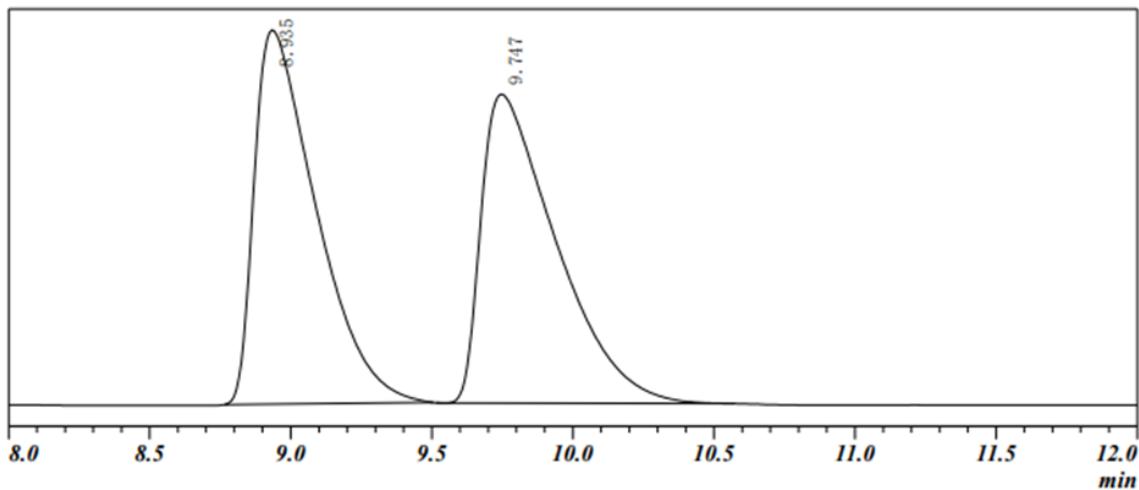
<峰表>

檢測器A Ch1 254nm

Peak NO.	Retention time	Area	Area%	Height	Height%
1	5.253	334	0.182	32	0.188
2	6.278	182937	99.818	16976	99.812
总计		183270	100.000	17008	100.000



[3fb]

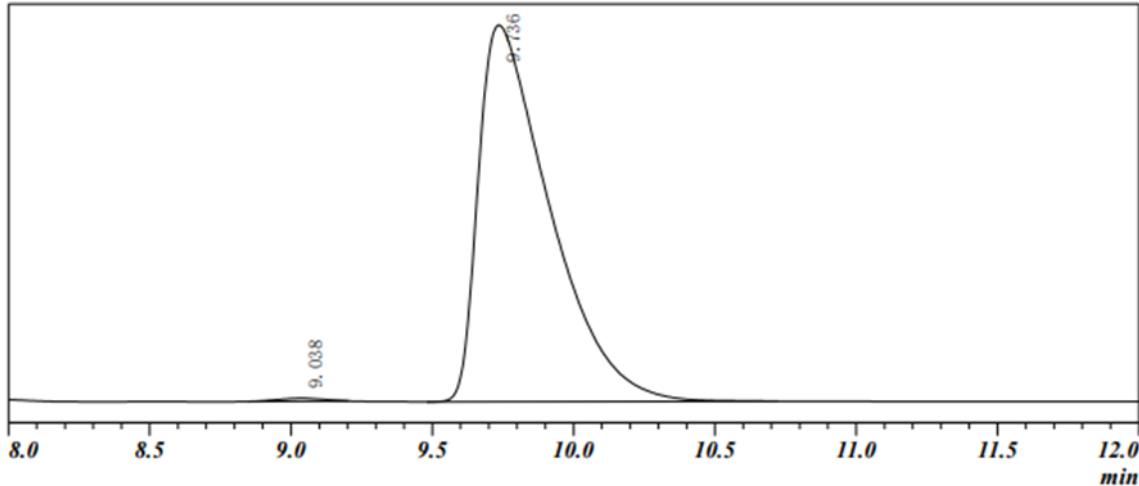


PEAK TABLE

<峰表>

检测器A Ch1 254nm

Peak NO.	Retention time	Area	Area%	Height	Height%
1	8.935	1249131	49.978	82062	54.789
2	9.747	1250224	50.022	67717	45.211
总计		2499355	100.000	149779	100.000

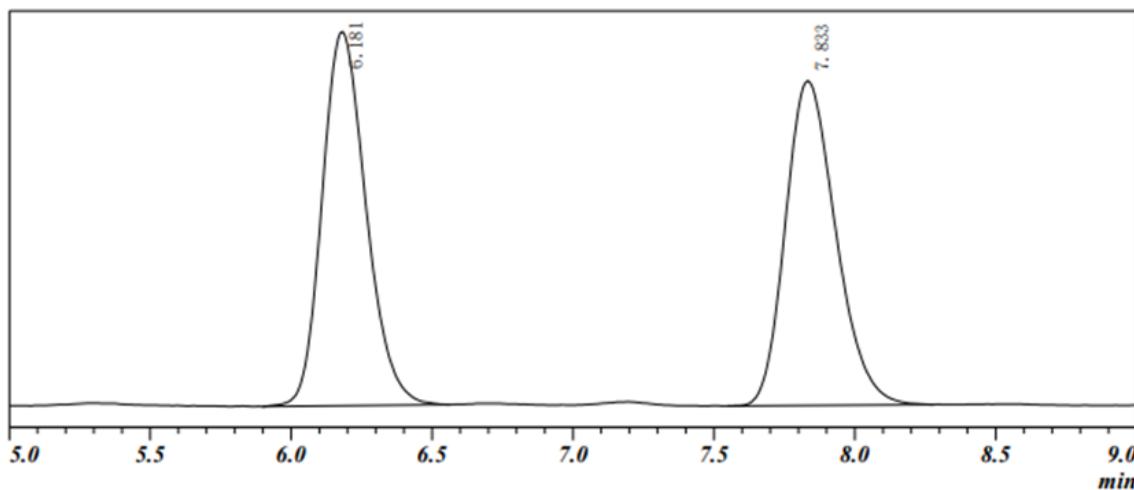
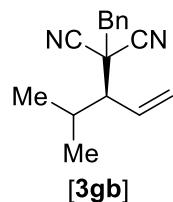


PEAK TABLE

<峰表>

检测器A Ch1 254nm

Peak NO.	Retention time	Area	Area%	Height	Height%
1	9.038	5008	0.490	456	0.782
2	9.736	1017316	99.510	57887	99.218
总计		1022325	100.000	58343	100.000

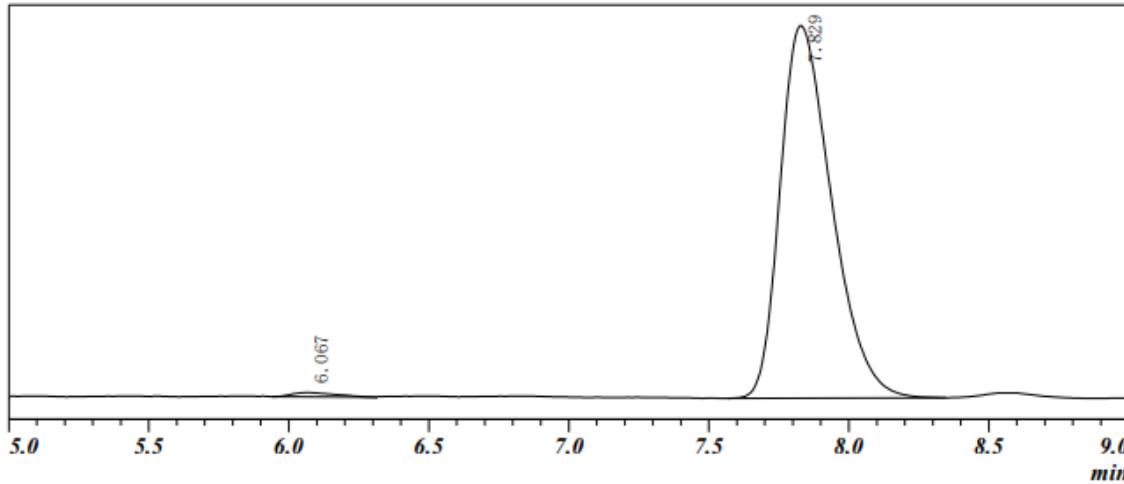


PEAK TABLE

<峰表>

檢測器A Ch1 254nm

Peak NO.	Retention time	Area	Area%	Height	Height%
1	6.181	98978	50.066	9126	53.522
2	7.833	98717	49.934	7925	46.478
总计		197695	100.000	17051	100.000

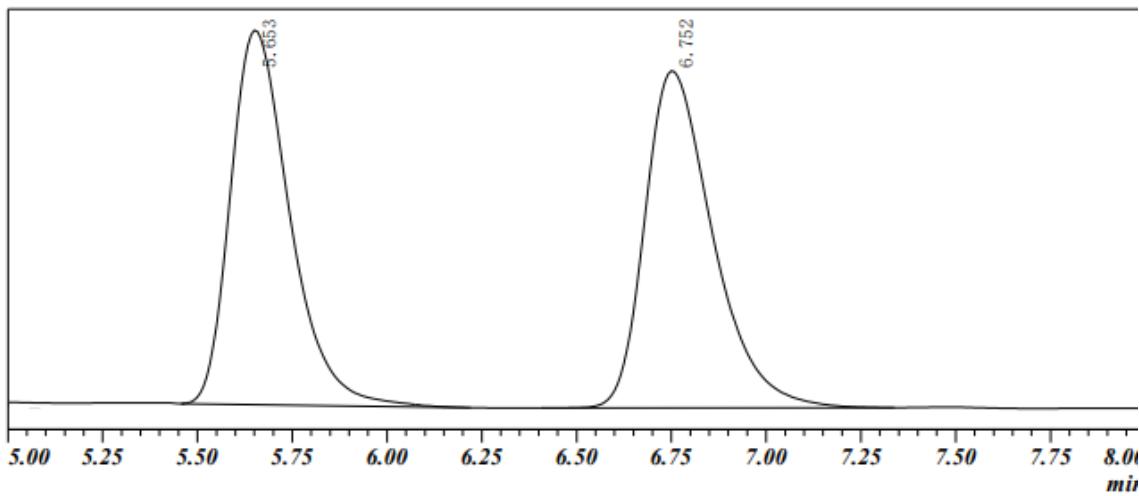
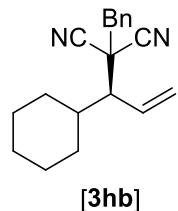


PEAK TABLE

<峰表>

檢測器A Ch1 254nm

Peak NO.	Retention time	Area	Area%	Height	Height%
1	6.067	3004	0.983	278	1.159
2	7.829	302511	99.017	23664	98.841
总计		305514	100.000	23942	100.000

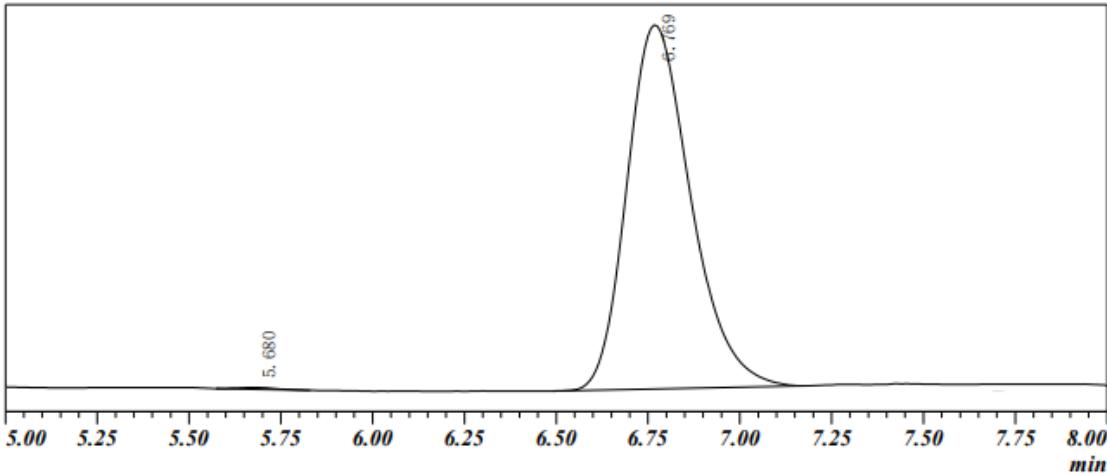


PEAK TABLE

<峰表>

检测器A Ch1 254nm

Peak NO.	Retention time	Area	Area%	Height	Height%
1	5.653	442411	49.206	40640	52.626
2	6.752	456690	50.794	36584	47.374
总计		899100	100.000	77224	100.000

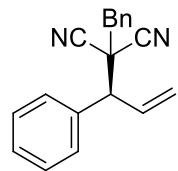


PEAK TABLE

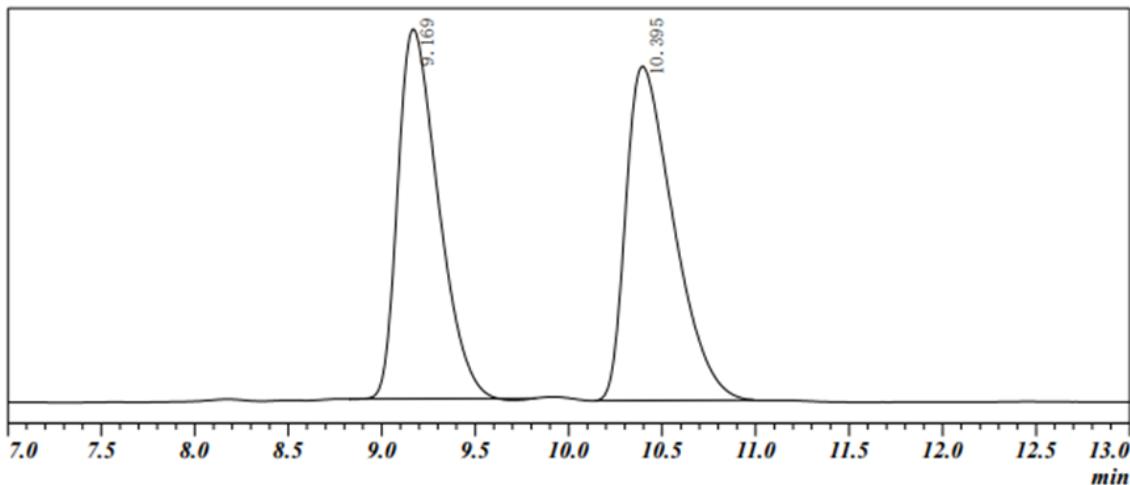
<峰表>

检测器A Ch1 254nm

Peak NO.	Retention time	Area	Area%	Height	Height%
1	5.680	415	0.278	61	0.497
2	6.769	149005	99.722	12294	99.503
总计		149420	100.000	12355	100.000



[3ib]

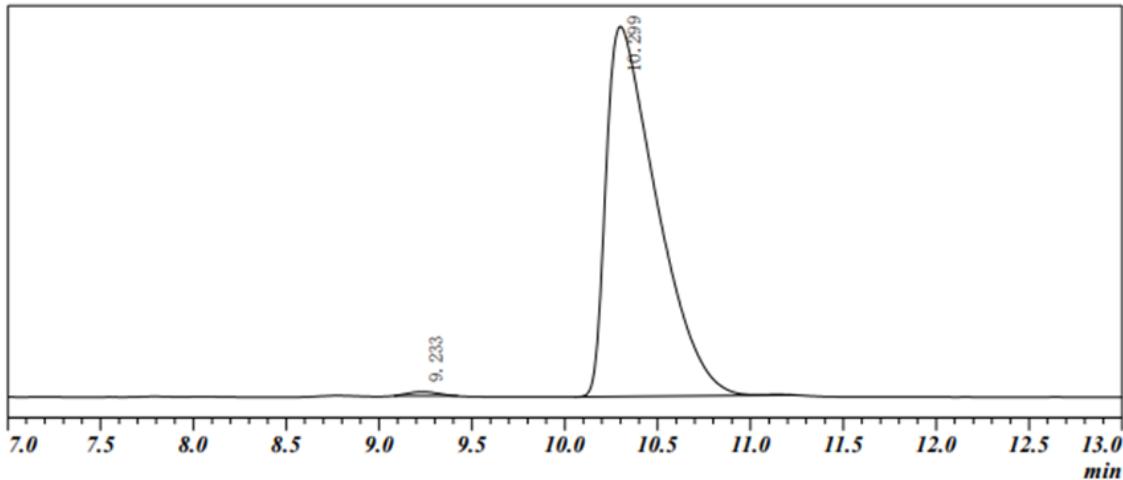


PEAK TABLE

<峰表>

檢測器A Ch1 254nm

Peak NO.	Retention time	Area	Area%	Height	Height%
1	9.169	1251278	48.382	83259	52.523
2	10.395	1334946	51.618	75259	47.477
总计		2586224	100.000	158518	100.000

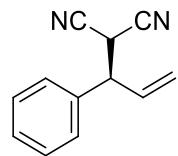


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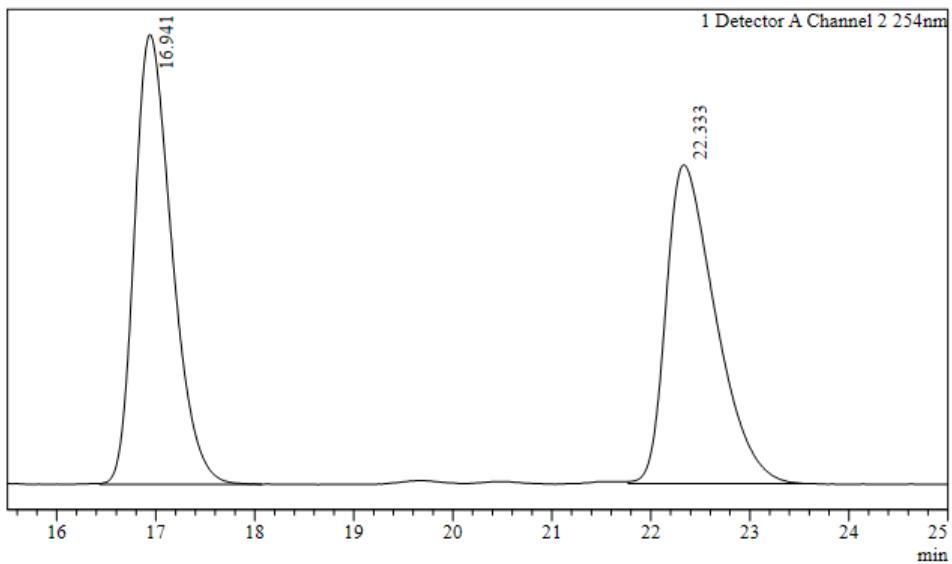
<峰表>

檢測器A Ch1 254nm

Peak NO.	Retention time	Area	Area%	Height	Height%
1	9.233	17864	0.627	1628	1.079
2	10.299	2832343	99.373	149195	98.921
总计		2850207	100.000	150823	100.000

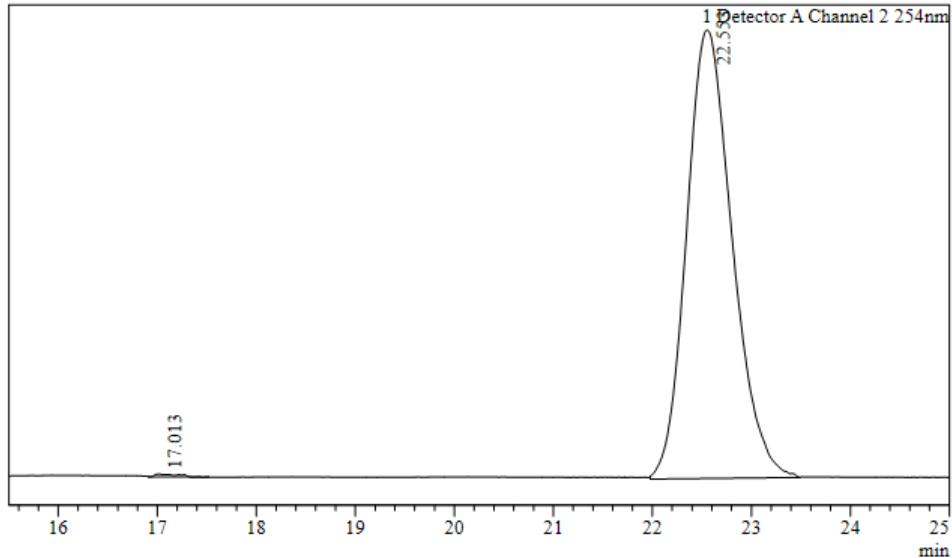


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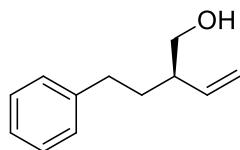
Peak Table

Detector A Channel 2 254nm					
Peak#	Ret. Time	Area	Area%	Height	Height%
1	16.941	457605	51.605	17411	58.517
2	22.333	429146	48.395	12343	41.483
Total		886751	100.000	29754	100.000

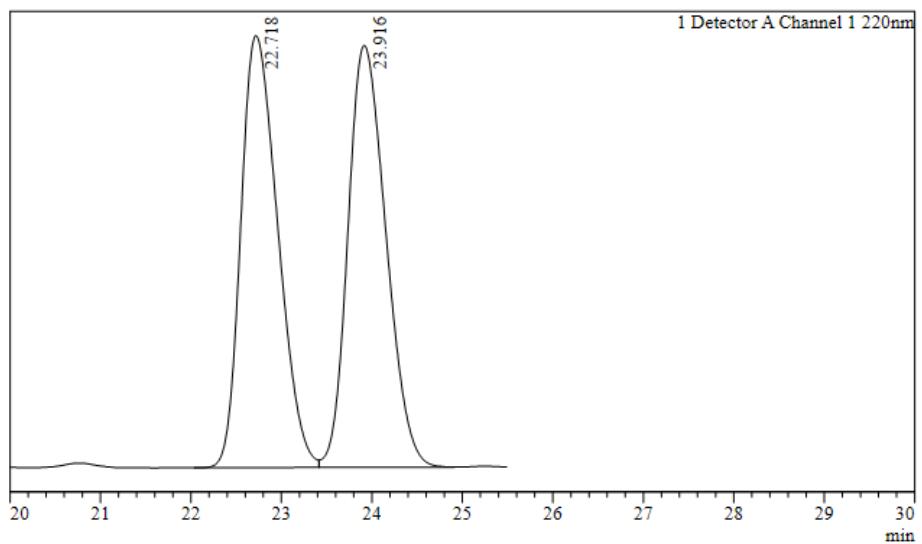


Peak Table

Detector A Channel 2 254nm					
Peak#	Ret. Time	Area	Area%	Height	Height%
1	17.013	238	0.297	14	0.578
2	22.553	79817	99.703	2490	99.422
Total		80055	100.000	2504	100.000

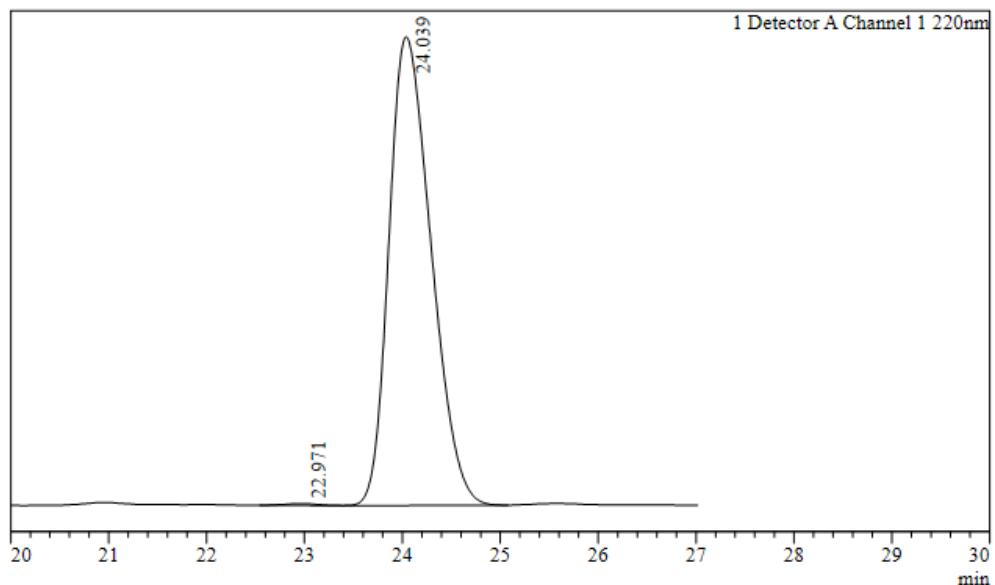


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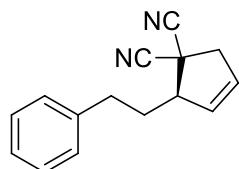
Peak Table

Detector A Channel 1 220nm						
Peak#	Ret. Time	Area	Height	Area%	Conc.	Name
1	22.718	2737301	96511	49.859	49.859	
2	23.916	2752797	94237	50.141	50.141	
Total		5490097	190748	100.000		

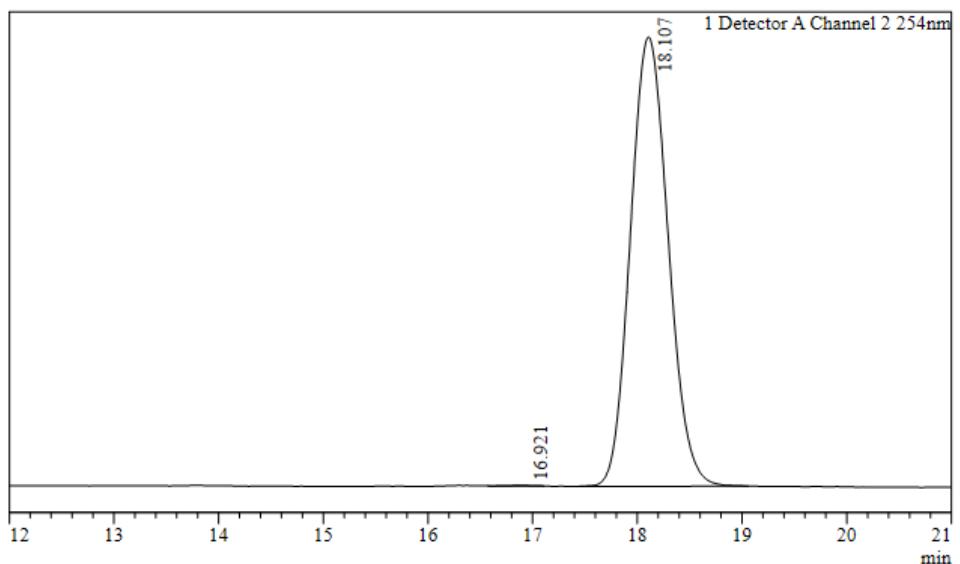
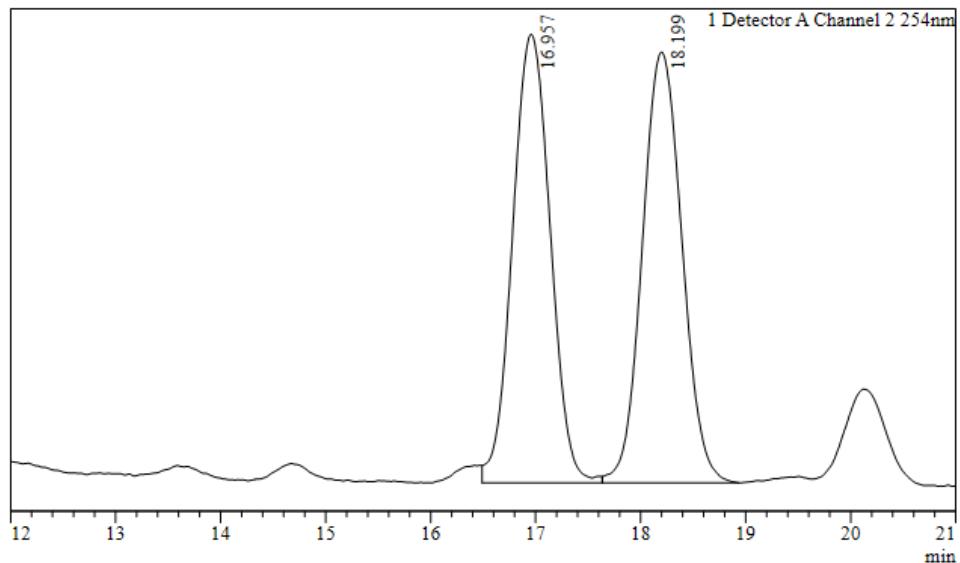


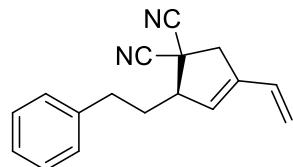
Peak Table

Detector A Channel 1 220nm						
Peak#	Ret. Time	Area	Height	Area%	Conc.	Name
1	22.971	9263	381	0.280	0.280	
2	24.039	3301121	110097	99.720	99.720	
Total		3310384	110477	100.000		

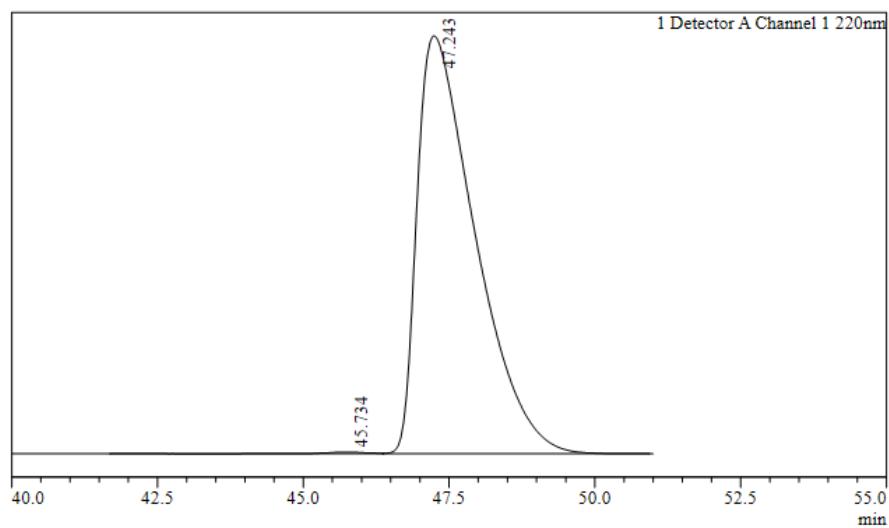
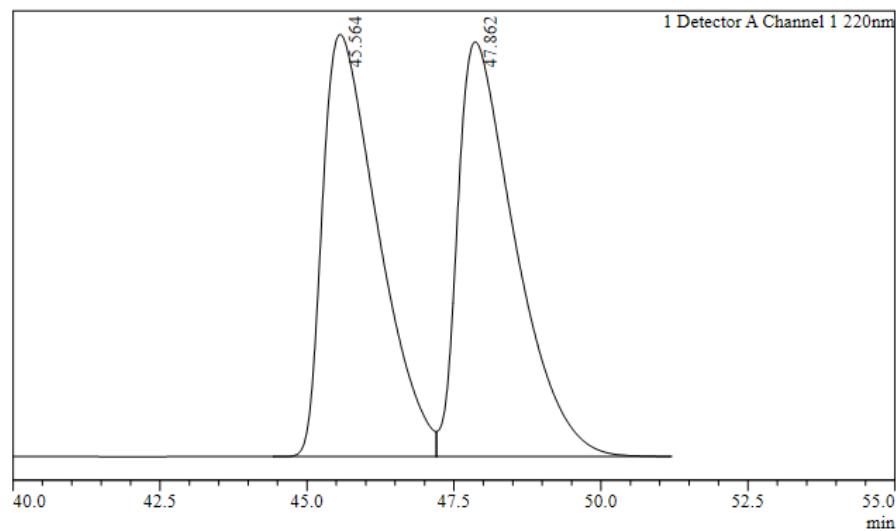


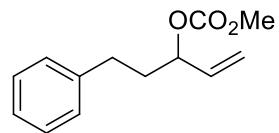
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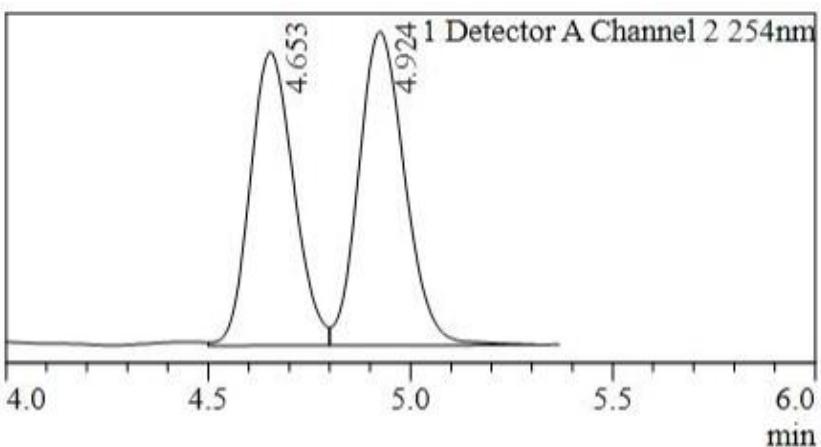
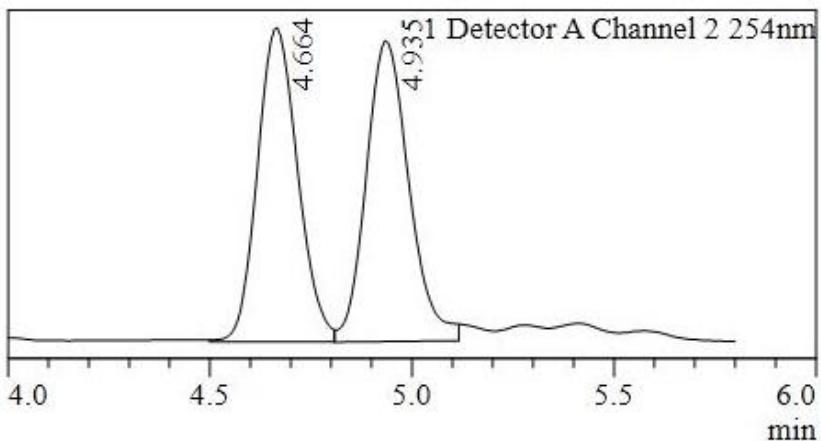


[6]





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16. References:

1. Ghorai, S.; Chirke, S. S.; Xu, W.-B.; Chen, J.-F.; Li, C. Cobalt-Catalyzed Regio- and Enantioselective Allylic Amination. *J. Am. Chem. Soc.* **2019**, *141*, 11430-11434.
2. Takizawa, K.; Sekino, T.; Sato, S.; Yoshino, T.; Kojima, M.; Matsunaga, S. Cobalt-Catalyzed Allylic Alkylation Enabled by Organophotoredox Catalysis. *Angew. Chem. Int. Ed.* **2019**, *58*, 9199-9203.
3. Tayyari, F.; Wood, D. E.; Fanwick, P. E.; Sammelson, R. E. Monosubstituted Malononitriles: Efficient One-Pot Reductive Alkylations of Malononitrile with Aromatic Aldehydes. *Synthesis*, **2008**, *2*, 279-285.
4. Ghorai, M. K.; Talukdar, R.; Tiwari, D. P. A Route to Highly Functionalized β -Enaminoesters via a Domino Ring-Opening Cyclization/Decarboxylative Tautomerization Sequence of Donor–Acceptor Cyclopropanes with Substituted Malononitriles. *Org. Lett.* **2014**, *16*, 2204-2207.
5. Grugel, C. P.; Breit, B. Rhodium-Catalyzed Asymmetric Allylation of Malononitriles as Masked Acyl Cyanide with Allenes: Efficient Access to β , γ -Unsaturated Carbonyls. *Chem. Eur. J.* **2018**, *24*, 15223-15226.
6. Stanley, L. M.; Bai, C.; Ueda, M.; Hartwig, J. F. Iridium-Catalyzed Kinetic Asymmetric Transformations of Racemic Allylic Benzoates. *J. Am. Chem. Soc.* **2010**, *132*, 8918-8920.