Isospecific Group Transfer Polymerization of Diethyl Vinylphosphonate and Multidimensional NMR Analysis of the Polymer Microstructure

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1. Experimental Section

Materials and Methods:

All reactions were carried out under argon atmosphere using standard Schlenk or glovebox techniques. All glassware was heat dried under vacuum prior to use. Unless otherwise stated, all chemicals were purchased from Sigma-Aldrich, VWR-International or ABCR and used as received. Toluene and tetrahydrofuran were dried using an MBraun SPS-800 solvent purification system. DEVP was dried over CaH₂ and destilled prior to use.

NMR spectra were recorded at 300 K on a Bruker AV-NEO 400 spectrometer equipped with a triple resonance TBI-probe, the $^{13}C\{^1H\}$ spectra on a Bruker AV-III-500 spectrometer equipped with a QNP-Cryoprobe and the $^1H\{^{31}P\}$ spectra on a Bruker AV-III-600 spectrometer equipped with a QCI-Cryoprobe. The details (acquisition parameters, pulse program) for the triple resonance experiments can be found in section 4. 1H NMR spectroscopic chemical shifts δ are reported in ppm relative to tetramethylsilane. $\delta(^1H)$ is calibrated to the residual proton signal of the solvent. Deuterated solvents were obtained from Eurisotop or Sigma Aldrich.

For the ESI-MS analysis a Varian 500 LC MS ion trap spectrometer was used with acetonitrile as carrier (20 μ l/min).

GPC was carried out on a Varian LC-920 equipped with two PL Polargel columns using tetrahydrofuran/water (1:1, 9 g/L tetrabutylammonium bromide) as eluent. Absolute molecular weights have been determined multiangle laser light scattering (MALLS) analysis (LC-920) using a Wyatt Dawn Heleos II in combination with a Wyatt Optilab rEX as concentration detector, coupled with GPC.

Elemental analysis was performed at the microanalytic laboratory of the Department of Inorganic Chemistry at the Technical University of Munich.

DSC measurements were carried out at a DSC G2000 of TA instruments with a heating rate of 1 °C/min in the first and 20 °C/min in the third cycle and a cooling rate of 5 °C/min.

Thermal gravimetric analysis (TGA) of the materials was carried out with a NETZSCH STA449 F5 Jupiter machine using aluminum oxide pans (70 µL with lid) with sample amounts of 1 to 5 mg. Temperature calibration of the oven cell was performed on the basis of the following metals: In, Sn, Bi, Zn, Al and Au. The baseline was corrected screening an empty sample pan with the respective measurement program prior to the experiment. The following thermal program was applied using argon gas (20 mL/min) for purging: At 27 °C isothermal

equilibration (30 min), ramp from 27 °C to 800 °C with 10 K/min, isothermal equilibrium for 5 minutes.

General procedure for the polymerization of DEVP:

Polymerizations were performed in 30 mL oven-dried glass reactors interfaced to a dual-manifold Schlenk line at various temperatures under argon atmosphere. A predetermined amount of a catalyst was first dissolved in toluene. Then the polymerization was started by addition of the monomer (1 mL) *via* a gastight syringe under vigorous stirring. After the measured time interval, a 0.2 mL aliquot was taken from the reaction mixture *via* a syringe and quickly quenched into a 4 mL vial containing 0.4 mL of undried "wet" MeOD-d₄. The quenched aliquots were later analyzed by ¹H-NMR and ³¹P-NMR to obtain the percent polymer yield data. The polymerization was immediately quenched after the removal of the aliquot by addition of 0.5 mL methanol. The solvents were evaporated from the quenched mixtures under reduced pressure, the polymers were purified by precipitation in pentane and dried in a vacuum oven at 50 °C overnight to a constant weight.

Activity measurements:

The stated amount of catalyst is dissolved in toluene and the reaction mixture is thermostated to the desired temperature. Then, the stated amount of monomer is added. During the course of the measurement, the temperature is monitored with a digital thermometer and aliquots (0.5 ml) are taken and quenched by addition to MeOD-d₄ (0.2 ml). After the stated reaction time, the reaction is quenched by addition of MeOH (0.5 ml). The procedure was performed at least twice for every polymerization to obtain accurate activity values The TOFs are calculated corresponding to the following equation:¹

$$TOF = \frac{n(Mon)}{n(Cat) \times t} = \frac{n(Mon)_0 \times X}{n(Cat) \times t}$$

Characterization of Synthesized Organometallic Compounds:

The ligands $(C_5Me_4H)CH_2CH_2NHtBu$, $(C_9H_7)SiMe_2NHtBu$, $(C_5Me_4H)SiMe_2NHtPh^4$ and $(C_5Me_4H)SiMe_2NHtBu^5$ were synthesized according to literature procedures.

LiCH2TMS 1.72 g Lithium granulate (249 mmol, 3.3 eq.) and 9.25 g chloromethyltrimethylsilane (75.0 mmol, 1 eq.) are suspended in 100 mL hexane and the mixture is heated at 35 °C for 24 h. Lithium is exempt of lithiumchloride several times using an ultrasonic bath. The supernatant solution is isolated using a filter cannula. The residue is

extracted three times with hexane (15 ml) and the solvent is removed in vacuo, yielding a white pyrophoric solid (6.25 g, 66.0 mmol, 88%). ¹H NMR (400 MHz, C_6D_6) $\delta = 0.16$ (s, 9H), -2.08 (s, 2H). ¹³C NMR (126 MHz, C_6D_6) $\delta = 3.6$, -4.7. elemental analysis calcd (%) for $C_4H_{11}LiSi$: C, 51.03; H, 11.78. Found: C, 51.14; H, 12.00.

YCl₃(thf)_{3.5} A glass thimble is charged inside a glovebox to 2/3 with yttrium(III) chloride and is placed in a Soxhlet extractor. All glass joints of the reaction setup are diligently sealed using Teflon grease. The Soxhlet extractor is attached to a Schlenk flask with 150 mL THF, a reflux condenser, and a pressure valve using a Schlenk line outside the glovebox. The oil bath is heated at 110 °C and THF is heated to reflux for 36 h under vigorous stirring. After cooling the Schlenk flask to room temperature and detaching it from the reaction setup, THF is removed in vacuo. The product is obtained as a white powder and the amount of coordinating THF is determined by elemental analysis. The yield is nearly quantitative if pure yttrium (III)chloride is used as a starting material. Elemental analysis calcd. for YCl₃(thf)_{3.5}: C 37.56, H 6.30. Found: C 37.45, H 6.33.

YCH₂TMS₃(thf)₂ 1.79 g YCl₃(thf)_{3.5} (4.00 mmol, 1 eq.) is suspended in 25 mL pentane. A solution of 1.13 g trimethylsilylmethyl lithium (12 mmol, 3 eq.) in 35 mL pentane is added dropwise at 0 °C and the reaction solution is stirred at 0 °C for 2 h. The supernatant solution is isolated using a filter cannula and the residue is extracted with pentane (2 × 10 mL). The solvent is removed in vacuo and the product is obtained as a white solid (1.78 g, 3.60 mmol, 90%). ¹H NMR (400 MHz, C_6D_6) $\delta = 4.01-3.87$ (m, 8 H, THF-H), 1.36–1.21 (m, 8 H, THF-H), 0.31 (s, 27 H, CH₂Si*Me*₃), -0.67 (d, 6 H, C*H*₂Si*Me*₃). ¹³C NMR (126 MHz, C_6D_6) $\delta = 69.8$, 33.8, 25.2, 4.5.

(C₅Me₄)Me₂SiN*t*BuYCH₂TMS(thf) was synthesized according to an literature procedure.⁶ Recrystallization from pentane yields a colorless powder (2.29 g, 4.60 mmol, 52%). ¹H NMR (400 MHz, C₆D₆) δ = 3.38–3.15 (m, 4H, THF-H), 2.21 (s, 12H, C*H*₃), 1.39 (s, 9H, NC(C*H*₃)₃), 1.03–0.96 (m, 4H, THF-H), 0.77 (s, 6H, SiMe₂), 0.31 (s, 9H, CH₂Si*Me*₃), -0.90 (d, ¹*J* = 3.2 Hz, 2H, YC*H*₂). ¹³C NMR (126 MHz, C₆D₆) δ = 126.4, 122.3, 106.6, 70.7, 54.0, 36.0, 26.2, 24.7, 14.0, 11.5, 8.4, 4.7. ²⁹Si NMR (99 MHz, C₆D₆) δ = -2.73, -25.06. Anal. Calcd for C₂₃H₄₆NOSi₂Y: C, 55.51; H, 9.32; N, 2.81. Found: C, 55.77; H, 9.50; N, 2.85.

(C₅Me₄)CH₂CH₂NtBuYCH₂TMS(thf) 988 mg Y(CH₂TMS)₃(thf)₂ (1.89 mmol, 1 eq.) is dissolved in 18 mL pentane and the mixture is cooled to 0 °C. A solution of 418 mg (C₅Me₄H)CH₂CH₂NHtBu (1.89 mmol, 1 eq.) in 7 mL pentane is added dropwise and the reaction solution is stirred for two hours at 0 °C. Volatile compounds are removed in vacuo,

resulting in a yellow solid. The crude product is washed with cold pentane and dried in vacuo. The product is purified by recrystallization from pentane and is obtained as a slightly yellow crystalline solid (787 mg, 1.68 mmol, 89%). 1 H NMR (400 MHz, $C_{6}D_{6}$) $\delta = 3.89-3.81$ (m, 2H, CH₂CH₂N), 3.60–3.20 (m, 4H, THF-H), 3.08 - 3.01 (t, 2H, CH₂CH₂N), 2.25–1.96 (m, 12H, CH₃), 1.34 (s, 9H, NC(CH₃)₃), 1.08–0.97 (m, 4H, THF-H), 0.36 (s, 9H, CH₂Si*Me*₃), -0.97 (d, 2H, YCH₂). 13 C NMR (126 MHz, $C_{6}D_{6}$) $\delta = 126.4$, 126.2, 126.0, 68.5, 53.7, 52.4, 28.2, 25.8, 23.2, 22.7, 9.1, 3.0, -2.0. 29 Si NMR (99 MHz, $C_{6}D_{6}$) $\delta = -2.86$. elemental analysis calcd (%) for C_{23} H₄₄NOSiY: C 59.08, H 9.48, N 3.00. Found: C 58.70, H 9.43, N 3.17.

(C₅Me₄)Me₂SiNPhYCH₂TMS(thf)₂ 959 mg Y(CH₂TMS)₃(thf)₂ (1.96 mmol, 1.1 eq.) is dissolved in 10 mL hexane and the mixture is cooled to 0 °C. A solution of 480 mg (C₅Me₄H)SiMe₂NHPh (1.77 mmol, 1 eq.) in 5 mL hexane is added dropwise and the reaction solution is stirred for two hours at r.t. After filtration the residue is washed with pentane (2 × 5 mL) and dried in vacuo. The product is purified by diffusion crystallization in toluene/pentane at –35 °C and obtained as a colorless crystalline solid (870 mg, 1.49 mmol, 84%). H NMR (400 MHz, C₆D₆) δ = 7.27-7.20 (m, 2H, Ph-H), 6.73-6.64 (m, 3H, Ph-H), 3.69–3.53 (m, 8H, THF-H), 2.11 (s, 6H, C*H*₃), 2.06 (s, 6H, C*H*₃), 1.42–1.31 (m, 8H, THF-H), 0.85 (s, 6H, SiMe₂), 0.25 (s, 9H, CH₂Si*Me*₃), –1.07 (d, ${}^{1}J$ = 2.6 Hz, 2H, YC*H*₂). 13 C NMR (126 MHz, C₆D₆) δ = 156.6, 129.4, 126.8, 122.6, 119.6, 114.5, 105.6, 68.2, 25.3, 21.4, 13.7, 11.4, 4.7, 4.3. 29 Si NMR (99 MHz, C₆D₆) δ = –2.44, –23.69. elemental analysis calcd (%) for C₂₉H₅₀NO₂Si₂Y: C, 59.06; H, 8.55; N, 2.37. Found: C, 58.75; H, 8.25; N, 2.29.

(C₉H₆)Me₂SiN*t*BuYCH₂TMS(thf) 2.10 g Y(CH₂TMS)₃(thf)₂ (4.24 mmol, 1. eq.) is dissolved in 50 mL pentane and the mixture is cooled to 0 °C. A solution of 1.04 mg (C₉H₇)SiMe₂NH*t*Bu (4.24 mmol, 1 eq.) in 8 mL pentane is added dropwise and the reaction solution is stirred over night at r.t. After filtration the filtrate is reduced to 10 mL and the product precipitates at -78 °C as a colorless crystalline solid (470 mg, 960 μmol, 23%). After drying the solid in vacuo, the known instability can be observed.⁷ ¹H NMR (400 MHz, C₆D₆) δ = 7.89-7.81 (m, 1H, H_{Ind}), 7.54 – 7.46 (m, 1H, H_{Ind}), 7.15 – 7.12 (m, 1H, H_{Ind}), 7.06 –7.01 (m, 1H, H_{Ind}), 6.79-6.70 (m, 2H, H_{Ind}), 2.95–2.82 (m, 4H, THF-H), 1.30 (s, 9H, NC(CH₃)₃), 0.91 (s, 3H, SiMe₂), 0.88–0.82 (m, 4H, THF-H), 0.80 (s, 3H, SiMe₂), 0.38 (s, 9H, CH₂Si*Me*₃), -0.79 (dd, ¹*J* = 3.5 Hz, 1H, YC*H*₂), -0.97 (dd, ¹*J* = 3.5 Hz, 1H, YC*H*₂). ¹³C NMR (126 MHz, C₆D₆) δ = 134.4, 131.8, 128.0, 123.4, 122.1, 120.8, 120.7, 104.2, 100.0, 70.3, 53.5, 35.5, 29.8, 24.3, 5.9, 4.3, 4.0. ²⁹Si NMR (99 MHz, C₆D₆) δ = -3.36, -24.34.

(C₅Me₄)SiMe₂N_tBuY(2,6-lutidinyl) (1) 2.20 g (C₅Me₄)SiMe₂N_tBuYCH₂TMS(thf) (4.40 mmol, 1 eq.) is dissolved in 100 mL pentane and 472 mg 2,6-lutidine (4.4 mmol, 1 eq.) in 20 mL pentane are added dropwise to the stirred solution. The mixture is stirred at room temperature for 30 minutes and volatile compounds are removed in vacuo. The residue is washed once with cold pentane and recrystallization from a saturated toluene-solution yields yellow crystals (1.78 g, 4.0 mmol, 91%). ¹H NMR (400 MHz, C₆D₆) δ = 6.76 (dd, ³*J* = 8.1 Hz, ³*J* = 7.7 Hz, 1H, H_{ar}), 6.46 (d, ³*J* = 8.1 Hz, 1H, H_{ar}), 5.98 (d, ³*J* = 7.7 Hz, 1H, H_{ar}), 2.52 (s, 3H, CH₃), 2.33 (s, 3H, CH₃), 2.15 (s, 3H, CH₃), 2.02 (s, 3H, CH₃), 1.85 (s, 3H, CH₃), 1.41 (s, 2H, YCH₂), 1.00 (s, 9H, NC(CH₃)₃), 0.83 (s, 3H, SiMe₂), 0.74 (s, 3H, SiMe₂). ¹³C NMR (126 MHz, C₆D₆) δ = 172.2, 155.0, 138.7, 127.2, 126.6, 122.8, 122.0, 114.3, 106.7, 54.4, 41.3, 33.7, 26.0, 15.6, 14.1, 11.4, 11.0, 8.3, 7.4. ²⁹Si NMR (99 MHz, C₆D₆) δ = -25.37. elemental analysis calcd (%) for C₂₂H₃₅N₂SiY: C, 59.44; H, 7.94; N, 6.30. Found: C, 59.66; H, 8.20; N, 6.22.

(C₅Me₄)CH₂CH₂NtBuY(2,6-lutidinyl) (2) 20.0 mg (C₅Me₄)CH₂CH₂NtBuYCH₂TMS(thf) (42.8 µmol, 1 eq.) is dissolved in 0.6 mL C₆D₆ and 4.58 mg 2,6-lutidine (42.8 µmol, 1 eq.) are added dropwise to the stirred solution. The mixture is stirred at room temperature for 30 minutes and NMR analysis shows quantitative conversion. ¹H NMR (400 MHz, C₆D₆) δ = 6.79-6.72 (m, 1H, H_{ar}), 6.45-6.39 (m, 1H, H_{ar}), 6.01 (d, 1H, H_{ar}), 3.61-3.57 (m, 2H, CH₂CH₂N), 3.11-3.06 (m, 2H, CH₂CH₂N), 2.45 (s, 3H, CH₃), 2.35 (s, 2H, YCH₂), 2.34 (s, 3H, CH₃), 2.20 (s, 3H, CH₃), 2.01 (s, 3H, CH₃), 1.11 (s, 9H, NC(CH₃)₃).

(C₉H₆)SiMe₂N/BuY(2,6-lutidinyl) (3) 20.0 mg (C₉H₆)SiMe₂N/BuYCH₂TMS(thf) (39.5 μmol, 1 eq.) is dissolved in 0.6 mL C₆D₆ and 4.23 mg 2,6-lutidine (39.5 μmol, 1 eq.) are added dropwise to the stirred solution. The mixture is stirred at room temperature for 30 minutes and NMR analysis shows quantitative conversion. Diffusion crystallization in C₆D₆/pentane at – 30 °C yields yellow crystals. ¹H NMR (400 MHz, C₆D₆) δ = 8.08-8.03 (m, 1H, H_{Ind}), 7.54–7.49 (m, 1H, H_{Ind}), 7.09-7.03 (m, 1H, H_{Ind}), 6.82-6.67 (m, 4H, H_{Ind}, H_{ar}), 6.18-6.14 (m, 1H, H_{ar}), 6.07-6.01 (m, 1H, H_{ar}), 2.41 (s, 6H, C*H*₃), 1.15 (s, 9H, NC(C*H*₃)₃), 1.00 (s, 3H, SiMe₂), 0.88 (s, 3H, SiMe₂), 0.29 (s, 2H, YC*H*₂). ¹³C NMR (126 MHz, C₆D₆) δ = 169.0, 164.7, 156.3, 136.3, 133.6, 131.6, 126.5, 124.4, 122.0, 120.6, 120.1, 115.7, 104.0, 54.2, 43.1, 34.5, 24.7, 6.4, 4.3. ²⁹Si NMR (99 MHz, C₆D₆) δ = -24.65. elemental analysis calcd (%) for C22H29YN2Si: C, 60.26; H, 6.67; N, 6.39. Found: C, 59.92; H, 7.04; N, 7.09.

(C₅Me₄)SiMe₂NPhY(2,6-lutidinyl) (4) 20.0 mg (C₅Me₄)Me₂SiNPhYCH₂TMS(thf)₂ (33.9 μ mol, 1 eq.) is dissolved in 0.6 mL C₆D₆ and 3.63 mg 2,6-lutidine (33.9 μ mol, 1 eq.) are added dropwise to the stirred solution. The mixture is stirred at room temperature for 30 minutes

and NMR analysis shows quantitative conversion. ¹H NMR (400 MHz, C_6D_6) $\delta = 7.29-7.17$ (m, 2H, Ph-H), 6.72 (t, 2H, Ph-H), 6.55 (d, 1H, Ph-H), 6.38 (s, 1H, H_{ar}), 6.29-6.22 (m, 1H, H_{ar}), 5.69 (s, 1H, H_{ar}), 2.17 (s, 6H, CH₃), 2.12 (s, 3H, CH₃), 2.10 (s, 2H, YCH₂), 2.04 (s, 3H, CH₃), 1.87 (s, 3H, CH₃), 0.97 (s, 6H, SiMe₂). ¹³C NMR (126 MHz, C_6D_6) $\delta = 165.9$, 156.5, 155.4, 147.1, 137.5, 129.2, 129.0, 128.2, 126.7, 125.3, 123.0, 119.4, 115.7, 115.2, 110.9, 106.9, 48.3, 31.6, 22.9, 14.0, 13.5, 11.1, 4.5. ²⁹Si NMR (99 MHz, C_6D_6) $\delta = -22.49$.

(C₅Me₄)CH₂CH₂NtBuMg · 2 THF 5.00 g (C₅Me₄H)CH₂CH₂NHtBu (22.6 mmol, 1.0 eq.) are dissolved in 100 ml toluene and 5 mL THF. The solution is cooled to 0 °C and 23.0 mL Dinbutylmagnesium (1.0 M in heptane, 3.14 g, 23.0 mmol, 1.0 eq.) is added within 30 minutes. After stirring the solution over night at r.t. the solvents are evaporated under high vacuum at 60 °C. Crystallization at –80 °C in THF yields a green solid (4.31 g, 17.7 mmol, 78%). ¹H NMR (400 MHz, C₆D₆) δ = 3.54 (m, 8H, THF-H), 3.15 (m, 1H, CH₂CH₂N), 2.70 (m, 1H, CH₂CH₂N), 2.65 (m, 1H, CH₂CH₂N), 2.48 (m, 1H, CH₂CH₂N), 2.24 (s, 3H, CH₃), 2.12 (s, 3H, CH₃), 2.06 (s, 3H, CH₃), 2.04 (s, 3H, CH₃), 1.33 – 1.49 (m, 8H, THF-H), 0.88 (s, 9H, NC(CH₃)₃). ¹³C NMR (126 MHz, C₆D₆) δ = 116.2, 111.9, 110.9, 109.7, 106.7, 68.3, 52.2, 50.3, 34.1, 30.1, 25.7, 13.0, 12.2, 11.4, 11.1.

(C₅Me₄)SiMe₂NtBuMg · 2 THF 4.00 g (C₅Me₄H)SiMe₂NHtBu (15.9 mmol, 1.0 eq.) are dissolved in 100 ml toluene and 5 mL THF. The solution is cooled to 0 °C and 16.2 mL Dinbutylmagnesium (1.0 M in heptane, 2.25 g, 16.2 mmol, 1.0 eq.) is added within 30 minutes. After stirring the solution over night at r.t. the solvents are evaporated under high vacuum at 60 °C. Crystallization at -80 °C in THF yields an orange solid (3.86 g, 11.2 mmol, 70%). ¹H NMR (400 MHz, C₆D₆) δ = 3.25 - 3.48 (m, 8H, THF-H), 2.52 (s, 6H, CH₃), 2.19 (s, 6H, CH₃), 1.42 (s, 9H, NC(CH₃)₃), 1.17 - 1.29 (m, 8H, THF-H), 0.98 (s, 6H, SiMe₂). ¹³C NMR (126 MHz, C₆D₆) δ = 116.0, 114.2, 108.4, 69.1, 50.8, 37.6, 24.8, 14.7, 12.0, 8.6.

(C₉H₆)SiMe₂NtBuMg · 2 THF 5.00 g (C₉H₇)SiMe₂NHtBu (20.4 mmol, 1.0 eq.) are dissolved in 60 ml toluene and 5 mL THF. The solution is cooled to 0 °C and 21.4 mL Dinbutylmagnesium (1.0 M in heptane, 2.96 g, 21.4 mmol, 1.0 eq.) is added within 30 minutes. After stirring the solution over night at r.t. the solvents are evaporated under high vacuum at 60 °C. Crystallization at -80 °C in THF yields a yellowish solid (4.38 g, 16.4 mmol, 70%). ¹H NMR (400 MHz, C₆D₆) δ = 8.18 (d, ³*J* = 7.9 Hz, 1H, H_{ar}), 8.01 (d, ³*J* = 7.9 Hz, 1H, H_{ar}), 7.31 (d, ³*J* = 3.4 Hz, 1H, H_{ar}), 7.14 (dd, ³*J* = 7.9 Hz, ³*J* = 6.7 Hz, 1H, H_{ar}), 7.07 (dd, ³*J* = 7.9 Hz, ³*J* = 6.7 Hz, 1H, H_{ar}), 6.89 (d, ³*J* = 3.5 Hz, 1H, H_{ar}), 3.15 (s, 8H, THF-H), 1.46 (s, 9H, NC(C*H*₃)₃),

1.12 (s, 8H, THF-H), 1.04 (s, 3H, SiMe₂), 0.91 (s, 3H, SiMe₂). 13 C NMR (126 MHz, C₆D₆) δ = 135.9, 135.1, 124.1, 120.8, 120.5, 116.7, 116.1, 103.5, 95.4, 69.7, 51.1., 38.1, 25.1, 6.6, 6.2.

(2,6-lutidinyl))**AlCl₂ · THF** 10.8 mL 2,6-lutidine (10.0 g, 93.3 mmol, 1.0 eq.) are dissolved in 100 mL THF and cooled to -78 °C. After slow addition of 39.2 mL *n*BuLi (2.5 M in pentane, 6.28 g, 1.05 eq.) a solution of 12.4 g aluminumchlorid (93.3 mmol, 1.0 eq.) in 300 mL THF is added within an hour. The mixture is carefully allowed to heat up to r.t. and the solvents are evaporated under reduced pressure. The residue is washed with 200 mL pentane and is suspended in 400 mL toluene after drying. The solvent of the filtrate is removed under vacuum. Crystallization at -90 °C in THF yields an off-white solid (18.8 g, 68.1 mmol, 73%). ¹H NMR (400 MHz, C₆D₆) δ = 6.91 (dd, ${}^{3}J$ = 7.6 Hz, ${}^{3}J$ = 7.7 Hz, 1H, H_{ar}), 6.76 (d, ${}^{3}J$ = 7.7 Hz, 1H, H_{ar}), 6.32 (d, ${}^{3}J$ = 7.6 Hz, 1H, H_{ar}), 3.78 - 3.91 (m, 4H, THF-H), 2.58 (s, 3H, C*H*₃), 1.97 (s, 2H, AlC*H*₂), 1.13 - 1.25 (m, 4H, THF-H). ¹³C NMR (126 MHz, C₆D₆) δ = 164.2, 155.1, 139.2, 121.7, 121.3, 70.4, 25.1, 21.0, 16.3. ²⁷Al NMR (78 MHz, C₆D₆) δ = 81.5.

(C₅Me₄)SiMe₂NtBuAl(2,6-lutidinyl) (5) 5.62 g (C₅Me₄)SiMe₂NtBuMg · 2 THF (16.3 mmol, 1.0 eq.) are dissolved in 150 mL THF and cooled to -10 °C. A solution of 4.49 g (2,6-lutidinyl))AlCl₂ · THF (16.3 mmol, 1.0 eq.) in 20 mL THF is added within an hour. The mixture is stirred over night at r.t. and the solvent is evaporated under reduced pressure. The residue is suspended in 200 mL pentane and the solvent of the filtrate is removed. Orange crystals could be received by recrystallization in pentane at -60 °C (5.14 g, 13.4 mmol, 83%). ¹H NMR (400 MHz, C₆D₆) δ = 6.67 (dd, ³*J* = 7.7 Hz, ³*J* = 7.7 Hz, 1H, H_{ar}), 6.50 (d, ³*J* = 7.7 Hz, 1H, H_{ar}), 6.15 (d, ³*J* = 7.7 Hz, 1H, H_{ar}), 2.40 (s, 6H, C*H*₃), 2.21 (s, 3H, C*H*₃), 2.14 – 1.70 (m, 6H, C*H*₃), 1.32 – 1.59 (m, 2H, AlC*H*₂), 1.25 (s, 9H, NC(C*H*₃)₃), 0.81 (s, 6H, SiMe₂). ¹³C NMR (126 MHz, C₆D₆) δ = 168.4, 154.3, 139.7, 122.7, 121.0, 108.3, 51.4, 35.7, 22.8, 19.1, 15.3, 11.6, 6.6. ²⁷Al NMR (78 MHz, C₆D₆) δ = 65.1. ²⁹Si NMR (60 MHz, C₆D₆) δ = –14.6. elemental analysis calcd (%) for C22H35AlN2Si: C, 69.06; H, 9.22; N, 7.32. Found: C, 69.28; H, 9.37; N, 7.10.

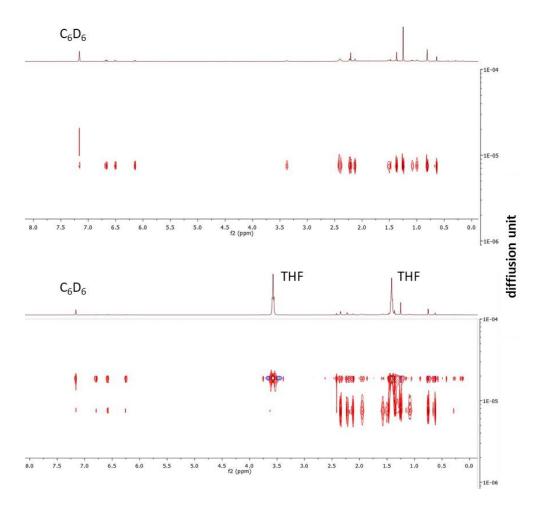


Figure S1. ¹H DOSY NMR spectra of **5** in C₆D₆ without (top) and with 10 eq. THF (bottom).

(C₅Me₄)CH₂CH₂NtBuAl(2,6-lutidinyl) (6) 1.25 g (C₅Me₄)CH₂CH₂NtBuMg · 2 THF (5.13 mmol, 1.0 Äq.) are dissolved in 50 mL THF and cooled to 0 °C. A solution of 1.42 g (2,6-lutidinyl))AlCl₂ · THF (5.13 mmol, 1.0 eq.) in 20 mL THF is added within 30 minutes. The mixture is stirred over night at r.t. and the solvent is evaporated under reduced pressure. The residue is suspended in 50 mL pentane and the solvent of the filtrate is removed. The red-brown solid could be received by precipitation in pentane -60 °C (1.02 g, 2.89 mmol, 56%). ¹H NMR (400 MHz, C₆D₆) δ = 6.73 (dd, ${}^{3}J$ = 7.5 Hz, ${}^{3}J$ = 7.6 Hz, 1H, H_{ar}), 6.60 (d, ${}^{3}J$ = 7.6 Hz, 1H, H_{ar}), 6.20 (d, ${}^{3}J$ = 7.5 Hz, 1H, H_{ar}), 3.46 (t, ${}^{3}J$ = 6.6 Hz, 2H, CH₂CH₂N), 3.05 (t, ${}^{3}J$ = 6.6 Hz, 2H, CH₂CH₂N), 2.21 (s, 3H, CH₃), 2.16 (s, 6H, CH₃), 2.14 – 1.88 (m, 6H, CH₃), 1.40 (s, 2H, AlCH₂), 1.15 (s, 9H, NC(CH₃)₃). ¹³C NMR (126 MHz, C₆D₆) δ = 168.3, 153.7, 139.1, 129.3, 124.9, 122.6, 120.8, 105.6, 51.7, 45.0, 31.1, 27.1, 22.6, 11.9, 11.8, 11.2.

(C₉H₆)SiMe₂NtBuAl(2,6-lutidinyl) (7) 3.00 g (C₉H₆)SiMe₂NtBuMg · 2 THF (11.2 mmol, 1.0 Äq.) are dissolved in 150 mL toluene and cooled to 0 °C. A solution of 3.09 g (2,6-lutidinyl))AlCl₂ · THF (11.2 mmol, 1.0 eq.) in 50 mL toluene is added within an hour. The mixture is stirred over night at r.t. and the solvent is evaporated under reduced pressure. The

residue is dissolved in 50 mL pentane and after one week at -45 °C, the solid is separated and dried. Yellow crystals could be received by recrystallization in toluene at -45 °C (3.29 g, 8.74 mmol, 78%). ¹H NMR (400 MHz, C₆D₆) δ = 7.62 (d, 1H, H_{Ind}), 7.09 – 7.16 (m, 1H, H_{Ind}), 6.82 – 6.88 (m, 2H, H_{Ind}), 6.67 – 6.76 (m, 2H, H_{ar}), 6.49 (d, ³*J* = 4.7 Hz, 1H, H_{Ind}), 6.37 (d, ³*J* = 4.7 Hz, 1H, H_{Ind}), 6.18 (d, 1H, H_{ar}), 2.86 (s, 6H, C*H*₃), 1.31 (s, 9H, NC(C*H*₃)₃), 1.28 (d, ²*J* = 12.0 Hz, 1H, AlC*H*₂), 0.43 (s, 3H, SiMe₂), 0.33 (s, 3H, SiMe₂), 1.04 (d, ²*J* = 12.0 Hz, 1H, AlC*H*₂). ¹³C NMR (126 MHz, C₆D₆) δ = 169.2, 155.1, 146.1, 143.0, 138.7, 136.8, 122.9, 120.9, 120.6, 120.4, 120.3, 119.0, 116.3, 110.4, 51.3, 35.5, 28.3, 24.7, 6.6, 4.6. elemental analysis calcd (%) for C22H29AlN2Si: C, 70.17; H, 7.76; N, 7.44. Found: C, 69.92; H, 8.04; N, 7.09.

2. Single Crystal X-Ray Diffraction Experiments

SC-XRD determination of compound 1 (CCDC 1936317)

A yellow fragment-like specimen of $C_{26}H_{43}N_2OSiY$, approximate dimensions 0.139 mm x 0.148 mm x 0.156 mm, was used for the X-ray crystallographic analysis. The X-ray intensity data were measured on a Bruker D8 Venture system equipped with a Helios optic monochromator and a Mo TXS rotating anode ($\lambda = 0.71073 \text{ Å}$).

A total of 1713 frames were collected. The total exposure time was 4.31 hours. The frames were integrated with the Bruker SAINT software package using a narrow-frame algorithm. The integration of the data using a monoclinic unit cell yielded a total of 60076 reflections to a maximum θ angle of 25.35° (0.83 Å resolution), of which 4876 were independent (average redundancy 12.321, completeness = 100.0%, $R_{int} = 6.84\%$, $R_{sig} = 2.79\%$) and 3942 (80.84%) were greater than $2\sigma(F^2)$. The final cell constants of a = 10.9805(4) Å, b = 20.9144(8) Å, c = $11.6587(4) \text{ Å}, \beta = 94.721(2)^{\circ}, \text{ volume} = 2668.34(17) \text{ Å}^3, \text{ are based upon the refinement of the}$ XYZ-centroids of 132 reflections above 20 $\sigma(I)$ with $4.202^{\circ} < 2\theta < 34.37^{\circ}$. Data were corrected for absorption effects using the Multi-Scan method (SADABS). The ratio of minimum to maximum apparent transmission was 0.866. The calculated minimum and maximum (based transmission coefficients crystal size) 0.7200 and 0.7450. on are

The final anisotropic full-matrix least-squares refinement on F^2 with 290 variables converged at R1=3.34%, for the observed data and wR2=8.15% for all data. The goodness-of-fit was 1.070. The largest peak in the final difference electron density synthesis was $0.712~e^{-}/Å^3$ and the largest hole was $-0.613~e^{-}/Å^3$ with an RMS deviation of $0.070~e^{-}/Å^3$. On the basis of the final model, the calculated density was $1.286~g/cm^3$ and F(000), $1096~e^{-}$.

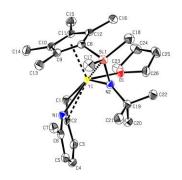


Figure S2. Ortep drawing with 50% ellipsoids for compound 1.

Table S1. Sample and crystal data for compound 1.

Identification codePahPh7 AP9248-100

Chemical formula C₂₆H₄₃N₂OSiY

Formula weight516.62Temperature100(2) KWavelength0.71073 Å

Crystal size 0.139 x 0.148 x 0.156 mm

Crystal habityellow fragmentCrystal systemmonoclinicSpace groupP 1 21/n 1

Unit cell dimensions a = 10.9805(4) Å $\alpha = 90^{\circ}$

b = 20.9144(8) Å $\beta = 94.721(2)^{\circ}$

c = 11.6587(4) Å $\gamma = 90^{\circ}$

Volume 2668.34(17) Å³

Z 4

Density (calculated)1.286 g/cm³**Absorption coefficient**2.250 mm⁻¹**F(000)**1096

Table S2. Data collection and structure refinement for compound 1.

DiffractometerBruker D8 VentureRadiation sourceTXS rotating anode, Mo

Theta range for data

collection 1.95 to 25.35°

Index ranges -13<=h<=13, -25<=k<=25, -14<=l<=14

100.0%

Reflections collected 60076

Independent reflections 4876 [R(int) = 0.0684]

Coverage of independent

reflections

Absorption correction Multi-Scan

Max. and min. transmission 0.7450 and 0.7200

Refinement method Full-matrix least-squares on F² **Refinement program** SHELXL-2014/7 (Sheldrick, 2014)

Function minimized $\Sigma \text{ w}(F_o^2 - F_c^2)^2$ Data / restraints / parameters 4876 / 0 / 290

Goodness-of-fit on F^2 1.070 Δ/σ_{max} 0.001

Final R indices 3942 data; R1 = 0.0334, wR2 = 0.0334, wR

 $I > 2\sigma(I)$ 0.0714

all data R1 = 0.0523, wR2 = 0.015

0.0815

Weighting scheme $W=1/[\sigma^2(F_0^2)+(0.0234P)^2+4.9294P]$

where $P=(F_0^2+2F_c^2)/3$

Largest diff. peak and hole $0.712 \text{ and } -0.613 \text{ eÅ}^{-3}$

R.M.S. deviation from mean 0.070 eÅ^{-3}

SC-XRD determination of compound 3 (CCDC 1936316)

A clear colourless fragment-like specimen of $C_{22}H_{29}N_2SiY$, approximate dimensions 0.126 mm x 0.178 mm x 0.234 mm, was used for the X-ray crystallographic analysis. The X-ray intensity data were measured on a Bruker D8 Venture Duo IMS system equipped with a Helios optic monochromator and a Mo IMS microsource ($\lambda = 0.71073 \text{ Å}$).

A total of 1563 frames were collected. The total exposure time was 0.82 hours. The frames were integrated with the Bruker SAINT software package using a narrow-frame algorithm. The integration of the data using a monoclinic unit cell yielded a total of 66552 reflections to a maximum θ angle of 29.57° (0.72 Å resolution), of which 5978 were independent (average redundancy 11.133, completeness = 100.0%, R_{int} = 9.34%, R_{sig} = 4.29%) and 4877 (81.58%) were greater than $2\sigma(F^2)$. The final cell constants of \underline{a} = 10.4940(5) Å, \underline{b} = 12.6978(6) Å, \underline{c} = 15.9846(7) Å, β = 91.044(2)°, volume = 2129.61(17) ų, are based upon the refinement of the XYZ-centroids of 9863 reflections above 20 $\sigma(I)$ with 4.682° < 20 < 61.07°. Data were corrected for absorption effects using the Multi-Scan method (SADABS). The ratio of minimum to maximum apparent transmission was 0.793. The calculated minimum and maximum transmission coefficients (based on crystal size) are 0.5600 and 0.7190.

The final anisotropic full-matrix least-squares refinement on F^2 with 249 variables converged at R1 = 3.48%, for the observed data and wR2 = 6.92% for all data. The goodness-of-fit was 1.041. The largest peak in the final difference electron density synthesis was 0.461 $e^-/Å^3$ and the largest hole was -0.386 $e^-/Å^3$ with an RMS deviation of 0.084 $e^-/Å^3$. On the basis of the final model, the calculated density was 1.368 g/cm³ and F(000), 912 e^- .

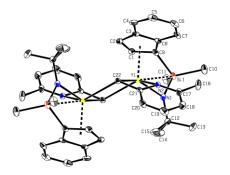


Figure S3. Ortep drawing with 50% ellipsoids for compound 3.

Table S3. Sample and crystal data for compound **3**.

Identification code PahPh6 AP8358-100

Chemical formula C₂₂H₂₉N₂SiY

Formula weight438.47Temperature100(2) KWavelength0.71073 Å

Crystal size 0.126 x 0.178 x 0.234 mm Crystal habit clear colourless fragment

Crystal system monoclinic **Space group** P 1 21/n 1

Unit cell dimensions a = 10.4940(5) Å $\alpha = 90^{\circ}$

b = 12.6978(6) Å $\beta = 91.044(2)^{\circ}$

c = 15.9846(7) Å $\gamma = 90^{\circ}$

Volume 2129.61(17) Å³

 \mathbf{Z}

Density (calculated) 1.368 g/cm³ **Absorption coefficient** 2.802 mm⁻¹

F(000) 912

Table S4. Data collection and structure refinement for compound 3.

Diffractometer Bruker D8 Venture Duo IMS

Radiation source IMS microsource, Mo

Theta range for data

collection 2.30 to 29.57°

Index ranges -14<=h<=14, -17<=k<=17, -22<=l<=22

100.0%

Reflections collected 66552

Independent reflections 5978 [R(int) = 0.0934]

Coverage of independent

reflections

Absorption correction Multi-Scan

Max. and min. transmission 0.7190 and 0.5600

Refinement method Full-matrix least-squares on F² **Refinement program** SHELXL-2014/7 (Sheldrick, 2014)

Function minimized $\Sigma \text{ w}(F_o^2 - F_c^2)^2$ **Data / restraints / parameters** 5978 / 0 / 249

Goodness-of-fit on F^2 1.041 Δ/σ_{max} 0.002

Final R indices 4877 data; R1 = 0.0348, wR2 = 0.0348

 $I > 2\sigma(I)$ 0.0645

all data R1 = 0.0534, wR2 = 0.0534

0.0692

Weighting scheme $W=1/[\sigma^2(F_0^2)+(0.0242P)^2+1.9235P]$

where $P = (F_0^2 + 2F_c^2)/3$

R.M.S. deviation from mean 0.084 eÅ^{-3}

SC-XRD determination of compound 5 (CCDC 1936318)

A clear yellow fragment-like specimen of $C_{44}H_{70}Al_2N_4Si_2$, approximate dimensions 0.136 mm x 0.137 mm x 0.203 mm, was used for the X-ray crystallographic analysis. The X-ray intensity data were measured on a Bruker D8 Venture system equipped with a Helios optic monochromator and a Mo TXS rotating anode ($\lambda = 0.71073 \text{ Å}$).

A total of 1443 frames were collected. The total exposure time was 3.56 hours. The frames were integrated with the Bruker SAINT software package using a narrow-frame algorithm. The integration of the data using a triclinic unit cell yielded a total of 42515 reflections to a maximum θ angle of 25.03° (0.84 Å resolution), of which 8029 were independent (average redundancy 5.295, completeness = 99.8%, R_{int} = 3.46%, R_{sig} = 2.41%) and 6684 (83.25%) were greater than $2\sigma(F^2)$. The final cell constants of \underline{a} = 11.8436(4) Å, \underline{b} = 12.0036(5) Å, \underline{c} = 16.2833(6) Å, α = 87.653(2)°, β = 80.177(2)°, γ = 88.234(2)°, volume = 2278.41(15) ų, are based upon the refinement of the XYZ-centroids of 9290 reflections above 20 $\sigma(I)$ with 4.930° < 2 θ < 51.39°. Data were corrected for absorption effects using the Multi-Scan method (SADABS). The ratio of minimum to maximum apparent transmission was 0.915. The calculated minimum and maximum transmission coefficients (based on crystal size) are 0.9700 and

The final anisotropic full-matrix least-squares refinement on F^2 with 698 variables converged at R1 = 4.64%, for the observed data and wR2 = 11.77% for all data. The goodness-of-fit was 1.040. The largest peak in the final difference electron density synthesis was $0.346 \ e^{-}/\text{Å}^3$ and the largest hole was $-0.339 \ e^{-}/\text{Å}^3$ with an RMS deviation of $0.048 \ e^{-}/\text{Å}^3$. On the basis of the final model, the calculated density was $1.115 \ g/\text{cm}^3$ and F(000), $832 \ e^{-}$.

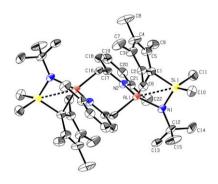


Figure S4. Ortep drawing with 50% ellipsoids for compound **5**.

Table S5. Sample and crystal data for compound **5**.

Identification code WegMi6 AP9203-100

Chemical formula $C_{44}H_{70}Al_2N_4Si_2\\$

Formula weight 255.06 100(2) K **Temperature** 0.71073 Å Wavelength

Crystal size 0.136 x 0.137 x 0.203 mm Crystal habit clear yellow fragment

Crystal system triclinic **Space group** P -1

Unit cell dimensions a = 11.8436(4) Å $\alpha = 87.653(2)^{\circ}$

> b = 12.0036(5) Å $\beta = 80.177(2)^{\circ}$ c = 16.2833(6) Å $\gamma = 88.234(2)^{\circ}$

Volume $2278.41(15) \text{ Å}^3$

 \mathbf{Z}

1.115 g/cm³ **Density (calculated)** 0.150 mm⁻¹ **Absorption coefficient**

F(000)832

Table S6. Data collection and structure refinement for compound 5.

Diffractometer Bruker D8 Venture Radiation source TXS rotating anode, Mo

Theta range for data

2.41 to 25.03° collection

Index ranges -14 <= h <= 14, -14 <= k <= 14, -19 <= l <= 19

Reflections collected 42515

Independent reflections 8029 [R(int) = 0.0346]

Coverage of independent

reflections

99.8%

Multi-Scan **Absorption correction**

Max. and min. transmission 0.9800 and 0.9700

Refinement method Full-matrix least-squares on F² Refinement program SHELXL-2014/7 (Sheldrick, 2014)

Function minimized $\Sigma \text{ w}(F_0^2 - F_c^2)^2$ **Data / restraints / parameters** 8029 / 262 / 698

Goodness-of-fit on F² 1.040 0.001 Δ/σ_{max}

6684 data; R1 = 0.0464, wR2 =Final R indices

0.1086 $I > 2\sigma(I)$

R1 = 0.0592, wR2 =all data

0.1177

 $w=1/[\sigma^2(F_0^2)+(0.0399P)^2+2.6556P]$ Weighting scheme

where $P=(F_0^2+2F_c^2)/3$

Largest diff. peak and hole 0.346 and -0.339 eÅ⁻³

R.M.S. deviation from mean 0.048 eÅ^{-3}

SC-XRD determination of compound 7 (CCDC 1936315)

A clear colourless fragment-like specimen of $C_{22}H_{29}AlN_2Si$, approximate dimensions 0.126 mm x 0.183 mm x 0.389 mm, was used for the X-ray crystallographic analysis. The X-ray intensity data were measured on a Bruker D8 Kappa Apex II system equipped with a Triumph monochromator monochromator and a Mo fine-focus sealed tube ($\lambda = 0.71073$ Å).

A total of 2682 frames were collected. The total exposure time was 3.73 hours. The frames were integrated with the Bruker SAINT software package using a narrow-frame algorithm. The integration of the data using a triclinic unit cell yielded a total of 35367 reflections to a maximum θ angle of 25.03° (0.84 Å resolution), of which 4254 were independent (average redundancy 8.314, completeness = 99.8%, R_{int} = 3.56%, R_{sig} = 2.35%) and 3524 (82.84%) were greater than $2\sigma(F^2)$. The final cell constants of \underline{a} = 9.558(2) Å, \underline{b} = 9.687(2) Å, \underline{c} = 13.866(3) Å, α = 106.254(9)°, β = 94.156(9)°, γ = 100.572(9)°, volume = 1201.1(5) ų, are based upon the refinement of the XYZ-centroids of 116 reflections above 20 $\sigma(I)$ with 4.362° < 20 < 47.95°. Data were corrected for absorption effects using the Multi-Scan method (SADABS). The ratio of minimum to maximum apparent transmission was 0.950. The calculated minimum and maximum transmission coefficients (based on crystal size) are 0.9470 and 0.9820.

The final anisotropic full-matrix least-squares refinement on F^2 with 241 variables converged at R1 = 3.46%, for the observed data and wR2 = 8.68% for all data. The goodness-of-fit was 1.036. The largest peak in the final difference electron density synthesis was $0.272 \text{ e}^{-}/\text{Å}^3$ and the largest hole was $-0.232 \text{ e}^{-}/\text{Å}^3$ with an RMS deviation of $0.040 \text{ e}^{-}/\text{Å}^3$. On the basis of the final model, the calculated density was 1.041 g/cm^3 and F(000), 404 e^{-} .

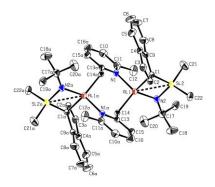


Figure S5. Ortep drawing with 50% ellipsoids for compound 7.

Table S7. Sample and crystal data for compound **7**.

Identification code SchFa2 AP7596-100

Chemical formula C₂₂H₂₉AlN₂Si

Formula weight376.54Temperature100(2) KWavelength0.71073 Å

Crystal size 0.126 x 0.183 x 0.389 mm Crystal habit clear colourless fragment

Crystal system triclinic **Space group** P -1

Unit cell dimensions a = 9.558(2) Å $\alpha = 106.254(9)^{\circ}$

b = 9.687(2) Å $\beta = 94.156(9)^{\circ}$ c = 13.866(3) Å $\gamma = 100.572(9)^{\circ}$

Volume 1201.1(5) Å³

 \mathbf{Z} 2

Density (calculated) 1.041 g/cm³ **Absorption coefficient** 0.141 mm⁻¹

F(000) 404

Table S8. Data collection and structure refinement for compound 7.

Diffractometer Bruker D8 Kappa Apex II **Radiation source** fine-focus sealed tube, Mo

Theta range for data

collection 2.19 to 25.03°

Index ranges -11<=h<=11, -11<=k<=11, -16<=l<=16

Reflections collected 35367

Independent reflections 4254 [R(int) = 0.0356]

Coverage of independent

reflections

99.8%

Absorption correction Multi-Scan

Max. and min. transmission 0.9820 and 0.9470

Refinement method Full-matrix least-squares on F² **Refinement program** SHELXL-2014/7 (Sheldrick, 2014)

Function minimized $\Sigma \text{ w}(F_o^2 - F_c^2)^2$ Data / restraints / parameters 4254 / 0 / 241

Goodness-of-fit on F^2 1.036

Final R indices 3524 data; R1 = 0.0346, wR2 = 0.0011

 $I > 2\sigma(I)$ 0.0811

all data R1 = 0.0447, wR2 =

0.0868

Weighting scheme $W=1/[\sigma^2(F_0^2)+(0.0349P)^2+0.6634P]$

where $P = (F_o^2 + 2F_c^2)/3$

Largest diff. peak and hole $0.272 \text{ and } -0.232 \text{ eÅ}^{-3}$

R.M.S. deviation from mean 0.040 eÅ⁻³

3. Polymerization Studies and Mechanism Elucidation

Further polymerization experiments in addition to Table 1:

Table S9. Catalytic conversion of DEVP with the synthesized CGCs. (a)

Run	Catalyst	V_{tol} / mL	t / min	Yield / % (b)	$M_n/~10^3g/mol~^{(c)}$	$\Theta\left(M_w/M_n\right)$	I / % $^{(d)}$	TOF / h^{-1}
1	1-CH ₂ TMS	20	1	100	150	1.19	32	18,000
2	2-CH ₂ TMS	20	1	100	450	1.26	11	18,000
3	5	3	45	91	160	1.83	28	360
4	6	3	45	38	160	1.67	10	150
5	7	3	45	98	100	2.28	48	400

 $^{^{(}a)}$ V_{DEVP} = 1.0 mL, T = 30 °C, [M]/[CGC] = 300/1, $^{(b)}$ measured gravimetrically and by NMR spectroscopy (^{31}P NMR spectroscopy), $^{(c)}$ determined by GPC-MALS in H_2O / THF (9 g/L tetrabutylammonium bromide), $^{(d)}$ initiator efficiency $(M_{n(theo.)}/M_{n(determ.)})$.

End group analysis via ESI-MS:

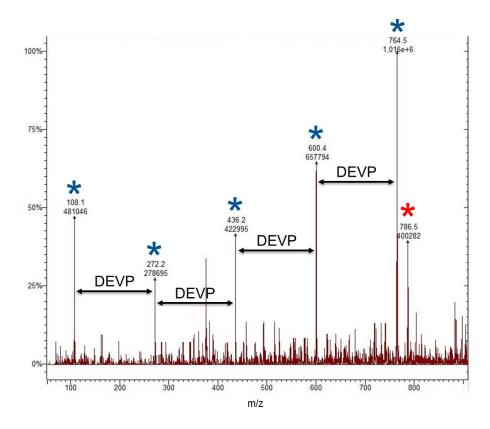


Figure S6. ESI MS spectrum of DEVP oligomers produced with **5**. One major series of peaks is evident: $m/z = n \times M_{Mon} + M_{H} + M_{Lut}$ (blue); $M_{Mon} = 164$, end groups: $M_{Lut} = 107$. Red peak: $m/z = n \times M_{Mon} + M_{Na} + M_{Lut}$; $M_{Mon} = 164$, end groups: $M_{Lut} = 107$.

Living polymerization behaviour (CGC 5):

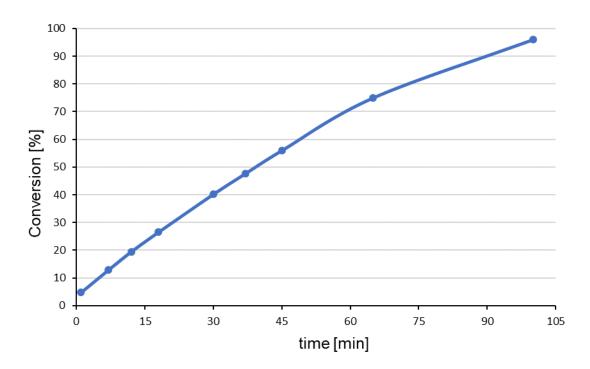


Figure S7. Time vs. conversion plot for polymerization of diethylvinylphosphonate with **5** (Monomer to catalyst ratio of 300/1, r.t., 5 mL toluene, 0.5 mL DEVP).

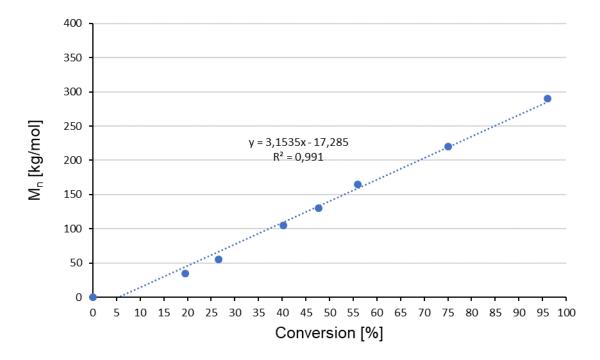


Figure S8. Linear growth of the molecular weight with increasing conversion.

Determination of catalyst order (differential method):

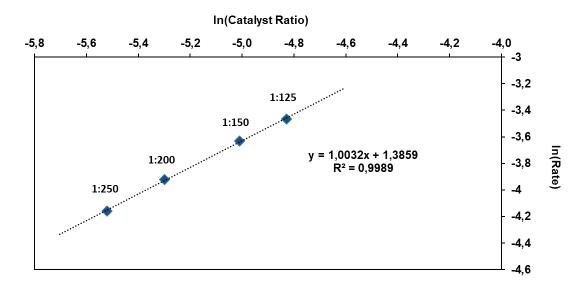


Figure S9. Determination of catalyst order: catalyst **5**, 0.15 mL DEVP, 0.75 mL C₆D₆, 25 °C.

Determination of monomer order (tangential method):

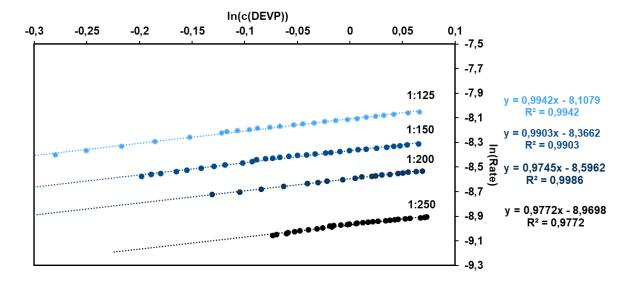


Figure S10. Determination of monomer order: catalyst **5**, 0.15 mL DEVP, 0.75 mL C₆D₆, 25 °C.

Living polymerization behaviour (CGC 1):

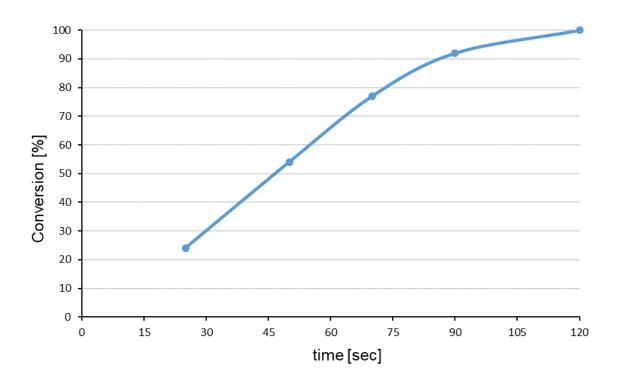


Figure S11. Time vs. conversion plot for polymerization of diethylvinylphosphonate with **1** (Monomer to catalyst ratio of 300/1, -30 °C, 20 mL toluene, 1 mL DEVP).

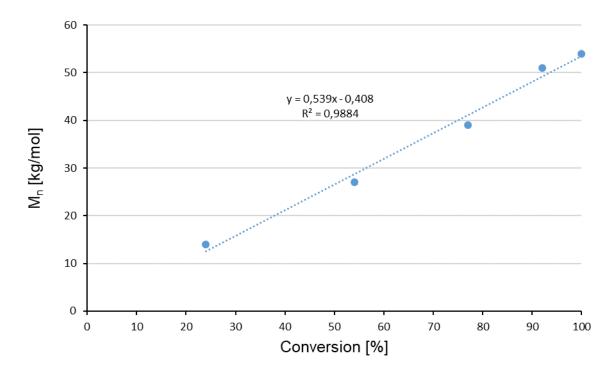


Figure S12. Linear growth of the molecular weight with increasing conversion.

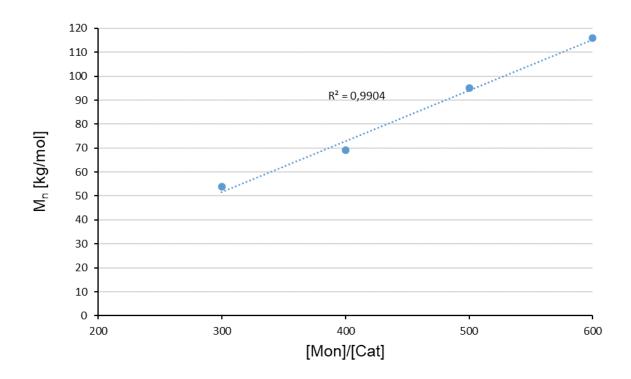


Figure S13. Molecular weight vs. [Mon]/[Cat] plot for polymerization of diethylvinylphosphonate with **1** ensuring quantitative conversions (–30 °C, 20 mL toluene, 1 mL DEVP).

Scheme S1. Proposed initiation reaction via nucleophilic transfer (six electron process, blue) or conjugate addition (eight electron process, red) and propagation via a repeated conjugate addition of the polymer chain to the coordinated monomer at the metal center by the example of catalyst **1**.

4. NMR, GPC and Thermal Analysis of the PDEVP Samples

Comparison of the ³¹P{¹H} NMR spectra:

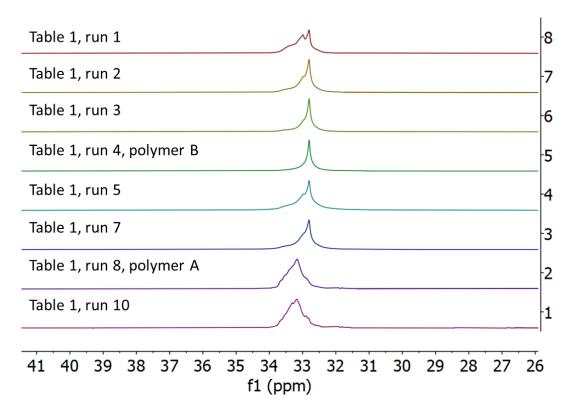


Figure S14. ³¹P{¹H} NMR spectra of the discussed polymers in MeOD₄.

NMR spectra, thermal characteristics & GPC trace of Table 1, run 4 (polymer B):

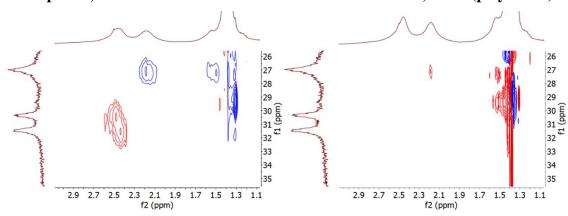


Figure S15. Phase-sensitive ${}^{1}\text{H-}{}^{13}\text{C}$ DEPT-edited HSQC vs. a ${}^{1}\text{H-}{}^{13}\text{C-}{}^{31}\text{P}$ triple resonance HCP 2D experiment edited by $J_{PC} = 20$ Hz (for details see Table S10).

Table S10. Pulseprogram parameters for $^{1}\text{H-}^{13}\text{C-}^{31}\text{P}$ triple resonance HCP 2D experiment edited by $J_{PC} = 20$ Hz (Figure S12).

General

PULPROG	na_hcpqetgpsi.1	
TD	2048	
SWH [Hz, ppm]	2380.95	5.94894
AQ [sec]	0.4300800	
RG	101	
DW [μsec]	210.000	
DE [μsec]	6.50	
CNST2 (J(HC))	140.0000000	
CNST5 (J(CP))	20.0000000	
CNST11	8.0000000	For multiplicity selection = 4 for CH, 8 for all
d0 [sec]	0.00000300	•
D1 [sec]	2.000000000	
d4 [sec]	0.00178571	
d11 [sec]	0.03000000	
d12 [sec]	0.00002000	
d13 [sec]	0.00000400	
D16 [sec]	0.000200000	
d20 [sec]	0.00500000	
d22 [sec]	0.00625000	
d24 [sec]	0.00089286	
DELTA1 [sec]	0.00505000	
DELTA2 [sec]	0.00499700	
DELTA3 [sec]	0.00120800	
DELTA4 [sec]	0.00500000	
DS	128	
in0 [sec]	0.00008280	
in20 [sec]	0.00008280	
INF1 [µsec]	165.60	
NS	256	
TAU [sec]	0.00002500	
TDav	0	
Channel f1		
SFO1 [MHz]	400.2312007	
O1 [Hz, ppm]	1200.69	3.000
NUC1	1H	
P1 [μsec]	10.000	
p2 [μsec]	20.00	
P28 [μsec]	1000.000	
PLW1 [W, dB]	9.604	-9.82
O2 [Hz, ppm]	4025.52	40.000

NUC2	13C	
CPDPRG 2	garp4	
P3 [μsec]	20.000	
p4 [μsec]	40.00	
PCPD2 [μsec]	80.00	
PLW2 [W, dB]	52.44	-17.20
PLW12 [W, dB]	3.2775	-5.16
Channel f3		
SFO3 [MHz]	162.0210965	
O3 [Hz, ppm]	5022.50	31.000
NUC3	31P	
CPDPRG 3	garp4	
P21 [μsec]	25.000	
p22 [μsec]	50.00	
PCPD3 [μsec]	100.00	
PLW3 [W, dB]	15.924	-12.02
PLW16 [W, dB]	0.99525	0.02
Gradient channel		
GPNAM 1	SMSQ10.100	
GPZ1 [%]	40.00	
GPNAM 2	SMSQ10.100	
GPZ2 [%]	25.00	
GPNAM 3	SMSQ10.100	
GPZ3 [%]	40.00	
GPNAM 4	SMSQ10.100	
GPZ4 [%]	20.10	
P16 [μsec]	1000.000	

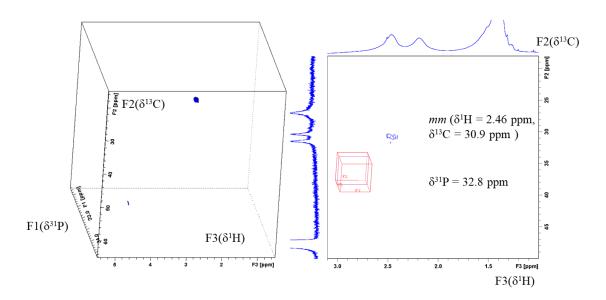


Figure S16. $^{1}\text{H}-^{13}\text{C}-^{31}\text{P}$ triple resonance HCP 3D experiment edited by $J_{PC}=110\,\text{Hz}$ (for details see Table S11, S12).

Table S11. Pulseprogram for ${}^{1}\text{H}$ - ${}^{13}\text{C}$ - ${}^{31}\text{P}$ triple resonance HCP 3D experiment edited by J_{PC} = 110 Hz (Figure S13, SX).

```
03.03.2019/gg
;hcp3d gg
ex hcacogp3d;
; avance-version (12/01/11)
;3D sequence with
    inverse correlation for triple resonance using multiple
        inept transfer steps
       F1(H) \rightarrow F2(C) \rightarrow F2'(P, t1) \rightarrow F2(C, t2) \rightarrow F1(H, t3)
;on/off resonance Ca and C=O pulses using shaped pulse
; phase sensitive (t1)
; phase sensitive using Echo/Antiecho-TPPI gradient selection (t2)
; (L.E. Kay et al., J. Magn. Reson 89, 496 - 514 (1990))
; $CLASS=HighRes
;$DIM=3D
; $TYPE=
;$SUBTYPE=
; $COMMENT=
prosol relations=<triple>
#include <Avance.incl>
#include <Grad.incl>
#include <Delay.incl>
"p2=p1*2"
"p4=p3*2"
"p22=p21*2"
```

```
"d11=30m"
"d12=20u"
"d13=4u"
"d22=0.25/cnst21"
"d0=3u"
"d10=3u"
"in0=inf1/2"
"in10=inf2/2"
"DELTA=d0*2+larger(p4,p2)"
"DELTA1=d22-d21-p2-p4/2-p16-4u"
"DELTA2=d22-p4/2-p16-4u"
"DELTA3=d22-p4/2"
"DELTA4=d22-d21-p2-p4/2-d10*2"
"DELTA5=d21-p16-d16"
"DELTA6=d4-p16-d16-8u"
agseg 321
1 d11 ze
  d11 pl1:f1
 d11 pl3:f3
2 d11 do:f2
3 d1
 p1 ph1
  d4 pl2:f2
  (center (p2 ph1) (p4 ph2):f2)
  d4
  (p1 ph2) (p3 ph3):f2
  d21 UNBLKGRAD
 p2 ph1
  p16:qp1
  DELTA1
  (center (p4 ph1):f2 (p22 ph1):f3)
  p16:gp2
  DELTA2
  (p3 ph1):f2
  d13
  (p21 ph4):f3
  (center (p2 ph1) (p4 ph1):f2)
  d0
  (p22 ph1):f3
  DELTA
  (p21 ph1):f3
  d13
```

```
(p3 ph5):f2
  DELTA3
  (p4 ph1):f2
  d10
  (p22 ph1):f3
  DELTA4
  p2 ph1
  p16:gp3*EA
  d16
  DELTA5
  d10
  (ralign (p1 ph1) (p3 ph6):f2 )
  (center (p2 ph1) (p4 ph1):f2 )
  4u
  p16:qp4
  d16
  DELTA6 pl12:f2
  4u BLKGRAD
  go=2 ph31 cpd2:f2
  d11 do:f2 mc #0 to 2
     F1PH(calph(ph4, +90), caldel(d0, +in0))
     F2EA(calgrad(EA), caldel(d10, +in10) & calph(ph5, +180) &
calph(ph31, +180))
exit
ph1 = 0
ph2 = 1
ph3 = 0 2
ph4 = 0 0 2 2
ph5 = 0 \ 0 \ 0 \ 2 \ 2 \ 2
ph6 = 0 0 0 0 0 0 0 0 2 2 2 2 2 2 2 2
ph31= 0 2 2 0 2 0 0 2 2 0 0 2 0 2 0 0
;pl1 : f1 channel - power level for pulse (default)
;pl2 : f2 channel - power level for pulse (default)
;pl3 : f3 channel - power level for pulse (default)
;pl12: f2 channel - power level for CPD/BB decoupling
;p1 : f1 channel - 90 degree high power pulse
;p2 : f1 channel - 180 degree high power pulse
;p3: f2 channel - 90 degree shaped pulse
;p4: f2 channel - 180 degree shaped pulse
;p16: homospoil/gradient pulse
                                                         [1 msec]
;p21: f3 channel - 90 degree high power pulse
;p22: f3 channel - 180 degree high power pulse
;d0 : incremented delay (F1 in 3D)
                                                        [3 usec]
;d1 : relaxation delay; 1-5 * T1
;d4 : 1/(4J(HC))
                                                        [1.7 msec]
;d10: incremented delay (F2 in 3D)
                                                         [3 usec]
;d11: delay for disk I/O
                                                         [30 msec]
;d12: delay for power switching
                                                        [20 usec]
;d13: short delay
                                                         [4 usec]
;d16: delay for homospoil/gradient recovery
;d21: 1/(4J)(HC) or 1/(6J)(HC) for CH or CH & CH2 [1.7 msec or 1.2
msec]
```

```
;d22: 1/(4J(13C,31P))
;cnst21: 1J(13C,31P)
; inf1: 1/SW(31P) = 2 * DW(31P)
; inf2: 1/SW(13C) = 2 * DW(13C)
;in0: 1/(2 * SW(31P) = DW(31P)
;nd0: 2
; in10: 1/(2 * SW(13C)) = DW(13C)
;nd10: 2
;ns: 16 * n
;ds: 32
;tdl: number of experiments in F1
;td2: number of experiments in F2
;FnMODE: States-TPPI (or TPPI) in F1
;FnMODE: echo-antiecho in F2
;cpd2: decoupling according to sequence defined by cpdprg2
;pcpd2: f2 channel - 90 degree pulse for decoupling sequence
;use gradient ratio: gp 1 : gp 2 : gp 3 : gp 4
                          3: 3:
                                      80 : 20.1
; for z-only gradients:
;gpz1: 3%
;gpz2: 3%
;gpz3: 80%
;gpz4: 20.1%
; use gradient files:
;gpnam1: SMSQ10.100
;gpnam2: SMSQ10.100
;gpnam3: SMSQ10.100
;gpnam4: SMSQ10.100
;$Id: hcacogp3d,v 1.9 2012/01/31 17:49:23 ber Exp $
```

Table S12. Pulseprogram parameters for ${}^{1}\text{H}-{}^{13}\text{C}-{}^{31}\text{P}$ triple resonance HCP 3D experiment edited by $J_{PC} = 110$ Hz (Figure S13, SX).

General		
PULPROG	hcp3d_gg	
TD	2048	
SWH [Hz, ppm]	2000.00	4.99711
AQ [sec]	0.5120000	
RG	101	
DW [μsec]	250.000	
DE [μsec]	6.50	
CNST21 (1J13C,31P)	110.0000000	
d0 [sec]	0.00000300	
D1 [sec]	2.000000000	
D4 [sec]	0.001700000	
d10 [sec]	0.00000300	

d11 [sec]	0.03000000	
d11 [sec]	0.00000400	
D16 [sec]	0.000200000	
D21 [sec]	0.001200000	
d22 [sec]	0.001200000	
= =	0.00227273	
DELTA [sec] DELTA1 [sec]	0.00004800	
DELTAT [sec] DELTA2 [sec]	0.0002873	
DELTA2 [sec] DELTA3 [sec]	0.00124873	
	0.00223273	
DELTA5 [sec]		
DELTA5 [sec]	0 0.00049200	
DELTA6 [sec] DS		
·-	32	
in0 [sec]	0.00102870	
in10 [sec]	0.00009940	
INF1 [µsec]	2057.40	
INF2 [µsec]	198.80	
NS TD	48	
TDav	0	
Channel f1		
SFO1 [MHz]	400.2312007	
O1 [Hz, ppm]	1200.69	3.000
NUC1	1H	
P1 [μsec]	10.000	
p2 [μsec]	20.00	
PLW1 [W, dB]	9.604	-9.82
Channel f2		
SFO2 [MHz]	100.6419395	
O2 [Hz, ppm]	4025.52	40.000
NUC2	13C	
CPDPRG 2	garp4	
P3 [μsec]	20.000	
p4 [μsec]	40.00	
PCPD2 [µsec]	80.00	
PLW2 [W, dB]	52.44	-17.20
PLW12 [W, dB]	3.2775	-5.16
Channel f3		
SFO3 [MHz]	162.0214529	
O3 [Hz, ppm]	5378.93	33.200
NUC3	31P	00.200
P21 [µsec]	25.000	
p22 [µsec]	50.00	
PLW3 [W, dB]	15.924	-12.02
Gradient channel		12.02
GPNAM 1	SMSQ10.100	
GPZ1 [%]	3.00	
O. 21 [/0]	5.00	

GPNAM 2	SMSQ10.100
GPZ2 [%]	3.00
GPNAM 3	SMSQ10.100
GPZ3 [%]	80.00
GPNAM 4	SMSQ10.100
GPZ4 [%]	20.10
P16 [μsec]	1000.000

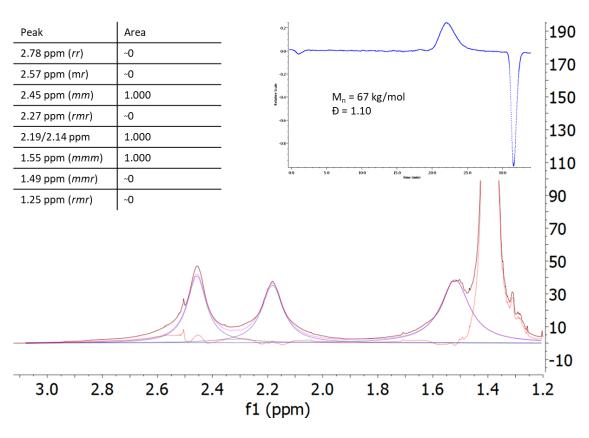


Figure S17. Quantification of the *mmm* tetrads with the line fitting method of MestReNova version 14.0.0-23239 and the GPC trace. Color coding: dark red – original spectrum, blue – peaks, pink – sum, red – residue.

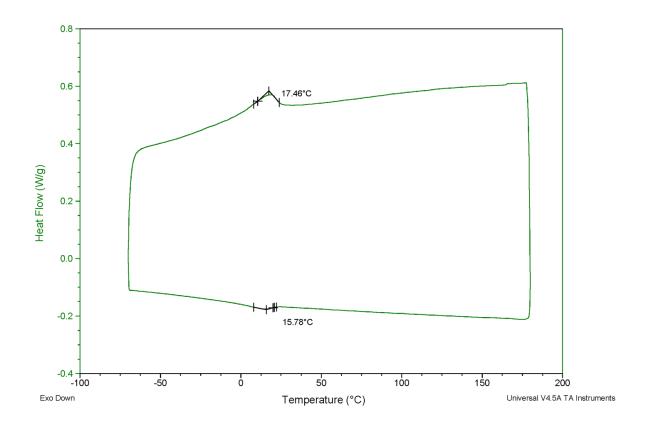


Figure S18. DSC analysis of polymer B showing a melting and crystallization point.

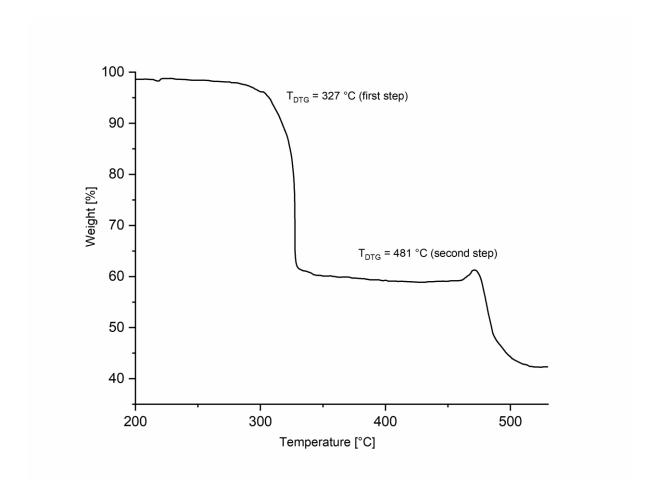


Figure S19. Thermogravimetric analysis of polymer B.

NMR spectra & GPC trace of Table 1, run 1:

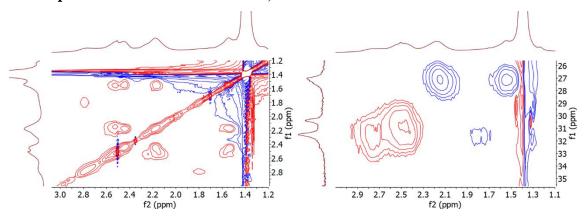


Figure S20. Phase-sensitive ¹H-¹³C DEPT-edited HSQC (right) and TOCSY{³¹P} NMR spectra in MeOD₄.

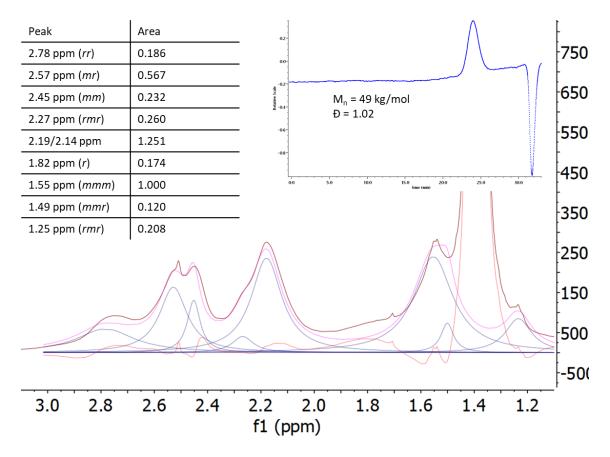


Figure S21. Quantification of the *mmm* tetrads with the line fitting method of MestReNova version 14.0.0-23239 and the GPC trace. Color coding: dark red – original spectrum, blue – peaks, pink – sum, red – residue.

NMR spectra & GPC trace of Table 1, run 2:

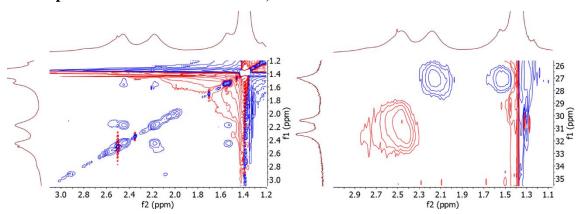


Figure S22. Phase-sensitive ¹H-¹³C DEPT-edited HSQC (right) and TOCSY{³¹P} NMR spectra in MeOD₄.

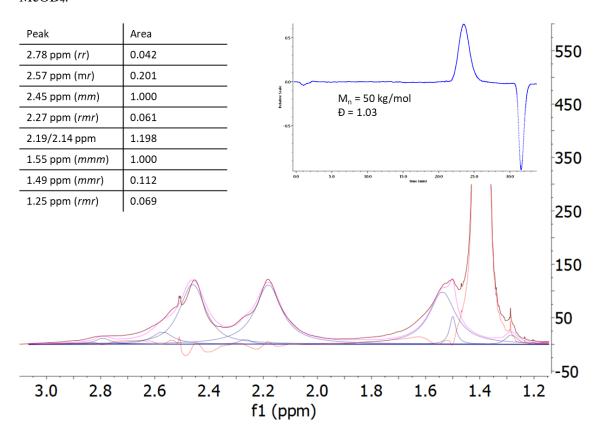


Figure S23. Quantification of the *mmm* tetrads with the line fitting method of MestReNova version 14.0.0-23239 and the GPC trace. Color coding: dark red – original spectrum, blue – peaks, pink – sum, red – residue.

NMR spectra & GPC trace of Table 1, run 3:

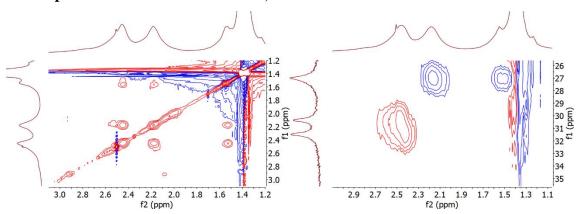


Figure S24. Phase-sensitive ¹H-¹³C DEPT-edited HSQC (right) and TOCSY{³¹P} NMR spectra in MeOD₄.

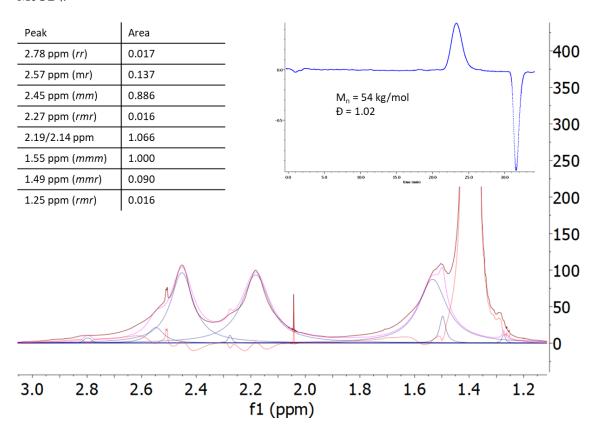


Figure S25. Quantification of the *mmm* tetrads with the line fitting method of MestReNova version 14.0.0-23239 and the GPC trace. Color coding: dark red – original spectrum, blue – peaks, pink – sum, red – residue.

NMR spectra & GPC trace of Table 1, run 5:

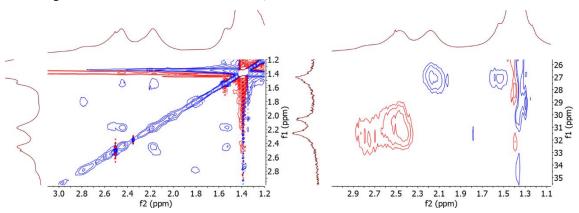


Figure S26. Phase-sensitive ¹H-¹³C DEPT-edited HSQC (right) and TOCSY{³¹P} NMR spectra in MeOD₄.

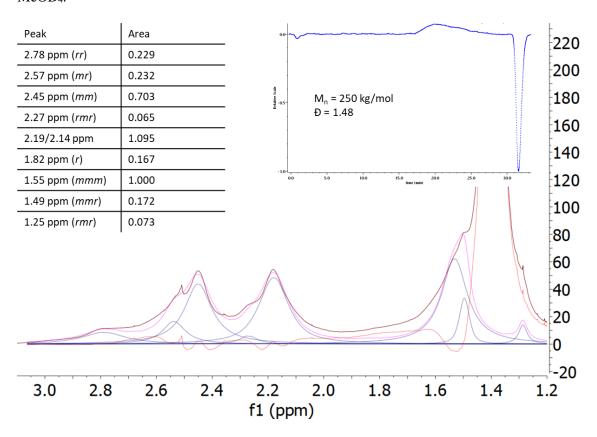


Figure S27. Quantification of the *mmm* tetrads with the line fitting method of MestReNova version 14.0.0-23239 and the GPC trace. Color coding: dark red – original spectrum, blue – peaks, pink – sum, red – residue.

NMR spectra & GPC trace of Table 1, run 7:

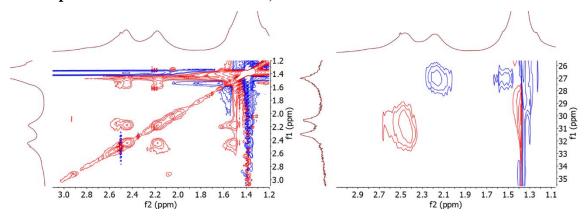


Figure S28. Phase-sensitive ¹H-¹³C DEPT-edited HSQC (right) and TOCSY{³¹P} NMR spectra in MeOD₄.

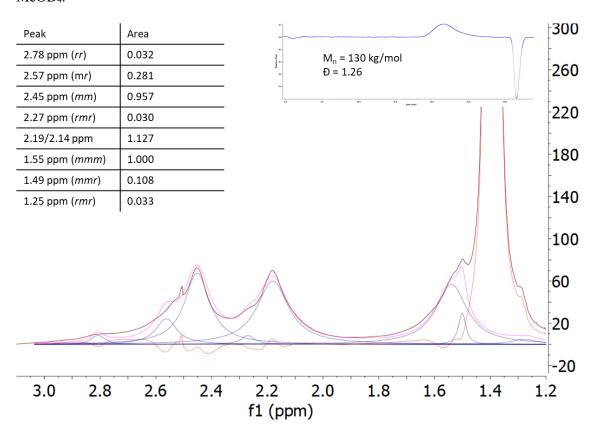


Figure S29. Quantification of the *mmm* tetrads with the line fitting method of MestReNova version 14.0.0-23239 and the GPC trace. Color coding: dark red – original spectrum, blue – peaks, pink – sum, red – residue.

NMR spectra, thermal characteristics & GPC trace of Table 1, run 8 (polymer B):

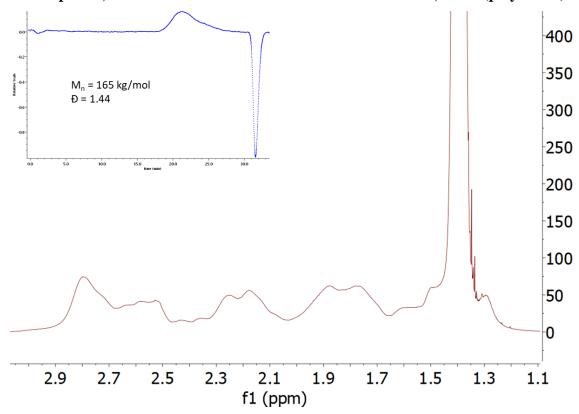


Figure S30. ¹H{³¹P} NMR spectrum and the GPC trace.

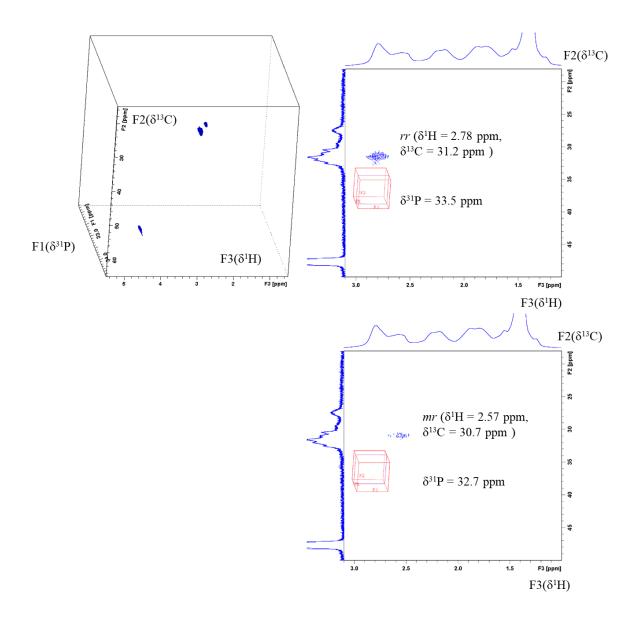


Figure S31. $^{1}\text{H}^{-13}\text{C}^{-31}\text{P}$ triple resonance HCP 3D experiment edited by $J_{PC} = 110 \text{ Hz}$ (for details see Table S11, S12).

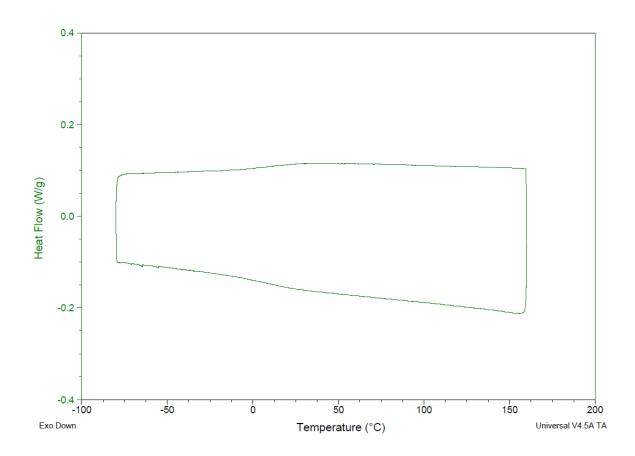


Figure S32. DSC analysis of polymer A showing no melting point.

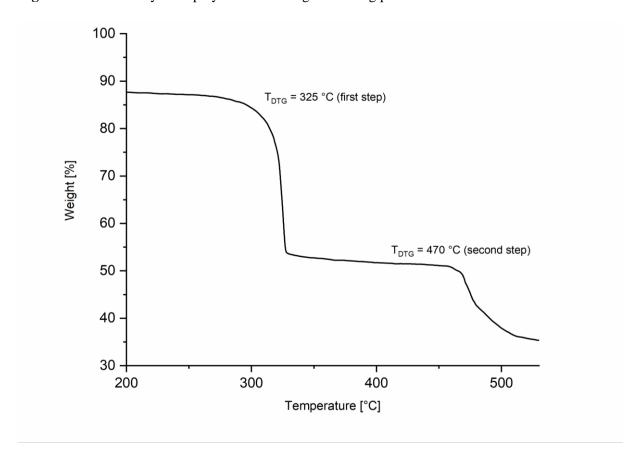


Figure S33. Thermogravimetric analysis of polymer A.

NMR spectra & GPC trace of Table 1, run 10:

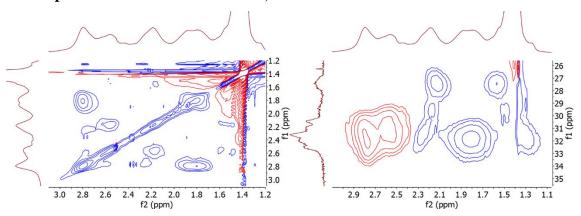


Figure S34. Phase-sensitive ¹H-¹³C DEPT-edited HSQC (right) and TOCSY{³¹P} NMR spectra in MeOD₄.

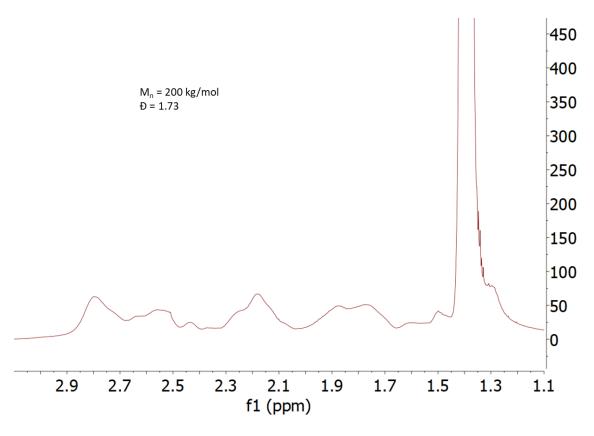


Figure S35. ¹H{³¹P} NMR spectrum and the GPC trace.

Saponification of Table 1, run 4 (polymer B):

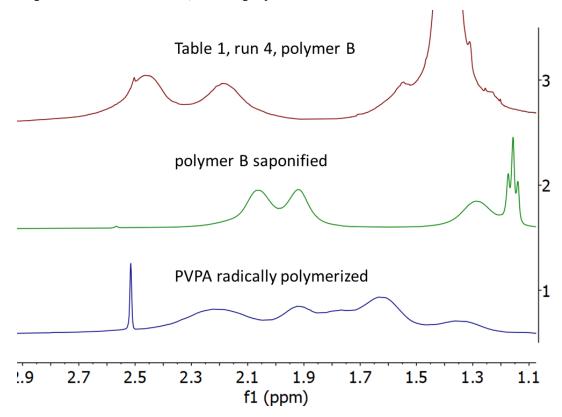


Figure S36. 1 H NMR spectra of polymer B in MeOD₄ and its saponification product in D₂O (1. 10 g/L DEVP in DCM + excess TMSBr 24 h reflux; 2. 1 m HCl 2 h r.t. and aqueous dialysis) compared to a radically produced PVPA in D₂O.

5. References

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